

Integrating the Neurobiology of Schizophrenia

EDITED BY

ANISSA ABI-DARGHAM

OLIVIER GUILLIN

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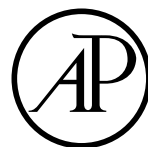
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PREFACE

“Integrating the Neurobiology of Schizophrenia” is meant to bring together the current knowledge implicating various neurotransmitter systems in the disease of schizophrenia while placing a big emphasis on their interactions. The goal is to build through each chapter one of the blocks leading to an integrative model showing how one neurochemical alteration could contribute to the final common pathway of another neurochemical dysregulation observed in this illness. It is intended to be a reference for clinicians, scientists, and students who want to learn more about the different neurotransmitters that may play a role in schizophrenia.

To anchor these discussions, we adopt the view that all neurotransmitter alterations may lead to the dopaminergic alterations observed in this illness. This point of view is an arbitrary oversimplification, but complex problems may be better addressed once broken down into simple questions. This view is suggested by the observation that alterations in dopamine transmission are most directly linked to the symptoms of the illness as well as the response of these symptoms to antipsychotic treatment. Recent research, mostly from imaging, over the last decade, has greatly advanced the field by providing strong evidence for few fundamental observations: studies have shown definitively that subcortical D2 hyperstimulation is associated with positive symptoms and more recent data has linked negative symptoms and cognitive disturbances with cortical dopamine dysfunction. Studies have also shown conclusively that all effective antipsychotics show significant D2 receptor occupancy. Despite the fact that there is no clear relationship between occupancy and clinical response, and there is no clear definition of the minimal occupancy needed to achieve therapeutic efficacy, this remains the most solid finding in antipsychotic therapy. We previously reported that elevated levels of striatal intrasynaptic DA concentration is predictive of fast response to antipsychotic drugs in patients with schizophrenia (Abi-Dargham *et al.*, 2000). Thus, the extent of the therapeutic response to D2 receptor antagonism is affected by the underlying pathology. Patients whose pathology is associated with excessive stimulation of D2 receptors by DA respond well to D2 receptor blockade. Conversely, patients who present a psychotic episode in the absence of detectable changes in synaptic DA levels are poor responders to current antipsychotic drugs. On the other hand, we did not observe a relationship between subcortical D2 hyperstimulation and the response of negative symptoms to treatment at 6 weeks. Overall these findings suggest that D2

hyperstimulation is relevant to the treatment of positive symptoms in most but not all patients with schizophrenia and is probably irrelevant to negative symptoms. This highlights the complexity of the problem. A provocative study by Seeman *et al.* (2006) has provided some support to the concept of common final pathway through hyperstimulation of D2 receptors leading to psychosis, essentially by increasing the high affinity states of dopamine D2 receptors.

In this book, we first present a comprehensive review of the alterations in dopamine (see the chapter by Guillin, Abi-Dargham, and Laruelle) and the underlying cellular and physiological events that may accompany them (see the chapter by Goto and Grace). Then we attempt to review most of the neurobiological alterations that may be implicated in schizophrenia and may contribute to the symptoms and their treatment, either by contributing to the dopaminergic alterations directly or indirectly, or by creating a different pathway to pathology. For each system, the main findings in schizophrenia are reviewed, followed by a discussion of how such findings may affect dopamine transmission, at least to the extent that these interactions are known. Whenever possible, inferences to treatment are made, resulting in a review of potential new therapeutic targets.

One possible conclusion that emerges from this review of various contributions of many systems to schizophrenia pathology is that the dysregulation in DA may be a consequence of other upstream events. New research has shed light on interactions with glutamate and a deficient NMDA system leading to both DA alterations observed in schizophrenia, including cortical deficit and subcortical excess. These are reviewed by Daniel C. Javitt in his chapter. Similarly NMDA antagonists have been recently shown to engender some of the alterations in a subset of GABA neurons in the dorsolateral prefrontal cortex that have been described in schizophrenia (see the chapter by Lewis and Hashimoto), leading to a deficit in the GABA_A α_2 subunit function and deficits in perisomatic inhibitory regulation of pyramidal neurons. Thus, these glutamatergic and GABAergic alterations are intimately linked and could lead to an inefficient control of cortical input onto subcortical striatal dopamine, as well as inefficient corticocortical connectivity and function. However, causality is difficult to assess in the presence of reciprocal regulations. Alterations in striatal dopamine transmission may itself affect cortical functioning by impairing glutamatergic flow of information in the corticostriatal-thalamocortical loops as illustrated recently by the genetically altered mice over-expressing striatal D2 leading to long-lasting cognitive deficits (Kellendonk *et al.*, 2006).

The role of 5-HT in schizophrenia is addressed by reviewing alterations in clinical studies: postmortem, pharmacological challenges and imaging studies (see the chapter by Abi-Dargham). As no clear patterns emerge for consistent findings, the emphasis seems to be better placed on the role that serotonin may play by modulating dopamine transmission in the critical brain regions implicated in schizophrenia and/or how it may affect response independently of its

role in modulating dopamine transmission. As reviewed by Marek in his chapter, DA–5-HT interactions in the brain are present at different anatomical levels, are mediated by different 5-HT receptor subtypes, and affect different aspects of DA function. This complexity leads to a rich potential pharmacology of cognitive enhancement with 5-HT_{1A} partial agonists, 5-HT_{2A} antagonists, 5-HT₄ partial agonists and 5-HT₆ antagonists.

While there have been many advances in the field of genetics and imaging contributing to our understanding of the basic pathophysiology of schizophrenia and its treatment, much remains to be discovered. Our therapeutic interventions are very limited in scope and in efficacy. The future seems to lie in all the unexplored potential targets that emerge from a systematic review of all systems, from D1 agonists, to GABA_A α_2 -specific allosteric modulators, to the many targets within the glutamate and serotonin systems. The modulation of the histaminergic system by antipsychotics and the antipsychotic-like properties of H₃-receptor antagonists/inverse agonists support a role of histamine neurons in schizophrenia (see the chapter by Arrang).

The following chapter reviews the evidence for a dysregulation of central cholinergic signaling in the pathophysiology of schizophrenia and suggests potential therapeutic roles for cholinergic targets (chapter by Berman, Talmage, and Role). This is followed by an in-depth review and analysis of one of these targets, the $\alpha 7$ nicotinic receptor (chapter by Martin and Freedman). The cannabinoid system and its relevance to schizophrenia is another emerging and exciting field, reviewed by D'Souza in his chapter. The connections between cannabinoids system and other neurotransmitters including dopamine are likely to be relevant as this system is present in all areas of the brain incriminated in schizophrenia, hippocampus, amygdala, prefrontal cortex, and striatum, where it exerts direct control over transmitter release. Endocannabinoids are released from lipid precursors in a receptor-dependent manner and serve as retrograde signaling messengers in GABAergic and glutamatergic synapses, as well as modulators of postsynaptic transmission, interacting with other neurotransmitters, including dopamine. A comprehensive review of clinical data linking alterations in neuropeptide systems to the etiology, pathophysiology, and treatment of schizophrenia in the chapter by Cáceda, Kinkead, and Nemeroff follows with a summary of potential therapeutic targets within this complex field. Genetic and postmortem evidence suggest a role for BDNF in the pathophysiology of schizophrenia. Interestingly, this neurotrophin controls the expression of the D3 receptor. The evidence supporting a role for BDNF in schizophrenia and the role of this neurotrophin at modulating dopamine transmission are reviewed in the chapter by Guillin, Demily, and Thibaut.

Finally, no review is complete without mention of the recent explosion of new candidate genes that may be affected in schizophrenia. Gogos in his chapter reviews the genetic contributions to schizophrenia and highlights the fact that

the neurobiology resulting from small variations in common genes is likely to be subtle and complex. This supports action at multiple molecular targets to reach an effective threshold in a large fraction of patients.

As many receptors modulate the same intracellular pathways, either synergistically or in an opposing manner, the final common pathway in schizophrenia may well be the signal transduction pathway(s) linked to these various neuroreceptor systems. Developing the appropriate tools to study intracellular targets in the living human brain is needed to understand how these alterations are integrated and lead to common symptomatology.

This book emphasizes what we know as well as all the many areas that need further research, that is, all the unknowns. In doing so, we hope it will provide a useful tool to the clinician and the researcher alike.

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References

- Abi-Dargham, A., Rodenhiser, J., Printz, D., Zea-Ponce, Y., Gil, R., Kegeles, L., Weiss, R., Cooper, T., Mann, J. J., Van Heertum, R., Gorman, J., and Laruelle, M. (2000). Increased baseline occupancy of D2 receptors by dopamine in schizophrenia. *Proc. Natl. Acad. Sci. USA* **97**, 8104–8109.
- Kellendonk, C., Simpson, E. H., Polan, H. J., Malleret, G., Vronskaya, S., Winiger, V., Moore, H., and Kandel, E. R. (2006). Transient and selective overexpression of dopamine D2 receptors in the striatum causes persistent abnormalities in prefrontal cortex functioning. *Neuron* **49**, 603–615.
- Seeman, P., Schwarz, J., Chen, J. F., Szechtman, H., Perreault, M., McKnight, G. S., Roder, J. C., Quirion, R., Boksa, P., Srivastava, L. K., Yanai, K., Weinshenker, D., *et al.* (2006). Psychosis pathways converge via D2 high dopamine receptors. *Synapse* **60**, 319–346.

NEUROBIOLOGY OF DOPAMINE IN SCHIZOPHRENIA

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- II. Dopaminergic System in the Brain
 - A. Dopaminergic Projections
 - B. Dopaminergic Receptors
- III. Evidence Supporting Alterations of DA Systems in Schizophrenia
 - A. Pharmacological Evidence
 - B. Postmortem Studies
 - C. Imaging Studies
- IV. Conclusions
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This chapter is an update on the dopamine (DA) imbalance in schizophrenia, including the evidence for subcortical hyperstimulation of D2 receptors underlying positive symptoms and cortical hypodopaminergia-mediating cognitive disturbances and negative symptoms. After a brief review of the anatomical neurocircuitry of this transmitter system as a background, we summarize the evidence for dopaminergic alterations deriving from pharmacological, postmortem, and imaging studies. This evidence supports a prominent role for D2 antagonism in the treatment of positive symptoms of schizophrenia and strongly suggests the need for alternative approaches to address the more challenging problem of negative symptoms and cognitive disturbances.

I. Introduction

Schizophrenia is a severe and chronic mental illness, associated with high prevalence (about 1% of the general population). Symptoms of schizophrenia typically emerge during adolescence or early adulthood. They are usually classified as positive, negative, or cognitive symptoms. Positive symptoms include: hallucinations, delusions, and severe thought disorganization. Negative symptoms

are a group of deficits comprising many dimensions such as affect (flattening), volition (apathy), speech (poverty), pleasure (anhedonia), and social life (withdrawal). Cognitive symptoms, such as deficits in attention and memory, are prominent features of the illness.

While the etiology and pathophysiology of schizophrenia remain unclear, a large body of evidence suggests that alterations in several neurotransmitter systems (e.g., dopamine, glutamate, GABA, serotonin, cholinergic system, and others) are involved in the pathophysiological processes leading to the expression of these symptoms. Among these, the dopamine (DA) system has received most attention.

The involvement of DA in the pathophysiology and treatment of schizophrenia has been the subject of intense research efforts over the last 50 years. The first formulation of the DA hypothesis of schizophrenia proposed that hyperactivity of DA transmission was responsible for the core or “positive” symptoms (hallucinations, delusions) observed in this disorder (Carlsson and Lindqvist, 1963). This hypothesis was based on the correlation between clinical doses of antipsychotic drugs and their potency to block DA D2 receptors (Creese *et al.*, 1976; Seeman and Lee, 1975) and by the psychotogenic effects of DA-enhancing drugs (for review see Angrist and van Kammen, 1984; Lieberman *et al.*, 1987a). Given the predominant localization of DA terminals and D2 receptors in subcortical regions such as the striatum and the nucleus accumbens, the classical DA hypothesis of schizophrenia was concerned mostly with these subcortical regions.

Over the years, the increasing awareness of the importance of enduring negative and cognitive symptoms in this illness and of their resistance to D2 receptor antagonism has led to a reformulation of this classical DA hypothesis. Functional brain imaging studies suggested that these symptoms might arise from altered prefrontal cortex (PFC) functions (for reviews see Knable and Weinberger, 1997). A wealth of preclinical studies emerged documenting the importance of prefrontal DA transmission at D1 receptors (the main DA receptor in the neocortex) for optimal PFC performance (for review see Goldman-Rakic *et al.*, 2000). Together, these observations led to the hypothesis that a deficit in DA transmission at D1 receptors in the PFC might be implicated in the cognitive impairments and negative symptoms of schizophrenia (Davis *et al.*, 1991; Weinberger, 1987).

Thus, the current predominant view in that DA systems in schizophrenia might be characterized by an imbalance between subcortical and cortical DA systems: subcortical mesolimbic DA projections might be hyperactive (resulting in hyperstimulation of D2 receptors and positive symptoms) while mesocortical DA projections to the PFC might be hypoactive (resulting in hypostimulation of D1 receptors, negative symptoms, and cognitive impairment). Furthermore, since the seminal work of Pycck *et al.* (1980), many laboratories have described reciprocal and opposite regulations between cortical and subcortical DA systems (for review see Tzschentke, 2001). An abundant literature suggests that prefrontal DA activity exerts an inhibitory influence on subcortical DA activity (Deutch *et al.*, 1990;

Karreman and Moghaddam, 1996; Kolachana *et al.*, 1995; Wilkinson, 1997). From these observations, it has been proposed that, in schizophrenia, both arms of the DA imbalance model might be related, inasmuch as a deficiency in mesocortical DA function might translate into disinhibition of mesolimbic DA activity (Weinberger, 1987).

Despite decades of effort to generate experimental data supporting these hypotheses, documentation of abnormalities of DA function in schizophrenia has been difficult. Postmortem studies measuring DA and its metabolites and DA receptors in the brains of patients with schizophrenia yielded inconsistent or inconclusive results (for review see Davis *et al.*, 1991). Over the last few years, the development of new brain imaging methods based on the principle of endogenous competition enabled direct measurement of DA transmission at D2 receptor in the striatum (for review see Laruelle, 2000a). Combined with studies that documented increased striatal [^{18}F]dopa accumulation in schizophrenia, application of these new techniques to the study of schizophrenia provided new information into subcortical DA function dysregulation in schizophrenia (for review see Weinberger and Laruelle, 2001). Imaging studies have consistently demonstrated that schizophrenia is associated with increased presynaptic activity of DA neurons projecting to the striatum. Thus, the first arm of the dopaminergic imbalance hypothesis (hyperactivity in subcortical territory) has received strong support from imaging studies.

On the other hand, the second arm of this hypothesis (DA deficit in cortical projections) is still largely based on inferences from preclinical model or indirect clinical evidence. In contrast to the striatum, presynaptic DA function in the PFC is not at present accessible to noninvasive imaging techniques. D1 receptor availability is the only parameter of prefrontal DA function that is currently quantifiable *in vivo* with adequate reliability. Despite the limited information that this parameter provides to characterize DA function, PET imaging studies have described interesting relationships between alterations of D1 receptor availability and cognitive functions in schizophrenia (Abi-Dargham *et al.*, 2002; Karlsson *et al.*, 2002; Okubo *et al.*, 1997).

The goal of this chapter is to review current evidence for DA dysregulation in schizophrenia. Following a brief review of dopaminergic systems and receptors, pharmacological, postmortem, and imaging data that implicate DA alterations in schizophrenia will be presented.

II. Dopaminergic System in the Brain

A. DOPAMINERGIC PROJECTIONS

Dopaminergic projections are classically divided in nigrostriatal, mesolimbic, and mesocortical systems (Lindvall and Björklund, 1983). The nigrostriatal system projects from the substantia nigra (SN) to the dorsal striatum and has been

classically involved in cognitive integration, habituation, sensorimotor coordination, and initiation of movement. The mesolimbic system projects from the ventral tegmental area (VTA) to limbic structures such as ventral striatum, hippocampus, and amygdala. The mesocortical system projects from the VTA to cortical regions, mostly orbitofrontal, medial prefrontal, and cingulate cortices, but also to the dorsolateral prefrontal cortex (DLPFC), temporal, and parietal cortex. The mesolimbic and mesocortical systems are involved in regulation of motivation, attention, and reward (Mogenson *et al.*, 1980).

Corticostriatal–thalamocortical loops are important targets of DA modulation (Fig. 1). The general scheme of these loops involves projections from the cortex to striatum to the internal segment of the globus pallidum (GPi) or the SN pars

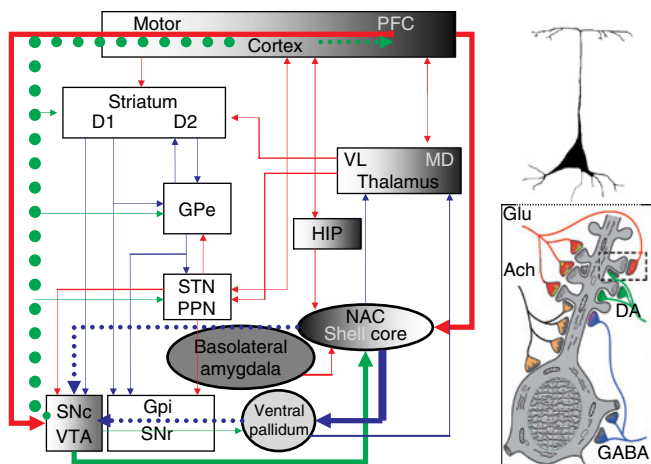


FIG. 1. Schematic representation of ventral limbic circuits implicated in the positive symptoms of schizophrenia. The primary neurotransmitter content and the major projections are represented by the colored arrows: blue, GABA; red, glutamate; green, DA. The nucleus accumbens (NAC) receives major excitatory inputs from prefrontal cortex (PFC), basolateral amygdala, and hippocampus (HIP) and DA input from ventral tegmental area (VTA). NAC sends inputs to the ventral pallidum (VP) which sends, among others, inputs to the mediodorsal nucleus (MD) of the thalamus. MD provides excitatory inputs back to PFC. The NAC is a crucial node in this circuit where inputs from the PFC are gated by excitatory projections from HIP and BLA and DA projections from VTA. It is proposed that, in schizophrenia, increased DA activity induces perturbation in the information flow within this circuit, in the modulation of this information by hippocampus and amygdala, and in the gating of sensory information by the thalamus. If sustained, the increase in DA activity might lead to neuroplastic changes in this circuit resulting in the emergence of psychotic experience. Antipsychotics are antagonists at the D2 receptor which is expressed in the motor circuits represented by the loops involving the striatum (caudate and putamen), the external (GPe) and internal (GPi) globus pallidus, the substantia nigra pars compacta (SNc) and pars reticulata (SNr), and the pedunculopontine nucleus (PPN) and subthalamic nucleus (STN), explaining some of their motor side effects.

reticulata (SNr) to thalamus and back to the cortex. These loops have been classified into “limbic” loops (medial prefrontal and orbitofrontal cortex–ventral striatum–ventral pallidum–mediodorsal thalamic nuclei–cortex), associative loops (DLPFC–head of the caudate–GPe/SNr–ventral anterior thalamic nuclei–cortex), and motor loops (premotor and motor areas–putamen and body of the caudate–GPe/SNr–ventral anterior thalamic nuclei back–cortex) (Alexander *et al.*, 1986; Ferry *et al.*, 2000; Hoover and Strick, 1993; Joel and Weiner, 2000; Parent and Hazrati, 1995). The amygdala and hippocampus provide significant inputs to the ventral striatum, contributing to information integration into the limbic loop (Everitt *et al.*, 1991; Grace, 2000; Kunishio and Haber, 1994; Pennartz *et al.*, 1994). Animal studies suggest that the nucleus accumbens is the critical region in which both typical and atypical antipsychotic drugs exert their antipsychotic effects (Chiodo and Bunney, 1983; Deutch *et al.*, 1991; Robertson *et al.*, 1994). It is important to note that these different corticostriatal–thalamocortical loops are not completely segregated parallel loops. While corticostriatal–thalamocortical loops do generally reenter the cortical area that provides input to the striatal subregions involved in these loops, thus formed closed circuits and serving segregating processes, they also project back to other areas of the cortex, forming open circuits and serving integrative processes (Joel and Weiner, 2000).

Within each loop, the striatum output reaches the GPe/SNr via a direct pathway and via an indirect pathway that travels along the external segment of the globus pallidus (GPe) and the subthalamic nuclei (STN), both pathways providing antagonistic inputs to the GPe/SNr (Albin *et al.*, 1989; DeLong *et al.*, 1985; Gerfen, 1992; Joel and Weiner, 2000). The view of the antagonistic nature of the direct/stimulatory pathway versus the indirect/inhibitory pathway has been criticized as oversimplistic (Parent and Hazrati, 1995). Nevertheless, it is important to keep in mind that activation of medium spiny GABAergic neurons in the striatum by corticostriatal glutamatergic afferents can provide both stimulatory or inhibitory influences on thalamocortical projection (Carlsson *et al.*, 2001).

DA modulates the flow of information within these loops. In primates, DA cells from the VTA project to the ventral striatum and cortex, the dorsal tier of the SN includes cells that project to all striatal regions and cortex, and the ventral tier of the SN projects widely throughout the dorsal striatum, but not to the cortex (for review see Haber and Fudge, 1997). The striatum provides GABA projections back to the VTA and SN. Projections from the VST to midbrain DA neurons are not restricted to the VTA and dorsal tier of the SN (where DA neurons projecting to the VST are located) but also terminate in the ventral tier of SN (where DA neurons projecting to the dorsal striatum are located). On the basis of these observations, Haber proposed that the DA system provides a bridge by which information circulating in the ventral limbic corticostriatal–thalamocortical loops spirals along nigrostriatal loops and feeds into the

cognitive and sensorimotor loops, translating drives into actions (Haber and Fudge, 1997; Haber *et al.*, 2000).

B. DOPAMINERGIC RECEPTORS

The advent of molecular biology techniques in the late 1980s enabled the cloning of these two receptors (Bunzow *et al.*, 1988; Dearry *et al.*, 1990; Monsma *et al.*, 1990; Zhou *et al.*, 1990), as well as three newer DA receptors, termed D3, D4, and D5 receptors (Sokoloff *et al.*, 1990; Sunahara *et al.*, 1991; Tiberi *et al.*, 1991; Van Tol *et al.*, 1991). On the basis of their sequence homologies, the five DA receptor subtypes were classified into two categories (Table I), a D1-like family (including D1 and D5 receptors), and a D2-like family (D2, D3, and D4 receptors) (for reviews see Civelli *et al.*, 1993; Gingrich and Caron, 1993; Sokoloff *et al.*, 1995). This classification is also coherent with the initial distinction of D1 and D2 receptors on the basis of their signaling system, that is, their coupling to Gs and Gi proteins and opposite effect on adenylyl cyclase (Kebabian and Calne, 1979; Spano *et al.*, 1978). D2-like family receptors are both postsynaptic and presynaptic autoreceptors (Diaz *et al.*, 2000; Missale *et al.*, 1998; Palermo-Neto, 1997).

DA receptors differ in their regional localization in the human brain (for reviews see Joyce and Meador-Woodruff, 1997; Meador-Woodruff *et al.*, 1996). D1 receptors show a widespread neocortical distribution, including the PFC, and are also present in high concentration in striatum. D5 receptors are concentrated in the hippocampus and entorhinal cortex (EC). D2 receptors are concentrated in the striatum, with low concentration in medial temporal structures (hippocampus, EC, amygdala) and thalamus. The concentration of D2 receptors in the PFC is extremely low. D3 receptors are present in the striatum, where their concentration is particularly high in the ventral striatum. D4 receptors are present in the PFC and hippocampus, but not detected in the striatum (Lahti *et al.*, 1998).

TABLE I
THE D1-LIKE AND D2-LIKE FAMILY OF DA RECEPTORS

Receptor	D1-like		D2-like		
	D1	D5	D2	D3	D4
Sequence homology	60%		50–70%		
Gene organization	Intronless genes		Genes with intron		
Transduction	Stimulate adenylyl cyclase		Inhibit adenylyl cyclase		
Pharmacology	Moderate to low affinity for antipsychotics		High to moderate affinity for antipsychotics		

In the striatum, D2 receptors are preferentially found in enkephalin-rich GABAergic neurons that participate in the indirect pathways, while D1 receptors are most abundant in dynorphin/substance P GABAergic neurons that contribute to the direct pathways (Gerfen, 1992; Hersch *et al.*, 1995; Le Moine *et al.*, 1990, 1991). In rodents, D3 receptors are expressed in the Island of Calleja and in medium-sized spiny neurons of the rostral and ventromedial shell of nucleus accumbens (Diaz *et al.*, 1995), while its distribution in the striatum is more widespread in humans (Gurevich *et al.*, 1997). The magnitude of the segregation versus coexpression of D1 and D2 receptors in striatal neurons is still a matter of debate (Surmeier *et al.*, 1992, 1996). In the VST, D3 receptors colocalize preferentially on neurons expressing D1 receptors, substance P, dynorphin, and/or neurotensin (Diaz *et al.*, 1995; Ridray *et al.*, 1998) and TrkB, the high-affinity site for the brain-derived neurotrophic factor (BDNF) (Guillin *et al.*, 2001). In the shell of the accumbens, activation of D1 and D3 receptors results in a synergistic enhancement of substance P gene expression (Ridray *et al.*, 1998). In view of the high degree of coexpression of the two receptor subtypes in medium-sized spiny neurons of this region, it seems likely that the synergism occurs at the single-cell level and reflects the participation of the MAP kinase pathway of D3 receptor signaling synergistically increased by the cAMP pathway of the D1 receptor. The segregation of D2 and D1 receptors on different and antagonistic pathways might account for the fact that activation of these receptors is often synergistic at the behavioral level (e.g., stimulation of both D1 and D2 receptors stimulate locomotion), while their effect on intracellular signaling (starting with adenylate cyclase activity) are opposite in many regards. For example, stimulation of D1 and D2 receptors increases or decreases DARP32 phosphorylation, induces or blocks c-fos expression, promotes or inhibits *N*-methyl-D-aspartate (NMDA) receptor function, respectively (Dunah and Standaert, 2001; Konradi, 1998; Leveque *et al.*, 2000; Nguyen *et al.*, 1992; Nishi *et al.*, 1997). Thus, activation of D2 receptors by DA might provide an inhibitory influence to the indirect pathway and activation of D1 receptors by DA might provide a stimulatory influence on the direct pathway. Both effects are expected to result in stimulation of thalamocortical neurons.

However, the action of DA on target neurons should not be viewed in terms of simple excitation or inhibition. Unlike classical "fast" transmitters, DA does not directly gate ion channels, but stimulation of DA G-protein-linked receptor induces a cascade of intracellular signaling that results in modifying the response of the cells to other transmitters. DA is neither "inhibitory" or "excitatory," but its action will depend on the state of the neurons at the time of the stimulation (Yang *et al.*, 1999). Cortical glutamatergic afferents and DA projections converge on GABAergic medium spiny neurons in the striatum, usually on dendritic shafts and spines (for review see Kotter, 1994; Smith and Bolam, 1990; Starr, 1995). At this convergence point, DA has potent modulatory effects on glutamate (Glu) transmission (for review see Cepeda and Levine, 1998; Konradi and

Heckers, 2003; Nicola *et al.*, 2000). Overall, D2 receptor stimulation inhibits NMDA-mediated Glu transmission and long-term potentiation (LTP), and D1 receptor stimulation facilitates Glu transmission and LTP (Centonze *et al.*, 2001; Levine *et al.*, 1996). The effect of D2 receptor stimulation on Glu transmission involves both pre- and postsynaptic effects: D2 stimulation inhibits Glu release and reduces the excitability of medium spiny neurons (Cepeda and Levine, 1998; Cepeda *et al.*, 2001; Leveque *et al.*, 2000; Nicola *et al.*, 2000; Onn *et al.*, 2000; Peris *et al.*, 1988; West and Grace, 2002). In contrast, D1 receptor stimulation generally promotes NMDA function and medium spiny neuron excitability, more specifically when the cells are in a depolarized “upstate,” due to the convergence of excitatory inputs (Dunah and Standaert, 2001; Flores-Hernandez *et al.*, 2002; Hernandez-Lopez *et al.*, 1997; Marti *et al.*, 2002; Morari *et al.*, 1994; West and Grace, 2002; Wilson and Kawaguchi, 1996; Fig. 2).

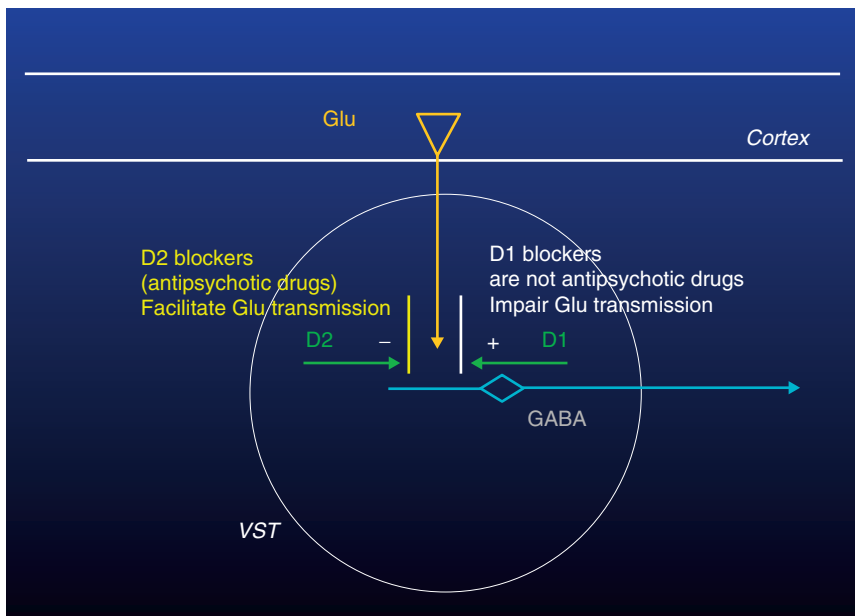


FIG. 2. Opposite modulations of NMDA transmission by D2 and D1 receptors in GABAergic medium spiny neurons in the striatum. D2 and D1 receptors inhibit and facilitate, respectively, Glu transmission. Thus, an excess of D2 receptor stimulation in schizophrenia would further impair NMDA-mediated information flow from the cortex into the striatum. By blocking D2 receptors, antipsychotic drugs promote NMDA transmission. Conversely, D1 receptor antagonists weaken NMDA transmission and are not antipsychotic drugs.

In the PFC, $D_{1/5}$ receptors are localized both on pyramidal cells (dendritic spines and shafts) and on axonal terminals of nondopaminergic neurons (Smiley *et al.*, 1994), while some data suggest that D_4 receptors might be localized on GABA interneurons (Mrzljak *et al.*, 1996). DA modulates pyramidal cell excitability, both directly and via GABAergic interneurons (Yang *et al.*, 1999). Recent data suggest that DA differently affect GABAergic activity in the PFC via D_1 - or D_2 -like mechanisms, whereas $D_{1/5}$ and $D_{2/4}$ receptor stimulation enhances or inhibits GABAergic activity, respectively. Here again, it has been proposed that DA increases the signal-to-noise ratio of glutamatergic afferents (Seamans *et al.*, 2001).

III. Evidence Supporting Alterations of DA Systems in Schizophrenia

A. PHARMACOLOGICAL EVIDENCE

1. *Aversive Pharmacological Effects*

The psychotogenic effect of amphetamine and other DA-enhancing drugs, such as methylphenidate and L-dopa, is a cornerstone of the classical DA hypothesis of schizophrenia. Two sets of observations are relevant to this issue. First, repeated exposure to high doses of psychostimulants in nonschizophrenic subjects might gradually induce paranoid psychosis. This well-documented observation shows that sustained increase in DA activity is psychotogenic. Second, low doses of psychostimulants that are not psychotogenic in healthy subjects might induce or worsen psychotic symptoms in patients with schizophrenia. This observation indicates that patients with schizophrenia have an increased vulnerability to the psychotogenic effects of DA-enhancing drugs.

a. Amphetamine-Induced Psychosis in Nonschizophrenic Subjects. Although mentioned in 1938 (Young and Scoville, 1938), amphetamine-induced psychosis was not clearly recognized as a possible consequence of chronic amphetamine use until 1958 on the publication of a 42-case monograph by Connell (1958). In this chapter, Connell provided the "classical" definition of amphetamine psychosis, as "*a paranoid psychosis with ideas of references, delusions of persecution, auditory and visual hallucinations in the setting of a clear sensorium*" and concluded that "*the mental picture may be indistinguishable from acute or chronic paranoid schizophrenia*" (Connell, 1958).

In the early 1970s, several studies experimentally induced amphetamine psychosis in nonschizophrenic amphetamine-abusers in order to better document the clinical pattern of this syndrome (Angrist and Gershon, 1970; Bell, 1973; Griffith *et al.*, 1968). These experiments formally established that sustained psychostimulant exposure can produce paranoid psychosis in nonschizophrenic

individuals. This reaction does not occur in the context of a delirium since subjects maintain a clear sensorium during the episode and are able to recollect the episode after its resolution. Since these studies were performed before the conceptualization of the symptoms of schizophrenia into positive and negative (Crow, 1980), they did not formally assess negative symptoms. These papers only include anecdotal reports of emotional blunting, withdrawal, or alogia, thereby, suggesting that sustained and excessive stimulation of DA systems does not consistently induce what is now defined as the “negative” symptoms of schizophrenia.

Ellinwood (Ellinwood 1967; Ellinwood *et al.*, 1973) provided one of the most insightful descriptions of amphetamine-induced psychosis by conceptualizing the condition as a continuum that evolves from the gradual onset of paranoid tendencies to delusional paranoia. The first step is characterized by stimulation of interpretative mental activities (great attention to details, intense feeling of curiosity, repetitive searching, and sorting behavior). Ellinwood sees in Sherlock Holmes, a regular cocaine user, a prototypical example of the endless search for meanings (*my mind rebels at stagnation*). With increased exposure, these paranoid tendencies and interests for the minutiae develop into an intermediate stage, which is characterized by marked enhancement of perceptual acuity, sustained “pleasurable” suspiciousness, and compulsive probing behavior. Finally, this inquisitive behavior is reversed and projected to others (persecution), leading to paranoia and ideas of references. The “enhancement of sensitive acuity” develops into hallucinations, initially auditory, then visual and tactile. The sensorium remains clear until toxic delirium is reached. Thought disorders might manifest toward the end of the continuum near the toxic stage. Kapur (2003) recently reformulated and modernized the Ellinwood “Sherlock Holmes” theory by defining schizophrenia psychosis as a state of aberrant salience.

Another important property of psychostimulants is their ability to induce reverse tolerance or “sensitization” (Kalivas *et al.*, 1993; Robinson and Becker, 1986). Long-term sensitization to psychostimulants is a process whereby repeated exposure to these drugs results in an enhanced response on subsequent exposures. The relevance of this process for the pathophysiology of schizophrenia has been reviewed (Laruelle, 2000b; Lieberman *et al.*, 1997). Subjects who abused psychostimulants and experienced stimulant-induced psychotic episodes are reported to remain vulnerable to low doses of psychostimulants (Connell, 1958; Ellinwood *et al.*, 1973; Sato *et al.*, 1983). In these subjects, exposure to psychostimulants at doses that do not normally produce psychotic symptoms can trigger a recurrence of these symptoms. The similarity between these patients and the patients with schizophrenia in terms of vulnerability to the psychotogenic effects of psychostimulants has led to the theorization that schizophrenia might be associated with an “endogenous” sensitization process (Glenthøj and Hemmingsen, 1997; Laruelle, 2000b; Lieberman *et al.*, 1990).

Considerable research efforts have been devoted to the identification of neuronal substrates involved in sensitization. Several studies have shown that sensitization is associated with increased stimulant-induced DA release in the axonal terminal fields (for references see Laruelle, 2000b). A brain imaging study confirmed that, in humans, sensitization to the effects of amphetamine involves increased amphetamine-induced DA release (Boileau *et al.*, 2003). The imaging studies reviewed below show that patients with schizophrenia display an enhanced amphetamine-induced DA release, supporting the notion of an endogenous sensitization process of subcortical DA system in schizophrenia.

b. Psychotogenic Effects of Amphetamine in Schizophrenic Patients. A number of studies reviewed by Lieberman *et al.* (1987b) provided evidence that patients with schizophrenia, as a group, display increased sensitivity to the psychotogenic effects of acute psychostimulant administration. In other terms, some, but not all patients with schizophrenia present emergence or worsening of psychotic symptoms after acute exposure to psychostimulants at doses that do not induced psychosis in healthy subjects. The psychotic response appears to be state dependent. First, patients who responded with a psychotic reaction to a psychostimulant challenge during an acute episode failed to show such a response when they were in remission. Second, the propensity to present a psychotic reaction to a psychostimulant challenge is predictive of relapse on antipsychotic discontinuation. Thus, the clinical response to stimulants might “reveal” an active phase of the illness that is not readily identifiable by the clinical symptomatology in the absence of a psychostimulant administration.

2. Therapeutic Pharmacological Effects

Since the recognition in 1952 of the antipsychotic properties of chlorpromazine (Delay *et al.*, 1952), antipsychotic medications have fundamentally altered the course and the prognosis of schizophrenia. They have proven effective at reducing the severity of symptoms and preventing episodes of illness exacerbation. To date, D2 receptor antagonism is the only pharmacological property shared by all antipsychotic drugs. The clinical dose of these drugs is related to their affinity for D2 receptors. D2 receptor antagonism appears both necessary and sufficient for antipsychotic action (as demonstrated by the selective D2 receptor antagonist amisulpride). The fact that patients with schizophrenia improve following administration of D2 receptor antagonists is one of the few irrefutable pieces of evidence in schizophrenia (Weinberger, 1987).

D2 receptor blockade by antipsychotic drugs has been confirmed by a large number of imaging studies (reviewed in Talbot and Laruelle, 2002). In general, these studies failed to observe a relationship between the degree of D2 receptor occupancy and the quality of the clinical response. However, most studies reported doses achieving more than 50% occupancy. The minimum occupancy required for a therapeutic response remains somewhat uncertain. Two studies

performed with low doses of relatively selective D2 receptor antagonists (haloperidol and raclopride) suggest that a minimum of 50% occupancy is required to observe a rapid clinical response (Kapur *et al.*, 2000; Nordstrom *et al.*, 1993). Imaging studies have repeatedly confirmed the existence of a striatal D2 receptor occupancy threshold (about 80%) above which extrapyramidal symptoms (EPS) are likely to occur (Farde *et al.*, 1992). Thus, these data suggest the existence of a therapeutic window between 50% and 80% striatal D2 receptor occupancy. Within this window, the relationship between occupancy and response is unclear, presumably because the variability in endogenous DA (Frankle *et al.*, 2004). Furthermore, the occupancy threshold required for therapeutic effects might differ among drugs.

The introduction of a second generation of antipsychotic (SGA) drugs since the early 1990s has not fundamentally altered the prominence of D2 receptor antagonism in the current treatment of schizophrenia. Most SGAs also potently interact with other receptors, such as the serotonin 5-HT_{2A} receptors, but the possibility to achieve an “atypical” profile with a pure D2 receptor antagonist, such as amisulpride, indicates that serotonin pharmacological effects are not absolutely required to produce this effect.

On the other hand, imaging studies have generally reported lower occupancies of striatal D2 receptors at therapeutic doses of SGAs compared to first generation antipsychotic drugs (FGAs). This seems to be especially true for amisulpride, clozapine, and quetiapine, which provide 50–60% D2 receptor occupancy range at clinically effective doses (for review and references see Abi-Dargham and Laruelle, 2005). In contrast, studies with FGAs often reported occupancies exceeding 75%. Thus, a parsimonious hypothesis to account for the SGA superiority is that, in general, clinical results obtained after moderate occupancies (50–75%) are better than after high occupancies (75–100%), and that, for a variety of reasons, SGAs tend to maintain lower occupancies than FGAs. The alternate hypothesis is that the D2 receptor occupancy required for therapeutic effects is lower in SGAs than FGAs. Should the alternate hypothesis be true, the mechanisms responsible for the gain in the occupancy–efficacy relationship of SGAs remain to be fully elucidated.

A potentially important synergistic effect of 5-HT_{2A} and D2 receptor antagonism is to increase prefrontal DA, an effect not observed with selective D2 or 5-HT_{2A} receptor antagonists administered alone (Gessa *et al.*, 2000; Ichikawa *et al.*, 2001; Melis *et al.*, 1999; Pehek and Yamamoto, 1994; Youngren *et al.*, 1999). This effect might be mediated by the stimulation of 5-HT_{1A} receptors: it is blocked by 5-HT_{1A} antagonists and is also observed following the combination of 5-HT_{1A} receptor agonism and D2 receptor antagonism (Ichikawa *et al.*, 2001; Rollema *et al.*, 2000). Aripiprazole, clozapine, quetiapine, and ziprasidone are also 5-HT_{1A} partial agonists, and this additional property might also contribute to their ability to increase prefrontal DA. As discussed in Section III.B, a decreased prefrontal

DA function might contribute to the cognitive deficits present in patients with schizophrenia, and it is possible that an increase in prefrontal DA induced by SGAs might mediate some of the modest cognitive improvements induced by these drugs (Keefe *et al.*, 1999). Yet, it is unclear whether this increase in prefrontal DA, documented as an acute response in animal studies, is sustained during the course of treatment in patients with schizophrenia.

B. POSTMORTEM STUDIES

The discovery of the antipsychotic effect of D2 receptor blockade inspired numerous postmortem studies seeking to determine whether schizophrenia was associated with altered parameters of DA transmission. These postmortem studies have for the most part failed to provide definitive answers, partly because of the confounding effects of antemortem antipsychotic treatment.

1. *Tissue DA and HVA*

Direct measures of tissue content of DA and its metabolites have failed to demonstrate consistent and reproducible abnormalities (for review see Davis *et al.*, 1991; Reynolds, 1989). It should be noted, however, that some studies have reported higher DA tissue levels in samples from patients with schizophrenia in subcortical regions such as caudate (Owen *et al.*, 1978), nucleus accumbens (Mackay *et al.*, 1982), or amygdala (Reynolds, 1983).

2. *D2 Receptors*

Increased density of striatal D2 receptors in patients with schizophrenia has been a consistent finding in a large number of postmortem studies (Cross *et al.*, 1983; Dean *et al.*, 1997; Hess *et al.*, 1987; Joyce *et al.*, 1988; Knable *et al.*, 1994; Lahti *et al.*, 1996; Lee *et al.*, 1978; Mackay *et al.*, 1982; Marzella *et al.*, 1997; Mita *et al.*, 1986; Owen *et al.*, 1978; Reynolds *et al.*, 1987; Ruiz *et al.*, 1992; Seeman *et al.*, 1984, 1987, 1993; Sumiyoshi *et al.*, 1995). Because chronic neuroleptic administration upregulates D2 receptor density (Burt *et al.*, 1977), it is likely that these postmortem findings are related to prior neuroleptic exposure rather than to the disease process per se. In light of these very consistent results with [^3H]spiperone, it is interesting to note that the striatal binding of [^3H]raclopride has been reported to increase in many studies (Dean *et al.*, 1997; Marzella *et al.*, 1997; Ruiz *et al.*, 1992; Sumiyoshi *et al.*, 1995), but normal in several others (Knable *et al.*, 1994; Lahti *et al.*, 1996; Seeman *et al.*, 1993), even in patients exposed to neuroleptic drugs prior to death. This observation suggests that the increase in [^3H]raclopride binding is of lower magnitude than the one of [^3H]spiperone binding. This discrepancy might simply reflect the observation that, for reasons that are not currently understood, antipsychotic drugs upregulate more [^3H]spiperone

than [^3H]raclopride binding to D2 receptors (Schoots *et al.*, 1995; Tarazi *et al.*, 1997).

3. D3 Receptors

A significant increase in D3 receptor number in VST samples from patients with schizophrenia who were off neuroleptics at the time of death has been reported in one study (Gurevich *et al.*, 1997). In contrast, in patients who had been treated with neuroleptics up to the time of death, D3 receptor levels did not differ significantly from those of controls (Gurevich *et al.*, 1997). These data were interpreted as indicating that antipsychotics downregulate the D3 receptor in schizophrenic patients who otherwise have a higher density of this receptor in the VST. The D3 receptor gene expression is under the control of a neurotrophin, called BDNF, that is synthesized either in the VTA and the PFC and released in the VST, where it maintains the expression of the D3 receptor (Guillin *et al.*, 2001). One study (Takahashi *et al.*, 2000) has shown increased and two decreased (Hashimoto *et al.*, 2005; Weickert *et al.*, 2003) of BDNF levels in the brain of patients with schizophrenia. D3 receptors are upregulated in the presence of hyperdopaminergic tone (Bordet *et al.*, 1997; Fauchey *et al.*, 2000; Guillin *et al.*, 2001; Le Foll *et al.*, 2002), under the control of the BDNF, whose synthesis is in turn under the control of the activity of neurons projecting from the PFC or the VTA in the VST.

4. D4 Receptors

On the basis of ligand subtraction techniques, several studies have reported increased D4-like receptors in schizophrenia (Marzella *et al.*, 1997; Murray *et al.*, 1995; Seeman *et al.*, 1993; Sumiyoshi *et al.*, 1995). These findings were not confirmed by other studies using the same technique (Lahti *et al.*, 1996; Reynolds and Mason, 1994), nor by a study using [^3H]NGD 94-1, a selective D4 ligand (Lahti *et al.*, 1998). Moreover, the hypothesis that clozapine might act by blocking the D4 receptor was not supported by a clinical trial with the D4 selective agent L745,870 (Kramer *et al.*, 1997).

5. D1 Receptors

Striatal D1 receptors have generally been reported to be unaltered in schizophrenia (Joyce *et al.*, 1988; Pimoule *et al.*, 1985; Reynolds and Czudek, 1988; Seeman *et al.*, 1987), although one study reported decreased density (Hess *et al.*, 1987). In the PFC, one study reported no changes (Laruelle *et al.*, 1990) and one reported a nonsignificant increase (Knable *et al.*, 1996).

6. DA Transporter

A large number of studies have reported unaltered DA transporter density (DAT) in the striatum of patients with schizophrenia (Chinaglia *et al.*, 1992;

Czudek and Reynolds, 1989; Hirai *et al.*, 1988; Joyce *et al.*, 1988; Knable *et al.*, 1994; Pearce *et al.*, 1990).

7. *Tyrosine Hydroxylase Immunolabeling*

A recent and interesting postmortem finding regarding DA parameters in patients with schizophrenia is the observation of decreased tyrosine hydroxylase (TH)-labeled axons in layers III and VI of the EC and in layer VI of the PFC, a finding suggesting that schizophrenia might be associated with deficit in DA transmission in the EC and PFC (Akil *et al.*, 1999, 2000). This finding was clearly unrelated to premortem neuroleptic exposure. Benes *et al.* (1997) observed no significant changes in TH-positive varicosities in the DLPFC. In the anterior cingulate region (layer II), these authors observed a significant shift in the distribution of TH varicosities from large neurons to small neurons.

In conclusion, postmortem measurements of indices of DA transmission generated a number of consistent observations in the striatum: (1) The binding of radioligand to D2-like receptors in the striatum of patients with schizophrenia is increased, but the magnitude of this increase varies with the type of radioligands used, and it is difficult to exclude the contribution of premortem antipsychotic exposure in this set of findings. (2) Striatal DAT and D1 receptors density is unaffected in schizophrenia. Several interesting observations such as increase in D3 receptors in the ventral striatum and alteration in TH immunolabeling in several cortical regions do not appear to be consequences of premortem neuroleptic exposure, but these findings have yet to be independently confirmed.

C. IMAGING STUDIES

1. *Striatal DA Function*

The development of PET and SPECT imaging techniques in the late 1980s made possible, for the first time, the examination of DA function *in vivo* in patients with schizophrenia never exposed to antipsychotic drugs (Fig. 3).

a. Striatal D2 and D1 Receptors. Striatal D2 receptor density in schizophrenia has been extensively studied with PET and SPECT imaging. In a meta-analysis (Weinberger and Laruelle, 2001), 17 imaging studies comparing D2 receptor parameters in patients with schizophrenia have been analyzed (included a total of 245 patients and 231 control subjects, Table II) (Abi-Dargham *et al.*, 1998, 2000b; Blin *et al.*, 1989; Breier *et al.*, 1997; Crawley *et al.*, 1986; Hietala *et al.*, 1994b; Knable *et al.*, 1997; Laruelle *et al.*, 1996; Martinot *et al.*, 1990, 1991; Pilowsky *et al.*, 1994; Wong *et al.*, 1986). Updated with a study (Yang *et al.*, 2004), this meta-analysis revealed a small (12%) but significant elevation of striatal D2 receptors in untreated patients with schizophrenia. No clinical correlates of increased D2 receptor-binding parameters could be identified. Studies performed with

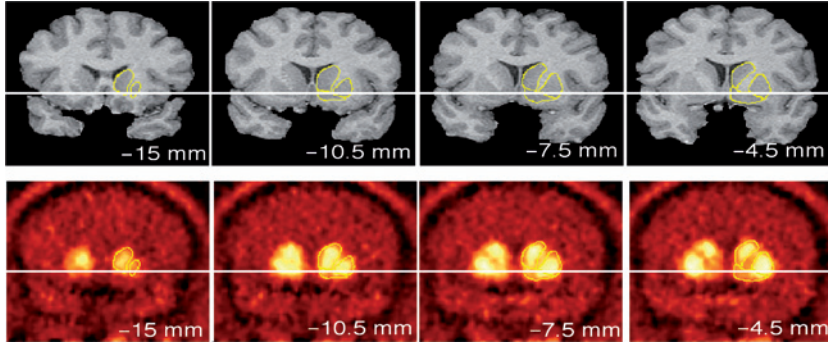


FIG. 3. D2 receptor-binding image with PET and [^{11}C]raclopride. MRI images (above) and PET images (below).

butyrophenones ($n = 7$) show an effect size of 0.96 ± 1.05 , significantly larger than the effect size observed with other ligands (benzamides and lisuride, $n = 11$, 0.19 ± 0.25 , $p = 0.02$). This difference might be due to differences in vulnerability of the binding of these tracers to endogenous DA and elevation of endogenous DA in schizophrenia (Seeman, 1988; Seeman *et al.*, 1989). Interestingly, the fact that D2 receptor levels are increased in healthy monozygotic twin compared to dizygotic twin of patients with schizophrenia has lead to the conclusion that the caudate DA D2 receptor upregulation is related to genetic risk for schizophrenia (Hirvonen *et al.*, 2005). Imaging studies of D1 receptors have consistently failed to detect abnormalities of D1 receptor availability in the striatum of patients with schizophrenia (Abi-Dargham *et al.*, 2002; Karlsson *et al.*, 2002; Okubo *et al.*, 1997).

b. Striatal Amphetamine-Induced DA Release. The decrease in [^{11}C]raclopride and [^{123}I]IBZM *in vivo* binding following acute amphetamine challenge has been well validated as a measure of the change in D2 receptor stimulation by DA due to amphetamine-induced DA release (Breier *et al.*, 1997; Laruelle *et al.*, 1997b; Villemagne *et al.*, 1999) (Table III).

Our results (Abi-Dargham *et al.*, 1998; Laruelle *et al.*, 1996), which have been independently replicated (Breier *et al.*, 1997), showed that the amphetamine-induced decrease in [^{11}C]raclopride or [^{123}I]IBZM binding is elevated in untreated patients with schizophrenia compared to well-matched controls (Fig. 4). A significant relationship was observed between the magnitude of this effect and transient induction or worsening of positive symptoms. This exaggerated response of the DA system to amphetamine was observed in both first episode/drug-naïve patients and previously treated patients (Laruelle *et al.*, 1999), but was larger in

TABLE II
IMAGING STUDIES OF STRIATAL D2 RECEPTOR PARAMETERS IN DRUG-NAIVE AND DRUG-FREE PATIENTS WITH SCHIZOPHRENIA

Class radiotracer	Radiotracer	Study	Controls, n	Patients, n (DN/DF) ^a	Method	Outcome	Controls (n .mean \pm SD) ^b	Patients (n .mean \pm SD) ^b	p	Effect size ^c	Ratio SD
Butyrophenones	[¹¹ C]NMSP	Wong <i>et al.</i> (1986)	11	15 (10/5)	Kinetic	B_{max}	100 \pm 50	253 \pm 105	<0.05	3.06	2.10
	[⁷⁶ Br]SPI	Crawley <i>et al.</i> (1986)	8	16 (12/4)	Ratio	S/C	100 \pm 14	111 \pm 12	<0.05	0.79	0.86
	[⁷⁶ Br]SPI	Blin <i>et al.</i> (1989)	8	8 (0/8)	Ratio	S/C	100 \pm 14	104 \pm 14	ns	0.28	1.00
	[⁷⁶ Br]SPI	Martinot <i>et al.</i> (1990)	12	12 (0/12)	Ratio	S/C	100 \pm 11	101 \pm 15	ns	0.14	1.41
	[¹¹ C]NMSP	Tunc <i>et al.</i> (1993)	17	10 (8/2)	Kinetic	B_{max}	100 \pm 80	173 \pm 143	0.08	0.91	1.79
	[¹¹ C]NMSP	Nordstrom <i>et al.</i> (1995)	7	7 (7/0)	Kinetic	B_{max}	100 \pm 25	133 \pm 63	ns	1.33	2.50
	[¹¹ C]NMSP	Okubo <i>et al.</i> (1997)	18	17(10/7)	Kinetic	k3	100 \pm 21	104 \pm 16	ns	0.19	0.74
Benzamides	[¹¹ C]raclopride	Farde <i>et al.</i> (1990)	20	18 (18/0)	Equilibrium	B_{max}	100 \pm 29	107 \pm 18	ns	0.23	0.63
	[¹¹ C]raclopride	Hietala <i>et al.</i> (1994a)	10	13 (0/13)	Equilibrium	B_{max}	100 \pm 22	112 \pm 43	ns	0.55	1.99
	[¹²³ I]IBZM	Pilowsky <i>et al.</i> (1994)	20	20 (17/3)	Ratio	S/FC	100 \pm 8	99 \pm 7	ns	-0.07	0.82

(Continued)

TABLE II (Continued)

Class radiotracer	Radiotracer	Study	Controls, n	Patients, n (DN/DF) ^a	Method	Outcome	Controls (n .mean \pm SD) ^b	Patients (n .mean \pm SD) ^b	p	Effect size ^c	Ratio SD
Ergot alk.	[¹²³ I]IBZM	Laruelle <i>et al.</i> (1996)	15	15 (1/14)	Equilibrium	BP	100 \pm 26	115 \pm 33	ns	0.56	1.25
	[¹²³ I]IBZM	Knable <i>et al.</i> (1997)	16	21 (1/20)	Equilibrium	BP	100 \pm 29	97 \pm 38	ns	-0.12	1.31
	[¹¹ C]raclopride	Breier <i>et al.</i> (1997)	12	11 (6/5)	Equilibrium	BP	100 \pm 18	100 \pm 30	ns	0.02	1.69
	[¹²³ I]IBZM	Abi-Dargham <i>et al.</i> (1998)	15	15 (2/13)	Equilibrium	BP	100 \pm 20	102 \pm 49	ns	0.09	2.50
	[¹²³ I]IBZM	Abi-Dargham <i>et al.</i> (2000b)	18	18 (8/10)	Equilibrium	BP	100 \pm 13	104 \pm 14	ns	0.33	1.11
	[¹²³ I]IBZM	Yang <i>et al.</i> (2004)	12	11(11/0)	Ratio	S/C	100 \pm 11	101 \pm 11	ns	0.09	1
	[⁷⁶ Br]lisuride	Martinot <i>et al.</i> (1991)	14	19 (10/9)	Ratio	S/C	100 \pm 10	104 \pm 12	ns	0.45	1.21
	[⁷⁶ Br]lisuride	Martinot <i>et al.</i> (1994)	10	10 (2/8)	Ratio	S/C	100 \pm 10	100 \pm 13	ns	0.00	1.29

^aDN, drug naive; DF, drug free.^bMean normalized to mean of control subjects.^cEffect size calculated as (mean patients - mean controls)/SD controls.

TABLE III
IMAGING STUDIES OF STRIATAL PRESYNAPTIC DA PARAMETERS IN DRUG-NAIVE AND DRUG-FREE PATIENTS WITH SCHIZOPHRENIA

Parameter	Study	Controls, n	Patients, n (DN/DF/T) ^a	Radiotracer (per challenge)	Method	Outcome	Controls (n , mean \pm SD) ^b	Patients (n , mean \pm SD) ^b	p	Effect size ^c
DOPA accumulation	Reith <i>et al.</i> (1994)	13	5 (4/0/1)	[¹⁸ F]DOPA	Kinetic	k3	100 \pm 23	120 \pm 15	<0.05	0.91
	Hietala <i>et al.</i> (1995)	7	7 (7/0/0)	[¹⁸ F]DOPA	Graphical	Ki	100 \pm 11	117 \pm 20	<0.05	1.54
	Dao-Castellana <i>et al.</i> (1997)	7	6 (2/4/0)	[¹⁸ F]DOPA	Graphical	Ki	100 \pm 11	103 \pm 40	ns	0.30
	Lindstrom <i>et al.</i> (1999)	10	12 (10/2)	[¹¹ C]DOPA	Graphical	Ki	100 \pm 17	113 \pm 12	<0.05	0.77
	Hietala <i>et al.</i> (1999)	13	10 (10/0)	[¹⁸ F]DOPA	Graphical	Ki	100 \pm 14	115 \pm 28	<0.05	1.09
	Elkashef <i>et al.</i> (2000)	13	19 (0/9/10)	[¹⁸ F]DOPA	Ratio	Ki	100 \pm 11.7	92.4 \pm 9.7	<0.05	-0.65
	Meyer-Lindenberg <i>et al.</i> (2002)	6	6 (0/6/0)	[¹⁸ F]DOPA	Graphical	Ki	100 \pm 9.7	119 \pm 9.7	<0.02	1.96
	McGowan <i>et al.</i> (2004)	12	16 (0/0/16)	[¹⁸ F]DOPA	Graphical	Ki	100 \pm 9.3	115 \pm 8.2	0.001	1.6
Amphetamine- induced DA release	Laruelle <i>et al.</i> (1996)	15	15 (2/13/0)	[¹²³ I]IBZM/ amphetamine	Equilibrium	Delta BP	100 \pm 113	271 \pm 221	<0.05	1.51
	Breier <i>et al.</i> (1997)	18	18 (8/10/0)	[¹¹ C]raclopride/ amphetamine	Equilibrium	Delta BP	100 \pm 43	175 \pm 82	<0.05	1.73
	Abi-Dargham <i>et al.</i> (1998)	16	21 (1/20/0)	[¹²³ I]IBZM/ amphetamine	Equilibrium	Delta BP	100 \pm 88	194 \pm 145	<0.05	1.07

(Continued)

TABLE III (Continued)

Parameter	Study	Controls, n	Patients, n (DN/DF/T) ^a	Radiotracer (per challenge)	Method	Outcome	Controls (n , mean \pm SD) ^b	Patients (n , mean \pm SD) ^b	p	Effect size ^c
Baseline DA concentration	Abi-Dargham <i>et al.</i> (2000b)	18	18 (8/10/0)	[¹²³ I]IBZM/ α MPT	Equilibrium	Delta BP	100 \pm 78	211 \pm 122	<0.05	1.43
DAT density	Laakso <i>et al.</i> (2000)	9	9 (9/0/0)	[¹⁸ F]CFT	Ratio	S/C	100 \pm 12	101 \pm 13	<0.05	0.11
	Laruelle <i>et al.</i> (2000a)	22	22 (2/20/0)	[¹²³ I]CIT	Equilibrium	BP	100 \pm 17	93 \pm 20	<0.05	-0.43
	Hsiao <i>et al.</i> (2003)	12	12 (12/0/0)	[^{99m} Tc]TRODAT	Ratio	S/Occ	100 \pm 18	104 \pm 21	ns	0.22

^aDN, drug naive; DF, drug free; T, treated with antipsychotics.

^bMean normalized to mean of control subjects.

^cEffect size calculated as (mean patients - mean controls)/SD controls.

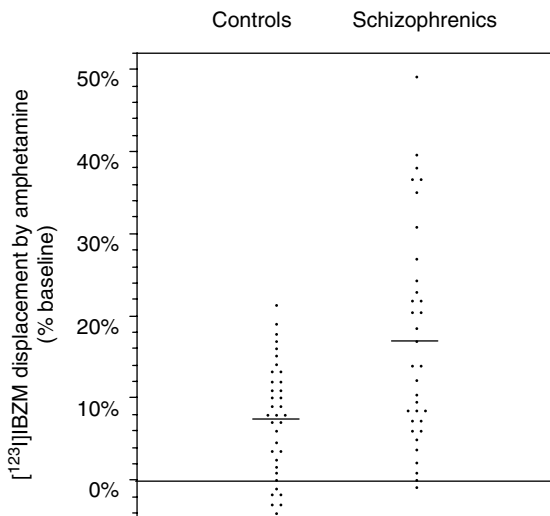


FIG. 4. Effect of amphetamine (0.3 mg/kg) on [^{123}I]IBZM binding in healthy controls and untreated patients with schizophrenia. The y axis shows the percentage decrease in [^{123}I]IBZM-binding potential induced by amphetamine, which is a measure of the increased occupancy of D2 receptors by DA following the challenge. Increased stimulation of D2 receptors in schizophrenia was associated with transient worsening or emergence of positive symptoms.

patients experiencing an episode of illness exacerbation than in patients in remission at the time of the scan (Laruelle *et al.*, 1999). This exaggerated DA reactivity did not appear to be a nonspecific effect of stress, as higher self-reports of anxiety before the experiments were not associated with larger effect of amphetamine on [^{123}I]IBZM binding. Furthermore, nonpsychotic subjects with unipolar depression, who reported levels of anxiety similar to the schizophrenic patients at the time of the scan, showed normal amphetamine-induced displacement of [^{123}I]IBZM (Parsey *et al.*, 2001).

These findings have generally been interpreted as reflecting an increase in synaptic DA following amphetamine in the schizophrenic group. Another interpretation of these observations would be that schizophrenia is associated with increased affinity of D2 receptors for DA.

c. DAT Transporters. Three imaging studies (listed in Table II) have confirmed the *in vitro* observation of normal striatal DAT density in schizophrenia (Laakso *et al.*, 2000; Laruelle *et al.*, 2000b). In addition, no association between amphetamine-induced DA release and DAT density was found (Laruelle *et al.*, 2000b), suggesting that the increased presynaptic output revealed by the studies reviewed above is not due to higher terminal density.

d. *Vesicular Monoamine Transporter*. Using the radiotracer [^{11}C]DTBZ (Taylor *et al.*, 2000) were not able to show any difference in vesicular monoamine transporter binding potential (BP) in patients with schizophrenia compared to healthy subjects.

e. *Baseline Occupancy of Striatal D2 Receptors by DA*. In rodents, acute depletion of synaptic DA is associated with an acute increase in the *in vivo* binding of [^{11}C]raclopride or [^{123}I]IBZM to D2 receptors (for review see Laruelle, 2000a). The increased binding is observed *in vivo* but not *in vitro*, indicating that it is not due to receptor upregulation (Laruelle *et al.*, 1997a) but to removal of endogenous DA and unmasking of D2 receptors previously occupied by DA. A similar acute DA depletion technique paired with D2 receptor imaging in humans using αHPT has been developed to assess the degree of occupancy of D2 receptors by DA (Laruelle *et al.*, 1997a). In schizophrenia, there was a higher occupancy of D2 receptors by DA in patients experiencing an episode of illness exacerbation, compared to healthy controls (Table III) (Abi-Dargham *et al.*, 2000b). Again assuming normal affinity of D2 receptors for DA, the data are consistent with higher DA synaptic levels in patients with schizophrenia. Higher synaptic DA levels in patients with schizophrenia were predictive of good therapeutic response of these symptoms following 6 weeks of treatment with atypical antipsychotic medications (Abi-Dargham *et al.*, 2000b).

f. *Striatal DOPA Decarboxylase Activity*. The eight studies which have reported rates of DOPA decarboxylase in patients with schizophrenia, using [^{18}F]DOPA or [^{11}C]DOPA are summarized in Table II. Six out of eight studies reported increased accumulation of DOPA in the striatum of patients with schizophrenia (Dao-Castellana *et al.*, 1997; Elkashef *et al.*, 2000; Hietala *et al.*, 1995, 1999; Lindstrom *et al.*, 1999; McGowan *et al.*, 2004; Meyer-Lindenberg *et al.*, 2002; Reith *et al.*, 1994), one reported no change (Dao-Castellana *et al.*, 1997) and one study reported reduced [^{18}F]DOPA striatal uptake (Elkashef *et al.*, 2000). Three studies involved first-episode schizophrenia, and all three showed an increase of DOPA in the striatum (Hietala *et al.*, 1995, 1999; Lindstrom *et al.*, 1999). Interestingly, a recent study observed a relationship between poor prefrontal activation during the Wisconsin Card Sorting task and elevated [^{18}F]DOPA accumulation in the striatum, suggesting a link between alteration of the dorsolateral PFC function and increased striatal DA activity in schizophrenia (Meyer-Lindenberg *et al.*, 2002). In rats as in anesthetized pigs, increases in aromatic L-amino acid decarboxylase (AADC) activity *in vitro* and *in vivo* have been reported following acute treatment with DA antagonists (Cho *et al.*, 1997; Danielsen *et al.*, 2001; Zhu *et al.*, 1993). Conversely, acute treatment with the DA agonist apomorphine decreases ^{11}C -DOPA influx in monkeys (Torstenson *et al.*, 1998). Evidence for such effects in humans, however, is extremely limited. Thus, in the only comprehensive study to date Grunder *et al.* (2003) reported a decrease in [^{18}F]DOPA uptake in nine

patients with schizophrenia following subchronic treatment with haloperidol, suggesting that chronic neuroleptic administration will tend to decrease AADC activity and hence DA synthesis. Interestingly, acute administration of antipsychotics increases DA neurons firing whereas chronic administration decreases the number of spontaneously active DA neurons in the rat SN (Grace, 1991), suggesting that the different effects of antipsychotics on AADC activity in the living brain could reflect such phenomena.

2. Prefrontal DA Function and Schizophrenia

Indirect evidence supports the hypothesis that a deficit in prefrontal DA function might contribute to prefrontal impairment in schizophrenia. Abundant preclinical evidences have documented the importance of prefrontal DA function for cognition (for review see Goldman-Rakic, 1994; Goldman-Rakic *et al.*, 2000). This important role has been recently confirmed in humans, by the repeated observation that the carriers of the high-activity allele of catechol-*O*-methyltransferase (COMT), an enzyme involved in DA metabolism, display lower performance in various cognitive tasks compared to carriers of the allele that induces lower concentration of DA in PFC (for review see Goldberg and Weinberger, 2004). Clinical studies have suggested a relationship between low cerebrospinal fluid homovanillic acid, a measure reflecting low DA activity in the PFC, and poor performance at tasks involving WM in schizophrenia (Kahn *et al.*, 1994; Weinberger *et al.*, 1988). Administration of DA agonists might have beneficial effects on the pattern of prefrontal activation measured with PET during these tasks (Daniel *et al.*, 1991; Dolan *et al.*, 1995). While these observations are consistent with the hypothesis of a hypodopaminergic state in the PFC of patients with schizophrenia, they do not constitute direct evidence.

The only parameter of DA transmission that is currently quantifiable using noninvasive *in vivo* studies is D1 receptor availability. Three PET studies of prefrontal D1 receptor availability in patients with schizophrenia have recently been published. Two studies were performed with [^{11}C]SCH 23390. The first reported decreased [^{11}C]SCH 23390 BP in the PFC (Okubo *et al.*, 1997) and the other reported no change (Karlsson *et al.*, 2002). One study was performed with [^{11}C]NNC 112 (Abi-Dargham *et al.*, 2002) and reported increased [^{11}C]NNC 112 BP in the DLPFC and no change in other regions of the PFC such as the medial prefrontal cortex (MPFC) or the orbitofrontal cortex. In patients with schizophrenia, increased [^{11}C]NNC 112 binding in the DLPFC was predictive of poor performance on a working memory task (Fig. 5; Abi-Dargham *et al.*, 2002). Many potential factors including patient heterogeneity and differences in the boundaries of the sampled regions might account for these discrepancies. However, severity of deficits at tasks involving WM were reported to be associated with both decreased

Thus, the increase in DLPFC [^{11}C]NNC 112 BP observed in schizophrenia might be related to a compensatory but inefficient upregulation of D1 receptors following sustained DA depletion, and it is conceivable that such an upregulation might not be detectable with [^{11}C]SCH 23390. Studies with both radiotracers on the same patients are required to clarify this issue.

IV. Conclusions

The development of new imaging methods aiming at measuring presynaptic activity in striatal DA afferents provided data consistent with the view that schizophrenia is associated with hyperactivity of subcortical transmission at D2 receptors (Fig. 6). These results are consistent with the known mode of action

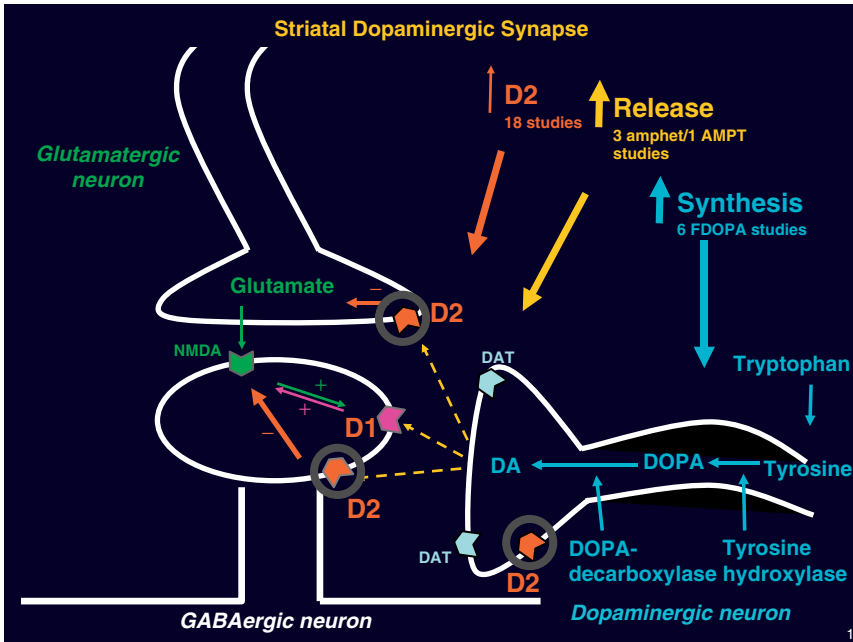


FIG. 6. Striatal dopaminergic synapse, summary of findings: evidence for excess DA transmission derives from pre- and postsynaptic studies. Excess DA transmission may impair glutamatergic NMDA transmission by a D2-mediated impaired presynaptic release of glutamate and an imbalance of D1/D2 opposing effects onto NMDA transmission. See text and tables for references.

of current antipsychotic treatment (D2 receptor blockade), with the psychotogenic effects of sustained stimulation of DA function by psychostimulants, and with the “classical” DA hypothesis of schizophrenia derived from these observations. In addition, these results suggest that the DA hyperactivity of subcortical systems is episodic in nature, and account for only some aspects of positive symptomatology. On the other hand, imaging methods might suggest that hypodopaminergia in the DLPFC participate to do pathophysiology of cognitive symptoms endured by patients with schizophrenia.

References

- Abi-Dargham, A., and Laruelle, M. (2005). Mechanisms of action of second generation antipsychotic drugs in schizophrenia: Insights from brain imaging studies. *Eur. Psychiatry* **20**, 15–27.
- Abi-Dargham, A., Gil, R., Krystal, J., Baldwin, R., Seibyl, J., Bowers, M., van Dyck, C., Charney, D., Innis, R., and Laruelle, M. (1998). Increased striatal dopamine transmission in schizophrenia: Confirmation in a second cohort. *Am. J. Psychiatry* **155**, 761–767.
- Abi-Dargham, A., Martinez, D., Mawlawi, O., Simpson, N., Hwang, D. R., Slifstein, M., Anjilvel, S., Pidcock, J., Guo, N. N., Lombardo, I., Mann, J. J., Van Heertum, R., *et al.* (2000a). Measurement of striatal and extrastriatal dopamine D1 receptor binding potential with [¹¹C] NNC 112 in humans: Validation and reproducibility. *J. Cereb. Blood Flow Metab.* **20**, 225–243.
- Abi-Dargham, A., Rodenhiser, J., Printz, D., Zea-Ponce, Y., Gil, R., Kegeles, L., Weiss, R., Cooper, T., Mann, J. J., Van Heertum, R., Gorman, J., and Laruelle, M. (2000b). Increased baseline occupancy of D2 receptors by dopamine in schizophrenia. *Proc. Natl. Acad. Sci. USA* **97**, 8104–8109.
- Abi-Dargham, A., Mawlawi, O., Lombardo, I., Gil, R., Martinez, D., Huang, Y., Hwang, D. R., Keilp, J., Kochan, L., Van Heertum, R., Gorman, J. M., and Laruelle, M. (2002). Prefrontal dopamine D1 receptors and working memory in schizophrenia. *J. Neurosci.* **22**, 3708–3719.
- Akil, M., Pierri, J. N., Whitehead, R. E., Edgar, C. L., Mohila, C., Sampson, A. R., and Lewis, D. A. (1999). Lamina-specific alterations in the dopamine innervation of the prefrontal cortex in schizophrenic subjects. *Am. J. Psychiatry* **156**, 1580–1589.
- Akil, M., Edgar, C. L., Pierri, J. N., Casali, S., and Lewis, D. A. (2000). Decreased density of tyrosine hydroxylase-immunoreactive axons in the entorhinal cortex of schizophrenic subjects. *Biol. Psychiatry* **47**, 361–370.
- Albin, R. L., Young, A. B., and Penney, J. B. (1989). The functional anatomy of basal ganglia disorders. *Trends Neurosci.* **12**, 366–375.
- Alexander, G. E., DeLong, M. R., and Strick, P. L. (1986). Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu. Rev. Neurosci.* **9**, 357–381.
- Angrist, B. M., and Gershon, S. (1970). The phenomenology of experimentally induced amphetamine psychosis: Preliminary observation. *Biol. Psychiatry* **2**, 95–107.
- Angrist, B. M., and van Kammen, D. P. (1984). CNS stimulants as a tool in the study of schizophrenia. *Trends Neurosci.* **7**, 388–390.
- Bell, D. S. (1973). The experimental reproduction of amphetamine psychosis. *Arch. Gen. Psychiatry* **29**, 35–40.

- Benes, F. M., Todtenkopf, M. S., and Taylor, J. B. (1997). Differential distribution of tyrosine hydroxylase fibres on small and large neurons in layer II of anterior cingulate cortex of schizophrenic brain. *Synapse* **25**(1), 80–92.
- Blin, J., Baron, J. C., Cambon, H., Bonnet, A. M., Dubois, B., Loc'h, C., Maziere, B., and Agid, Y. (1989). Striatal dopamine D2 receptors in tardive dyskinesia: PET study. *J. Neurol. Neurosurg. Psychiatry* **52**, 1248–1252.
- Boileau, I., Dagher, A., Leyton, M., Gunn, R., Diksic, M., and Benkelfat, C. (2003). Sensitization to psychostimulants: A PET/[11C]-raclopride study in healthy volunteers. *ACNP Annual Meeting Abstracts*.
- Bordet, R., Ridray, S., Carboni, S., Diaz, J., Sokoloff, P., and Schwartz, J. C. (1997). Induction of dopamine D3 receptor expression as a mechanism of behavioral sensitization to levodopa. *Proc. Natl. Acad. Sci. USA* **94**, 3363–3367.
- Breier, A., Su, T. P., Saunders, R., Carson, R. E., Kolachana, B. S., de Bartolomeis, A., Weinberger, D. R., Weisenfeld, N., Malhotra, A. K., Eckelman, W. C., and Pickar, D. (1997). Schizophrenia is associated with elevated amphetamine-induced synaptic dopamine concentrations: Evidence from a novel positron emission tomography method. *Proc. Natl. Acad. Sci. USA* **94**, 2569–2574.
- Bunzow, J. R., Van Tol, H. H., Grandy, D. K., Albert, P., Salon, J., Christie, M., Machida, C. A., Neve, K. A., and Civelli, O. (1988). Cloning and expression of a rat D2 dopamine receptor cDNA. *Nature* **336**, 783–787.
- Burt, D. R., Creese, I., and Snyder, S. H. (1977). Antischizophrenic drugs: Chronic treatment elevates dopamine receptor binding in brain. *Science* **196**, 326–328.
- Carlsson, A., and Lindqvist, M. (1963). Effect of chlorpromazine or haloperidol on formation of 3-methoxytyramine and normetanephrine in mouse brain. *Acta Pharmacol. Toxicol.* **20**, 140–144.
- Carlsson, A., Waters, N., Holm-Waters, S., Tedroff, J., Nilsson, M., and Carlsson, M. L. (2001). Interactions between monoamines, glutamate, and GABA in schizophrenia: New evidence. *Annu. Rev. Pharmacol. Toxicol.* **41**, 237–260.
- Centonze, D., Picconi, B., Gubellini, P., Bernardi, G., and Calabresi, P. (2001). Dopaminergic control of synaptic plasticity in the dorsal striatum. *Eur. J. Neurosci.* **13**, 1071–1077.
- Cepeda, C., and Levine, M. S. (1998). Dopamine and N-methyl-D-aspartate receptor interactions in the neostriatum. *Dev. Neurosci.* **20**, 1–18.
- Cepeda, C., Hurst, R. S., Altemus, K. L., Flores-Hernandez, J., Calvert, C. R., Jokel, E. S., Grandy, D. K., Low, M. J., Rubinstein, M., Ariano, M. A., and Levine, M. S. (2001). Facilitated glutamatergic transmission in the striatum of D2 dopamine receptor-deficient mice. *J. Neurophysiol.* **85**, 659–670.
- Chinaglia, G., Alvarez, F. J., Probst, A., and Palacios, J. M. (1992). Mesostriatal and mesolimbic dopamine uptake binding sites are reduced in Parkinson's disease and progressive supranuclear palsy: A quantitative autoradiographic study using [3H]mazindol. *Neuroscience* **49**, 317–327.
- Chiodo, L. A., and Bunney, B. S. (1983). Typical and atypical neuroleptics: Differential effects of chronic administration on the activity of A9 and A10 midbrain dopaminergic neurons. *J. Neurosci.* **3**, 1607–1619.
- Cho, S., Neff, N. H., and Hadjiconstantinou, M. (1997). Regulation of tyrosine hydroxylase and aromatic L-amino acid decarboxylase by dopaminergic drugs. *Eur. J. Pharmacol.* **323**, 149–157.
- Civelli, O., Bunzow, J. R., and Grandy, D. K. (1993). Molecular diversity of the dopamine receptors. *Annu. Rev. Pharmacol. Toxicol.* **33**, 281–307.
- Connell, P. H. (1958). "Amphetamine Psychosis." Chapman and Hill, London.
- Crawley, J. C., Owens, D. G., Crow, T. J., Poulter, M., Johnstone, E. C., Smith, T., Oldland, S. R., Veall, N., Owen, F., and Zanelli, G. D. (1986). Dopamine D2 receptors in schizophrenia studied *in vivo*. *Lancet* **2**, 224–225.

- Creese, I., Burt, D. R., and Snyder, S. H. (1976). Dopamine receptor binding predicts clinical and pharmacological potencies of antischizophrenic drugs. *Science* **19**, 481–483.
- Cross, A. J., Crow, T. J., Ferrier, I. N., Johnstone, E. C., McCreadie, R. M., Owen, F., Owens, D. G., and Poulter, M. (1983). Dopamine receptor changes in schizophrenia in relation to the disease process and movement disorder. *J. Neural. Transm. Suppl.* **18**, 265–272.
- Crow, T. J. (1980). Molecular pathology of schizophrenia: More than one disease process? *Br. Med. J.* **280**, 66–68.
- Czudek, C., and Reynolds, G. P. (1989). [3H] GBR 12935 binding to the dopamine uptake site in post-mortem brain tissue in schizophrenia. *J. Neural. Transm.* **77**, 227–230.
- Daniel, D. G., Weinberger, D. R., Jones, D. W., Zigun, J. R., Coppola, R., Handel, S., Bigelow, L. B., Goldberg, T. E., Berman, K. F., and Kleinman, J. E. (1991). The effect of amphetamine on regional cerebral blood flow during cognitive activation in schizophrenia. *J. Neurosci.* **11**, 1907–1917.
- Danielsen, E. H., Smith, D., Hermansen, F., Gjedde, A., and Cumming, P. (2001). Acute neuroleptic stimulates DOPA decarboxylase in porcine brain *in vivo*. *Synapse* **41**, 172–175.
- Dao-Castellana, M. H., Paillere-Martinot, M. L., Hantraye, P., Attar-Levy, D., Remy, P., Crouzel, C., Artiges, E., Feline, A., Syrota, A., and Martinot, J. L. (1997). Presynaptic dopaminergic function in the striatum of schizophrenic patients. *Schizophr. Res.* **23**, 167–174.
- Davis, K. L., Kahn, R. S., Ko, G., and Davidson, M. (1991). Dopamine in schizophrenia: A review and reconceptualization. *Am. J. Psychiatry* **148**, 1474–1486.
- Dean, B., Pavey, G., and Opeskin, K. (1997). [3H]raclopride binding to brain tissue from subjects with schizophrenia: Methodological aspects. *Neuropharmacology* **36**, 779–786.
- Dearry, A., Gingrich, J. A., Falardeau, P., Freneau, R. T., Jr., Bates, M. D., and Caron, M. G. (1990). Molecular cloning and expression of the gene for a human D1 dopamine receptor. *Nature* **347**, 72–76.
- Delay, J., Deniker, P., and Harl, J. (1952). Therapeutic use in psychiatry of phenothiazine of central elective action (4560 RP). *Ann. Med. Psychol. Paris* **110**, (2:1), 112–117.
- DeLong, M. R., Crutcher, M. D., and Georgopoulos, A. P. (1985). Primate globus pallidus and subthalamic nucleus: Functional organization. *J. Neurophysiol.* **53**, 530–543.
- Deutch, A. Y., Clark, W. A., and Roth, R. H. (1990). Prefrontal cortical dopamine depletion enhances the responsiveness of the mesolimbic dopamine neurons to stress. *Brain Res.* **521**, 311–315.
- Deutch, A. Y., Moghaddam, B., Innis, R. B., Krystal, J. H., Aghajanian, G. K., Bunney, B. S., and Charney, D. S. (1991). Mechanisms of action of atypical antipsychotic drugs. Implications for novel therapeutic strategies for schizophrenia. *Schizophr. Res.* **4**, 121–156.
- Diaz, J., Levesque, D., Lammers, C. H., Griffon, N., Martres, M. P., Schwartz, J. C., and Sokoloff, P. (1995). Phenotypical characterization of neurons expressing the dopamine D3 receptor in the rat brain. *Neuroscience* **65**, 731–745.
- Diaz, J., Pilon, C., Le Foll, B., Gros, C., Triller, A., Schwartz, J. C., and Sokoloff, P. (2000). Dopamine D3 receptors expressed by all mesencephalic dopamine neurons. *J. Neurosci.* **20**, 8677–8684.
- Dolan, R. J., Fletcher, P., Frith, C. D., Friston, K. J., Frackowiak, R. S., and Grasby, P. M. (1995). Dopaminergic modulation of impaired cognitive activation in the anterior cingulate cortex in schizophrenia. *Nature* **378**, 180–182.
- Dumartin, B., Jaber, M., Gonon, F., Caron, M. G., Giros, B., and Bloch, B. (2000). Dopamine tone regulates D1 receptor trafficking and delivery in striatal neurons in dopamine transporter-deficient mice. *Proc. Natl. Acad. Sci. USA* **97**, 1879–1884.
- Dunah, A. W., and Standaert, D. G. (2001). Dopamine D1 receptor-dependent trafficking of striatal NMDA glutamate receptors to the postsynaptic membrane. *J. Neurosci.* **21**, 5546–5558.

- Elkashef, A. M., Doudet, D., Bryant, T., Cohen, R. M., Li, S. H., and Wyatt, R. J. (2000). 6-(18) F-DOPA PET study in patients with schizophrenia. Positron emission tomography. *Psychiatry Res.* **100**, 1–11.
- Ellinwood, E. H., Jr. (1967). Amphetamine psychosis: I. Description of the individuals and process. *J. Nerv. Ment. Dis.* **144**, 273–283.
- Ellinwood, E. H., Jr., Sudilovsky, A., and Nelson, L. M. (1973). Evolving behavior in the clinical and experimental amphetamine model psychosis. *Am. J. Psychiatry* **130**, 1088–1093.
- Everitt, B. J., Morris, K. A., O'Brien, A., and Robbins, T. W. (1991). The basolateral amygdala-ventral striatal system and conditioned place preference: Further evidence of limbic-striatal interactions underlying reward-related processes. *Neuroscience* **42**, 1–18.
- Farde, L., Wiesel, F., Stone-Elander, S., Halldin, C., Nordsröm, A. L., Hall, H., and Sedvall, G. (1990). D2 dopamine receptors in neuroleptic-naïve schizophrenic patients. A positron emission tomography study with [¹¹C]raclopride. *Arch. Gen. Psychiatry* **47**, 213–219.
- Farde, L., Nordsröm, A. L., Wiesel, F. A., Pauli, S., Halldin, C., and Sedvall, G. (1992). Positron emission tomography analysis of central D1 and D2 dopamine receptor occupancy in patients treated with classical neuroleptics and clozapine. *Arch. Gen. Psychiatry* **49**, 538–544.
- Fauchey, V., Jaber, M., Caron, M. G., Bloch, B., and Le Moine, C. (2000). Differential regulation of the dopamine D1, D2 and D3 receptor gene expression and changes in the phenotype of the striatal neurons in mice lacking the dopamine transporter. *Eur. J. Neurosci.* **12**, 19–26.
- Ferry, A. T., Ongur, D., An, X., and Price, J. L. (2000). Prefrontal cortical projections to the striatum in macaque monkeys: Evidence for an organization related to prefrontal networks. *J. Comp. Neurol.* **425**, 447–470.
- Flores-Hernandez, J., Cepeda, C., Hernandez-Echeagaray, E., Calvert, C. R., Jokel, E. S., Fienberg, A. A., Greengard, P., and Levine, M. S. (2002). Dopamine enhancement of NMDA currents in dissociated medium-sized striatal neurons: Role of D1 receptors and DARPP-32. *J. Neurophysiol.* **88**, 3010–3020.
- Frankle, W. G., Gil, R., Hackett, E., Mawlawi, O., Zea-Ponce, Y., Zhu, Z., Kochan, L. D., Cangiano, C., Slifstein, M., Gorman, J. M., Laruelle, M., and Abi-Dargham, A. (2004). Occupancy of dopamine D2 receptors by the atypical antipsychotic drugs risperidone and olanzapine: Theoretical implications. *Psychopharmacology (Berl.)* **175**, 473–480.
- Gerfen, C. R. (1992). The neostriatal mosaic: Multiple levels of compartmental organization in the basal ganglia. *Annu. Rev. Neurosci.* **15**, 285–320.
- Gessa, G. L., Devoto, P., Diana, M., Flore, G., Melis, M., and Pistis, M. (2000). Dissociation of haloperidol, clozapine, and olanzapine effects on electrical activity of mesocortical dopamine neurons and dopamine release in the prefrontal cortex. *Neuropsychopharmacology* **22**, 642–649.
- Gingrich, J. A., and Caron, M. G. (1993). Recent advances in the molecular biology of dopamine receptors. *Annu. Rev. Neurosci.* **16**, 299–321.
- Glenthøj, B. Y., and Hemmingsen, R. (1997). Dopaminergic sensitization: Implications for the pathogenesis of schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **21**, 23–46.
- Goldberg, T. E., and Weinberger, D. R. (2004). Genes and the parsing of cognitive processes. *Trends Cogn. Sci.* **8**, 325–335.
- Goldman-Rakic, P. (1994). Working memory dysfunction in schizophrenia. *J. Neuropsychiatry Clin. Neurosci.* **6**, 348.
- Goldman-Rakic, P. S., Muly, E. C., III, E. C., and Williams, G. V. (2000). D(1) receptors in prefrontal cells and circuits. *Brain Res. Brain Res. Rev.* **31**, 295–301.
- Grace, A. A. (1991). Phasic versus tonic dopamine release and the modulation of dopamine system responsivity: A hypothesis for the etiology of schizophrenia. *Neuroscience* **41**, 1–24.
- Grace, A. A. (2000). Gating of information flow within the limbic system and the pathophysiology of schizophrenia. *Brain Res. Brain Res. Rev.* **31**, 330–341.

- Griffith, J. J., Oates, J., and Cavanaugh, J. (1968). Paranoid episodes induced by drugs. *J. Am. Med. Assoc.* **205**, 39.
- Grunder, G., Vernaleken, I., Muller, M. J., Davids, E., Heydari, N., Buchholz, H. G., Bartenstein, P., Munk, O. L., Stoeter, P., Wong, D. F., Gjedde, A., and Cumming, P. (2003). Subchronic haloperidol downregulates dopamine synthesis capacity in the brain of schizophrenic patients *in vivo*. *Neuropsychopharmacology* **28**, 787–794.
- Guillin, O., Diaz, J., Carroll, P., Griffon, N., Schwartz, J. C., and Sokoloff, P. (2001). BDNF controls dopamine D3 receptor expression and triggers behavioural sensitization. *Nature* **411**, 86–89.
- Guo, N., Hwang, D., Abdellhadi, S., Abi-Dargham, A., Zarahn, E., and Laruelle, M. (2001). The effect of chronic DA depletion on D1 ligand binding in rodent brain. *Soc. Neurosci. Abst.* **27**.
- Gurevich, E. V., Bordelon, Y., Shapiro, R. M., Arnold, S. E., Gur, R. E., and Joyce, J. N. (1997). Mesolimbic dopamine D3 receptors and use of antipsychotics in patients with schizophrenia. A postmortem study. *Arch. Gen. Psychiatry* **54**, 225–232.
- Haber, S. N., and Fudge, J. L. (1997). The primate substantia nigra and VTA: Integrative circuitry and function. *Crit. Rev. Neurobiol.* **11**, 323–342.
- Haber, S. N., Fudge, J. L., and McFarland, N. R. (2000). Striatonigrostriatal pathways in primates form an ascending spiral from the shell to the dorsolateral striatum. *J. Neurosci.* **20**, 2369–2382.
- Hashimoto, T., Bergen, S. E., Nguyen, Q. L., Xu, B., Monteggia, L. M., Pierri, J. N., Sun, Z., Sampson, A. R., and Lewis, D. A. (2005). Relationship of brain-derived neurotrophic factor and its receptor TrkB to altered inhibitory prefrontal circuitry in schizophrenia. *J. Neurosci.* **25**, 372–383.
- Hernandez-Lopez, S.,argas, J., Surmeier, D. J., Reyes, A., and Galarraga, E. (1997). D1 receptor activation enhances evoked discharge in neostriatal medium spiny neurons by modulating an L-type Ca²⁺ conductance. *J. Neurosci.* **17**, 3334–3342.
- Hersch, S. M., Ciliax, B. J., Gutekunst, C. A., Rees, H. D., Heilman, C. J., Yung, K. K., Bolam, J. P., Ince, E., Yi, H., and Levey, A. I. (1995). Electron microscopic analysis of D1 and D2 dopamine receptor proteins in the dorsal striatum and their synaptic relationships with motor corticostriatal afferents. *J. Neurosci.* **15**, 5222–5237.
- Hess, E. J., Bracha, H. S., Kleinman, J. E., and Creese, I. (1987). Dopamine receptor subtype imbalance in schizophrenia. *Life Sci.* **40**, 1487–1497.
- Hietala, J., Syvälahti, E., Vuorio, K., Nagren, K., Lehtikoinen, P., Ruotsalainen, U., Rääkköläinen, V., Lehtinen, V., and Wegelius, U. (1994a). Striatal D2 receptor characteristics in neuroleptic-naïve schizophrenic patients studied with positron emission tomography. *Arch. Gen. Psychiatry* **51**, 116–123.
- Hietala, J., West, C., Syvälahti, E., Nagren, K., Lehtikoinen, P., Sonninen, P., and Ruotsalainen, U. (1994b). Striatal D2 dopamine receptor binding characteristics *in vivo* in patients with alcohol dependence. *Psychopharmacology (Berl.)* **116**, 285–290.
- Hietala, J., Syvälahti, E., Vuorio, K., Rakkolainen, V., Bergman, J., Haaparanta, M., Solin, O., Kuoppamäki, M., Kirvelä, M., Ruotsalainen, U., and Salokangas, R. K. R. (1995). Presynaptic dopamine function in striatum of neuroleptic-naïve schizophrenic patients. *Lancet* **346**, 1130–1131.
- Hietala, J., Syvälahti, E., Vilkkumäki, H., Vuorio, K., Rakkolainen, V., Bergman, J., Haaparanta, M., Solin, O., Kuoppamäki, M., Eronen, E., Ruotsalainen, U., and Salokangas, R. K. (1999). Depressive symptoms and presynaptic dopamine function in neuroleptic-naïve schizophrenia. *Schizophr. Res.* **35**, 41–50.
- Hirai, M., Kitamura, N., Hashimoto, T., Nakai, T., Mita, T., Shirakawa, O., Yamadori, T., Amano, T., Noguchi-Kuno, S. A., and Tanaka, C. (1988). [³H]GBR-12935 binding sites in human

- striatal membranes: Binding characteristics and changes in parkinsonians and schizophrenics. *Jpn. J. Pharmacol.* **47**, 237–243.
- Hirvonen, J., van Erp, T. G., Huttunen, J., Aalto, S., Nagren, K., Huttunen, M., Lonnqvist, J., Kaprio, J., Hietala, J., and Cannon, T. D. (2005). Increased caudate dopamine D2 receptor availability as a genetic marker for schizophrenia. *Arch. Gen. Psychiatry* **62**, 371–378.
- Hoover, J. E., and Strick, P. L. (1993). Multiple output channels in the basal ganglia. *Science* **259**, 819–821.
- Hsiao, M. C., Lin, K. J., Liu, C. Y., Tzen, K. Y., and Yen, T. C. (2003). Dopamine transporter change in drug-naïve schizophrenia: An imaging study with 99mTc-TRODAT-1. *Schizophr. Res.* **65**, 39–46.
- Ichikawa, J., Ishii, H., Bonaccorso, S., Fowler, W. L., O'Laughlin, I. A., and Meltzer, H. Y. (2001). 5-HT(2A) and D(2) receptor blockade increases cortical DA release via 5-HT(1A) receptor activation: A possible mechanism of atypical antipsychotic-induced cortical dopamine release. *J. Neurochem.* **76**, 1521–1531.
- Joel, D., and Weiner, I. (2000). The connections of the dopaminergic system with the striatum in rats and primates: An analysis with respect to the functional and compartmental organization of the striatum. *Neuroscience* **96**, 451–474.
- Joyce, J. N., and Meador-Woodruff, J. H. (1997). Linking the family of D-2 receptors to neuronal circuits in human brain: Insights into schizophrenia. *Neuropsychopharmacology* **16**, 375–384.
- Joyce, J. N., Lexow, N., Bird, E., and Winokur, A. (1988). Organization of dopamine D1 and D2 receptors in human striatum: Receptor autoradiographic studies in Huntington's disease and schizophrenia. *Synapse* **2**, 546–557.
- Kahn, R. S., Harvey, P. D., Davidson, M., Keefe, R. S., Apter, S., Neale, J. M., Mohs, R. C., and Davis, K. L. (1994). Neuropsychological correlates of central monoamine function in chronic schizophrenia: Relationship between CSF metabolites and cognitive function. *Schizophr. Res.* **11**, 217–224.
- Kalivas, P. W., Sorg, B. A., and Hooks, M. S. (1993). The pharmacology and neural circuitry of sensitization to psychostimulants. *Behav. Pharmacol.* **4**, 315–334.
- Kapur, S. (2003). Psychosis as a state of aberrant salience: A framework linking biology, phenomenology, and pharmacology in schizophrenia. *Am. J. Psychiatry* **160**, 13–23.
- Kapur, S., Zipursky, R., Jones, C., Remington, G., and Houle, S. (2000). Relationship between dopamine D(2) occupancy, clinical response, and side effects: A double-blind PET study of first-episode schizophrenia. *Am. J. Psychiatry* **157**, 514–520.
- Karlsson, P., Farde, L., Halldin, C., and Sedvall, G. (2002). PET study of D(1) dopamine receptor binding in neuroleptic-naïve patients with schizophrenia. *Am. J. Psychiatry* **159**, 761–767.
- Karreman, M., and Moghaddam, B. (1996). The prefrontal cortex regulates the basal release of dopamine in the limbic striatum: An effect mediated by ventral tegmental area. *J. Neurochem.* **66**, 589–598.
- Kebabian, J. W., and Calne, D. B. (1979). Multiple receptors for dopamine. *Nature* **277**, 93–96.
- Keefe, R. S., Silva, S. G., Perkins, D. O., and Lieberman, J. A. (1999). The effects of atypical antipsychotic drugs on neurocognitive impairment in schizophrenia: A review and meta-analysis. *Schizophr. Bull.* **25**, 201–222.
- Knable, M. B., and Weinberger, D. R. (1997). Dopamine, the prefrontal cortex and schizophrenia. *J. Psychopharmacol.* **11**, 123–131.
- Knable, M. B., Hyde, T. M., Herman, M. M., Carter, J. M., Bigelow, L., and Kleinman, J. E. (1994). Quantitative autoradiography of dopamine-D1 receptors, D2 receptors, and dopamine uptake sites in postmortem striatal specimens from schizophrenic patients. *Biol. Psychiatry* **36**, 827–835.

- Knable, M. B., Hyde, T. M., Murray, A. M., Herman, M. M., and Kleinman, J. E. (1996). A postmortem study of frontal cortical dopamine D1 receptors in schizophrenics, psychiatric controls, and normal controls. *Biol. Psychiatry* **40**, 1191–1199.
- Knable, M. B., Egan, M. F., Heinz, A., Gorey, J., Lee, K. S., Coppola, R., and Weinberger, D. R. (1997). Altered dopaminergic function and negative symptoms in drug-free patients with schizophrenia. [123I]-iodobenzamide SPECT study. *Br. J. Psychiatry* **171**, 574–577.
- Kolachana, B. S., Saunders, R., and Weinberger, D. (1995). Augmentation of prefrontal cortical monoaminergic activity inhibits dopamine release in the caudate nucleus: An *in vivo* neurochemical assessment in the rhesus monkey. *Neuroscience* **69**, 859–868.
- Konradi, C. (1998). The molecular basis of dopamine and glutamate interactions in the striatum. *Adv. Pharmacol.* **42**, 729–733.
- Konradi, C., and Heckers, S. (2003). Molecular aspects of glutamate dysregulation: Implications for schizophrenia and its treatment. *Pharmacol. Ther.* **97**, 153–179.
- Kotter, R. (1994). Postsynaptic integration of glutamatergic and dopaminergic signals in the striatum. *Prog. Neurobiol.* **44**, 163–196.
- Kramer, M. S., Last, B., Getson, A., and Reines, S. A. (1997). The effects of a selective D4 dopamine receptor antagonist (L-745,870) in acutely psychotic inpatients with schizophrenia. D4 Dopamine Antagonist Group. *Arch. Gen. Psychiatry* **54**, 567–572.
- Kunishio, K., and Haber, S. N. (1994). Primate cingulostriatal projection: Limbic striatal versus sensorimotor striatal input. *J. Comp. Neurol.* **350**, 337–356.
- Laakso, A., Vilkmann, H., Alakare, B., Haaparanta, M., Bergman, J., Solin, O., Peurasaari, J., Rakkolainen, V., Syvalahti, E., and Hietala, J. (2000). Striatal dopamine transporter binding in neuroleptic-naïve patients with schizophrenia studied with positron emission tomography. *Am. J. Psychiatry* **157**, 269–271.
- Lahti, R. A., Roberts, R. C., Conley, R. R., Cochrane, E. V., Mutin, A., and Tamminga, C. A. (1996). D2-type dopamine receptors in postmortem human brain sections from normal and schizophrenic subjects. *Neuroreport* **7**, 1945–1948.
- Lahti, R. A., Roberts, R. C., Cochrane, E. V., Primus, R. J., Gallager, D. W., Conley, R. R., and Tamminga, C. A. (1998). Direct determination of dopamine D4 receptors in normal and schizophrenic postmortem brain tissue: A [3H]NGD-94-1 study. *Mol. Psychiatry* **3**, 528–533.
- Laruelle, M. (2000a). Imaging synaptic neurotransmission with *in vivo* binding competition techniques: A critical review. *J. Cereb. Blood Flow Metab.* **20**, 423–451.
- Laruelle, M. (2000b). The role of endogenous sensitization in the pathophysiology of schizophrenia: Implications from recent brain imaging studies. *Brain Res. Rev.* **31**, 371–384.
- Laruelle, M., Casanova, M., Weinberger, D., and Kleinman, J. (1990). Postmortem study of the dopaminergic D1 receptors in the dorsolateral prefrontal cortex of schizophrenics and controls. *Schizophr. Res* **3**, 30–31.
- Laruelle, M., Abi-Dargham, A., van Dyck, C. H., Gil, R., D'Souza, C. D., Erdos, J., McCance, E., Rosenblatt, W., Fingado, C., Zoghbi, S. S., Baldwin, R. M., Seibyl, R. M., *et al.* (1996). Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug-free schizophrenic subjects. *Proc. Natl. Acad. Sci. USA* **93**, 9235–9240.
- Laruelle, M., D'Souza, C. D., Baldwin, R. M., Abi-Dargham, A., Kanes, S. J., Fingado, C. L., Seibyl, J. P., Zoghbi, S. S., Bowers, M. B., Jatlow, P., Charney, D. S., and Innis, R. B. (1997a). Imaging D-2 receptor occupancy by endogenous dopamine in humans. *Neuropsychopharmacology* **17**, 162–174.
- Laruelle, M., Iyer, R. N., al-Tikriti, M. S., Zea-Ponce, Y., Malison, R., Zoghbi, S. S., Baldwin, R. M., Kung, H. F., Charney, D. S., Hoffer, P. B., Innis, R. B., and Bradberry, C. W. (1997b). Microdialysis

- and SPECT measurements of amphetamine-induced dopamine release in nonhuman primates. *Synapse* **25**, 1–14.
- Laruelle, M., Abi-Dargham, A., Gil, R., Kegeles, L., and Innis, R. (1999). Increased dopamine transmission in schizophrenia: Relationship to illness phases. *Biol. Psychiatry* **46**, 56–72.
- Laruelle, M., Abi-Dargham, A., van Dyck, C., Gil, R., D'Souza, D. C., Krystal, J., Seibyl, J., Baldwin, R., and Innis, R. (2000a). Dopamine and serotonin transporters in patients with schizophrenia: An imaging study with [123 I]beta-CIT. *Biol. Psychiatry* **47**, 371–379.
- Laruelle, M., Abi-Dargham, A., van Dyck, C., Gil, R., D'Souza, D. C., Krystal, J., Seibyl, J., Baldwin, R., and Innis, R. (2000b). Dopamine and serotonin transporters in patients with schizophrenia: An imaging study with [(123)I]beta-CIT. *Biol. Psychiatry* **47**, 371–379.
- Le Foll, B., Frances, H., Diaz, J., Schwartz, J. C., and Sokoloff, P. (2002). Role of the dopamine D3 receptor in reactivity to cocaine-associated cues in mice. *Eur. J. Neurosci.* **15**, 2016–2026.
- Le Moine, C., Tison, F., and Bloch, B. (1990). D2 dopamine receptor gene expression by cholinergic neurons in the rat striatum. *Neurosci. Lett.* **117**, 248–252.
- Le Moine, C., Normand, E., and Bloch, B. (1991). Phenotypical characterization of the rat striatal neurons expressing the D1 dopamine receptor gene. *Proc. Natl. Acad. Sci. USA* **88**, 4205–4209.
- Lee, T., Seeman, P., Tourtellotte, W. W., Farley, I. J., and Hornykeiwicz, O. (1978). Binding of 3H-neuroleptics and 3H-apomorphine in schizophrenic brains. *Nature* **274**, 897–900.
- Leveque, J. C., Macias, W., Rajadhyaksha, A., Carlson, R. R., Barczak, A., Kang, S., Li, X. M., Coyle, J. T., Huganir, R. L., Heckers, S., and Konradi, C. (2000). Intracellular modulation of NMDA receptor function by antipsychotic drugs. *J. Neurosci.* **20**, 4011–4020.
- Levine, M. S., Li, Z., Cepeda, C., Cromwell, H. C., and Altemus, K. L. (1996). Neuromodulatory actions of dopamine on synaptically-evoked neostriatal responses in slices. *Synapse* **24**, 65–78.
- Lieberman, J. A., Kane, J. M., and Alvir, J. (1987a). Provocative tests with psychostimulant drugs in schizophrenia. *Psychopharmacology* **91**, 415–433.
- Lieberman, J. A., Kane, J. M., Sarantakos, S., Gadaleta, D., Woerner, M., Alvir, J., and Ramos-Lorenzi, J. (1987b). Prediction of relapse in schizophrenia. *Arch. Gen. Psychiatry* **44**, 597–603.
- Lieberman, J. A., Kinon, B. L., and Loebel, A. D. (1990). Dopaminergic mechanisms in idiopathic and drug-induced psychoses. *Schizophr. Bull.* **16**, 97–110.
- Lieberman, J. A., Sheitman, B. B., and Kinon, B. J. (1997). Neurochemical sensitization in the pathophysiology of schizophrenia: Deficits and dysfunction in neuronal regulation and plasticity. *Neuropsychopharmacology* **17**, 205–229.
- Lindstrom, L. H., Gefvert, O., Hagberg, G., Lundberg, T., Bergstrom, M., Hartvig, P., and Langstrom, B. (1999). Increased dopamine synthesis rate in medial prefrontal cortex and striatum in schizophrenia indicated by L-(beta-11C) DOPA and PET. *Biol. Psychiatry* **46**, 681–688.
- Lindvall, O., and Björklund, A. (1983). Dopamine- and norepinephrine-containing neuron systems: Their anatomy in the rat brain. In “Chemical Neuroanatomy” (P. Emson, Ed.), pp. 229–255. Raven Press, New York.
- Mackay, A. V., Iversen, L. L., Rossor, M., Spokes, E., Bird, E., Arregui, A., Creese, I., and Synder, S. H. (1982). Increased brain dopamine and dopamine receptors in schizophrenia. *Arch. Gen. Psychiatry* **39**, 991–997.
- Marti, M., Mela, F., Bianchi, C., Beani, L., and Morari, M. (2002). Striatal dopamine-NMDA receptor interactions in the modulation of glutamate release in the substantia nigra pars reticulata *in vivo*: Opposite role for D1 and D2 receptors. *J. Neurochem.* **83**, 635–644.
- Martinot, J.-L., Peron-Magnan, P., Huret, J.-D., Mazoyer, B., Baron, J.-C., Boulenger, J.-P. C., Lh, B. M., Caillard, V. H. L., and Syrota, A. (1990). Striatal D2 dopaminergic receptors assessed

- with positron emission tomography and 76-Br-bromospiperone in untreated patients. *Am. J. Psychiatry* **147**, 346–350.
- Martinot, J. L., Paillère-Martinot, M. L., Loch'h, C., Hardy, P., Poirier, M. F., Mazoyer, B., Beaufile, B., Mazière, B., Alliaire, J. F., and Syrota, A. (1991). The estimated density of D2 striatal receptors in schizophrenia. A study with positron emission tomography and 76Br-bromolisuride. *Br. J. Psychiatry* **158**, 346–350.
- Martinot, J. L., Paillère-Martinot, M. L., Loch'h, C., Lecrubier, Y., Dao-Castellana, M. H., Aubin, F., Alliaire, J. F., Mazoyer, B., Mazière, B., and Syrota, A. (1994). Central D2 receptors and negative symptoms of schizophrenia. *Br. J. Pharmacol.* **164**, 27–34.
- Marzella, P. L., Hill, C., Keks, N., Singh, B., and Copolov, D. (1997). The binding of both [3H] nemonapride and [3H]raclopride is increased in schizophrenia. *Biol. Psychiatry* **42**, 648–654.
- McGowan, S. W., Lawrence, A., Sale, T., Qusted, D., and Grasby, P. M. (2004). Presynaptic dopaminergic dysfunction in medicated schizophrenic patients. *Arch. Gen. Psychiatry* **61**, 134–142.
- Meador-Woodruff, J. H., Damask, S. P., Wang, J., Haroutunian, V., Davis, K. L., and Watson, S. J. (1996). Dopamine receptor mRNA expression in human striatum and neocortex. *Neuropsychopharmacology* **15**, 17–29.
- Melis, M., Diana, M., and Gessa, G. L. (1999). Clozapine potently stimulates mesocortical dopamine neurons. *Eur. J. Pharmacol.* **366**, R11–R13.
- Meyer-Lindenberg, A., Miletich, R. S., Kohn, P. D., Esposito, G., Carson, R. E., Quarantelli, M., Weinberger, D. R., and Berman, K. F. (2002). Reduced prefrontal activity predicts exaggerated striatal dopaminergic function in schizophrenia. *Nat. Neurosci.* **5**, 267–271.
- Missale, C., Nash, S. R., Robinson, S. W., Jaber, M., and Caron, M. G. (1998). Dopamine receptors: From structure to function. *Physiol. Rev.* **78**, 189.
- Mita, T., Hanada, S., Nishino, N., Kuno, T., Nakai, H., Yamadori, T., Mizoi, Y., and Tanaka, C. (1986). Decreased serotonin S2 and increased dopamine D2 receptors in chronic schizophrenics. *Biol. Psychiatry* **21**, 1407–1414.
- Mogenson, G. J., Jones, D. L., and Yim, C. Y. (1980). From motivation to action: Functional interface between the limbic system and the motor system. *Prog. Neurobiol.* **14**, 69–97.
- Monsma, F. J., Jr., Mahan, L. C., McVittie, L. D., Gerfen, C. R., and Sibley, D. R. (1990). Molecular cloning and expression of a D1 dopamine receptor linked to adenyl cyclase activation. *Proc. Natl. Acad. Sci. USA* **87**, 6723–6727.
- Morari, M., O'Connor, W. T., Ungerstedt, U., and Fuxe, K. (1994). Dopamine D1 and D2 receptor antagonism differentially modulates stimulation of striatal neurotransmitter levels by N-methyl-D-aspartic acid. *Eur. J. Pharmacol.* **256**, 23–30.
- Mrzljak, L., Bergson, C., Pappy, M., Huff, R., Levenson, R., and Goldman-Rakic, P. S. (1996). Localization of dopamine D4 receptors in GABAergic neurons of the primate brain. *Nature* **381**, 245–248.
- Murray, A. M., Hyde, T. M., Knable, M. B., Herman, M. M., Bigelow, L. B., Carter, J. M., Weinberger, D. R., and Kleinman, J. E. (1995). Distribution of putative D4 dopamine receptors in postmortem striatum from patients with schizophrenia. *J. Neurosci.* **15**, 2186–2191.
- Nguyen, T. V., Kosofsky, B. E., Birnbaum, R., Cohen, B. M., and Hyman, S. E. (1992). Differential expression of c-fos and zif268 in rat striatum after haloperidol, clozapine, and amphetamine. *Proc. Natl. Acad. Sci. USA* **89**, 4270–4274.
- Nicola, S. M., Surmeier, J., and Malenka, R. C. (2000). Dopaminergic modulation of neuronal excitability in the striatum and nucleus accumbens. *Annu. Rev. Neurosci.* **23**, 185–215.
- Nishi, A., Snyder, G. L., and Greengard, P. (1997). Bidirectional regulation of DARPP-32 phosphorylation by dopamine. *J. Neurosci.* **17**, 8147–8155.

- Nordstrom, A. L., Farde, L., Wiesel, F. A., Forslund, K., Pauli, S., Halldin, C., and Uppfeldt, G. (1993). Central D2-dopamine receptor occupancy in relation to antipsychotic drug effects: A double-blind PET study of schizophrenic patients. *Biol. Psychiatry* **33**, 227–235.
- Nordstrom, A. L., Farde, L., Eriksson, L., and Halldin, C. (1995). No elevated D2 dopamine receptors in neuroleptic-naïve schizophrenic patients revealed by positron emission tomography and [¹¹C]N-methylspiperone [see comments]. *Psychiatry Res.* **61**, 67–83.
- Okubo, Y., Suhara, T., Suzuki, K., Kobayashi, K., Inoue, O., Terasaki, O., Someya, Y., Sassa, T., Sudo, Y., Matsushima, E., Iyo, M., Tateno, Y., *et al.* (1997). Decreased prefrontal dopamine D1 receptors in schizophrenia revealed by PET. *Nature* **385**, 634–636.
- Onn, S. P., West, A. R., and Grace, A. A. (2000). Dopamine-mediated regulation of striatal neuronal and network interactions. *Trends Neurosci.* **23**, S48–S56.
- Owen, F., Cross, A. J., Crow, T. J., Longden, A., Poulter, M., and Riley, G. J. (1978). Increased dopamine-receptor sensitivity in schizophrenia. *Lancet* **2**, 223–226.
- Palermo-Neto, J. (1997). Dopaminergic systems. Dopamine receptors. *Psychiatr. Clin. North Am.* **20**, 705–721.
- Parent, A., and Hazrati, L. N. (1995). Functional anatomy of the basal ganglia. I. The cortico-basal ganglia-thalamo-cortical loop. *Brain Res. Brain Res. Rev.* **20**, 91–127.
- Parsey, R. V., Oquendo, M. A., Zea-Ponce, Y., Rodenhiser, J., Kegeles, L. S., Prata, M., Cooper, T. B., Van Heertum, R., Mann, J. J., and Laruelle, M. (2001). Dopamine D(2) receptor availability and amphetamine-induced dopamine release in unipolar depression. *Biol. Psychiatry* **50**, 313–322.
- Pearce, R. K., Seeman, P., Jellinger, K., and Tourtellotte, W. W. (1990). Dopamine uptake sites and dopamine receptors in Parkinson's disease and schizophrenia. *Eur. Neurol.* **30**(Suppl. 1), 9–14.
- Pehek, E. A., and Yamamoto, B. K. (1994). Differential effects of locally administered clozapine and haloperidol on dopamine efflux in the rat prefrontal cortex and caudate-putamen. *J. Neurochem.* **63**, 2118–2124.
- Pennartz, C. M., Groenewegen, H. J., and Lopes da Silva, F. H. (1994). The nucleus accumbens as a complex of functionally distinct neuronal ensembles: An integration of behavioural, electrophysiological and anatomical data. *Prog. Neurobiol.* **42**, 719–761.
- Peris, J., Dwoskin, L. P., and Zahniser, N. R. (1988). Biphasic modulation of evoked [³H]D-aspartate release by D-2 dopamine receptors in rat striatal slices. *Synapse* **2**, 450–456.
- Pilowsky, L. S., Costa, D. C., Ell, P. J., Verhoeff, N. P., Murray, R. M., and Kerwin, R. W. (1994). D2 dopamine receptor binding in the basal ganglia of antipsychotic-free schizophrenic patients. An 123I-IBZM single photon emission computerised tomography study. *Br. J. Psychiatry* **164**, 16–26.
- Pimoule, C., Schoemaker, H., Reynolds, G. P., and Langer, S. Z. (1985). [³H]SCH 23390 labeled D1 dopamine receptors are unchanged in schizophrenia and Parkinson's disease. *Eur. J. Pharmacol.* **114**, 235–237.
- Pycock, C. J., Kerwin, R. W., and Carter, C. J. (1980). Effect of lesion of cortical dopamine terminals on subcortical dopamine receptors in rats. *Nature* **286**, 74–77.
- Reith, J., Benkelfat, C., Sherwin, A., Yasuhara, Y., Kuwabara, H., Andermann, F., Bachneff, S., Cumming, P., Diksic, M., Dyve, S. E., Etienne, P., Evans, A. C., *et al.* (1994). Elevated dopa decarboxylase activity in living brain of patients with psychosis. *Proc. Natl. Acad. Sci. USA* **91**, 11651–11654.
- Reynolds, G. P. (1983). Increased concentrations and lateral asymmetry of amygdala dopamine in schizophrenia. *Nature* **305**, 527–529.
- Reynolds, G. P. (1989). Beyond the dopamine hypothesis. The neurochemical pathology of schizophrenia. *Br. J. Psychiatry* **155**, 305–316.

- Reynolds, G. P., and Czudek, C. (1988). Status of the dopaminergic system in post-mortem brain in schizophrenia. *Psychopharmacol. Bull.* **24**, 345–347.
- Reynolds, G. P., and Mason, S. L. (1994). Are striatal dopamine D4 receptors increased in schizophrenia? *J. Neurochem.* **63**, 1576–1577.
- Reynolds, G. P., Czudek, C., Bzowej, N., and Seeman, P. (1987). Dopamine receptor asymmetry in schizophrenia. *Lancet* **1**, 979.
- Ridray, S., Griffon, N., Mignon, V., Souil, E., Carboni, S., Diaz, J., Schwartz, J. C., and Sokoloff, P. (1998). Coexpression of dopamine D1 and D3 receptors in islands of Calleja and shell of nucleus accumbens of the rat: Opposite and synergistic functional interactions. *Eur. J. Neurosci.* **10**, 1676–1686.
- Robertson, G. S., Matsumura, H., and Fibiger, H. C. (1994). Induction patterns of Fos-like immunoreactivity in the forebrain as predictors of atypical antipsychotic activity. *J. Pharmacol. Exp. Ther.* **271**, 1058–1066.
- Robinson, T. E., and Becker, J. B. (1986). Enduring changes in brain and behavior produced by chronic amphetamine administration: A review and evaluation of animal models of amphetamine psychosis. *Brain Res. Rev.* **11**, 157–198.
- Rollema, H., Lu, Y., Schmidt, A. W., Sprouse, J. S., and Zorn, S. H. (2000). 5-HT(1A) receptor activation contributes to ziprasidone-induced dopamine release in the rat prefrontal cortex. *Biol. Psychiatry* **48**, 229–237.
- Ruiz, J., Gabilondo, A. M., Meana, J. J., and Garcia-Sevilla, J. A. (1992). Increased [3H] raclopride binding sites in postmortem brains from schizophrenic violent suicide victims. *Psychopharmacology (Berl.)* **109**, 410–414.
- Sato, M., Chen, C. C., Akiyama, K., and Otsuki, S. (1983). Acute exacerbation of paranoid psychotic state after long-term abstinence in patients with previous methamphetamine psychosis. *Biol. Psychiatry* **18**, 429–440.
- Schoots, O., Seeman, P., Guan, H. C., Paterson, A. D., and Van Tol, H. H. (1995). Long-term haloperidol elevates dopamine D4 receptors by 2-fold in rats. *Eur. J. Pharmacol.* **289**, 67–72.
- Scott, L., Kruse, M. S., Forssberg, H., Brismar, H., Greengard, P., and Aperia, A. (2002). Selective up-regulation of dopamine D1 receptors in dendritic spines by NMDA receptor activation. *Proc. Natl. Acad. Sci. USA* **99**, 1661–1664.
- Seeman, P. (1988). Brain dopamine receptors in schizophrenia: PET problems. *Arch. Gen. Psychiatry* **45**, 598–600.
- Seeman, P., and Lee, T. (1975). Antipsychotic drugs: Direct correlation between clinical potency and presynaptic action on dopamine neurons. *Science* **188**, 1217–1219.
- Seeman, P., Ulpian, C., Bergeron, C., Riederer, P., Jellinger, K., Gabriel, E., Reynolds, G. P., and Tourtellotte, W. W. (1984). Bimodal distribution of dopamine receptor densities in brains of schizophrenics. *Science* **225**, 728–731.
- Seeman, P., Bzowej, N. H., Guan, H. C., Bergeron, C., Reynolds, G. P., Bird, E. D., Riederer, P., Jellinger, K., and Tourtellotte, W. W. (1987). Human brain D1 and D2 dopamine receptors in schizophrenia, Alzheimer's, Parkinson's, and Huntington's diseases. *Neuropsychopharmacology* **1**, 5–15.
- Seeman, P., Guan, H. C., and Niznik, H. B. (1989). Endogenous dopamine lowers the dopamine D2 receptor density as measured by [3H]raclopride: Implications for positron emission tomography of the human brain. *Synapse* **3**, 96–97.
- Seeman, P., Guan, H. C., and Van Tol, H. H. (1993). Dopamine D4 receptors elevated in schizophrenia. *Nature* **365**, 441–445.

- Seamans, J. K., Gorelova, N., Durstewitz, D., and Yang, C. R. (2001). Bidirectional dopamine modulation of GABAergic inhibition in prefrontal cortical pyramidal neurons. *J. Neurosci.* **21**, 3628–3638.
- Smiley, J. F., Levey, A. I., Ciliax, B. J., and Goldman-Rakic, P. S. (1994). D1 dopamine receptor immunoreactivity in human and monkey cerebral cortex: Predominant and extrasynaptic localization in dendritic spines. *Proc. Natl. Acad. Sci. USA* **91**, 5720–5724.
- Smith, A. D., and Bolam, J. P. (1990). The neural network of the basal ganglia as revealed by the study of synaptic connections of identified neurones. *Trends Neurosci.* **13**, 259–265.
- Sokoloff, P., Giros, B., Martres, M. P., Bouthenet, M. L., and Schwartz, J. C. (1990). Molecular cloning and characterization of a novel dopamine receptor (D3) as a target for neuroleptics. *Nature* **347**, 146–151.
- Sokoloff, P., Diaz, J., Levesque, D., Pilon, C., Dimitriadou, V., Griffon, N., Lammers, C. H., Martres, M. P., and Schwartz, J. C. (1995). Novel dopamine receptor subtypes as targets for antipsychotic drugs. *Ann. N. Y. Acad. Sci.* **757**, 278–292.
- Spano, P. F., Govoni, S., and Trabucchi, M. (1978). Studies on the pharmacological properties of dopamine receptors in various areas of the central nervous system. *Adv. Biochem. Psychopharmacol.* **19**, 155–165.
- Starr, M. S. (1995). Glutamate/dopamine D1/D2 balance in the basal ganglia and its relevance to Parkinson's disease. *Synapse* **19**, 264–293.
- Sumiyoshi, T., Stockmeier, C. A., Overholser, J. C., Thompson, P. A., and Meltzer, H. Y. (1995). Dopamine D4 receptors and effects of guanine nucleotides on [3H]raclopride binding in postmortem caudate nucleus of subjects with schizophrenia or major depression. *Brain Res.* **681**, 109–116.
- Sunahara, R. K., Guan, H. C., O'Dowd, B. F., Seeman, P., Laurier, L. G., Ng, G., George, S. R., Torchia, J., Van Tol, H. H., and Niznik, H. B. (1991). Cloning of the gene for a human dopamine D5 receptor with higher affinity for dopamine than D1. *Nature* **350**, 614–619.
- Surmeier, D. J., Eberwine, J., Wilson, C. J., Cao, Y., Stefani, A., and Kitai, S. T. (1992). Dopamine receptor subtypes colocalize in rat striatonigral neurons. *Proc. Natl. Acad. Sci. USA* **89**, 10178–10182.
- Surmeier, D. J., Song, W. J., and Yan, Z. (1996). Coordinated expression of dopamine receptors in neostriatal medium spiny neurons. *J. Neurosci.* **16**, 6579–6591.
- Takahashi, M., Shirakawa, O., Toyooka, K., Kitamura, N., Hashimoto, T., Maeda, K., Koizumi, S., Wakabayashi, K., Takahashi, H., Someya, T., and Nawa, H. (2000). Abnormal expression of brain-derived neurotrophic factor and its receptor in the corticolimbic system of schizophrenic patients. *Mol. Psychiatry* **5**, 293–300.
- Talbot, P. S., and Laruelle, M. (2002). The role of *in vivo* molecular imaging with PET and SPECT in the elucidation of psychiatric drug action and new drug development. *Eur. Neuropsychopharmacol.* **12**, 503–511.
- Tarazi, F. I., Florijn, W. J., and Creese, I. (1997). Differential regulation of dopamine receptors after chronic typical and atypical antipsychotic drug treatment. *Neuroscience* **78**, 985–996.
- Taylor, S. F., Koeppe, R. A., Tandon, R., Zubieta, J. K., and Frey, K. A. (2000). *In vivo* measurement of the vesicular monoamine transporter in schizophrenia. *Neuropsychopharmacology* **23**(6), 667–675.
- Tiberi, M., Jarvie, K. R., Silvia, C., Falardeau, P., Gingrich, J. A., Godinot, N., Bertrand, L., Yang-Feng, T. L., Freneau, R. T., Jr., and Caron, M. G. (1991). Cloning, molecular characterization, and chromosomal assignment of a gene encoding a second D1 dopamine receptor subtype: Differential expression pattern in rat brain compared with the D1A receptor. *Proc. Natl. Acad. Sci. USA* **88**, 7491–7495.

- Torstenson, R., Hartvig, P., Langstrom, B., Bastami, S., Antoni, G., and Tedroff, J. (1998). Effect of apomorphine infusion on dopamine synthesis rate relates to dopaminergic tone. *Neuropharmacology* **37**, 989–995.
- Tune, L. E., Dean, F., Wong, D. F., Pearlson, G., Strauss, M., Young, T., Shaya, E. K., Dannals, R. F., Wilson, A. A., Ravert, H. T., Sapp, J., Cooper, T., *et al.* (1993). Dopamine D2 receptor density estimates in schizophrenia: A positron emission tomography study with ¹¹C-N-methylspiperone. *Psychiatry Res.* **49**, 219–237.
- Tzschentke, T. M. (2001). Pharmacology and behavioral pharmacology of the mesocortical dopamine system. *Prog. Neurobiol.* **63**, 241–320.
- Van Tol, H. H., Bunzow, J. R., Guan, H. C., Sunahara, R. K., Seeman, P., Niznik, H. B., and Civelli, O. (1991). Cloning of the gene for a human dopamine D4 receptor with high affinity for the antipsychotic clozapine. *Nature* **350**, 610–614.
- Villemagne, V. L., Wong, D. F., Yokoi, F., Stephane, M., Rice, K. C., Matecka, D., Clough, D. J., Dannals, R. F., and Rothman, R. B. (1999). GBR12909 attenuates amphetamine-induced striatal dopamine release as measured by [¹¹C]raclopride continuous infusion PET scans. *Synapse* **33**, 268–273.
- Weickert, C. S., Hyde, T. M., Lipska, B. K., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2003). Reduced brain-derived neurotrophic factor in prefrontal cortex of patients with schizophrenia. *Mol. Psychiatry* **8**, 592–610.
- Weinberger, D., and Laruelle, M. (2001). Neurochemical and neuropharmacological imaging in schizophrenia. In “Neuropsychopharmacology: The Fifth Generation of Progress” (K. L. Davis, D. S. Charney, J. T. Coyle, and Charles Nemeroff, Eds.), pp. 833–856. Lippincott, Williams, and Wilkins. New York.
- Weinberger, D. R. (1987). Implications of the normal brain development for the pathogenesis of schizophrenia. *Arch. Gen. Psychiatry* **44**, 660–669.
- Weinberger, D. R., Berman, K. F., and Chase, T. N. (1988). Mesocortical dopaminergic function and human cognition. *Ann. N. Y. Acad. Sci.* **537**, 330–338.
- West, A. R., and Grace, A. A. (2002). Opposite influences of endogenous dopamine D1 and D2 receptor activation on activity states and electrophysiological properties of striatal neurons: Studies combining *in vivo* intracellular recordings and reverse microdialysis. *J. Neurosci.* **22**, 294–304.
- Wilkinson, L. S. (1997). The nature of interactions involving prefrontal and striatal dopamine systems. *J. Psychopharmacol.* **11**, 143–150.
- Wilson, C. J., and Kawaguchi, Y. (1996). The origins of two-state spontaneous membrane potential fluctuations of neostriatal spiny neurons. *J. Neurosci.* **16**, 2397–2410.
- Wong, D. F., Wagner, H. N., Jr., Tune, L. E., Dannals, R. F., Pearlson, G. D., Links, J. M., Tamminga, C. A., Broussolle, E. P., Ravert, H. T., Wilson, A. A., Toung, J. K., Malat, J., *et al.* (1986). Positron emission tomography reveals elevated D2 dopamine receptors in drug-naïve schizophrenics. *Science* **234**, 1558–1563.
- Yang, C. R., Seamans, J. K., and Gorelova, N. (1999). Developing a neuronal model for the pathophysiology of schizophrenia based on the nature of electrophysiological actions of dopamine in the prefrontal cortex. *Neuropsychopharmacology* **21**, 161–194.
- Yang, Y. K., Yu, L., Yeh, T. L., Chiu, N. T., Chen, P. S., and Lee, I. H. (2004). Associated alterations of striatal dopamine D2/D3 receptor and transporter binding in drug-naïve patients with schizophrenia: A dual-isotope SPECT study. *Am. J. Psychiatry* **161**, 1496–1498.
- Young, D., and Scoville, W. B. (1938). Paranoid psychosis in narcolepsy and the possible dangers of benzedrine treatment. *Med. Clin. North Am.* **22**, 637.

- Youngren, K. D., Inglis, F. M., Pivrotto, P. J., Jedema, H. P., Bradberry, C. W., Goldman-Rakic, P. S., Roth, R. H., and Moghaddam, B. (1999). Clozapine preferentially increases dopamine release in the rhesus monkey prefrontal cortex compared with the caudate nucleus. *Neuropsychopharmacology* **20**, 403–412.
- Zhou, Q. Y., Grandy, D. K., Thambi, L., Kushner, J. A., Van Tol, H. H., Cone, R., Pribnow, D., Salon, J., Bunzow, J. R., and Civelli, O. (1990). Cloning and expression of human and rat D1 dopamine receptors. *Nature* **347**, 76–80.
- Zhu, M. Y., Juorio, A. V., Paterson, I. A., and Boulton, A. A. (1993). Regulation of striatal aromatic L-amino acid decarboxylase: Effects of blockade or activation of dopamine receptors. *Eur. J. Pharmacol.* **238**, 157–164.

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THE DOPAMINE SYSTEM AND THE PATHOPHYSIOLOGY OF SCHIZOPHRENIA: A BASIC SCIENCE PERSPECTIVE

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- I. Introduction
- II. Neuroanatomy of DA Systems
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The dopamine system has been a subject of intense investigation due to its role in a number of normal functions and its disruption in pathological conditions. Thus, the dopamine system has been shown to play a major role in cognitive, affective, and motor functions, and its disruption has been proposed to underlie the pathophysiology of several major psychiatric and neurological disorders, including schizophrenia, Parkinson's disease, drug abuse, and attention deficit/hyperactivity disorder. Although these studies have continued to define the basic functional principles of the dopamine system in the mammalian brain, we are still at the initial stages in unraveling the complex role of this transmitter system in regulating behavioral processes. Accumulating evidence suggests that dopamine modulates excitatory and inhibitory neurotransmission, and moreover affects synaptic plasticity induced within the circuits of its target brain regions. It is this role in synaptic plasticity that has associated the dopamine system with aspects of cognitive function involving learning and memory. In this chapter, we summarize recent findings relevant to the role of the dopamine system in psychiatric disorders at cellular, anatomical, and functional levels. In particular, we will focus on the regulation of dopamine neuron activity states and how this impacts dopamine release in cortical and subcortical systems, and the physiological and behavioral impact of dopamine receptor stimulation in the postsynaptic targets of these neurons. A brief summary of recent findings regarding the development and maturation of DA

system and how this relates to the pathophysiology of psychiatric disorders are given, and finally models of dopamine system disruption in schizophrenia and how therapeutic approaches impact on dopamine system dynamics is presented.

I. Introduction

Dopamine (DA) is the most basic of the catecholamine neurotransmitters in the central nervous system. Since the identification of DA as an independent neurotransmitter in the brain (Carlsson, 1959; Carlsson *et al.*, 1958), a large number of studies from molecular to behavioral levels has been done to understand the functional roles of DA. However, there is general agreement from research to suggest that the role of DA is not to mediate direct synaptic driving of neurotransmission in the brain, but instead to modulate excitatory and inhibitory neurotransmission (Kupfermann, 1979). Therefore, DA is now considered to be a neuromodulator.

The reason for this substantial level of interest is due to its involvement in a number of neurological and psychiatric disorders including Parkinson's disease (Hornykiewicz, 1971; Lloyd and Hornykiewicz, 1970) and schizophrenia (Faurbye, 1968; Fischer, 1970). The loss of DA neurons in the nigrostriatal system has been shown to underlie the symptoms of Parkinson's disease (Hornykiewicz, 1971; Lloyd and Hornykiewicz, 1970), whereas excessive DA release has been suggested in the pathophysiology of schizophrenia. The latter is based on studies showing that DA agonists such as amphetamine induce psychosis similar to that of schizophrenia patients (Connell, 1958; Snyder, 1972) as well as the fact that antipsychotic drugs used for the treatment of schizophrenia are DA receptor antagonists (Carlsson, 1974; Seeman, 1987). These studies suggest that proper DA signaling in the brain is critical for cognitive and affective functions as well as motor control, which are disrupted in these neurological and psychiatric disorders.

In this chapter, we summarize recent findings of the DA system at cellular, pharmacological, physiological, and functional levels, and how these findings relate DA system function to the pathophysiology of schizophrenia.

II. Neuroanatomy of DA Systems

DA neurons are located in the mesencephalon and project into the forebrain along three major pathways.

The nigrostriatal system is composed of DA neurons located in the substantia nigra pars compacta (SNc) that project into the dorsal striatum (Anden *et al.*, 1964;

Bedard *et al.*, 1969). This is the system that is believed to have a principal involvement in motor control, since degeneration of DA neurons in the SNc is the primary pathology of Parkinson's disease (Hornykiewicz, 1971; Lloyd and Hornykiewicz, 1970). Of particular importance in the functional significance of nigrostriatal DA projections is its involvement in motor learning and habit formation (Graybiel, 1998; Jog *et al.*, 1999). However, animal and human studies have also revealed that the nigrostriatal DA projection is also involved in non-motor cognitive functions (Carbon and Marie, 2003; Graybiel, 1997; Schultz, 2002).

The mesolimbic DA system is composed of DA neurons located in the ventral tegmental area (VTA) that project to the ventral striatum (VS) including the nucleus accumbens, olfactory tubercle, and limbic structures such as the basolateral amygdala and hippocampus (HPC) (Brinley-Reed and McDonald, 1999; Fallon and Moore, 1978; Fallon *et al.*, 1978; Scatton *et al.*, 1980; Voorn *et al.*, 1986). DA release in the amygdala and HPC are thought to be involved in emotional learning (Bissiere *et al.*, 2003; Jellestad *et al.*, 1986; Rosenkranz and Grace, 2002) and long-term memory (Li *et al.*, 2003; Lisman and Grace, 2005; Otmakhova and Lisman, 1996), respectively. The VS is the brain region where limbic and cortical inputs converge, and information encoded on these brain structures is integrated to organize goal-directed behavior (Groenewegen *et al.*, 1996; Mogenson *et al.*, 1980). As such, the mesolimbic DA projections to the VS modulate this information integration, and thereby influence motor behavior. In particular, these DA projections into the VS are considered to be crucial for motivation and reward seeking (Everitt and Robbins, 2005; Iversen, 1984).

Finally, the mesocortical DA system is composed of DA neurons also located in the VTA that project to the prefrontal cortex (PFC), anterior cingulate, and entorhinal cortex in rodents (Fallon *et al.*, 1978; Thierry *et al.*, 1973) as well as additional neocortical and even cerebellar areas in primates and humans (De Keyser *et al.*, 1989; Lewis *et al.*, 1987; Melchitzky and Lewis, 2000; Moore *et al.*, 2003). The PFC is considered to be the highest center of cognition (Funahashi, 2001; Fuster, 1997; Goldman-Rakic, 1995; Knight *et al.*, 1995; Robbins, 2000; Shimamura, 2000), and DA release in the PFC is essential for its function. As such, a number of cognitive functions such as short-term memory (Funahashi *et al.*, 1993; Goldman-Rakic, 1995), attention (Gorenstein *et al.*, 1989; Knight *et al.*, 1995; Muir *et al.*, 1996), future planning (Baker *et al.*, 1996; Ingvar, 1985; Owen *et al.*, 1990), and set shifting (Milner, 1963; Owen *et al.*, 1993) have been proposed to be mediated by the PFC, and interruption of the mesocortical DA innervation of the PFC is known to produce impairments in these functions (Floresco *et al.*, 2006; Ragozzino, 2002; Sawaguchi and Goldman-Rakic, 1994; Seamans *et al.*, 1998).

The VTA is a heterogeneous structure consisting of DA neurons and gamma-aminobutyric acid (GABA) neurons. Some of these GABA neurons are interneurons, whereas other GABA neurons are projection neurons that innervate both the

PFC and VS. Anatomical and physiological studies (Carr and Sesack, 2000; Floresco *et al.*, 2001, 2003; Lisman and Grace, 2005; Sesack and Carr, 2002) have shown that the mesolimbic and mesocortical DA pathways are organized into two independent closed loops.

In the mesocortical DA system, PFC pyramidal neurons (in layers V and VI) are reported to project selectively onto VTA DA neurons that project back to the PFC (Fig. 1; Carr and Sesack, 2000; Sesack and Carr, 2002). In addition, PFC afferents are also shown to target GABA interneurons that in turn regulate the activity of DA neurons that project to the VS as well as GABA neurons that project directly to the VS (Fig. 1; Carr and Sesack, 2000; Sesack and Carr, 2002). Consequently, this anatomically closed loop induces higher DA release in the PFC during periods of PFC activation, while at the same time it suppresses DA release in the VS.

In contrast, the mesolimbic DA system also forms another closed loop circuit with the HPC and VS (Fig. 2). Thus, the HPC send excitatory projections into the VS (Groenewegen *et al.*, 1987; Kelley and Domesick, 1982), which in turn regulates other basal ganglia nuclei including the ventral pallidum and pedunculo-pontine tegmentum (Floresco *et al.*, 2003). Through this loop, activity within the HPC efferents is positioned to control DA release in the VS (Floresco *et al.*, 2001).

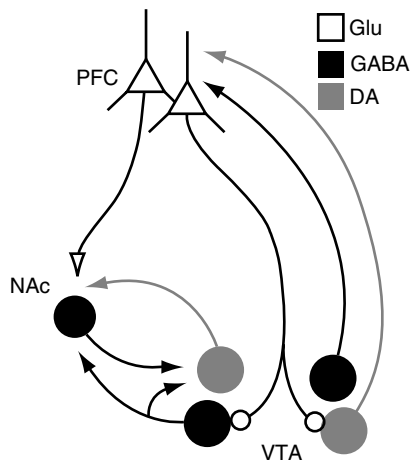


FIG. 1. Schematic diagram of the relationship between the PFC, VS, and the DA system. The reciprocal interaction between the PFC and VTA would allow PFC activation to selectively induce DA release in the PFC and suppress DA release in the VS simultaneously. Glu and NAc denote glutamate and nucleus accumbens, respectively. Reprinted from Sesack and Carr, copyright (2002), with permission from Elsevier.

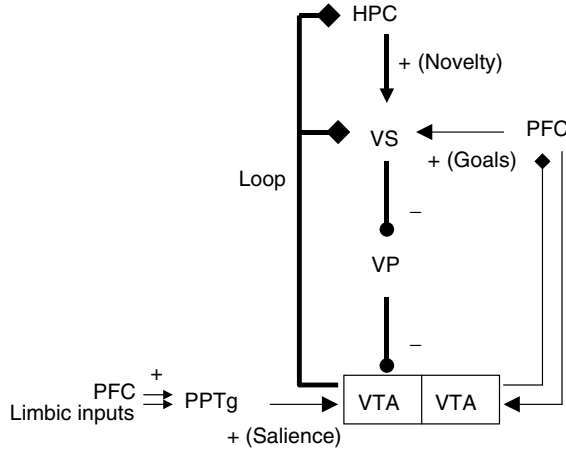


FIG. 2. Functional interactions between the HPC and the VTA that can mediate novelty detection and incorporation into memory. It is proposed that the incorporation of an object or an event into memory is dependent on two aspects: novelty and salience. In this model, HPC afferents regulate activity within the VS and its afferent basal ganglia nuclei including the ventral pallidum (VP). These structures, in turn, can modulate VTA DA neuron activity. It is proposed that novelty detection in the HPC will cause excitation of VTA DA neurons via this pathway. When the HPC novelty-dependent excitation of the VTA occurs in coincidence with salience-dependent activation of pedunculopontine tegmental (PPTg) input to the VTA neurons, the resultant enhanced VTA DA input to the HPC will trigger incorporation into memory. Triangles, circles, and diamonds indicate glutamate, GABA, and DA projections, respectively. Adapted from Lisman and Grace (2005).

III. DA Neuron Activity and Release

DA neurons are known to exhibit two types of spike firing patterns: tonic and burst spike firing (Grace and Bunney, 1984a,b). Tonic spike firing is the baseline spontaneous activity state that is driven by an endogenous pacemaker conductance (Grace and Bunney, 1984b) and does not depend on an excitatory driving force. Thus, tonic spike firing is still observed in an *in vitro* brain slice preparation in which the afferent inputs onto the DA neurons are transected. In contrast, transient, burst spike firing is known to be triggered by external stimuli, especially those associated with the presentation of unexpected reward or sensory signals that predict rewards (Schultz *et al.*, 1993). Furthermore, DA neurons exhibit transient suppression of tonic spike firing with aversive stimuli or omission of expected rewards (Tobler *et al.*, 2003; Ungless *et al.*, 2004). However, it is still controversial regarding whether only reward-associated stimuli can evoke burst spike firing in DA neurons (e.g., burst spike firing evoked by aversive stimuli; Guarraci and Kapp, 1999; Mantz *et al.*, 1989). Phasic burst spike firing is regulated

by excitatory glutamatergic inputs onto DA neurons arising from other brain structures including the PFC (Au-Young *et al.*, 1999; Gariano and Groves, 1988), subthalamic nucleus (Chergui *et al.*, 1994; Smith and Grace, 1992), and glutamatergic/cholinergic afferents from the pedunculo pontine tegmentum onto DA neurons (Floresco *et al.*, 2003; Futami *et al.*, 1995). Furthermore, the ability of DA neurons to fire in bursts is dependent on inputs from the lateral dorsal tegmental nucleus (Lodge and Grace, 2006a); interruption of this source of afferents prevents glutamate-driven burst firing, causing the DA neurons to discharge in a manner similar to that observed *in vitro* (Grace and Onn, 1989).

DA is released from DA terminals located in the targeted brain areas as a function of spike firing patterns of DA neurons (Fig. 3). Tonic spike firing of DA neurons induces tonic, low concentration of DA release (i.e., nanomolar; Floresco *et al.*, 2003; Grace, 1991). *In vivo*, the level of tonic DA release is regulated by powerful GABAergic inhibitory inputs arising from the ventral pallidum that innervate DA neurons in the VTA. Thus, when the ventral pallidum is inactivated, the number of spontaneously firing DA neurons increases and tonic DA release in the VS is increased (Floresco *et al.*, 2003). It is hypothesized that such tonic DA release can escape from the reuptake system in the synaptic cleft of the release sites, and therefore determines the basal concentration of extracellular DA level within these regions (Fig. 3; Grace, 1991). Tonic DA release is too low in concentration to effectively stimulate the postsynaptic targets of these neurons; however, it appears to be sufficient in concentration to stimulate presynaptic DA receptors, including those on corticostriatal projections (O'Donnell and Grace, 1994; West and Grace, 2002) and the autoreceptors on the DA terminals themselves, which provide a local regulation of DA synthesis and release (Nowycky and Roth, 1978; Starke *et al.*, 1978; Wolf and Roth, 1990). In contrast, burst spike firing of DA neurons induces a substantially higher amplitude (i.e., micromolar to millimolar; Floresco *et al.*, 2003; Grace, 1991) of phasic DA release within the synaptic cleft of the DA terminals, where it can stimulate postsynaptic DA receptors. This high-amplitude phasically released DA is then subject to immediate reuptake into DA terminals via the dopamine transporter (DAT) in the striatum, where it is then recycled or metabolized by monoamine oxidase (Fig. 3; Grace, 1991; Kuhar *et al.*, 1990; Molinoff and Axelrod, 1971). This contrasts with the PFC, in which DAT is low, and DA is believed to be removed primarily by actions of metabolic enzyme catechol-O-methyltransferase (COMT) (Fig. 3; Karoum *et al.*, 1994; Molinoff and Axelrod, 1971; Sesack *et al.*, 1998; White, 1996) or by reuptake into noradrenergic terminals (Moron *et al.*, 2002). Indeed, DAT knockout mice show a greater DA overflow in the striatum and hyperlocomotion, but the amount of DA reuptake is not changed in the PFC of these mice (Gogos *et al.*, 1998). Recent studies combining human imaging studies with genetic analysis have revealed that single nucleotide polymorphism (SNP) of genes encoding COMT affects PFC function. For example, human subjects with the Val/Val allele at position 158 in COMT exhibit significantly

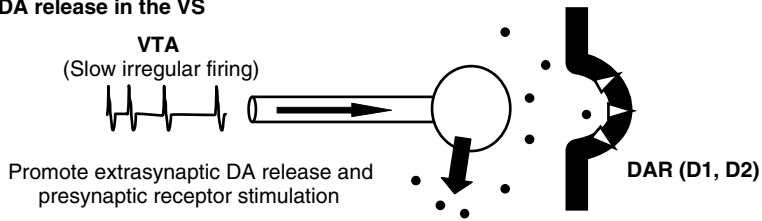
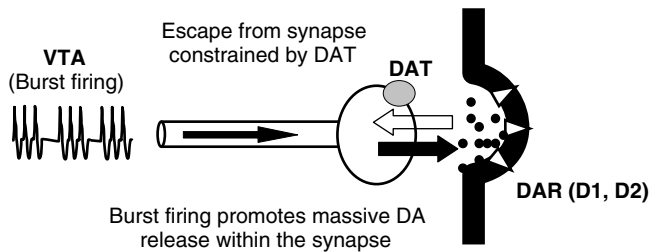
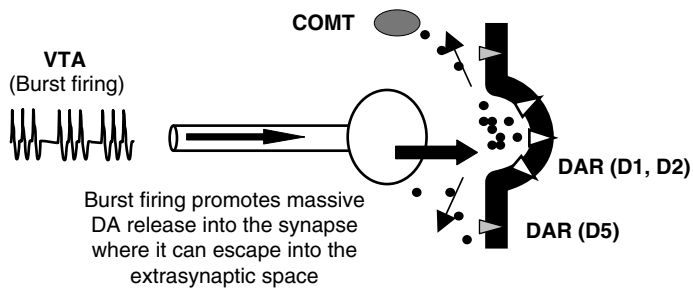
A Tonic DA release in the VS**B Phasic DA release in the VS****C Phasic DA release in the PFC**

FIG. 3. Tonic and phasic DA release in the PFC and striatum is regulated by COMT and DAT. (A) The slow, irregular baseline spike firing of DA neurons induces steady-state tonic DA release in the postsynaptic target regions. DA released in this manner is not strongly attenuated by the reuptake system, and hence can diffuse into the extrasynaptic space. (B) When burst spike firing occurs in DA neurons, massive, phasic DA release is produced in the striatum. Phasic DA release is subject to rapid removal by the DA uptake system, DAT, before it can escape the synaptic cleft. Therefore, phasic DA release is transient and spatially constrained to the vicinity of the synaptic cleft. (C) In contrast, in the PFC where there are very low levels of DAT, the primary means for inactivation of released DA is via the metabolizing enzyme COMT. As a result, both tonic and phasic DA released from terminals in the PFC is allowed to diffuse into the extrasynaptic space. One of the targets that extrasynaptic DA can stimulate is the D5 receptor, which is selectively located in this compartment. Adapted from Bilder *et al.* (2004).

lower working memory and PFC activity than those having the Met/Met allele. Since Val/Val COMT metabolizes DA at a faster rate than Met/Met COMT, this difference is believed to affect DA dynamics within the PFC (Egan *et al.*, 2001; Weinberger *et al.*, 2001). Given that COMT cannot inactivate DA actions as efficiently as that produced by DAT in the subcortical area, it is likely that tonic and phasic DA release play substantially different roles between the PFC and the VS, with phasic DA release underlying extrasynaptic DA concentrations that are regulated by COMT activity (Fig. 3; Bilder *et al.*, 2004). The increased area of diffusion of extracellular DA in the PFC, in addition to affecting a number of neuronal elements, may also lead to a selective stimulation of a particular subtype of DA receptor. Thus, a study has shown that the DA D5 receptor subtype is located exclusively at extrasynaptic sites of PFC neurons (Paspalas and Goldman-Rakic, 2004). This would position the D5 receptor to respond preferentially to tonic DA that has diffused away from its release site.

Studies have found that tonic and phasic DA release can actually regulate the balance between PFC and limbic inputs into the VS (Goto and Grace, 2005b). It was found that inactivation of the ventral pallidum to disinhibit spontaneous DA neuron firing and consequently increase tonic DA release induces attenuation of PFC responses in the VS, which is also mimicked by local infusion of a D2 agonist. In contrast, activation of the ventral pallidum to decrease tonic DA release causes a selective increase of PFC responses in the VS, which is also mimicked by local infusion of a D2 antagonist. Therefore, as observed *in vitro* (O'Donnell and Grace, 1994) and *in vivo* (Brady and O'Donnell, 2004; West and Grace, 2002), the DA system exerts a bimodal regulation over the PFC afferents to the VS, and this modulation is controlled by tonic DA release caused by spontaneous DA neuron spike firing (Goto and Grace, 2005b). In contrast, when the DA neurons are caused to burst firing by activation of the pedunculopontine tegmental glutamatergic afferent drive to the VTA, the resultant phasic DA release was found to selectively augment HPC afferent drive of the VS without affecting PFC inputs. This action can be mimicked by local infusion of a D1 agonist. However, unlike the D2 system, there does not appear to be a baseline D1 modulation of HPC synaptic drive on VS neurons.

The information gleaned from the studies above shows that the tonic and phasic DA system can exert unique effects on afferent integration within the VS. However, it is likely that the tonic and phasic DA system work in concert to modulate VS function, rather than as independent entities. Indeed, we have found that these systems can function in an integrated manner to augment DA function. Thus, we have found that burst firing can only be induced in DA neurons that are exhibiting spontaneous spike firing. This is likely due to the fact that burst firing is dependent on N-methyl-D-aspartate (NMDA) receptor stimulation (Overton and Clark, 1992, 1997), and in a hyperpolarized, nonfiring neuron, NMDA channels are not closed by a magnesium block (Mayer *et al.*, 1984; Nowak *et al.*, 1984). This

hyperpolarized state can be relieved by decreasing the massive bombardment of DA neurons by GABAergic inhibition (Grace and Bunney, 1979, 1985) that hold these neurons in the inactive state. When tonic DA spike firing is activated by a decrease in ventral pallidal inhibitory control, pedunculo-pontine tegmental glutamatergic inputs are capable of causing a much larger population of DA neurons to enter the burst firing mode (Lodge and Grace, 2006b). With the tonic and phasic system acting together, DA transmission in the VS would exert two effects: a tonic DA D2-mediated attenuation of PFC drive and a phasic D1-mediated potentiation of HPC drive over VS neurons. Thus, increasing DA transmission will shift the balance of inputs to the VS away from PFC drive, favoring limbic drive. Conversely, a decrease in DA system function, as may occur during cognitive states or as a consequence of antipsychotic drug-induced depolarization block as described below (Section VII; Grace *et al.*, 1997), would shift the balance toward PFC predominance over the limbic input.

IV. Cellular Actions of DA

There are two major classes of DA receptors that mediate the actions of DA: the D1 family and the D2 family. The D1 receptor family has two subclasses, D1 and D5, whereas D2 receptor family is composed of three subclasses, D2, D3, and D4 receptors (Jackson and Westlind-Danielsson, 1994; Seeman and Van Tol, 1993). These receptors are located at both pre- and postsynaptic sites. D1 and D2 receptors are coupled to Gs and Gi proteins and, as a consequence, mediate activation and inhibition of second messenger cascades, respectively (Greengard *et al.*, 1999; Neve *et al.*, 2004). This leads to phosphorylation and dephosphorylation of a number of channels and receptors, modulating cell excitability (Cepeda *et al.*, 1993, 1995; Gorelova and Yang, 2000; Gullledge and Jaffe, 1998; Tseng and O'Donnell, 2004; Wang and Goldman-Rakic, 2004), glutamate and GABA neurotransmission (Gao *et al.*, 2001; Seamans *et al.*, 2001a,b), and synaptic plasticity (Bissiere *et al.*, 2003; Centonze *et al.*, 2001; Otani *et al.*, 2003; Otmakhova and Lisman, 1996; see references such as Nicola *et al.*, 2000 and Seamans and Yang, 2004 for detailed summary of channels and receptors modulated by DA). The most consistent finding with respect to DA modulation of channels and receptors in the PFC and striatum is the D1-mediated facilitation of calcium influx into neurons via interactions with NMDA channels (Cepeda *et al.*, 1993; Tseng and O'Donnell, 2004) and L-type calcium (Surmeier *et al.*, 1995; Tseng and O'Donnell, 2004). In contrast, although it is still controversial, the D2 receptors have been suggested to decrease AMPA in the PFC and striatum (Cepeda *et al.*, 1993; Tseng and O'Donnell, 2004). DA also seems to increase or decrease GABA release via D1 and D2 receptors, respectively, located on interneurons

in the PFC (Gorelova *et al.*, 2002; Seamans *et al.*, 2001b). However, another study shows that D2 receptor activation has the opposite action: an increase in GABA release in the PFC (Tseng and O'Donnell, 2004). Anatomical studies have shown that D1 receptors are found on presynaptic sites of terminals within the PFC (Paspalas and Goldman-Rakic, 2005), and stimulation of presynaptic D1 receptors have been reported to decrease glutamate release in the PFC (Gao *et al.*, 2001; Seamans *et al.*, 2001a). In contrast, D1 receptors have not been found to be present presynaptically on terminals within the striatum (Hara and Pickel, 2005). Instead, in the striatum only the D2 DA receptor has been localized to presynaptic sites (Fisher *et al.*, 1994; Wang and Pickel, 2002). Although the role of presynaptic DA receptors in controlling glutamate and/or GABA release has not been fully characterized, within the striatum, stimulation of presynaptic D2 receptors has been shown to decrease glutamate release (Bamford *et al.*, 2004; Umemiya and Raymond, 1997) and to attenuate PFC-evoked excitatory postsynaptic potentials (EPSPs) in the striatum (Brady and O'Donnell, 2004; Goto and Grace, 2005b; O'Donnell and Grace, 1994; West and Grace, 2002).

It is difficult to examine the cellular effects of DA transmission *in vivo*, given the potent regulation of the system and the multiple sites of action of DA release. In order to examine how DA is modulating activity within the striatum, studies were conducted in which DA antagonists were infused locally into the striatum via reverse microdialysis to achieve local blockade of receptors while performing *in vivo* electrophysiological recordings from neurons adjacent to the probe, through which the effects produced by attenuating endogenous DA transmission can be observed (West and Grace, 2002). Using this methodology, it was found that striatal neurons *in vivo* are potently modulated by DA D1 and D2 receptors, with D1 receptors controlling neuronal excitability and D2 receptors modulating baseline activity and cortical afferent drive (West and Grace, 2002).

Functional studies into the significance of DA modulation of neuronal activity points to the involvement of DA in synaptic plasticity and learning and memory associated with it (Schultz, 2002). Synaptic plasticity has been most commonly tested by induction of long-term potentiation (LTP) and depression (LTD; Abraham and Tate, 1997; Bear and Malenka, 1994). Since LTP and LTD induction is closely linked with intracellular calcium influx (Abraham and Tate, 1997; Bear and Malenka, 1994), D1-mediated NMDA and L-type calcium channel phosphorylation have a critical impact on them (Greengard *et al.*, 1999). Indeed, synaptic plasticity induction in brain regions receiving DA innervation, including the PFC (Gurden *et al.*, 1999; Otani *et al.*, 1998), striatum (Arbuthnott *et al.*, 2000; Centonze *et al.*, 2001; Reynolds *et al.*, 2001), HPC (Li *et al.*, 2003; Otmakhova and Lisman, 1996), and amygdala (Bissiere *et al.*, 2003), is dependent on DA release. For example, D1 antagonist is known to prevent LTP induced at HPC afferents into the PFC (Gurden *et al.*, 2000).

Furthermore, LTD induction within PFC circuitry is shown to depend on both D1 and D2 receptor activation (Otani *et al.*, 1998). Similarly, D1- but not D2-dependent LTP has been found in the HPC (Otmakhova and Lisman, 1996) and amygdala (Bissiere *et al.*, 2003). However, LTP and LTD induction in the striatum is somewhat controversial. DA-dependent plasticity that involves both D1 and D2 receptors has been reported with respect to LTP and LTD induction in the dorsal striatum (Calabresi *et al.*, 1992; Spencer and Murphy, 2000), whereas despite the presence of dense DA projections, synaptic plasticity in the VS has been reported to be DA independent (Pennartz *et al.*, 1993). In contrast, studies have described DA-dependent plasticity in the VS (Goto and Grace, 2005a; Thomas *et al.*, 2001). Some studies have shown that DA affects NMDA and AMPA receptor trafficking (Malenka, 2003; Wolf *et al.*, 2004). D1 receptor activation stimulates cell surface expression of GluR1 subunit of NMDA receptors in the PFC (Sun *et al.*, 2005) and VS (Mangiacavalli and Wolf, 2004), resulting in increased calcium influx into the neurons.

Overall, DA appears to control the balance of excitatory and inhibitory drive of neural activity within its projection sites, and to mediate synaptic plasticity associated with learning and memory.

V. Roles of DA on Cognitive and Affective Functions

DA is involved in the induction of synaptic plasticity in the brain regions receiving DA neuron projections, and therefore is believed to play a pivotal role in learning and memory processes (Schultz, 2002). An elegant study of electrophysiological recordings from DA neurons in primates done by Schultz and colleagues (Waelti *et al.*, 2001) have revealed that the activation of DA neuron spike discharge during learning trials is consistent with that predicted by learning theory (Rescorla–Wagner rule of classical conditioning; Rescorla and Wagner, 1972), suggesting that DA signals can be used to prime selective brain areas for learning (Schultz, 2002). DA signals are used to mediate different types of learning and memory depending on the specific regions involved (Schultz *et al.*, 2000, 2003). Thus, DA projections into the dorsal striatum are critical for motor learning and habit formation such as playing musical instruments or riding a bicycle (Graybiel, 1998). Moreover, studies have shown that the DA innervation of limbic structures plays a different role in mediating learning processes. Thus, the DA innervation of the HPC appears to be involved in the formation of long-term memory (Lisman and Grace, 2005), whereas DA projections into the amygdala mediate emotional memory such as aversive conditioning (Nader and LeDoux, 1999; Rodrigues *et al.*, 2004; Rosenkranz and Grace, 2002). The role of the DA innervation of the VS in learning processes is less clear. However,

the VS is the brain region where limbic and cortical inputs converge to integrate context- and emotion-associated goal-directed behavior (Mogenson *et al.*, 1980). Therefore, it is likely that mesolimbic DA projections to the VS are involved in learning processes that could occur in acquiring these aspects of goal-directed motor control (Everitt and Robbins, 2005; Kelley and Berridge, 2002). Given that the DA system is also believed to provide a “learning signal” to the limbic system (Schultz, 2002), we examined the ability of DA transmission to exert short-term modulation of the PFC and HPC drive within the VS, and how this relates to the processing of goal-directed behavior (Goto and Grace, 2005b). Disconnection of the HPC inputs from the VS by unilateral injection of lidocaine into the HPC while infusing D1 antagonists into the contralateral VS was found to interfere with the efficient acquisition of goal-directed behavior. In contrast, when the PFC was disconnected from the VS by unilateral infusion of lidocaine into the PFC while injecting a D2 agonist into the contralateral VS, there was no interference in the acquisition of an initial discrimination task. Instead, when the response strategy was altered (i.e., when the rat had to change from a visually guided response strategy to a direction-guided response strategy), the rats exhibited perseveration, using the previous response strategy rather than switching to the new paradigm. This suggests that D1-dependent HPC-VS information processing mediates learning of a response strategy, whereas D2-dependent PFC-VS information processing is crucial for flexible switching of this response strategy in guiding goal-directed behavior (Goto and Grace, 2005b). This type of disruption of limbic and cortical balance in regulating information processing in the VS for goal-directed behavior is consistent with the type of behavioral disruption proposed to occur in drug addiction (Everitt and Wolf, 2002; Kelley and Berridge, 2002; Ridley, 1994). Thus, in drug-addicted subjects, a loss of cortically driven behavioral flexibility in favor of HPC-driven perseveration in drug-seeking behavior could be one of the mechanisms by which drug-addicted individuals lose their ability to modify their behavior in favor of more behaviorally effective strategies. Indeed, we also found that repeated cocaine treatment could induce abnormal strengthening of HPC inputs, with LTP induction in the HPC inputs and LTD attenuation of PFC inputs, resulting in an imbalance of limbic and cortical drive of VS activity (Goto and Grace, 2005a).

In contrast, the role of the mesocortical DA innervation into the PFC for learning and memory is not readily apparent from what we know about this system. Thus, the functions associated with the PFC, such as short-term storage of memory (few seconds to up to minutes; Funahashi *et al.*, 1993; Goldman-Rakic, 1995), flexible switching of response strategy (Milner, 1963; Owen *et al.*, 1993), attention (Gorenstein *et al.*, 1989; Knight *et al.*, 1995; Muir *et al.*, 1996), and future planning (Baker *et al.*, 1996; Ingvar, 1985; Owen *et al.*, 1990), are considered to be independent of what has been typically associated with more standard learning and memory paradigms, and may not involve synaptic plasticity

such as LTP and LTD induction in this region. Nevertheless, the PFC is known to exhibit synaptic plasticity in its network (Herry and Garcia, 2002; Laroche *et al.*, 1990; Otani *et al.*, 2003), and mesocortical DA release is essential for induction of such synaptic plasticity (Gurden *et al.*, 1999; Otani *et al.*, 1998), suggesting that any cognitive functions requiring proper DA release in the PFC could involve DA-dependent synaptic plasticity. However, the exact roles of such synaptic plasticity in the PFC have yet to be determined.

With respect to working memory, it has been reported that DA effects within the PFC are bimodal in function. Thus, there is an optimal level of DA stimulation required for proper PFC functioning, with either over- or understimulation of D1 receptors leading to dysfunctional states (Granon *et al.*, 2000; Zahrt *et al.*, 1997). This is an example of the classic “inverted U”-shaped relationship known as the Yerkes–Dodson curve (Yerkes and Dodson, 1908). In primate studies, it has been shown that D1 receptors play a crucial role in short-term memory functions (Goldman-Rakic, 1995). This is particularly pertinent for spatial working memory. Thus, when a monkey is required to hold the location of an object in memory for a short period of time in order to guide a subsequent behavior, PFC neurons associated with that special location become activated, and remain so until the task is completed (Funahashi *et al.*, 1993). Such sustained spike firing of PFC neurons during the time in which information is held in memory is disrupted by a D1 receptor antagonist (Sawaguchi and Goldman-Rakic, 1994). Whether D2 receptors also exhibit a U-shaped functional relationship in the PFC is not known. In primate electrophysiological recording studies, evidence for an involvement of D2 receptors in the temporal retention of information during short time periods has not been reported (Sawaguchi and Goldman-Rakic, 1994; Wang *et al.*, 2004). Nonetheless, evidence suggests that short-term memory can be affected by administration of D2 agonists or antagonists into human subjects (Kimberg *et al.*, 1997; Mehta *et al.*, 2001). Although this supports a D2 receptor involvement in short-term memory in humans, the location of this D2 action is not known.

VI. Development and Maturation of the DA System

There is increasing evidence that many major psychiatric disorders have their origin in a disruption occurring during development of the nervous system. Therefore, understanding the development and maturation of DA systems is essential for a more complete comprehension of the etiology and pathophysiology of a number of major psychiatric disorders such as schizophrenia and attention deficit/hyperactivity disorder (ADHD) in which neurodevelopmental compromises in the DA system have been implicated (Castellanos, 1997; Eells, 2003;

Harrison, 1999; Heyman and Murray, 1992; Nicoullon, 2002; Sonuga-Barke, 2005; Weinberger, 1987).

DA neurons are born in the midbrain of rats at around embryonic day (ED) 12–16, with a peak occurring at ED13 (Lauder and Bloom, 1974). This is followed by programmed cell death of subsets of DA neurons that is initiated around postnatal day (PD) 2 and PD14 (Jackson-Lewis *et al.*, 2000; Oo and Burke, 1997). Studies have shown that growth factors released in the areas targeted by DA neuron terminals such as the striatum appear to regulate this programmed cell death (Breiter *et al.*, 1997; Hyman *et al.*, 1991; Poulsen *et al.*, 1994). This pruning of the DA cell population in the midbrain continues to occur until about PD20 to arrive at the final adult population of midbrain DA neurons (Jackson-Lewis *et al.*, 2000; Oo and Burke, 1997). In contrast, the DA collateralization into the targeted areas continues to increase until adolescence begins. This occurs in concert with an increase in the number of DA receptors expressed in postsynaptic target areas. During puberty, a second wave of pruning of the DA innervation is initiated, but DA receptor pruning occurs differentially depending on the target region. Thus, there is substantial pruning of both D1 and D2 receptors in the PFC (Andersen *et al.*, 2000) and dorsal striatum (Teicher *et al.*, 1995) during adolescence and young adulthood. Indeed, it has been shown that the maturation of the DA system has functional significance with respect to D1-mediated facilitation of NMDA currents (Tseng and O'Donnell, 2004) and D2 receptor modulation of interneuron activity (Tseng and O'Donnell, 2004) in the PFC. In contrast, the reorganization of DA receptor expression in the VS during adolescence is not as prominent (Teicher *et al.*, 1995). Nevertheless, it appears that the mesolimbic DA system in the VS does not seem to be fully matured after puberty, since for example, drug sensitization produced by repeated psychostimulant administration induces significantly different effects in pre- and midpubertal animals (Tirelli *et al.*, 2003; Ujike *et al.*, 1995); a process that in the adult animal is known to involve sensitization of DA release in the VS (but see Borgland *et al.*, 2006 for evidence of sensitization in prepubertal animals). DA system maturation during adolescence is of particular interest with respect to the pathophysiology of schizophrenia, given that brain compromises at the second trimester of pregnancy have been suggested to occur in the brains of schizophrenia patients, whereas the onset of psychotic symptoms are typically delayed until late adolescence to early adulthood (Harrison, 1999; Weinberger, 1987). With respect to disorders such as ADHD, a similar type of disruption of mesocortical DA system function has also been proposed (Castellanos, 1997; Heyman and Murray, 1992; Nicoullon, 2002; Sonuga-Barke, 2005). However, the time course underlying the origin of ADHD symptoms appears to be substantially different from that of schizophrenia, since in schizophrenia the DA deficit occurs during maturation, whereas in ADHD the symptomatology is already present at a very early age.

VII. DA Deficits in Schizophrenia

Since the first description of amphetamine induction of schizophrenia-like symptoms in normals (Connell, 1958) and the observation that DA D2 antagonists are effective in the treatment of this disorder (Carlsson, 1974), a dysfunction within the DA system has been implicated in schizophrenia (Seeman, 1987). These classical studies suggested that schizophrenia symptoms may be caused by an excess in DA release; however, more recent studies suggest that this may be an oversimplification. Thus, results show that there is an augmentation of DA release in the striatum only during specific types of system activation, and this is correlated with the positive psychotic symptoms of this disorder (Laruelle *et al.*, 1999). In contrast, it has been proposed that the functional deficits observed in the PFC of schizophrenia patients could be due to a deficit in DA activity, which may underlie the negative or deficit state of schizophrenia (Abi-Dargham *et al.*, 2002; Davis *et al.*, 1991).

A potential relationship that has been advanced to account for the disturbed DA system in schizophrenia is an opposing relationship between PFC and striatal DA release (Fig. 4). Thus, primate (Kolachana *et al.*, 1995; Saunders *et al.*, 1998), rodent (Jackson *et al.*, 2001), and human studies (Meyer-Lindenberg *et al.*, 2002) have shown that the attenuated PFC activity in schizophrenia may be correlated with an exaggerated DA release in the striatum. This can be accounted for by an examination of the anatomical organization of PFC-VTA and HPC-VTA loops (Figs. 1 and 2; Lisman and Grace, 2005; Sesack and Carr, 2002). PFC projections onto GABA interneurons in the VTA can suppress spike firing in the DA neurons that project into the striatum. Therefore, with PFC deficits, the PFC drive of VTA DA neurons is diminished, leading to abnormally augmented DA release in the striatum in schizophrenia. Our study has revealed that increasing DA release in the VS facilitates limbic inputs and attenuates PFC inputs, whereas decreasing DA release shifts the balance in favor of the PFC inputs (Goto and Grace, 2005b), suggesting that DA maintains the balance between limbic and PFC drive of VS neurons. As such, a combination of abnormally attenuated PFC drive of VS neurons with augmented DA release in the VS that is shown to occur in schizophrenia patients (Laruelle *et al.*, 1999; Meyer-Lindenberg *et al.*, 2002) could cause inappropriate limbic drive of the VS and disruption of goal-directed behavior. A key mechanism for the inverse relationship between PFC activity and striatal DA release may be drawn from the known reciprocal interactions between the HPC and PFC (Fig. 4). Thus, the HPC sends direct projections into the PFC (Fuster, 1997; Jay *et al.*, 1989), whereas the PFC sends indirect projections into the HPC through the temporal cortex (Fuster, 1997; Groenewegen and Uylings, 2000; Kyd and Bilkey, 2003). Since it has been suggested that the PFC exerts an inhibitory influence over limbic structures (Fuster, 1997; Grace and Rosenkranz, 2002;

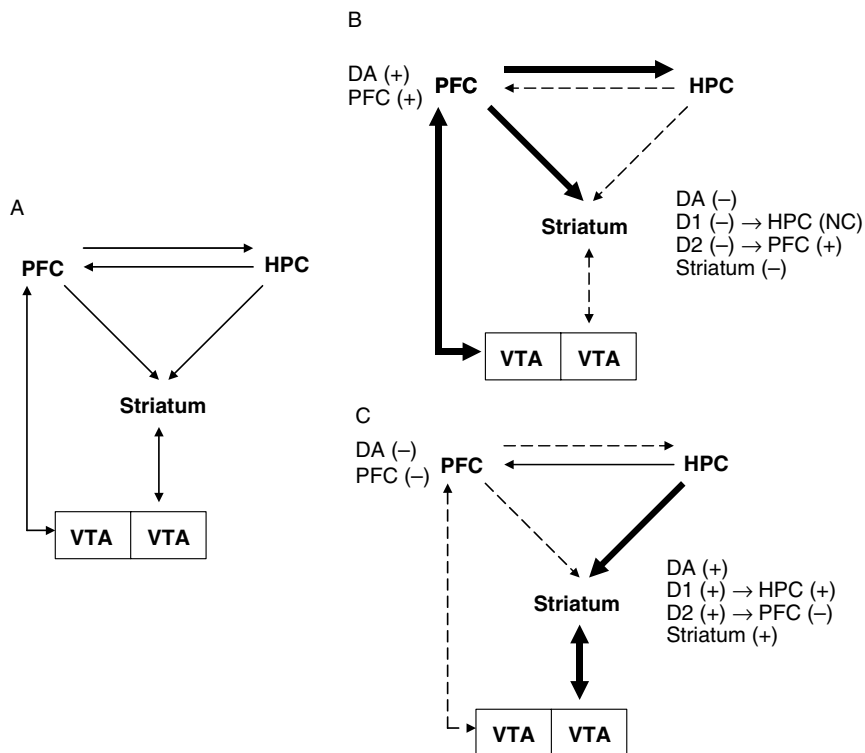


FIG. 4. Schematic diagrams of the mesocortical and mesolimbic DA system and its regulation by interactions among the PFC, HPC, and striatum. (A) The reciprocal interaction between the PFC and HPC maintains the balance of mesocortical and mesolimbic DA release. (B) When PFC activity is stronger than HPC activity, mesocortical DA release is increased in the PFC, which would further facilitate PFC activity. Given our observation that the PFC will attenuate HPC activity, this would lead to an attenuation of HPC drive on the striatum which, in turn, will lead to a decrease in mesolimbic DA release. With a fall in mesolimbic DA levels, the PFC input to the striatum will be further facilitated due to the decreased stimulation of presynaptic D2 receptors on the PFC afferents. (C) In contrast, when PFC activity is abnormally attenuated, as may occur in schizophrenia, there would be a reduction in mesocortical DA release as well as a decreased PFC-mediated inhibition of HPC activity. As a consequence, there would be an increase in HPC drive of striatal neuron activity and mesolimbic DA release. The increase in striatal DA D1 and D2 receptor stimulation would then cause a further attenuation of PFC input and facilitation of HPC input to the striatum. Administration of a DA antagonist would then serve to restore the HPC/PFC balance. (+), (-), and (NC) denotes increase, decrease, and no change, respectively.

Shimamura, 2000; Zironi *et al.*, 2001), when there is a higher degree of PFC activity in relation to the HPC, DA release in the VS could be suppressed, whereas higher activity within the HPC would lead to augmented DA release in the VS. Therefore, in schizophrenia, one would postulate that attenuated PFC function

could result in augmented HPC activity, which in turn would cause exaggerated DA release in the VS and further shift toward limbic predominance over PFC control (Fig. 4; Goto and Grace, 2005b; Meyer-Lindenberg *et al.*, 2002, 2005).

Since the discovery that schizophrenia symptoms respond to treatment by D2 antagonists (Carlsson, 1974), the therapeutic approach to treating schizophrenia by antipsychotic drugs has targeted brain DA systems. Nevertheless, examination into a biological basis for this purported hyper-DA state via testing for alterations in D2 receptors has not produced consistent results. Postmortem tissue and imaging studies reporting increased D2 density in schizophrenia have been controversial, with many studies reporting no difference (Farde *et al.*, 1987; Nordstrom *et al.*, 1995). In contrast, given that one of the core deficits in schizophrenia is cognitive dysfunctions that have been associated with PFC activity (Weinberger *et al.*, 1994), studies have focused on alterations of D1 receptors in the PFC of schizophrenia patients. Indeed, although the results are still incomplete, alterations, that is increases (Okubo *et al.*, 1997) or decreases (Abi-Dargham *et al.*, 2002), of D1 receptors in the PFC of schizophrenia patients have been reported. Given that D1 receptor stimulation facilitates calcium influx via NMDA channels (Greengard *et al.*, 1999; Tseng and O'Donnell, 2004), alterations in D1 receptors are also consistent with the hypo-NMDA function theory of schizophrenia pathophysiology (Coyle *et al.*, 2003; Goff and Coyle, 2001; Jentsch and Roth, 1999). This model is based on observations that NMDA antagonists can lead to schizophrenia-like symptoms in normal patients, and moreover can precipitate a recurrence of symptoms in schizophrenia patients that is indistinguishable from that of a relapse (Coyle *et al.*, 2003; Goff and Coyle, 2001; Jentsch and Roth, 1999). Since it is known that DA stimulation of D1 receptors in the PFC exhibits an inverted U-shaped relationship, with optimal DA release required for mediating effective cognitive functions (Lidow *et al.*, 1998; Robbins, 2005), increases or decreases in D1 receptors would shift the relationship between optimal DA release and over- or understimulation of D1 receptors and their impact on PFC function (Callicott *et al.*, 2003; Goto *et al.*, 2004; Manoach, 2003).

The exact mechanism by which D2 antagonism achieves therapeutic efficacy in schizophrenia is still unclear. However, a number of possibilities have been suggested. First, D2 antagonists increase glutamate release from the PFC afferents into the striatum (Brady and O'Donnell, 2004; Goto and Grace, 2005b; O'Donnell and Grace, 1994), facilitating corticostriatal information processing. Alternately, D2 antagonism could affect D2 receptors located within the PFC (Seamans *et al.*, 2001b; Tseng and O'Donnell, 2004; Wang and Goldman-Rakic, 2004; Wang *et al.*, 2004), which may facilitate PFC activity. These two possibilities predict that D2 antagonists should have immediate effects on schizophrenia symptoms. Although studies have suggested that antipsychotic drugs may have an immediate effect on schizophrenia symptoms (Ngan *et al.*, 2002), these results are confounded by the fact that the antipsychotic drugs were tested in patients that

had been withdrawn from treatment. It is known from animal models that prior antipsychotic drug treatment will sensitize the DA system to subsequent administration, dramatically shortening the time required to achieve DA neuron inactivation (Moore *et al.*, 1998). In contrast, most clinical trials of drug-naïve patients show a delayed onset of therapeutic actions (Hamill and Fontana, 1975; Johnstone *et al.*, 1978), suggesting other possible mechanisms may be involved. One mechanism that has been consistent with both the pharmacology and time course of antipsychotic drug action is the induction of depolarization block of DA neurons by repeated antipsychotic drug treatment (Grace *et al.*, 1997). It has been shown that repeated administration of D2 antagonists for 3 weeks induces substantial membrane potential depolarization of DA neurons, leading to a cessation of spike discharge (Grace and Bunney, 1986). As a consequence, these DA neurons become unable to evoke spike firing secondary to an overdepolarized membrane potential state. Therefore, unlike the blockade of postsynaptic receptors produced by acute administration of these drugs (that could be overcome by compensatory changes), an abnormal excitatory drive of DA neurons would be incapable of increasing DA release. Therefore, depolarization block of DA neurons could reduce the amount of DA release in the striatum produced in an event-related manner.

VIII. Conclusions

One thing that is clear from the above review is that the DA system exerts complex, multifaceted actions within several interrelated systems of the mammalian brain. It has a role in motor function, motivation and reward, attention, and learning and memory. Therefore, the widespread but anatomically discrete projections of the DA system are positioned to coordinate functions that can have a major impact on cognition and goal-directed behavior. While such diverse functions illuminate the many-faceted disruptions that can occur within this system to lead to a variety of psychiatric disturbances, it also highlights the difficulty in specifically targeting therapeutic agents to single DA systems. A better understanding of the factors that control the DA system, and how they may differentially affect specific DA circuits, may provide the type of insight required to effectively target therapeutic agents.

References

- Abi-Dargham, A., Mawlawi, O., Lombardo, I., Gil, R., Martinez, D., Huang, Y., Hwang, D. R., Keilp, J., Kochan, L., Van Heertum, R., Gorman, J. M., and Laruelle, M. (2002). Prefrontal dopamine D1 receptors and working memory in schizophrenia. *J. Neurosci.* **22**, 3708–3719.

- Abraham, W. C., and Tate, W. P. (1997). Metaplasticity: A new vista across the field of synaptic plasticity. *Prog. Neurobiol.* **52**, 303–323.
- Anden, N. E., Carlsson, A., Dahlstroem, A., Fuxe, K., Hillarp, N. A., and Larsson, K. (1964). Demonstration and mapping out of nigro-neostriatal dopamine neurons. *Life Sci.* **3**, 523–530.
- Andersen, S. L., Thompson, A. T., Rutstein, M., Hostetter, J. C., and Teicher, M. H. (2000). Dopamine receptor pruning in prefrontal cortex during the periadolescent period in rats. *Synapse* **37**, 167–169.
- Arbuthnott, G. W., Ingham, C. A., and Wickens, J. R. (2000). Dopamine and synaptic plasticity in the neostriatum. *J. Anat.* **196**(Pt. 4), 587–596.
- Au-Young, S. M., Shen, H., and Yang, C. R. (1999). Medial prefrontal cortical output neurons to the ventral tegmental area (VTA) and their responses to burst-patterned stimulation of the VTA: Neuroanatomical and *in vivo* electrophysiological analyses. *Synapse* **34**, 245–255.
- Baker, S. C., Rogers, R. D., Owen, A. M., Frith, C. D., Dolan, R. J., Frackowiak, R. S., and Robbins, T. W. (1996). Neural systems engaged by planning: A PET study of the Tower of London task. *Neuropsychologia* **34**, 515–526.
- Bamford, N. S., Zhang, H., Schmitz, Y., Wu, N. P., Cepeda, C., Levine, M. S., Schmauss, C., Zakharenko, S. S., Zablow, L., and Sulzer, D. (2004). Heterosynaptic dopamine neurotransmission selects sets of corticostriatal terminals. *Neuron* **42**, 653–663.
- Bear, M. F., and Malenka, R. C. (1994). Synaptic plasticity: LTP and LTD. *Curr. Opin. Neurobiol.* **4**, 389–399.
- Bedard, P., Larochelle, L., Parent, A., and Poirier, L. J. (1969). The nigrostriatal pathway: A correlative study based on neuroanatomical and neurochemical criteria in the cat and the monkey. *Exp. Neurol.* **25**, 365–377.
- Bilder, R. M., Volavka, J., Lachman, H. M., and Grace, A. A. (2004). The catechol-O-methyltransferase polymorphism: Relations to the tonic-phasic dopamine hypothesis and neuropsychiatric phenotypes. *Neuropsychopharmacology* **29**, 1943–1961.
- Bissiere, S., Humeau, Y., and Luthi, A. (2003). Dopamine gates LTP induction in lateral amygdala by suppressing feedforward inhibition. *Nat. Neurosci.* **6**, 587–592.
- Borgland, S. L., Taha, S. A., Sarti, F., Fields, H. L., and Bonci, A. (2006). Orexin A in the VTA is critical for the induction of synaptic plasticity and behavioral sensitization to cocaine. *Neuron* **49**, 589–601.
- Brady, A. M., and O'Donnell, P. (2004). Dopaminergic modulation of prefrontal cortical input to nucleus accumbens neurons *in vivo*. *J. Neurosci.* **24**, 1040–1049.
- Breiter, H. C., Gollub, R. L., Weisskoff, R. M., Kennedy, D. N., Makris, N., Berke, J. D., Goodman, J. M., Kantor, H. L., Gastfriend, D. R., Riorden, J. P., Mathew, R. T., Rosen, B. R., *et al.* (1997). Acute effects of cocaine on human brain activity and emotion. *Neuron* **19**, 591–611.
- Brinley-Reed, M., and McDonald, A. J. (1999). Evidence that dopaminergic axons provide a dense innervation of specific neuronal subpopulations in the rat basolateral amygdala. *Brain Res.* **850**, 127–135.
- Calabresi, P., Maj, R., Mercuri, N. B., and Bernardi, G. (1992). Coactivation of D1 and D2 dopamine receptors is required for long-term synaptic depression in the striatum. *Neurosci. Lett.* **142**, 95–99.
- Callicott, J. H., Mattay, V. S., Verchinski, B. A., Marengo, S., Egan, M. F., and Weinberger, D. R. (2003). Complexity of prefrontal cortical dysfunction in schizophrenia: More than up or down. *Am. J. Psychiatry* **160**, 2209–2215.
- Carbon, M., and Marie, R. M. (2003). Functional imaging of cognition in Parkinson's disease. *Curr. Opin. Neurol.* **16**, 475–480.
- Carlsson, A. (1959). Detection and assay of dopamine. *Pharmacol. Rev.* **11**, 300–304.
- Carlsson, A. (1974). Antipsychotic drugs and catecholamine synapses. *J. Psychiatr. Res.* **11**, 57–64.
- Carlsson, A., Lindqvist, M., Magnusson, T., and Waldeck, B. (1958). On the presence of 3-hydroxytyramine in brain. *Science* **127**, 471.

- Carr, D. B., and Sesack, S. R. (2000). Projections from the rat prefrontal cortex to the ventral tegmental area: Target specificity in the synaptic associations with mesoaccumbens and mesocortical neurons. *J. Neurosci.* **20**, 3864–3873.
- Castellanos, F. X. (1997). Toward a pathophysiology of attention-deficit/hyperactivity disorder. *Clin. Pediatr. (Phila.)* **36**, 381–393.
- Centonze, D., Picconi, B., Gubellini, P., Bernardi, G., and Calabresi, P. (2001). Dopaminergic control of synaptic plasticity in the dorsal striatum. *Eur. J. Neurosci.* **13**, 1071–1077.
- Cepeda, C., Buchwald, N. A., and Levine, M. S. (1993). Neuromodulatory actions of dopamine in the neostriatum are dependent upon the excitatory amino acid receptor subtypes activated. *Proc. Natl. Acad. Sci. USA* **90**, 9576–9580.
- Cepeda, C., Chandler, S. H., Shumate, L. W., and Levine, M. S. (1995). Persistent Na^+ conductance in medium-sized neostriatal neurons: Characterization using infrared videomicroscopy and whole cell patch-clamp recordings. *J. Neurophysiol.* **74**, 1343–1348.
- Chergui, K., Akaoka, H., Charlety, P. J., Saunier, C. F., Buda, M., and Chouvet, G. (1994). Subthalamic nucleus modulates burst firing of nigral dopamine neurones via NMDA receptors. *Neuroreport* **5**, 1185–1188.
- Connell, P. H. (1958). "Amphetamine Psychosis." Oxford University Press, London.
- Coyle, J. T., Tsai, G., and Goff, D. (2003). Converging evidence of NMDA receptor hypofunction in the pathophysiology of schizophrenia. *Ann. NY Acad. Sci.* **1003**, 318–327.
- Davis, K. L., Kahn, R. S., Ko, G., and Davidson, M. (1991). Dopamine in schizophrenia: A review and reconceptualization. *Am. J. Psychiatry* **148**, 1474–1486.
- De Keyser, J., Ebinger, G., and Vauquelin, G. (1989). Evidence for a widespread dopaminergic innervation of the human cerebral neocortex. *Neurosci. Lett.* **104**, 281–285.
- Eells, J. B. (2003). The control of dopamine neuron development, function and survival: Insights from transgenic mice and the relevance to human disease. *Curr. Med. Chem.* **10**, 857–870.
- Egan, M. F., Goldberg, T. E., Kolachana, B. S., Callicott, J. H., Mazzanti, C. M., Straub, R. E., Goldman, D., and Weinberger, D. R. (2001). Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia. *Proc. Natl. Acad. Sci. USA* **98**, 6917–6922.
- Everitt, B. J., and Robbins, T. W. (2005). Neural systems of reinforcement for drug addiction: From actions to habits to compulsion. *Nat. Neurosci.* **8**, 1481–1489.
- Everitt, B. J., and Wolf, M. E. (2002). Psychomotor stimulant addiction: A neural systems perspective. *J. Neurosci.* **22**, 3312–3320.
- Fallon, J. H., and Moore, R. Y. (1978). Catecholamine innervation of the basal forebrain. III. Olfactory bulb, anterior olfactory nuclei, olfactory tubercle and piriform cortex. *J. Comp. Neurol.* **180**, 533–544.
- Fallon, J. H., Koziell, D. A., and Moore, R. Y. (1978). Catecholamine innervation of the basal forebrain. II. Amygdala, suprarhinal cortex and entorhinal cortex. *J. Comp. Neurol.* **180**, 509–532.
- Farde, L., Wiesel, F. A., Hall, H., Halldin, C., Stone-Elander, S., and Sedvall, G. (1987). No D2 receptor increase in PET study of schizophrenia. *Arch. Gen. Psychiatry* **44**, 671–672.
- Faurbye, A. (1968). The role of amines in the etiology of schizophrenia. *Compr. Psychiatry* **9**, 155–177.
- Fischer, E. (1970). Biogenic amines and schizophrenia. *Psychosomatics* **11**, 495.
- Fisher, R. S., Levine, M. S., Sibley, D. R., and Ariano, M. A. (1994). D2 dopamine receptor protein location: Golgi impregnation-gold toned and ultrastructural analysis of the rat neostriatum. *J. Neurosci. Res.* **38**, 551–564.
- Floresco, S. B., Todd, C. L., and Grace, A. A. (2001). Glutamatergic afferents from the hippocampus to the nucleus accumbens regulate activity of ventral tegmental area dopamine neurons. *J. Neurosci.* **21**, 4915–4922.
- Floresco, S. B., West, A. R., Ash, B., Moore, H., and Grace, A. A. (2003). Afferent modulation of dopamine neuron firing differentially regulates tonic and phasic dopamine transmission. *Nat. Neurosci.* **6**, 968–973.

- Floresco, S. B., Magyar, O., Ghods-Sharifi, S., Vexelman, C., and Tse, M. T. (2006). Multiple dopamine receptor subtypes in the medial prefrontal cortex of the rat regulate set-shifting. *Neuropsychopharmacology* **31**, 297–309.
- Funahashi, S. (2001). Neuronal mechanisms of executive control by the prefrontal cortex. *Neurosci. Res.* **39**, 147–165.
- Funahashi, S., Chafee, M. V., and Goldman-Rakic, P. S. (1993). Prefrontal neuronal activity in rhesus monkeys performing a delayed anti-saccade task. *Nature* **365**, 753–756.
- Fuster, J. M. (1997). “The Prefrontal Cortex: Anatomy, Physiology, and Neuropsychology of the Frontal Lobe.” Lippincott-Raven, Philadelphia.
- Futami, T., Takakusaki, K., and Kitai, S. T. (1995). Glutamatergic and cholinergic inputs from the pedunculopontine tegmental nucleus to dopamine neurons in the substantia nigra pars compacta. *Neurosci. Res.* **21**, 331–342.
- Gao, W. J., Krimer, L. S., and Goldman-Rakic, P. S. (2001). Presynaptic regulation of recurrent excitation by D1 receptors in prefrontal circuits. *Proc. Natl. Acad. Sci. USA* **98**, 295–300.
- Gariano, R. F., and Groves, P. M. (1988). Burst firing induced in midbrain dopamine neurons by stimulation of the medial prefrontal and anterior cingulate cortices. *Brain Res.* **462**, 194–198.
- Goff, D. C., and Coyle, J. T. (2001). The emerging role of glutamate in the pathophysiology and treatment of schizophrenia. *Am. J. Psychiatry* **158**, 1367–1377.
- Gogos, J. A., Morgan, M., Luine, V., Santha, M., Ogawa, S., Pfaff, D., and Karayiorgou, M. (1998). Catechol-O-methyltransferase-deficient mice exhibit sexually dimorphic changes in catecholamine levels and behavior. *Proc. Natl. Acad. Sci. USA* **95**, 9991–9996.
- Goldman-Rakic, P. S. (1995). Cellular basis of working memory. *Neuron* **14**, 477–485.
- Gorelova, N., Seamans, J. K., and Yang, C. R. (2002). Mechanisms of dopamine activation of fast-spiking interneurons that exert inhibition in rat prefrontal cortex. *J. Neurophysiol.* **88**, 3150–3166.
- Gorelova, N. A., and Yang, C. R. (2000). Dopamine D1/D5 receptor activation modulates a persistent sodium current in rat prefrontal cortical neurons *in vitro*. *J. Neurophysiol.* **84**, 75–87.
- Gorenstein, E. E., Mammato, C. A., and Sandy, J. M. (1989). Performance of inattentive-overactive children on selected measures of prefrontal-type function. *J. Clin. Psychol.* **45**, 619–632.
- Goto, Y., and Grace, A. A. (2005a). Dopamine-dependent interactions between limbic and prefrontal cortical plasticity in the nucleus accumbens: Disruption by cocaine sensitization. *Neuron* **47**, 255–266.
- Goto, Y., and Grace, A. A. (2005b). Dopaminergic modulation of limbic and cortical drive of nucleus accumbens in goal-directed behavior. *Nat. Neurosci.* **8**, 805–812.
- Goto, Y., Tseng, K. Y., Lewis, B. L., and O'Donnell, P. (2004). Dopamine modulation of prefrontal cortical neural ensembles and synaptic plasticity: Potential involvement in schizophrenia. In “Prefrontal Cortex: From Synaptic Plasticity to Cognition” (S. Otani, Ed.), pp. 61–84. Kluwer Academic Press, New York.
- Grace, A. A. (1991). Phasic versus tonic dopamine release and the modulation of dopamine system responsivity: A hypothesis for the etiology of schizophrenia. *Neuroscience* **41**, 1–24.
- Grace, A. A., and Bunney, B. S. (1979). Paradoxical GABA excitation of nigral dopaminergic cells: Indirect mediation through reticulata inhibitory neurons. *Eur. J. Pharmacol.* **59**, 211–218.
- Grace, A. A., and Bunney, B. S. (1984a). The control of firing pattern in nigral dopamine neurons: Burst firing. *J. Neurosci.* **4**, 2877–2890.
- Grace, A. A., and Bunney, B. S. (1984b). The control of firing pattern in nigral dopamine neurons: Single spike firing. *J. Neurosci.* **4**, 2866–2876.
- Grace, A. A., and Bunney, B. S. (1985). Opposing effects of striatonigral feedback pathways on midbrain dopamine cell activity. *Brain Res.* **333**, 271–284.
- Grace, A. A., and Bunney, B. S. (1986). Induction of depolarization block in midbrain dopamine neurons by repeated administration of haloperidol: Analysis using *in vivo* intracellular recording. *J. Pharmacol. Exp. Ther.* **238**, 1092–1100.

- Grace, A. A., and Onn, S. P. (1989). Morphology and electrophysiological properties of immunocytochemically identified rat dopamine neurons recorded *in vitro*. *J. Neurosci.* **9**, 3463–3481.
- Grace, A. A., and Rosenkranz, J. A. (2002). Regulation of conditioned responses of basolateral amygdala neurons. *Physiol. Behav.* **77**, 489–493.
- Grace, A. A., Bunney, B. S., Moore, H., and Todd, C. L. (1997). Dopamine-cell depolarization block as a model for the therapeutic actions of antipsychotic drugs. *Trends Neurosci.* **20**, 31–37.
- Granon, S., Passetti, F., Thomas, K. L., Dalley, J. W., Everitt, B. J., and Robbins, T. W. (2000). Enhanced and impaired attentional performance after infusion of D1 dopaminergic receptor agents into rat prefrontal cortex. *J. Neurosci.* **20**, 1208–1215.
- Graybiel, A. M. (1997). The basal ganglia and cognitive pattern generators. *Schizophr. Bull.* **23**, 459–469.
- Graybiel, A. M. (1998). The basal ganglia and chunking of action repertoires. *Neurobiol. Learn. Mem.* **70**, 119–136.
- Greengard, P., Allen, P. B., and Nairn, A. C. (1999). Beyond the dopamine receptor: The DARPP-32/protein phosphatase-1 cascade. *Neuron* **23**, 435–447.
- Groenewegen, H. J., and Uylings, H. B. (2000). The prefrontal cortex and the integration of sensory, limbic and autonomic information. *Prog. Brain Res.* **126**, 3–28.
- Groenewegen, H. J., Vermeulen-Van der Zee, E., te Kortschot, A., and Witter, M. P. (1987). Organization of the projections from the subiculum to the ventral striatum in the rat. A study using anterograde transport of Phaseolus vulgaris leucoagglutinin. *Neuroscience* **23**, 103–120.
- Groenewegen, H. J., Wright, C. I., and Beijer, A. V. (1996). The nucleus accumbens: Gateway for limbic structures to reach the motor system? *Prog. Brain Res.* **107**, 485–511.
- Guarraci, F. A., and Kapp, B. S. (1999). An electrophysiological characterization of ventral tegmental area dopaminergic neurons during differential pavlovian fear conditioning in the awake rabbit. *Behav. Brain Res.* **99**, 169–179.
- Gulledge, A. T., and Jaffe, D. B. (1998). Dopamine decreases the excitability of layer V pyramidal cells in the rat prefrontal cortex. *J. Neurosci.* **18**, 9139–9151.
- Gurden, H., Takita, M., and Jay, T. M. (2000). Essential role of D1 but not D2 receptors in the NMDA receptor-dependent long-term potentiation at hippocampal-prefrontal cortex synapses *in vivo*. *J. Neurosci.* **20**, RC106.
- Gurden, H., Tassin, J.-P., and Jay, T. M. (1999). Integrity of the mesocortical dopaminergic system is necessary for complete expression of *in vivo* hippocampal-prefrontal cortex long-term potentiation. *Neuroscience* **94**, 1019–1027.
- Hamill, W. T., and Fontana, A. F. (1975). The immediate effects of chlorpromazine in newly admitted schizophrenic patients. *Am. J. Psychiatry* **132**, 1023–1026.
- Hara, Y., and Pickel, V. M. (2005). Overlapping intracellular and differential synaptic distributions of dopamine D1 and glutamate N-methyl-D-aspartate receptors in rat nucleus accumbens. *J. Comp. Neurol.* **492**, 442–455.
- Harrison, P. J. (1999). The neuropathology of schizophrenia. A critical review of the data and their interpretation. *Brain* **122**(Pt. 4), 593–624.
- Herry, C., and Garcia, R. (2002). Prefrontal cortex long-term potentiation, but not long-term depression, is associated with the maintenance of extinction of learned fear in mice. *J. Neurosci.* **22**, 577–583.
- Heyman, I., and Murray, R. M. (1992). Schizophrenia and neurodevelopment. *J. R. Coll. Physicians Lond.* **26**, 143–146.
- Hornykiewicz, O. (1971). Pharmacology and pathophysiology of dopaminergic neurons. *Adv. Cytopharmacol.* **1**, 369–377.
- Hyman, C., Hofer, M., Barde, Y. A., Juhasz, M., Yancopoulos, G. D., Squinto, S. P., and Lindsay, R. M. (1991). BDNF is a neurotrophic factor for dopaminergic neurons of the substantia nigra. *Nature* **350**, 230–232.

- Ingvar, D. H. (1985). "Memory of the future": An essay on the temporal organization of conscious awareness. *Hum. Neurobiol.* **4**, 127–136.
- Iversen, S. D. (1984). Behavioural effects of manipulation of basal ganglia neurotransmitters. *Ciba Found. Symp.* **107**, 183–200.
- Jackson, D. M., and Westlind-Danielsson, A. (1994). Dopamine receptors: Molecular biology, biochemistry and behavioural aspects. *Pharmacol. Ther.* **64**, 291–370.
- Jackson, M. E., Frost, A. S., and Moghaddam, B. (2001). Stimulation of prefrontal cortex at physiologically relevant frequencies inhibits dopamine release in the nucleus accumbens. *J. Neurochem.* **78**, 920–923.
- Jackson-Lewis, V., Vila, M., Djaldetti, R., Guegan, C., Liberatore, G., Liu, J., O'Malley, K. L., Burke, R. E., and Przedborski, S. (2000). Developmental cell death in dopaminergic neurons of the substantia nigra of mice. *J. Comp. Neurol.* **424**, 476–488.
- Jay, T. M., Glowinski, J., and Thierry, A. M. (1989). Selectivity of the hippocampal projection to the prelimbic area of the prefrontal cortex in the rat. *Brain Res.* **505**, 337–340.
- Jellestad, F. K., Markowska, A., Bakke, H. K., and Walther, B. (1986). Behavioral effects after ibotenic acid, 6-OHDA and electrolytic lesions in the central amygdala nucleus of the rat. *Physiol. Behav.* **37**, 855–862.
- Jentsch, J. D., and Roth, R. H. (1999). The neuropsychopharmacology of phencyclidine: From NMDA receptor hypofunction to the dopamine hypothesis of schizophrenia. *Neuropsychopharmacology* **20**, 201–225.
- Jog, M. S., Kubota, Y., Connolly, C. I., Hillegaart, V., and Graybiel, A. M. (1999). Building neural representations of habits. *Science* **286**, 1745–1749.
- Johnstone, E. C., Crow, T. J., Frith, C. D., Carney, M. W., and Price, J. S. (1978). Mechanism of the antipsychotic effect in the treatment of acute schizophrenia. *Lancet* **1**, 848–851.
- Karoum, F., Chrapusta, S. J., and Egan, M. F. (1994). 3-Methoxytyramine is the major metabolite of released dopamine in the rat frontal cortex: Reassessment of the effects of antipsychotics on the dynamics of dopamine release and metabolism in the frontal cortex, nucleus accumbens, and striatum by a simple two pool model. *J. Neurochem.* **63**, 972–979.
- Kelley, A. E., and Berridge, K. C. (2002). The neuroscience of natural rewards: Relevance to addictive drugs. *J. Neurosci.* **22**, 3306–3311.
- Kelley, A. E., and Domesick, V. B. (1982). The distribution of the projection from the hippocampal formation to the nucleus accumbens in the rat: An anterograde- and retrograde-horseradish peroxidase study. *Neuroscience* **7**, 2321–2335.
- Kimberg, D. Y., D'Esposito, M., and Farah, M. J. (1997). Effects of bromocriptine on human subjects depend on working memory capacity. *Neuroreport* **8**, 3581–3585.
- Knight, R. T., Grabowecky, M. F., and Scabini, D. (1995). Role of human prefrontal cortex in attention control. *Adv. Neurol.* **66**, 21–34; discussion 34–36.
- Kolachana, B. S., Saunders, R. C., and Weinberger, D. R. (1995). Augmentation of prefrontal cortical monoaminergic activity inhibits dopamine release in the caudate nucleus: An *in vivo* neurochemical assessment in the rhesus monkey. *Neuroscience* **69**, 859–868.
- Kuhar, M. J., Sanchez-Roa, P. M., Wong, D. F., Dannals, R. F., Grigoriadis, D. E., Lew, R., and Milberger, M. (1990). Dopamine transporter: Biochemistry, pharmacology, and imaging. *Eur. Neurol.* **30**(Suppl. 1), 15–20.
- Kupfermann, I. (1979). Modulatory actions of neurotransmitters. *Annu. Rev. Neurosci.* **2**, 447–465.
- Kyd, R. J., and Bilkey, D. K. (2003). Prefrontal cortex lesions modify the spatial properties of hippocampal place cells. *Cereb. Cortex* **13**, 444–451.
- Laroche, S., Jay, T. M., and Thierry, A. M. (1990). Long-term potentiation in the prefrontal cortex following stimulation of the hippocampal CA1/subicular region. *Neurosci. Lett.* **114**, 184–190.
- Laruelle, M., Abi-Dargham, A., Gil, R., Kegeles, L., and Innis, R. (1999). Increased dopamine transmission in schizophrenia: Relationship to illness phases. *Biol. Psychiatry* **46**, 56–72.

- Lauder, J. M., and Bloom, F. E. (1974). Ontogeny of monoamine neurons in the locus coeruleus, Raphe nuclei and substantia nigra of the rat. I. Cell differentiation. *J. Comp. Neurol.* **155**, 469–481.
- Lewis, D. A., Campbell, M. J., Foote, S. L., Goldstein, M., and Morrison, J. H. (1987). The distribution of tyrosine hydroxylase-immunoreactive fibers in primate neocortex is widespread but regionally specific. *J. Neurosci.* **7**, 279–290.
- Li, S., Cullen, W. K., Anwyl, R., and Rowan, M. J. (2003). Dopamine-dependent facilitation of LTP induction in hippocampal CA1 by exposure to spatial novelty. *Nat. Neurosci.* **6**, 526–531.
- Lidow, M. S., Williams, G. V., and Goldman-Rakic, P. S. (1998). The cerebral cortex: A case for a common site of action of antipsychotics. *Trends Pharmacol. Sci.* **19**, 136–140.
- Lisman, J. E., and Grace, A. A. (2005). The hippocampal-VTA loop: Controlling the entry of information into long-term memory. *Neuron* **46**, 703–713.
- Lloyd, K., and Hornykiewicz, O. (1970). Parkinson's disease: Activity of L-dopa decarboxylase in discrete brain regions. *Science* **170**, 1212–1213.
- Lodge, D. J., and Grace, A. A. (2006a). The laterodorsal tegmental nucleus is essential for burst firing of dopamine neurons in the rat VTA. *Proc. Natl. Acad. Sci. USA* **103**, 5167–5172.
- Lodge, D. J., and Grace, A. A. (2006b). The hippocampus modulates dopamine neuron responsivity by regulating the intensity of phasic neuron activation. *Neuropsychopharmacology* **31**, 1356–1361.
- Malenka, R. C. (2003). Synaptic plasticity and AMPA receptor trafficking. *Ann. NY Acad. Sci.* **1003**, 1–11.
- Mangiavacchi, S., and Wolf, M. E. (2004). D1 dopamine receptor stimulation increases the rate of AMPA receptor insertion onto the surface of cultured nucleus accumbens neurons through a pathway dependent on protein kinase A. *J. Neurochem.* **88**, 1261–1271.
- Manoach, D. S. (2003). Prefrontal cortex dysfunction during working memory performance in schizophrenia: Reconciling discrepant findings. *Schizophr. Res.* **60**, 285–298.
- Mantz, J., Thierry, A. M., and Glowinski, J. (1989). Effect of noxious tail pinch on the discharge rate of mesocortical and mesolimbic dopamine neurons: Selective activation of the mesocortical system. *Brain Res.* **476**, 377–381.
- Mayer, M. L., Westbrook, G. L., and Guthrie, P. B. (1984). Voltage-dependent block by Mg^{2+} of NMDA responses in spinal cord neurones. *Nature* **309**, 261–263.
- Mehta, M. A., Swanson, R., Ogilvie, A. D., Sahakian, J., and Robbins, T. W. (2001). Improved short-term spatial memory but impaired reversal learning following the dopamine D(2) agonist bromocriptine in human volunteers. *Psychopharmacology (Berl.)* **159**, 10–20.
- Melchitzky, D. S., and Lewis, D. A. (2000). Tyrosine hydroxylase- and dopamine transporter-immunoreactive axons in the primate cerebellum. Evidence for a lobular- and laminar-specific dopamine innervation. *Neuropsychopharmacology* **22**, 466–472.
- Meyer-Lindenberg, A., Miletich, R. S., Kohn, P. D., Esposito, G., Carson, R. E., Quarantelli, M., Weinberger, D. R., and Berman, K. F. (2002). Reduced prefrontal activity predicts exaggerated striatal dopaminergic function in schizophrenia. *Nat. Neurosci.* **5**, 267–271.
- Meyer-Lindenberg, A. S., Olsen, R. K., Kohn, P. D., Brown, T., Egan, M. F., Weinberger, D. R., and Berman, K. F. (2005). Regionally specific disturbance of dorsolateral prefrontal-hippocampal functional connectivity in schizophrenia. *Arch. Gen. Psychiatry* **62**, 379–386.
- Milner, B. (1963). Effects of different brain lesions on card sorting. *Arch. Neurol.* **9**, 90–100.
- Mogenson, G. J., Jones, D. L., and Yim, C. Y. (1980). From motivation to action: Functional interface between the limbic system and the motor system. *Prog. Neurobiol.* **14**, 69–97.
- Molinoff, P. B., and Axelrod, J. (1971). Biochemistry of catecholamines. *Annu. Rev. Biochem.* **40**, 465–500.
- Moore, H., Todd, C. L., and Grace, A. A. (1998). Striatal extracellular dopamine levels in rats with haloperidol-induced depolarization block of substantia nigra dopamine neurons. *J. Neurosci.* **18**, 5068–5077.

- Moore, R. Y., Whone, A. L., McGowan, S., and Brooks, D. J. (2003). Monoamine neuron innervation of the normal human brain: An 18F-DOPA PET study. *Brain Res.* **982**, 137–145.
- Moron, J. A., Brockington, A., Wise, R. A., Rocha, B. A., and Hope, B. T. (2002). Dopamine uptake through the norepinephrine transporter in brain regions with low levels of the dopamine transporter: Evidence from knock-out mouse lines. *J. Neurosci.* **22**, 389–395.
- Muir, J. L., Everitt, B. J., and Robbins, T. W. (1996). The cerebral cortex of the rat and visual attentional function: Dissociable effects of mediofrontal, cingulate, anterior dorsolateral, and parietal cortex lesions on a five-choice serial reaction time task. *Cereb. Cortex* **6**, 470–481.
- Nader, K., and LeDoux, J. E. (1999). Inhibition of the mesoamygdala dopaminergic pathway impairs the retrieval of conditioned fear associations. *Behav. Neurosci.* **113**, 891–901.
- Neve, K. A., Seamans, J. K., and Trantham-Davidson, H. (2004). Dopamine receptor signaling. *J. Recept. Signal Transduct. Res.* **24**, 165–205.
- Ngan, E. T., Lane, C. J., Ruth, T. J., and Liddle, P. F. (2002). Immediate and delayed effects of risperidone on cerebral metabolism in neuroleptic naive schizophrenic patients: Correlations with symptom change. *J. Neurol. Neurosurg. Psychiatry* **72**, 106–110.
- Nicola, S. M., Surmeier, J., and Malenka, R. C. (2000). Dopaminergic modulation of neuronal excitability in the striatum and nucleus accumbens. *Annu. Rev. Neurosci.* **23**, 185–215.
- Nieoullon, A. (2002). Dopamine and the regulation of cognition and attention. *Prog. Neurobiol.* **67**, 53–83.
- Nordstrom, A. L., Farde, L., Eriksson, L., and Halldin, C. (1995). No elevated D2 dopamine receptors in neuroleptic-naïve schizophrenic patients revealed by positron emission tomography and [¹¹C]N-methylspiperone. *Psychiatry Res.* **61**, 67–83.
- Nowak, L., Bregestovski, P., Ascher, P., Herbert, A., and Prochiantz, A. (1984). Magnesium gates glutamate-activated channels in mouse central neurones. *Nature* **307**, 462–465.
- Nowicky, M. C., and Roth, R. H. (1978). Dopaminergic neurons: Role of presynaptic receptors in the regulation of transmitter biosynthesis. *Prog. Neuropsychopharmacol.* **2**, 139–158.
- O'Donnell, P., and Grace, A. A. (1994). Tonic D2-mediated attenuation of cortical excitation in nucleus accumbens neurons recorded *in vitro*. *Brain Res.* **634**, 105–112.
- Okubo, Y., Suhara, T., Suzuki, K., Kobayashi, K., Inoue, O., Terasaki, O., Someya, Y., Sassa, T., Sudo, Y., Matsushima, E., Iyo, M., Tateno, Y., *et al.* (1997). Decreased prefrontal dopamine D1 receptors in schizophrenia revealed by PET. *Nature* **385**, 634–636.
- Oo, T. F., and Burke, R. E. (1997). The time course of developmental cell death in phenotypically defined dopaminergic neurons of the substantia nigra. *Brain Res. Dev. Brain Res.* **98**, 191–196.
- Otani, S., Blond, O., Desce, J. M., and Crepel, F. (1998). Dopamine facilitates long-term depression of glutamatergic transmission in rat prefrontal cortex. *Neuroscience* **85**, 669–676.
- Otani, S., Daniel, H., Roisin, M. P., and Crepel, F. (2003). Dopaminergic modulation of long-term synaptic plasticity in rat prefrontal neurons. *Cereb. Cortex* **13**, 1251–1256.
- Otmakhova, N. A., and Lisman, J. E. (1996). D1/D5 dopamine receptor activation increases the magnitude of early long-term potentiation at CA1 hippocampal synapses. *J. Neurosci.* **16**, 7478–7486.
- Overton, P., and Clark, D. (1992). Ionophoretically administered drugs acting at the N-methyl-D-aspartate receptor modulate burst firing in A9 dopamine neurons in the rat. *Synapse* **10**, 131–140.
- Overton, P. G., and Clark, D. (1997). Burst firing in midbrain dopaminergic neurons. *Brain Res. Rev.* **25**, 312–334.
- Owen, A. M., Downes, J. J., Sahakian, B. J., Polkey, C. E., and Robbins, T. W. (1990). Planning and spatial working memory following frontal lobe lesions in man. *Neuropsychologia* **28**, 1021–1034.
- Owen, A. M., Roberts, A. C., Hodges, J. R., Summers, B. A., Polkey, C. E., and Robbins, T. W. (1993). Contrasting mechanisms of impaired attentional set-shifting in patients with frontal lobe damage or Parkinson's disease. *Brain* **116**, 1159–1175.

- Paspalas, C. D., and Goldman-Rakic, P. S. (2004). Microdomains for dopamine volume neurotransmission in primate prefrontal cortex. *J. Neurosci.* **24**, 5292–5300.
- Paspalas, C. D., and Goldman-Rakic, P. S. (2005). Presynaptic D1 dopamine receptors in primate prefrontal cortex: Target-specific expression in the glutamatergic synapse. *J. Neurosci.* **25**, 1260–1267.
- Pennartz, C. M., Ameerun, R. F., Groenewegen, H. J., and Lopes da Silva, F. H. (1993). Synaptic plasticity in an *in vitro* slice preparation of the rat nucleus accumbens. *Eur. J. Neurosci.* **5**, 107–117.
- Poulsen, K. T., Armanini, M. P., Klein, R. D., Hynes, M. A., Phillips, H. S., and Rosenthal, A. (1994). TGF beta 2 and TGF beta 3 are potent survival factors for midbrain dopaminergic neurons. *Neuron* **13**, 1245–1252.
- Ragozzino, M. E. (2002). The effects of dopamine D(1) receptor blockade in the prelimbic-infralimbic areas on behavioral flexibility. *Learn. Mem.* **9**, 18–28.
- Rescorla, R. A., and Wagner, A. R. (1972). A theory of Pavlovian conditioning: Variations in the effectiveness of reinforcement and non reinforcement. In “Classical Conditioning II” (A. H. Black and W. F. Prokasy, Eds.), pp. 64–99. Appleton-Century-Croft, New York.
- Reynolds, J. N., Hyland, B. I., and Wickens, J. R. (2001). A cellular mechanism of reward-related learning. *Nature* **413**, 67–70.
- Ridley, R. M. (1994). The psychology of perseverative and stereotyped behaviour. *Prog. Neurobiol.* **44**, 221–231.
- Robbins, T. W. (2000). From arousal to cognition: The integrative position of the prefrontal cortex. *Prog. Brain Res.* **126**, 469–483.
- Robbins, T. W. (2005). Chemistry of the mind: Neurochemical modulation of prefrontal cortical function. *J. Comp. Neurol.* **493**, 140–146.
- Rodrigues, S. M., Schafe, G. E., and LeDoux, J. E. (2004). Molecular mechanisms underlying emotional learning and memory in the lateral amygdala. *Neuron* **44**, 75–91.
- Rosenkranz, J. A., and Grace, A. A. (2002). Dopamine-mediated modulation of odour-evoked amygdala potentials during Pavlovian conditioning. *Nature* **417**, 282–287.
- Saunders, R. C., Kolachana, B. S., Bachevalier, J., and Weinberger, D. R. (1998). Neonatal lesions of the medial temporal lobe disrupt prefrontal cortical regulation of striatal dopamine. *Nature* **393**, 169–171.
- Sawaguchi, T., and Goldman-Rakic, P. S. (1994). The role of D1-dopamine receptor in working memory: Local injections of dopamine antagonists into the prefrontal cortex of rhesus monkeys performing an oculomotor delayed-response task. *J. Neurophysiol.* **71**, 515–528.
- Scatton, B., Simon, H., Le Moal, M., and Bischoff, S. (1980). Origin of dopaminergic innervation of the rat hippocampal formation. *Neurosci. Lett.* **18**, 125–131.
- Schultz, W. (2002). Getting formal with dopamine and reward. *Neuron* **36**, 241–263.
- Schultz, W., Apicella, P., and Ljungberg, T. (1993). Responses of monkey dopamine neurons to reward and conditioned stimuli during successive steps of learning a delayed response task. *J. Neurosci.* **13**, 900–913.
- Schultz, W., Tremblay, L., and Hollerman, J. R. (2000). Reward processing in primate orbitofrontal cortex and basal ganglia. *Cereb. Cortex* **10**, 272–284.
- Schultz, W., Tremblay, L., and Hollerman, J. R. (2003). Changes in behavior-related neuronal activity in the striatum during learning. *Trends Neurosci.* **26**, 321–328.
- Seamans, J. K., and Yang, C. R. (2004). The principal features and mechanisms of dopamine modulation in the prefrontal cortex. *Prog. Neurobiol.* **74**, 1–58.
- Seamans, J. K., Floresco, S. B., and Phillips, A. G. (1998). D1 receptor modulation of hippocampal-prefrontal cortical circuits integrating spatial memory with executive functions in the rat. *J. Neurosci.* **18**, 1613–1621.

- Seamans, J. K., Durstewitz, D., Christie, B. R., Stevens, C. F., and Sejnowski, T. J. (2001a). Dopamine D1/D5 receptor modulation of excitatory synaptic inputs to layer V prefrontal cortex neurons. *Proc. Natl. Acad. Sci. USA* **98**, 301–306.
- Seamans, J. K., Gorelova, N., Durstewitz, D., and Yang, C. R. (2001b). Bidirectional dopamine modulation of GABAergic inhibition in prefrontal cortical pyramidal neurons. *J. Neurosci.* **21**, 3628–3638.
- Seeman, P. (1987). Dopamine receptors and the dopamine hypothesis of schizophrenia. *Synapse* **1**, 133–152.
- Seeman, P., and Van Tol, H. H. (1993). Dopamine receptor pharmacology. *Curr. Opin. Neurol. Neurosurg.* **6**, 602–608.
- Sesack, S. R., and Carr, D. B. (2002). Selective prefrontal cortex inputs to dopamine cells: Implications for schizophrenia. *Physiol. Behav.* **77**, 513–517.
- Sesack, S. R., Hawrylak, V. A., Guido, M. A., and Levey, A. I. (1998). Cellular and subcellular localization of the dopamine transporter in rat cortex. *Adv. Pharmacol.* **42**, 171–174.
- Shimamura, A. P. (2000). The role of the prefrontal cortex in dynamic filtering. *Psychobiology* **28**, 207–218.
- Smith, I. D., and Grace, A. A. (1992). Role of the subthalamic nucleus in the regulation of nigral dopamine neuron activity. *Synapse* **12**, 287–303.
- Snyder, S. H. (1972). Catecholamines in the brain as mediators of amphetamine psychosis. *Arch. Gen. Psychiatry* **27**, 169–179.
- Sonuga-Barke, E. J. (2005). Causal models of attention-deficit/hyperactivity disorder: From common simple deficits to multiple developmental pathways. *Biol. Psychiatry* **57**, 1231–1238.
- Spencer, J. P., and Murphy, K. P. (2000). Bi-directional changes in synaptic plasticity induced at corticostriatal synapses *in vitro*. *Exp. Brain Res.* **135**, 497–503.
- Starke, K., Reimann, W., Zumstein, A., and Hertting, G. (1978). Effect of dopamine receptor agonists and antagonists on the release of dopamine in the rabbit caudate nucleus *in vitro*. *Naunyn Schmiedeberg's Arch. Pharmacol.* **305**, 27–36.
- Sun, X., Zhao, Y., and Wolf, M. E. (2005). Dopamine receptor stimulation modulates AMPA receptor synaptic insertion in prefrontal cortex neurons. *J. Neurosci.* **25**, 7342–7351.
- Surmeier, D. J., Bargas, J., Hemmings, H. C. J., Nairn, A. C., and Greengard, P. (1995). Modulation of calcium currents by a D1 dopaminergic protein kinase/phosphatase cascade in rat neostriatal neurons. *Neuron* **14**, 385–397.
- Teicher, M. H., Andersen, S. L., and Hostetter, J. C., Jr. (1995). Evidence for dopamine receptor pruning between adolescent and adulthood in striatum but not nucleus accumbens. *Dev. Brain Res.* **89**, 167–172.
- Thierry, A. M., Blanc, G., Sobel, A., Stinus, L., and Golwinski, J. (1973). Dopaminergic terminals in the rat cortex. *Science* **182**, 499–501.
- Thomas, M. J., Beurrier, C., Bonci, A., and Malenka, R. C. (2001). Long-term depression in the nucleus accumbens: A neural correlate of behavioral sensitization to cocaine. *Nat. Neurosci.* **4**, 1217–1223.
- Tirelli, E., Laviola, G., and Adriani, W. (2003). Ontogenesis of behavioral sensitization and conditioned place preference induced by psychostimulants in laboratory rodents. *Neurosci. Biobehav. Rev.* **27**, 163–178.
- Tobler, P. N., Dickinson, A., and Schultz, W. (2003). Coding of predicted reward omission by dopamine neurons in a conditioned inhibition paradigm. *J. Neurosci.* **23**, 10402–10410.
- Tseng, K. Y., and O'Donnell, P. (2004). Dopamine-glutamate interactions controlling prefrontal cortical pyramidal cell excitability involve multiple signaling mechanisms. *J. Neurosci.* **24**, 5131–5139.
- Ujike, H., Tsuchida, K., Akiyama, K., Fujiwara, Y., and Kuroda, S. (1995). Ontogeny of behavioral sensitization to cocaine. *Pharmacol. Biochem. Behav.* **50**, 613–617.

- Umemiya, M., and Raymond, L. A. (1997). Dopamine modulation of excitatory postsynaptic currents in rat neostriatal neurons. *J. Neurophysiol.* **78**, 1248–1255.
- Ungless, M. A., Magill, P. J., and Bolam, J. P. (2004). Uniform inhibition of dopamine neurons in the ventral tegmental area by aversive stimuli. *Science* **303**, 2040–2042.
- Voorn, P., Jorritsma-Byham, B., Van Dijk, C., and Buijs, R. M. (1986). The dopaminergic innervation of the ventral striatum in the rat: A light- and electron-microscopical study with antibodies against dopamine. *J. Comp. Neurol.* **251**, 84–99.
- Waelti, P., Dickinson, A., and Schultz, W. (2001). Dopamine responses comply with basic assumptions of formal learning theory. *Nature* **412**, 43–48.
- Wang, H., and Pickel, V. M. (2002). Dopamine D2 receptors are present in prefrontal cortical afferents and their targets in patches of the rat caudate-putamen nucleus. *J. Comp. Neurol.* **442**, 392–404.
- Wang, M., Vijayraghavan, S., and Goldman-Rakic, P. S. (2004). Selective D2 receptor actions on the functional circuitry of working memory. *Science* **303**, 853–856.
- Wang, Y., and Goldman-Rakic, P. S. (2004). D2 receptor regulation of synaptic burst firing in prefrontal cortical pyramidal neurons. *Proc. Natl. Acad. Sci. USA* **101**, 5093–5098.
- Weinberger, D. R. (1987). Implications of normal brain development for the pathogenesis of schizophrenia. *Arch. Gen. Psychiatry* **44**, 660–669.
- Weinberger, D. R., Aloia, M. S., Goldberg, T. E., and Berman, K. F. (1994). The frontal lobes and schizophrenia. *J. Neuropsychiatry Clin. Neurosci.* **6**, 419–427.
- Weinberger, D. R., Egan, M. F., Bertolino, A., Callicott, J. H., Mattay, V. S., Lipska, B. K., Berman, K. F., and Goldberg, T. E. (2001). Prefrontal neurons and the genetics of schizophrenia. *Biol. Psychiatry* **50**, 825–844.
- West, A. R., and Grace, A. A. (2002). Opposite influences of endogenous dopamine D1 and D2 receptor activation on activity states and electrophysiological properties of striatal neurons: Studies combining *in vivo* intracellular recordings and reverse microdialysis. *J. Neurosci.* **22**, 294–304.
- White, F. J. (1996). Synaptic regulation of mesocorticolimbic dopamine neurons. *Annu. Rev. Neurosci.* **19**, 405–436.
- Wolf, M. E., and Roth, R. H. (1990). Autoreceptor regulation of dopamine synthesis. *Ann. NY Acad. Sci.* **604**, 323–343.
- Wolf, M. E., Sun, X., Mangiavacchi, S., and Chao, S. Z. (2004). Psychomotor stimulants and neuronal plasticity. *Neuropharmacology* **47**(Suppl. 1), 61–79.
- Yerkes, R. M., and Dodson, J. D. (1908). The relation of strength of stimulus to rapidity of habit-formation. *J. Comp. Neurol. Psychol.* **18**, 459–482.
- Zahrt, J., Taylor, J. R., Mathew, R. G., and Arnsten, A. F. (1997). Supranormal stimulation of D1 dopamine receptors in the rodent prefrontal cortex impairs spatial working memory performance. *J. Neurosci.* **17**, 8528–8535.
- Zironi, I., Iacovelli, P., Aicardi, G., Liu, P., and Bilkey, D. K. (2001). Prefrontal cortex lesions augment the location-related firing properties of area TE/perirhinal cortex neurons in a working memory task. *Cereb. Cortex* **11**, 1093–1100.

GLUTAMATE AND SCHIZOPHRENIA: PHENCYCLIDINE, N-METHYL-D-ASPARTATE RECEPTORS, AND DOPAMINE–GLUTAMATE INTERACTIONS

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Schizophrenia is a serious mental disorder that affects up to 1% of the population worldwide. As of yet, neurochemical mechanisms underlying schizophrenia remain unknown. To date, the most widely considered neurochemical hypothesis of schizophrenia is the dopamine hypothesis, which postulates that symptoms of schizophrenia may result from excess dopaminergic neurotransmission particularly in striatal brain regions, along with dopaminergic deficits in prefrontal brain regions. Alternative neurochemical models of schizophrenia, however, have been proposed

involving glutamatergic mechanisms in general and *N*-methyl-D-aspartate (NMDA) receptors in particular. A potential role for glutamatergic mechanisms in schizophrenia was first proposed ~15 years ago based on the observation that the psychotomimetic agents phencyclidine (PCP) and ketamine induce psychotic symptoms and neurocognitive disturbances similar to those of schizophrenia by blocking neurotransmission at NMDA-type glutamate receptors. Since that time, significant additional evidence has accumulated supporting a role for NMDA hypofunction in the pathophysiology of schizophrenia. Clinical challenge studies with PCP and ketamine have confirmed the close resemblance between NMDA antagonist-induced symptoms and neurocognitive deficits and those observed in schizophrenia, and suggest that NMDA dysfunction may lead to secondary dopaminergic dysregulation in striatal and prefrontal brain regions. As compared to dopaminergic agents, NMDA antagonists induce negative and cognitive symptoms of schizophrenia, as well as positive symptoms. Treatment studies with NMDA modulators, such as glycine, D-serine, and glycine transport inhibitors (GTIs), have yielded encouraging findings, although results remain controversial. Finally, genetic linkage and *in vivo* neurochemical studies in schizophrenia highlight potential etiological mechanisms giving rise to glutamatergic/NMDA dysfunction in schizophrenia.

I. Introduction

Schizophrenia is a serious mental disorder that affects up to 1% of the population worldwide, and is one of the leading causes of chronic disability. The first effective treatments for schizophrenia were discovered fortuitously in the late 1950s, and subsequently shown to mediate their effects at dopamine D2 receptors. Since that time, dopamine has been the primary neurotransmitter implicated in schizophrenia, and the majority of neurochemical studies of schizophrenia continue to focus on dopaminergic mechanisms (Carlsson, 1988).

Neurochemical models of schizophrenia based on dopaminergic theories have had substantial heuristic value in explaining key symptoms of schizophrenia, in particular, positive symptoms, and in guiding treatment considerations. For example, all antipsychotics are effective at doses that occupy ~80% of brain D2 receptors (Kapur and Remington, 2001). Further, individuals with schizophrenia do show enhanced striatal dopamine release to amphetamine challenge at least during the acute stage of illness (Laruelle, 1998). Nevertheless, significant limitations with regard to the dopamine hypothesis remain. First, no intrinsic deficits have been observed within the dopamine system to account for the presumed hyperdopaminergia associated with schizophrenia. Second, reconceptualizations

of the dopamine hypothesis propose that subcortical hyperdopaminergia may coexist with cortical hypodopaminergia (Davis *et al.*, 1991), although mechanisms underlying the differential cortical and subcortical abnormalities remain to be determined. Finally, dopaminergic dysfunction, in general, accounts poorly for symptom classes in schizophrenia other than positive symptoms, and for the pattern of neurocognitive dysfunction associated with schizophrenia. Thus, alternative conceptual models of schizophrenia are required.

An alternative to the dopamine model was first proposed in the early 1990s, based on the observation that phencyclidine (PCP) and similarly acting psychotomimetic compounds induced their unique behavioral effects by blocking neurotransmission at *N*-methyl-D-aspartate (NMDA)-type glutamate receptors (Javitt, 1987; Javitt and Zukin, 1991). The ability of these compounds to transiently reproduce key symptoms of schizophrenia by blocking NMDA receptors led to the concept that symptoms in schizophrenia may reflect underlying dysfunction or dysregulation of NMDA receptor-mediated neurotransmission.

Over the past 15 years, convergent evidence has accumulated to support a primary role for glutamatergic dysfunction in the pathophysiology of schizophrenia (Abi-Saab *et al.*, 1998; Coyle, 1996; Olney *et al.*, 1999; Tamminga *et al.*, 1995). In particular, studies have documented a close congruence between symptomatic and neurocognitive effects induced by NMDA antagonists such as PCP and the related drug ketamine, and the pattern observed in schizophrenia. Further, both genetic and neurochemical studies have begun to identify pathogenetic events that may impact on glutamatergic neurotransmission, and provide plausible bases for underlying NMDA dysfunction. Finally, evidence from both animal and human studies suggest that the hyperdopaminergia associated with schizophrenia may, in fact, result from underlying dysfunction of NMDA-related neuromodulatory feedback mechanisms. Overall, these findings suggest new etiological and psychotherapeutic conceptualizations of schizophrenia.

II. Glutamatergic Physiology

Glutamate is the primary excitatory neurotransmitter in brain, accounting for roughly 60% of neurons and 40% of synapses. Virtually all cortical pyramidal neurons use glutamate as their primary excitatory neurotransmitter. Glutamate is synthesized in brain from glutamine, which is transported across the blood-brain barrier with high affinity and present at high concentration in extracellular brain fluid and cerebrospinal fluid. Following release, glutamate is reabsorbed by both neuronal and glial glutamate transporters via an energy-dependent transport process. Much of the brain energy demand relates to glutamate homeostasis.

A. GLUTAMATE–DOPAMINE COMPARISONS

To date, dopaminergic models of schizophrenia have emphasized primary dysfunction of circumscribed brain pathways, such as the mesocortical and mesolimbic dopamine systems, as being the primary pathophysiological events in schizophrenia, with other types of deficits deriving secondarily from such disturbances. Given the widespread distribution of glutamate in the brain, however, glutamatergic models start from a different conceptual framework.

In glutamatergic models, it is presumed that similar functional disturbances are present throughout cortex and that such deficits may even involve subcortical glutamatergic pathways. Thus, glutamatergic models differ from dopaminergic models in that they predict widespread, rather than circumscribed, patterns of cortical dysfunction in schizophrenia. Nevertheless, not all glutamate receptors appear to be involved equally. In particular, clinical features of schizophrenia are most consistent with circumscribed dysfunction NMDA receptors, and new treatment approaches for schizophrenia are increasingly targeting these receptors. NMDA receptors, however, work in association with other glutamatergic and nonglutamatergic receptors. Such interactions may play a critical role as well in both etiopathology and treatment of schizophrenia.

B. GLUTAMATE RECEPTORS

Receptors for glutamate are divided into two broad families. Ionotropic receptors are differentiated based on sensitivity to the synthetic glutamate derivatives NMDA, AMPA, and kainate. Metabotropic receptors, which are G-protein coupled and mediate longer-term neuromodulatory effects of glutamate, are divided into groups on the basis of effector coupling and ligand sensitivity. Despite the differential sensitivity of these receptors to specific synthetic ligands, the endogenous neurotransmitter for all receptors is glutamate, and, to a lesser extent, the closely related amino acid aspartate.

C. NMDA RECEPTORS

NMDA receptors (Fig. 1) are the most complex of the ionotropic receptors. In addition to the recognition site for glutamate, NMDA receptors contain an allosteric modulatory site that binds the endogenous brain amino acids glycine and D-serine. This glycine-binding site, like the benzodiazepine site of the GABA_A receptor, regulates channel open time and desensitization rate in the presence of agonist (glutamate), but does not, of itself, induce channel opening. Like the benzodiazepine site, therefore, this may be an ideal target for drug development.

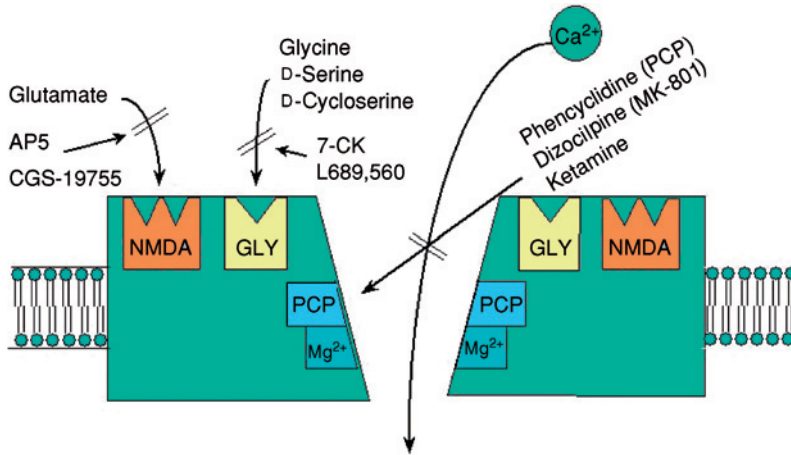


FIG. 1. Schematic model of the NMDA receptor complex.

Both glycine and D-serine are present in high concentration in brain. However, NMDA receptors appear to be protected from circulating glycine/D-serine level by the presence of amino acid transporters that are colocalized with NMDA receptors. Glycine type I (GLYT1) transporters may play a key role, although other small neutral amino acid transporters (SNATs) may also contribute (Javitt *et al.*, 2005a). As with the glycine site itself, these transporters have become a prominent target for drug development.

In addition to the glycine modulatory site, the NMDA receptor complex contains regulatory sites that are sensitive to polyamines, Zn^{2+} , protons, and redox agents such as glutathione. The multiple influences that converge on NMDA receptors speak to the critical role played by these receptors in a multitude of brain processes and suggest additional potential sites for therapeutic intervention.

NMDA receptors are blocked in a voltage sensitive fashion by Mg^{2+} , which binds to a site within the NMDA ion channel. As a result, NMDA receptors are uniquely voltage- as well as ligand (glutamate)-sensitive. This property permits NMDA receptors to play a unique role in the regulation of connection strength between neurons through a process known as long-term potentiation (LTP), and to act in a “Hebbian” fashion to integrate input from multiple independent pathways. In addition, because NMDA receptors can be turned “on” or “off” simply by varying the membrane voltage, they serve as a key elements in circuits related to attention, gating, and feedback regulation.

NMDA receptors are composed of multiple subunits, including at least one NR1 subunit and one or more modulatory subunits from the NR2

(NR2A–NR2D) and/or NR3 (NR3A, NR3B) families. These subunits significantly alter the functional properties of native NMDA receptors, including their voltage sensitivity, peak conductance, and degree to which they are influenced by the endogenous modulators glycine and D-serine. Interestingly, while the modulatory agents glycine and D-serine have similar, excitatory effects on NMDA receptors containing NR2 subunits, they have opposite effects on receptors containing NR3 subunits, with glycine serving to activate NR3-containing receptors and D-serine to inhibit them (Chatterton *et al.*, 2002).

NMDA receptors are blocked in a noncompetitive fashion by PCP, ketamine, and other agents such as dizocilpine (MK-801), which bind to a site (PCP receptor) located within the ion channel formed by the NMDA complex. The ability of NMDA antagonists to induce schizophrenia-like psychotic symptoms is among the strongest evidence to date linking glutamatergic dysfunction to the pathophysiology of schizophrenia.

D. AMPA/KAINATE RECEPTORS

AMPA/kainate receptors are a second class of ionotropic receptors for the neurotransmitter glutamate. AMPA receptors are composed of combinations of GluR1–4 subunits, while kainate receptors are composed of GluR5–7 and KA1 and KA2 subunits. Both receptor types interact closely with NMDA receptors, although at present the role of AMPA receptors is better understood, especially with regard to LTP. LTP is a fundamental process in the brain by which the strength of connections between neurons is modulated. It is thus the basis for much of learning, memory, and synaptic plasticity. Modulation of connection strength between neurons has long been known to be initiated by Ca^{2+} flux through open, unblocked NMDA channels. More recently, the interplay among glutamate receptors that permits such alterations in connections strength has also been evaluated.

Mature AMPA receptors containing the GluR2 subunit are Ca^{2+} impermeant (Tanaka *et al.*, 2000), and thus do not directly trigger LTP. Nevertheless, AMPA receptors provide the primary depolarization necessary to unblock NMDA receptors and to permit calcium entry into the cell. Ca^{2+} entry through unblocked NMDA receptors, in turn, triggers AMPA insertion into the postsynaptic density and synaptic strengthening. Thus, activity at AMPA and NMDA receptors is needed for coordinated glutamatergic neurotransmission.

The inverse of the synergistic relationship is that dysfunction of either AMPA or NMDA receptors may lead to the phenomenon of the silent synapse. AMPA receptors are continuously recycled, leading to gradual synaptic weakening. If AMPA density falls below a critical threshold, levels of depolarization are insufficient to unblock NMDA channels, preventing postsynaptic depolarization

or Ca^{2+} influx. The lack of Ca^{2+} influx precludes subsequent AMPA receptor insertion into the postsynaptic membrane. Thus, such synapses, despite containing histologically identifiable NMDA receptors, are functionally silent and cannot be recovered by electrical stimulation alone (Isaac *et al.*, 1999). To the extent that it occurs in schizophrenia, the silencing of synapses may limit the degree of recovery to be expected even if normal glutamatergic functioning could be restored.

E. METABOTROPIC RECEPTORS

As opposed to ionotropic receptors, which are linked directly to ion channels, metabotropic receptors are linked to second messenger systems and affect neuronal metabolism. A particular role of glutamatergic metabotropic receptors is regulation of presynaptic glutamate release and postsynaptic sensitivity. Metabotropic receptors are divided into three groups based on functional activity. Group I (types 1 and 4) receptors function predominantly to potentiate both presynaptic glutamate release and postsynaptic NMDA neurotransmission. In contrast, Group II (types 2 and 3) and Group III (types 5–7) receptors serve to limit glutamate release, particularly during conditions of glutamate spillover from the synaptic cleft. Thus, Group I agonists would be expected to stimulate neurotransmission mediated by ionotropic glutamate receptors, whereas agonists for Group II/III receptors would be expected to have opposite effects.

III. Glutamatergic Models of Schizophrenia

The strongest evidence linking glutamate in general and NMDA receptors in particular to the pathophysiology comes from studies of PCP and other “dissociative anesthetics” such as ketamine. Although the overall similarity between NMDA antagonist-induced psychosis has been appreciated since the early 1960s, studies continue to refine the relationships between the two clinical states.

Symptoms of schizophrenia are currently divided into at least three independent factors, labeled positive, negative, and cognitive or disorganized symptoms, respectively on the basis of rating scales such as the Psychiatric Rating Scale (BPRS) (Overall and Gorham, 1961) or the Positive and Negative Symptom Scale (PANSS) (Kay *et al.*, 1987). The positive factor consists of items such as hallucinations, paranoia, and agitation, while the negative factor consists of items such as apathy, motor retardation, and emotional withdrawal. The “cognitive” or “disorganized” factor, while more variable across studies, tends to include items such as conceptual disorganization, difficulties in abstract thinking, mannerisms,

and poor attention. Although the absolute level of symptoms varies across subjects, factor scores tend to be correlated both cross-sectionally and longitudinally. Thus, all patients with schizophrenia show at least some degree of negative and cognitive symptoms, along with positive.

In addition to symptoms, patients with schizophrenia show a pattern of neurocognitive dysfunction that represents a core feature of the disorder. Neurocognitive deficits are present at first episode and cannot be attributed to effects of medication. At present, it appears that patients who subsequently develop schizophrenia may have some level of neurocognitive dysfunction even during childhood but that further neurocognitive decline occurs in the years immediately preceding psychotic decompensation (Lieberman *et al.*, 2001). As with symptoms, the underlying neurochemical basis of neurocognitive dysfunction in schizophrenia has not been determined.

Finally, over recent years, specific neurochemical features of schizophrenia, including subcortical hyperdopaminergia and white matter degeneration, have been defined based on *in vivo* imaging and postmortem analysis. As the schizophrenia phenotype has been increasingly well characterized, it has been possible to assess with increasing precision the degree to which NMDA dysfunction may account for specific symptoms and features of schizophrenia.

A. SYMPTOM PATTERNS FOLLOWING NMDA ANTAGONIST ADMINISTRATION

In initial studies with PCP and ketamine in the early 1960s, it was noted that both agents produced what would now be considered positive, negative, and cognitive symptoms of schizophrenia (Javitt and Zukin, 1991; Luby *et al.*, 1962). At the time, however, no formal rating scales were used. In subsequent studies using such rating scales, however, significant increases are observed not only in positive symptoms but also in negative symptoms and disorganization (Krystal *et al.*, 1994; Lahti *et al.*, 2001; Malhotra *et al.*, 1996; Newcomer *et al.*, 1999). Levels of symptoms during acute ketamine challenge tend to show a similar pattern across factors as they do in schizophrenia. When patients with schizophrenia are exposed to ketamine, they also show increases in positive symptoms, as well as negative symptoms (Lahti *et al.*, 2001; Malhotra *et al.*, 1996). Ketamine induced symptoms respond poorly to conventional antipsychotics, but may be reversed particularly by clozapine treatment (Malhotra *et al.*, 1997a).

Despite the overall similarity between ketamine-induced symptoms and those of schizophrenia, there are some potentially informative differences. In particular, patients with schizophrenia often report hearing voices, while during ketamine administration such symptoms are rare. In addition, visual perceptual distortions are common during ketamine infusion but rare in established cases of schizophrenia. Nevertheless, the pattern of auditory and visual disturbances seen during

ketamine administration does resemble the pattern observed early in the course in schizophrenia (McGhie and Chapman, 1961) where both auditory and visual perceptual disturbances are common, and auditory hallucinations have not yet crystallized to the point of being identifiable as speech. Thus, acute ketamine challenge may be viewed as a model of acute incipient schizophrenia, rather than later, more chronic, phases. In patients with established schizophrenia, increases in hallucinatory activity are observed during ketamine challenge (Lahti *et al.*, 2001; Malhotra *et al.*, 1997b). Further, in primates, hallucinatory behavior is not observed during acute PCP treatment but does emerge during chronic administration (Linn *et al.*, 1999). In humans, for obvious ethical reasons, effects of chronic ketamine or PCP treatment are not well characterized.

Although symptomatic effects of ketamine and amphetamine have been extensively investigated in separate investigations, relatively few studies have tested both sets of compounds within the same volunteer subjects. A study, however, confirms results of prior independent investigations (Krystal *et al.*, 2005). In that study, effects of ketamine and amphetamine were assessed both individually and in combination. Consistent with prior studies, ketamine induced both positive and negative symptoms in approximately equal proportions—that is, similar to the type of levels seen in schizophrenia—whereas amphetamine induced only positive symptoms with no significant effect on negative symptoms.

The specific pattern of symptoms induced by amphetamine and ketamine differed as well. Thus, amphetamine induced significant increases in grandiosity, but not delusions, whereas ketamine induced significant increases in delusions but not grandiosity. Ketamine also produced greater degrees of hallucinatory behavior than did amphetamine. Additive effects between amphetamine and ketamine were seen only in the case of hallucinations, suggesting that the circuitry underlying hallucinations may have unique sensitivity to both glutamatergic and dopaminergic dysfunctions.

Similarly, although both amphetamine and ketamine produced increases in the cognitive factor score, the pattern of symptoms induced by the two compounds differed significantly. While amphetamine increased scoring primarily on one element of the cognitive symptom factor, conceptual disorganization, it had no effect on other cognitive symptoms. In contrast, ketamine significantly increased schizophrenia-like deficits not only in conceptual disorganization but also in difficulties in abstract thinking, mannerisms, and poor attention. Although some additivity between amphetamine and ketamine effects was observed, the degree of interaction did not reach statistical significance.

Overall, these findings continue to support the close similarity between NMDA antagonist-induced symptoms and those observed in schizophrenia. In contrast, patterns of symptoms induced by dopaminergic agonists, such as amphetamine, differ markedly from those seen in schizophrenia. Although there is a tendency to attribute positive symptoms in schizophrenia to dopaminergic hyperactivity and

negative symptoms to disturbances in other brain pathways, ketamine challenge studies to date suggest that NMDA dysfunction, by itself, would be sufficient to account for pathogenesis of all three symptom dimensions (positive, negative, cognitive) most associated with schizophrenia.

B. COGNITIVE DEFICITS FOLLOWING NMDA ANTAGONIST TREATMENT

In addition to symptoms, schizophrenia is associated with a pattern of neurocognitive dysfunction that is highly characteristic of the disorder (Bilder *et al.*, 1991; Gold *et al.*, 1999). Based in part on the influence of dopaminergic models of schizophrenia, a great number of cognitive studies in schizophrenia have focused on dysfunction of specific brain regions such as prefrontal dysfunction. However, neurocognitive deficits in schizophrenia are in no ways limited to prefrontal function. In studies that have utilized comprehensive neuropsychological batteries, similar levels of deficit have been observed across widespread neurocognitive domains, with preferential deficits, if any, observed in learning and declarative memory formation (Bilder *et al.*, 2000; Dickinson *et al.*, 2006; Keefe *et al.*, 2006; Saykin *et al.*, 1991).

A challenge in the neuropsychological literature has been to define a single brain abnormality that could account for the complex array of neuropsychological deficits seen in schizophrenia. Thus, while patients have many deficits attributable to prefrontal dysfunction in some tests (MacDonald *et al.*, 2005), on other prefrontal tests the pattern of deficit is incompatible with local structural disturbance (Shurman *et al.*, 2005). Further, patients show deficits even in simple visual (Butler *et al.*, 2005) and auditory (Strous *et al.*, 1995; Wexler *et al.*, 1998) sensory processing, with pattern of deficit indicating dysfunction within primary sensory regions (Rabinowicz *et al.*, 2000). Thus, no single structural deficit within cortex can give rise to the complex pattern of neurocognitive dysfunction seen in schizophrenia.

Glutamatergic models provide a potential alternative framework from which to view the pattern of neuropsychological dysfunction associated with schizophrenia. Although glutamatergic systems are widespread, within each brain region NMDA receptors participate in only a subset of processes. For example, in hippocampus and cortex, NMDA receptor activation is required for the initiation, but not maintenance of LTP (Miyamoto, 2006). The observation that patients with schizophrenia (as opposed to those with the amnesic syndrome) show deficits in memory formation (Hartvig *et al.*, 1995; Krystal *et al.*, 2005; Morgan *et al.*, 2004a; Newcomer *et al.*, 1999; Parwani *et al.*, 2005; Radant *et al.*, 1998; Rowland *et al.*, 2005), but not retention, is thus consistent with an NMDA pattern of dysfunction within hippocampal regions, rather than structural damage to the hippocampus itself.

To date, a substantial literature has accumulated comparing effects of NMDA antagonists to those observed in schizophrenia, using paradigms sensitive to both sensory and cognitive aspects of information processing dysfunction in schizophrenia. Deficits have been observed across widespread neuropsychological domains, including working memory (Honey *et al.*, 2003; Morgan *et al.*, 2004a), response inhibition (Morgan *et al.*, 2004b), procedural memory (Morgan *et al.*, 2004a), and executive processing (Krystal *et al.*, 1994; Umbricht *et al.*, 2000). Further, deficits are observed in sensory processing as well. For example, ketamine administration inhibits generation of mismatch negativity (MMN), an event-related potential (ERP) component that reflects impaired information processing at the level of auditory cortex (Umbricht *et al.*, 2000), and alters proprioceptive performance (Oye *et al.*, 1992). Similarly, administration of NMDA antagonists in rodents produces a pattern of visual ERP deficit similar to that observed in schizophrenia (Butler *et al.*, 2005). Ketamine infusion also reproduces both the severity and type of thought disorder seen in schizophrenia with both, for example, being associated with high levels of poverty of speech, circumstantiality and loss of goal, and relatively low levels of distractive or stilted speech or paraphasias (Adler *et al.*, 1999). Thus, reduction in NMDA functioning within brain could serve as a single unifying feature to account for the otherwise complex pattern of deficit observed in the disorder.

As opposed to ketamine, administration of dopaminergic agonists such as amphetamine does not reproduce the pattern of deficit observed in schizophrenia. Further, several recent studies have assessed the ability of amphetamine to improve neurocognitive performance in schizophrenia, on tasks such as the Stroop test. In this test, patients showed a characteristic pattern of deficit characterized by increased facilitation of response by stimulus congruence. Although amphetamine improved overall performance in this task in both normal and schizophrenia subjects, it nonetheless failed to reverse the specific pattern of neurocognitive dysfunction associated with schizophrenia (Barch and Carter, 2005). On the basis of these findings, dysfunction of dopaminergic systems appears neither necessary nor sufficient to account for the overall pattern of neuropsychological disturbance in schizophrenia, although interactions between dopaminergic and glutamatergic systems may occur.

C. *In Vivo* FINDINGS IN SCHIZOPHRENIA BASED ON DOPAMINE RECEPTOR OCCUPANCY

Along with neurocognitive studies, which provide insights into patterns of cortical dysfunction in schizophrenia, positron emission (PET) and single photon emission (SPECT) *in vivo* tomographic studies provide insights into patterns of dopaminergic dysfunction in schizophrenia. In such studies, D2 agonists are tagged

with appropriate radionuclides (e.g., [14C], [123I]) and pattern of displacement is evaluated following administration of a dopaminergic agent. Increased synaptic dopamine levels are associated with reduced binding potential of D2 ligands. Striatal and cortical dopaminergic circuits are known to be under regulatory control by glutamatergic systems (Carlsson, 2006; Javitt and Zukin, 1991; Kulagina *et al.*, 2001). Such studies permit *in vivo* assessment of dopamine–glutamate interactions.

Both amphetamine (Breier *et al.*, 1998; Laruelle *et al.*, 1995) and ketamine (Breier *et al.*, 1998; Smith *et al.*, 1998; Vollenweider *et al.*, 2000) decrease striatal binding potential of D2 ligands following acute administration in humans, suggesting that both increase striatal dopamine levels. Thus, the presumed subcortical hyperdopaminergia of schizophrenia could result from either underlying dopaminergic hyperactivity or NMDA hypoactivity. Dissociative effects of ketamine, however, have been observed even under administration conditions that do not acutely affect striatal dopamine levels (Kegeles *et al.*, 2002), suggesting that psychotomimetic effects of ketamine cannot be attributed to alterations in dopaminergic function alone.

Objective assessment of dopamine function has been operationalized in schizophrenia using amphetamine challenge. Across cohorts, patients with acute schizophrenia show enhanced striatal dopamine release to amphetamine challenge, consistent with presumed dysregulation of subcortical dopamine circuits (Laruelle *et al.*, 1999). Levels of dopamine increase, moreover, correlate with severity of amphetamine-induced positive symptoms (Laruelle *et al.*, 1996). Deficits similar to those observed in schizophrenia are observed as well in normal volunteers undergoing ketamine infusion (Kegeles *et al.*, 2000), and in rodents treated acutely (Miller and Abercrombie, 1996) or subchronically (Balla *et al.*, 2001) with NMDA receptor antagonists. In nonhuman primates, the metabotropic agonist LY354740 also potentiates amphetamine-induced dopamine release (van Berckel *et al.*, 2006), by inhibiting presynaptic glutamate release, further suggesting that deficits in glutamatergic functioning may underlie dopaminergic hyperreactivity in schizophrenia.

Although initial *in vivo* studies in schizophrenia focused primarily on striatal functioning, dopamine receptor occupancy studies have now been performed in cortex as well. For example, Aalto *et al.* (2005) have demonstrated that acute ketamine administration increases dopamine release in cortex, as well as in striatum, and that the increase in release correlates with severity of ketamine-induced psychotic symptoms. Narendran *et al.* (2005) have demonstrated increases in D1 receptor binding in chronic ketamine abusers, suggesting also that ketamine may modulate prefrontal dopaminergic neurotransmission. Similar effects are observed in primates, in which chronic treatment with NMDA antagonists reduced tonic dopamine levels and D1 receptor upregulation, along with deficits in working memory (Tsukada *et al.*, 2005). In rodents, subchronic

treatment with NMDA antagonists induces enhanced amphetamine-induced dopamine release in prefrontal cortex (Balla *et al.*, 2003), suggesting that schizophrenia may be associated with reduced tonic dopamine levels as well with prefrontal hyperdopaminergia.

At present, radioligand binding studies of D1 receptor binding in schizophrenia have yielded conflicting results, with individual studies showing decreases (Okubo *et al.*, 1997), no change (Karlsson *et al.*, 2002) and increases (Abi-Dargham, 2003) in schizophrenia subjects, with discrepancy between studies most likely reflecting differences in patient and receptor ligand binding characteristics.

D. POSTMORTEM FINDINGS

Postmortem studies provide a final source of information that can be used to evaluate potential explanatory value of glutamatergic models. Schizophrenia is associated with complex patterns of alterations in protein and gene expression that cannot be easily explained based on dopaminergic models alone. For example, robust and reproducible deficits in parvalbumin and GAD67 expression are observed in postmortem hippocampus and prefrontal cortex in schizophrenia subjects (Reynolds *et al.*, 2004; Torrey *et al.*, 2005), although underlying mechanisms are unknown. NMDA receptors regulate both parvalbumin and GAD67 expression in cultured GABAergic interneurons, with ketamine leading to reduced expression of both agents (Kinney *et al.*, 2006). Similarly, subchronic treatment with PCP in rodents leads to downregulation of parvalbumin expression *in vivo* (Abdul-Monim *et al.*, 2006; Reynolds *et al.*, 2004). Other deficits, such as altered levels of *N*-acetylaspartate (NAA), *N*-acetylaspartylglutamate (NAAG) (Tsai *et al.*, 1995), and carboxypeptidase (Ghose *et al.*, 2004) may also be reproduced by subchronic PCP administration (Flores and Coyle, 2003; Reynolds *et al.*, 2005). Effects of NMDA antagonists have only been tested on a minority of postmortem markers of schizophrenia, and effects of antipsychotic agents in postmortem studies cannot always be excluded. Nevertheless, studies performed to date suggest that glutamatergic models may be able to explain postmortem as well as *in vivo* findings in schizophrenia.

IV. Clinical Studies with NMDA Agonists

To date, all approved agents for treatment of schizophrenia function by blocking neurotransmission at D2-type dopamine receptors. Given the hypothesis that NMDA dysfunction may underlie both clinical symptoms and neurocognitive dysfunction associated with schizophrenia, a critical issue is whether glutamate

TABLE I
SUMMARY OF CLINICAL FINDINGS WITH THE FULL *N*-METHYL-D-ASPARTATE RECEPTOR GLYCINE-SITE AGONISTS GLYCINE AND D-SERINE AND THE PARTIAL AGONIST D-CYCLOSERINE IN COMBINATION WITH TYPICAL, ATYPICAL, OR MIXED ANTIPSYCHOTICS IN SCHIZOPHRENIA

Study	Agonist ^a	Antipsychotic ^b	<i>N</i>	Negative		Cognitive		Positive	
				Change (%)	<i>p</i>	Change (%)	<i>p</i>	Change (%)	<i>p</i>
Heresco-Levy <i>et al.</i> (1999)	GLY	Mixed	22 ^c	−39	<0.001	−24	0.01	−20	NS
Javitt <i>et al.</i> (2001)	GLY	Mixed	12 ^c	−34	<0.05	−11.9	0.1	−11	0.08
Heresco-Levy <i>et al.</i> (2004)	GLY	Olz/Risp	17 ^c	−23	<0.0001	−9.2	0.02	−11.4	0.006
Evins <i>et al.</i> (2000)	GLY	Clozapine	27	−4	NS	—	—	−7	NS
Tsai <i>et al.</i> (1998)	DSER	Mixed	29	−20	<0.001	−17.7	0.004	−21.9	0.004
Heresco-Levy <i>et al.</i> (2005)	DSER	Olz/Risp	39 ^c	−16	<0.001	−11.7	0.001	−13	0.001
Tsai <i>et al.</i> (1999)	DSER	Clozapine	20	−2.5	NS	−0.8	NS	−3.6	NS
Tsai <i>et al.</i> (2005)	D-Alanine	Mixed	32	−17	<.0001	−13	<.0001	−13	.0002
Goff <i>et al.</i> (1999b)	DCS	Conventional	47	−23	<.02 ^b	—	—	—	NS
Heresco-Levy <i>et al.</i> (2002)	DCS	Mixed	21 ^c	−14	<.05	—	—	—	NS
Goff <i>et al.</i> (1999a)	DCS	Clozapine	17 ^c	+13 ^d	<.005	—	—	—	NS
Tsai <i>et al.</i> (2004a)	Sarcosine	Mixed	38	−14	<.0001	−17	<.0001	−13	<.0001
Lane <i>et al.</i> (2006)	Sarcosine	Clozapine	20	−2.1	NS	−9.1	NS	−6.6	NS

agonists can ameliorate persistent symptoms of schizophrenia. The glutamate-binding site of the NMDA and AMPA receptors cannot easily be targeted because of fear of seizures, excitotoxicity, and other evidence of cortical hyperexcitability. Further, these sites are not designed to be tonically occupied by agonist but instead must be physically activated analogously to other ionotropic receptors (e.g., nicotinic cholinergic, GABA_A). Instead, current strategies have focused on mechanisms for increasing efficiency of both NMDA- and AMPA-mediated neurotransmissions without altering levels of glutamate itself. For NMDA receptors, studies have targeted primarily the glycine modulatory site, in part because of the availability of naturally occurring compounds that permitted early stage clinical investigations. A “second generation” approach has been the use of glycine (GLYT1) transport inhibitors, which lead to increases in brain glycine levels by blocking its removal from the synaptic space. Finally, early stage trials have been conducted with allosteric AMPA receptor modulators (AMPAkines), as well as metabotropic receptor agonists and antagonists.

A. NMDA RECEPTOR GLYCINE-SITE AGONISTS

To date, the majority of clinical studies based on the glutamate hypothesis have been conducted with positive allosteric modulators of the NMDA receptor complex. Three separate agents have been used for these studies: glycine and D-serine, which function as full agonists, and D-cycloserine, which functions as a partial agonist. Glycine has been used primarily at doses of ~0.4–0.8 g/kg/day (~30–60 g/day); D-serine, at a dose of 30 mg/kg/day (~2.1 g/day) and D-cycloserine at a dose of 50 mg/day. In addition, one study has been performed with D-alanine, a close structural analogue of D-serine, used at a dose of 100 mg/kg/day (~7 g/day) (Table I). With glycine and D-serine, effectiveness of higher doses has not been explored so that maximal benefit obtainable from glycine-site stimulation is unknown. With D-cycloserine, doses in excess of 100 mg cause symptom exacerbation due to emergent NMDA receptor antagonist effects, producing a narrow therapeutic window (van Berckel *et al.*, 1999).

To date, 11 studies have been performed involving over 250 subjects (Table I). All studies involved patients with persistent negative symptoms while on stable

^aGLY, glycine (0.4–0.8 g/kg/day); DSER, D-serine (30 mg/kg/day); DALA, D-alanine (100 mg/kg/day); DCS, D-cycloserine (50 mg/day); SARC, sarcosine (30 mg/kg/day).

^bOLZ, olanzapine; RISP, risperidone.

^cCrossover study.

^dSignificant difference with SANS only; PANSS difference not significant, positive value represents significant worsening of symptoms.

–, Not determined; NS, not significant.

dose of antipsychotic medication. NMDA agonists were added as adjunctive medication while patients remained on prior antipsychotic regimen. All studies published to date have demonstrated large effect-size (0.9–2.1 SD units) improvement in negative and cognitive symptoms when these agents are added to typical antipsychotics, or newer atypicals. Percentage improvement in negative symptoms range from 16% to 39% (weighted mean 22.5%) for trials in the range of 6–12 weeks. Whether greater reduction occurs during longer-term treatment, or whether tolerance develops, is currently unknown.

The level of cognitive and positive symptom improvement, across studies, is roughly 15%, suggesting the possibility of adjunctive benefit in combination with antipsychotics. Given the ability of NMDA antagonists to induce positive as well as negative symptoms during acute challenge studies, it is not clear whether the greater effect of NMDA agonists on negative, as compared with positive, symptoms is due to an intrinsic property of the approach, or rather to the fact that positive symptoms are already partially treated by antipsychotic medication. Ultimately, monotherapy or antipsychotic withdrawal studies will be required to address these possibilities.

In some, but not all studies with glycine, the degree of negative symptoms improvement has correlated significantly with baseline glycine levels, suggesting that patients with lowest pretreatment levels respond best to NMDA receptor agonist treatment (Heresco-Levy *et al.*, 1999). With glycine, the critical plasma level for therapeutic response appears to be in the range of 600–1000 μM versus a basal level of ~ 200 μM . Similar levels have been observed in animal studies (Javitt *et al.*, 2004). The mean percentage change associated with glycine treatment in these studies was $\sim 30\%$, suggesting that if these findings could be replicated results would be clinically meaningful.

Partial agonist effects: As compared with full glycine-site agonists, the partial agonist D-cycloserine has proven less efficacious across clinical sites (Table I), and across studies within clinical site (Heresco-Levy *et al.*, 1998). A meta-analysis of clinical studies performed through 2004 found evidence of significant beneficial effect of full glycine-site agonists across studies, but not of D-cycloserine (Tuominen *et al.*, 2005). Despite its overall lack of efficacy, however, D-cycloserine was reported to increase temporal lobe activation during a word recall task, with effects correlating with degree of reduction in negative symptoms (Yurgelun-Todd *et al.*, 2005). Thus, specific beneficial effects may occur over short-term treatment, although it is postulated that tolerance may occur during longer term trials.

NMDA agonists in combination with clozapine: Relative to effects in combination with typical or newer atypical antipsychotics, glycine-site agonists have proven less effective when combined with clozapine. In double blind, placebo-controlled studies in which glycine (Evins *et al.*, 2000) or D-serine (Tsai *et al.*, 1999) have been added to clozapine, no significant beneficial response has been observed,

while D-cycloserine is reported to lead to worsening of symptoms when used in combination with clozapine (Goff *et al.*, 1996).

D-Cycloserine functions as a glycine-site agonist in the presence of high glycine concentrations, and as an antagonist in the presence of high concentrations (Hood *et al.*, 1989). A parsimonious explanation for the differential effects of NMDA agonists in combination with clozapine versus other antipsychotic agents, therefore, is that clozapine may already increase synaptic glycine levels through as yet unknown mechanisms. Recently, clozapine has recently been shown to block glycine and glutamine transport mediated by SNAT2-like synaptosomal transporters, providing a potential mechanism for both the differential therapeutic effects of clozapine and the differential effects of NMDA receptor modulators in the presence of clozapine versus other antipsychotics (Javitt *et al.*, 2005a). This finding may also account for the reported ability of clozapine to increase serum glutamate levels (Evins *et al.*, 1997) and downregulate central glutamate transport (Melone *et al.*, 2003; Pietraszek *et al.*, 2002).

CONSIST: In addition to published studies, one additional study (CONSIST) has been presented to date only in abstract form (Buchanan *et al.*, 2004). In that study, glycine (60 g/day) and D-cycloserine (50 mg/day) were compared versus placebo as adjunctive medication for persistent negative symptoms, using inclusion/exclusion criteria designed to enrich recruitment for individuals meeting criteria for the deficit syndrome (Carpenter *et al.*, 1988). Study duration was 16 weeks. Patients were on stable doses of antipsychotics other than clozapine. In that study, no significant beneficial effects were observed for either glycine or D-cycloserine, although subgroup analyses showed significant beneficial effects of glycine in inpatients and in patients receiving conventional medications only. No significant beneficial effects on cognition were observed in any group. Overall, larger trials are required, possibly with enriched inpatient populations.

Use of glycine in the schizophrenic prodrome: The majority of studies with NMDA agonists have focused on individuals well advanced in their illness. Recently, however, glycine was used in an open label monotherapy study in 10 individuals showing prodromal signs of schizophrenia. Although the number of subjects was limited, three met early remission criteria, one other showed substantial improvement, and two showed moderate improvement. Across all subjects, large effect size changes were observed across both positive and negative domains. Effects of glycine were more pronounced than those that had been observed in a prior double blind study of olanzapine (Woods *et al.*, 2004). These data, if confirmed, would indicate that NMDA agonists might have a primary role in the earliest stages of schizophrenia psychosis, with potential impact across symptomatic domains.

B. GLYCINE TRANSPORT INHIBITORS

Both glycine and D-serine appear to be effective when used in treatment resistant schizophrenia. However, both must be given at gram-level doses in order to significantly elevate CNS levels. An alternative approach to increasing CNS levels is use of glycine transport inhibitors (GTIs), which raise synaptic glycine levels by preventing its removal from the synaptic cleft. Use of GTIs to augment NMDA functioning is analogous to use of selective serotonin reuptake inhibitors (SSRIs) to raise synaptic serotonin levels in depression.

Initial studies were performed using the relatively nonselective glycine transport antagonist, glycyldodecylamide (GDA). This drug was shown to inhibit glycine transport in cortical (Javitt and Frusciante, 1997) or hippocampal (Harsing *et al.*, 2001) synaptosomes, and inhibit amphetamine-induced dopamine release (Javitt *et al.*, 2000) and PCP-induced hyperactivity in rodents (Javitt *et al.*, 1997, 1999; Toth *et al.*, 1986). Studies have been performed with selective, high-affinity GTIs such as *N*[3-(4'-fluorophenyl)-3-(4'-phenylphenoxy)propyl]sarcosine (NFPS; Aubrey and Vandenberg, 2001), Org 24598 (Brown *et al.*, 2001), CP-802,079 (Martina *et al.*, 2004), or SSR504734 (Depoortere *et al.*, 2005).

As with GDA, high-affinity GTIs have been found to reverse PCP-induced hyperactivity (Harsing *et al.*, 2003) and dopaminergic hyperreactivity (Javitt *et al.*, 2004) in rodents, and to potentiate NMDA responses in hippocampal slices *in vitro* (Bergeron *et al.*, 1998; Depoortere *et al.*, 2005; Kinney *et al.*, 2003b; Martina *et al.*, 2004) and prefrontal cortical neurons *in vivo* (Chen *et al.*, 2003). Glycine transport inhibitors also reverse PPI abnormalities in DBA/2J mice (Depoortere *et al.*, 2005; Kinney *et al.*, 2003b) and rats with neonatal hippocampal lesions (Le Pen *et al.*, 2003), supporting a potential role of GTIs in treatment of schizophrenia. In striatal dopamine assays, GLYT1 inhibitors reduce amphetamine-induced dopamine release *in vivo* (Javitt *et al.*, 2004) and NMDA-stimulated release *in vitro* (Bennett and Gronier, 2005; Javitt *et al.*, 2005b), suggesting a likely effect on positive, as well as negative, symptoms of schizophrenia.

At saturating doses, effective GLYT1 inhibitors produce approximately two- to threefold increases in extracellular glycine concentrations (Depoortere *et al.*, 2005; Martina *et al.*, 2004). Significantly, however, positive effects on NMDA receptor-mediated neurotransmission occur at concentrations two to three orders of magnitude lower than those needed to significantly increase extracellular glycine levels. Further, an inverted U-shape curve has been observed in several studies where beneficial effects of GLYT1 antagonists on NMDA function may be diminished or lost at the highest doses used. These findings are consistent with a model in which GLYT1 inhibitors primarily affect glycine concentrations within the synaptic cleft (Fig. 2), which represents a separate brain compartment from the overall extracellular space or cerebrospinal fluid. Increases in extracellular glycine levels would occur only as a consequence of diffusion of glycine from

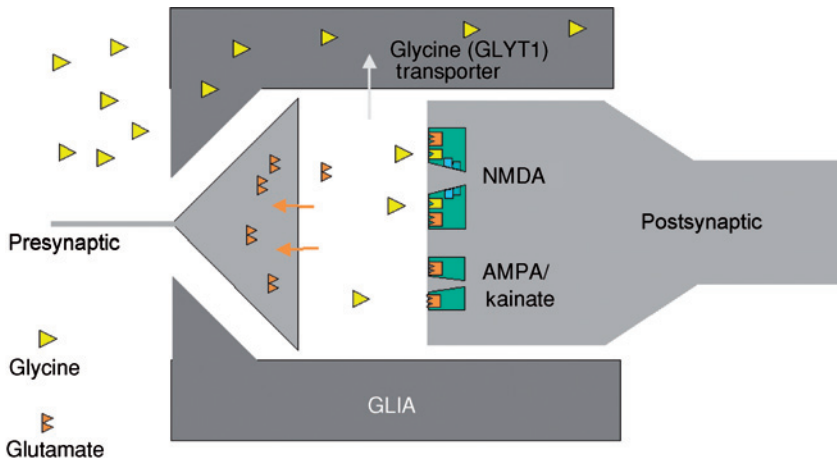


FIG. 2. Schematic model of synaptic glycine regulation by glycine transport inhibitors.

the synaptic to the general extracellular space, a process that would occur only at very high glycine concentrations. At extremely high concentrations, glycine may induce internalization of NMDA receptors, leading to loss of facilitatory glycine effects on NMDA transmission (Martina *et al.*, 2004). In rodents, effects of GLYT1 inhibitors have been found to be similar to those of clozapine (Lipina *et al.*, 2005), suggesting potential overlap of cognition enhancing mechanisms.

Further support for use of GLYT1 antagonists comes from studies of GLYT1 knockdown mice. GLYT1^{-/-} mice cannot be developed due to neonatal lethality (loss of breathing) (Tsai *et al.*, 2004b). Nevertheless, alternative strategies have been employed. For example, GLYT1^{+/-} heterozygote mice show a significant decrease in GLYT1 expression throughout brain and an enhancement of hippocampal NMDA activity consistent with GLYT1 inhibitor studies (Gabernet *et al.*, 2005; Martina *et al.*, 2005; Tsai *et al.*, 2004b). Similarly, selective forebrain knockouts show reductions in frontal glycine transport and potentiation of hippocampal NMDA responses as well as procognitive ability on several learning/memory paradigms (Yee *et al.*, 2006). As with animals treated with GLYT1 inhibitors, GLYT1^{+/-} heterozygote mice show some evidence of NMDA internalization, particularly behavioral hypersensitivity to the NMDA antagonist MK-801 (Tsai *et al.*, 2004b). Nevertheless, on balance, the phenotype supports a net potentiation of NMDA neurotransmission.

Sarcosine (*N*-methylglycine) is a naturally occurring GLYT1 antagonist that has been used for preliminary proof-of-principle studies. To date, clinical studies with sarcosine have been conducted only in Taiwan due to its regulatory status in

the United States. In one study (Tsai *et al.*, 2004a) in which it was combined with mixed typical and atypical antipsychotics, sarcosine, at a dose of 30 mg/kg/day (2.1 g/day) produced clinical effects extremely similar to those of glycine and D-serine (Table I). In contrast, when combined with clozapine (Lane *et al.*, 2006), sarcosine had no significant effect on symptoms, also consistent with prior glycine/D-serine studies. Interestingly, in one study where sarcosine and D-serine were added to risperidone in acutely relapsing subjects.

C. OTHER IONOTROPIC TARGETS

Because AMPA receptors function in concert with NMDA receptors, they have been proposed as alternative therapeutic targets in schizophrenia. AMPA-kinases function as positive allosteric modulators of AMPA receptor-mediated neurotransmission, and facilitate learning and memory in both human (Ingvar *et al.*, 1997) and animal (Hampson *et al.*, 1998a,b) models. Further, these drugs act synergistically with antipsychotics to reverse amphetamine-induced hyperactivity (Johnson *et al.*, 1999).

In a pilot study, the AMPA-kinase CX-516 induced significant improvements in memory and attention when added to clozapine, despite lack of symptomatic improvement (Goff *et al.*, 2001). However, these results were not confirmed in a larger confirmation study (Goff *et al.*, 2005), nor were beneficial effects observed in a small monotherapy trial (Marenco *et al.*, 2002). Although downregulation of AMPA receptors is less with AMPA-kinases than with direct agonists, there is some concern that downregulation may nonetheless occur and may limit long-term treatment strategies (Jourdi *et al.*, 2005).

Lamotrigine, an antiepileptic that reduces presynaptic glutamate release, has also been proposed as a potential adjunctive medication in schizophrenia, based on the theory that detrimental effects of NMDA antagonists on cognitive functioning may be due to glutamatergic rebound (Anand *et al.*, 2000; Dursun *et al.*, 1999). In a clinical challenge study, lamotrigine prevented acute psychotomimetic effects of ketamine, with greater effects on positive than negative symptoms (Anand *et al.*, 2000) supporting potential therapeutic efficacy. Improvements in positive and general symptoms were reported as well in small-scale studies of lamotrigine in clozapine-treated patients with persistent clinical symptoms (Dursun and Deakin, 2001; Tiihonen *et al.*, 2003). However, effects failed to reach statistical significance in a subsequent double blind study (Kremer *et al.*, 2004). Further, an industry-sponsored multicenter controlled study also did not show significant benefit (<http://ctr.gsk.co.uk/Summary/lamotrigine/studylist.asp>). Thus, as of yet limited clinical evidence is available to support the efficacy of either AMPA agonists or general glutamate antagonists.

D. METABOTROPIC RECEPTORS

Metabotropic modulators are currently in an early stage of development for treatment of schizophrenia. Studies attempting to validate metabotropic receptors as therapeutic targets in schizophrenia have been based on two alternative conceptualizations of the disorder. Group I receptors potentiate presynaptic glutamate release and NMDA receptor-mediated neurotransmission. Therapeutic effectiveness of Group I agonists is therefore predicted based on models which postulate low NMDA receptor activity and/or glutamate levels as being pathophysiological in schizophrenia. In contrast, Group II/III agonists inhibit glutamate release. Use of these agents follows models, which postulate that glutamatergic hyperactivity may be pathophysiological.

E. GROUP I RECEPTORS

Group I includes both mGluR1 and mGluR5 receptors, both of which stimulate NMDA receptors via differential second messenger cascades (Benquet *et al.*, 2002; Heidinger *et al.*, 2002). Preclinical studies have evaluated the ability of Group I antagonists to induce schizophrenia-like behavioral effects, and Group I agonists to reverse effects of amphetamine, PCP and other psychotomimetics. The most widely used mGluR5 antagonist, 2-methyl-6-(phenylethynyl)pyridine (MPEP), does not affect locomotor activity or PPI by itself but potentiates PCP-induced increases in locomotor activity and disruption of PPI (Henry *et al.*, 2002; Kinney *et al.*, 2003a). Similar effects have been observed with the more recently developed compound 3-[(2-methyl-1,3-thiazol-4-yl)ethynyl]-pyridine (MTEP) (Cosford *et al.*, 2003; Pilc *et al.*, 2002). Finally, mGluR1 (Brody *et al.*, 2003a) and mGluR5 (Kinney *et al.*, 2003a) knockout mice show disruptions of PPI, that respond poorly to known treatments for schizophrenia (Brody *et al.*, 2003b), supporting a potential role of Group I receptors as therapeutic targets in schizophrenia. Group I antagonists also produce anxiolytic-like effects in several animal models of anxiety, suggesting that they may be independent targets for the treatment of anxiety disorders (Chojnacka-Wojcik *et al.*, 2001).

Studies with Group I agonists have also been supportive of potential therapeutic effectiveness, but are more limited. For example, the mGluR5 agonist 2-chloro-5-hydroxyphenylglycine (CHPG) has been found to reverse PPI-disruptive effects of amphetamine in rodents (Kinney *et al.*, 2003a). Similarly, both nonselective and Group I selective agonists inhibit PCP-induced dopamine release in rodent prefrontal cortex (Maeda *et al.*, 2003). An issue in the use of direct agonists is rapid receptor desensitization, preventing chronic use. An alternative approach is the use of positive allosteric modulators, which, do not bind directly to

the agonist-binding site. Positive modulators, in general, have proven to be lipophilic and centrally acting, making them attractive as potential pharmacological agents (Pin and Acher, 2002).

Despite some encouraging results with Group I agonists in animal models, clinical data remain lacking. Further, Group I receptors have a markedly different cellular distribution in primates than rodents (Muly *et al.*, 2003; Paquet and Smith, 2003). Thus, primate studies and eventual clinical trials will be needed to validate this target for treatment of neuropsychiatric disorders.

F. GROUP II METABOTROPIC AGONISTS

Groups II and III metabotropic receptors are negatively linked to glutamate release, and may limit endogenous release under conditions of glutamate excess. Use of Group II/III agonists in schizophrenia is therefore based on the hypothesis that increased glutamate levels may be pathophysiological. Several high-affinity agonists have been developed over recent years, including (-)-2-oxa-4-aminobicyclo[3.1.0.]hexane-4,6-dicarboxylate (LY379268) and the related compound LY354740, permitting characterization of effects of Group II agonists in both preclinical and clinical studies (Schoepp and Marek, 2002).

An initial study with LY379268 demonstrated its ability to block PCP-induced increases in prefrontal glutamate, along with PCP-induced impairments in working memory, suggesting a role of glutamatergic hyperactivity in at least some forms of prefrontal dysfunction (Lorrain *et al.*, 2003; Moghaddam and Adams, 1998). Similarly, LY3279268 has been shown by a variety of groups to inhibit PCP-induced hyperactivity during both acute (Clark *et al.*, 2002; Makino *et al.*, 2003) and repeated (Cartmell *et al.*, 2000) administration, and reverse PCP-induced behaviors in monoamine depleted mice (Swanson and Schoepp, 2002). Finally, LY354740 has been found to reverse effects of NMDA antagonists in both rodents and humans (Krystal *et al.*, 2004; Moghaddam and Adams, 1998). Despite these intriguing results, however, clinical data with metabotropic agonists or antagonists are yet to be reported.

V. Potential Causes of Glutamatergic Dysfunction in Schizophrenia

The observation that NMDA antagonists induce both symptoms and neurocognitive deficits closely resembling those of schizophrenia (at least the early stages), suggests strongly that dysfunction or dysregulation of NMDA receptor-mediated neurotransmission may contribute heavily to the pathophysiology of schizophrenia. As of yet, however, the basis for NMDA dysfunction has yet to be

determined. Given dopaminergic theories of the disorder, one issue concerns whether glutamatergic deficits in schizophrenia may be secondary to primary disturbances in dopaminergic mechanisms. Additionally, schizophrenia is generally conceived as having both genetic and environmental components, with each set of factors contributing approximately half to overall risk. Over recent years, candidate genetic and environment mechanisms have been proposed. Several of these mechanisms significantly impact glutamatergic neurotransmission, and may account for the apparent NMDA dysregulation observed in schizophrenia.

A. DOPAMINE-GLUTAMATE INTERACTIONS

At present, dopamine D2 antagonists are the mainstay treatments for schizophrenia. Their efficacy depends on the presumed dysregulation of dopaminergic systems in schizophrenia, although objective evidence of such dysregulation is observed primarily during acute phases of the illness (Laruelle *et al.*, 1999). As such, the issue arises as to how dopaminergic treatments affect NMDA receptor-mediated phenomena, and whether primary disturbances in dopaminergic neurotransmission may contribute to secondary dysregulation of NMDA activation.

The strongest evidence for primary dopaminergic dysfunction in schizophrenia, at present, comes from studies of polymorphisms of the catechol-*O*-methyltransferase (COMT) gene. A common SNP in the COMT gene causes a Val to Met transition at AA158/AA108 (Val158Met), resulting in reduced COMT activity in Met allele carriers, and thus increased dopamine levels in prefrontal cortex. In initial studies, increased presence of the Val (high activity) allele was associated with increased risk for schizophrenia as well as impaired prefrontal function (Weinberger *et al.*, 2001), suggesting that low prefrontal dopamine levels may mediate both sets of effect.

However, considerable controversy has arisen concerning the linkage to schizophrenia (Meyer-Lindenberg *et al.*, 2006), with several studies failing to find an association with schizophrenia independent of effects on cognition (Ehlis *et al.*, 2006). It has also been suggested that COMT polymorphisms are not associated with schizophrenia itself, but with manic (Derosse *et al.*, 2006), as well as aggression (Lachman *et al.*, 1998) symptoms within schizophrenia, which may lead to overrepresentation in specific clinical populations. To the extent that COMT polymorphisms are associated with schizophrenia, they suggest that low, rather than high, dopamine levels in PFC may be pathogenic. Whether the association with schizophrenia is ultimately supported, however, remains to be determined.

Other evidence for potential etiological involvement of dopamine in schizophrenia derives from convergences between brain glutamatergic and dopaminergic systems. One primary site of convergence of glutamatergic and dopaminergic systems is on dendritic shafts and spines of striatal GABAergic medium spiny

interneurons (for review see Kotter, 1994; Smith and Bolam, 1990; Starr, 1995). In striatum, NMDA and D2 receptors produce opposite effects, with NMDA receptors producing net stimulation of striatal interneurons, and D2 receptors producing net inhibition (Cepeda and Levine, 1998; Cepeda *et al.*, 2001; Leveque *et al.*, 2000; Nicola *et al.*, 2000; Onn *et al.*, 2000; Peris *et al.*, 1988; West and Grace, 2002). Thus, in striatum, NMDA antagonists and dopamine agonists produce similar inhibition of GABAergic outflow, the first by decreasing excitation and the second by increasing inhibition.

Dopamine is not needed to mediate the behavioral effects of NMDA antagonists in rodents (Chartoff *et al.*, 2005). Nevertheless, dopamine may enhance these effects, and D2 antagonists may normalize GABAergic tone in striatum regardless of whether the primary deficit consists of dopaminergic hyperactivity or glutamatergic hypoactivity. Both glycine and GTIs have been shown to increase NMDA-stimulated GABA release in isolated rodent striatum, while decreasing NMDA-stimulated dopamine release (Javitt *et al.*, 2005b), consistent with a facilitatory effect on NMDA activation of GABAergic striatal interneurons. GABAergic effects, in turn, are mediated most likely through GABA_B receptors on presynaptic dopaminergic terminals (Javitt *et al.*, 2005b).

In contrast to D2 receptors, which mediate opposite effects to those of NMDA, D1 receptors show complex but primarily facilitatory interactions (Cepeda and Levine, 2006; Missale *et al.*, 2006). First, D1 receptors potentiate NMDA receptor-mediated responses via stimulation of cyclic AMP (cAMP)-protein kinase A pathway leading to DARPP-32 phosphorylation and subsequent phosphorylation of the NMDA receptor. Second, D1 receptor stimulation induces translocation of NMDA supporting a potential therapeutic role of D1 agonists. Second, D1 receptor activation results in rapid translocation of NMDA receptors to postsynaptic membranes, which, in turn, recruits D1 receptors to the membrane and enhances D1 receptor-mediated cAMP, leading to a positive feedback loop. Finally, direct protein-protein coupling between D1 and NMDA receptors may occupy, primarily involving NR1 and NR2A, but not NR2B, subunits. The interaction between D1 and NR2A receptors may attenuate NMDA response. Thus, depending on which effects predominate, either D1 agonists or antagonists might produce psychotherapeutic effects in schizophrenia.

Convergence between D1 and NMDA receptors occurs as well in cortex, where D1 receptors predominate over D2. In rodents and primates, chronic exposure to the NMDA receptors antagonists PCP and MK-801 results in decreased DA levels in the PFC (Jentsch and Roth, 1999; Jentsch *et al.*, 1997, 1998; Tsukada *et al.*, 2005). Further, in monkeys (Jentsch and Roth, 1999; Jentsch *et al.*, 1997, 1998; Tsukada *et al.*, 2005), chronic exposure to the NMDA antagonist MK-801 has been found to gradually lower DA levels in the PFC, and gradually upregulate binding of the D1 ligand [¹¹C]NNC 112. This finding was highly reminiscent of the increased

[¹¹C]NNC 112 binding observed in patients with schizophrenia in the DLPFC (Abi-Dargham *et al.*, 2002).

Furthermore, in the Tsukada *et al.* (2005) monkey study, upregulated prefrontal [¹¹C]NNC 112 BP was associated with impaired WM performance, a relationship observed in patients with schizophrenia as well (Abi-Dargham *et al.*, 2002). These data supported the hypothesis that, in schizophrenia, increased [¹¹C]NNC 112 BP is a compensatory response to a sustained deficit in prefrontal DA function stemming from a sustained deficit in NMDA transmission. Of note, however, alterations in D1 binding in both monkeys (Jentsch and Roth, 1999; Jentsch *et al.*, 1997, 1998; Tsukada *et al.*, 2005) and humans (Abi-Dargham *et al.*, 2002) are restricted to prefrontal cortex, suggesting that such interactions cannot easily account for cognitive deficits in schizophrenia that localize elsewhere in brain. In cortex, D2 receptor stimulation inhibits NMDA responses similarly to that observed in striatum (Tseng and O'Donnell, 2004). Although D2 receptor density in cortex is generally low, this interaction would be consistent with procognitive effects of D2 antagonists under hypofunctional NMDA conditions.

A final site of convergence is through glutamatergic projections from prefrontal cortex to midbrain dopaminergic nuclei. These projections modulate activity of midbrain DA neurons via an activating pathway, which has been termed the “accelerator” and an inhibitory pathway that has been called the “brake.” This dual excitatory–inhibitory interaction potentially permits the prefrontal cortex to fine-tune dopaminergic activity and produce regionally dichotomous effects. AMPA and NMDA receptors also serve different roles in this system, with AMPA receptors subserving tonic inhibitory regulation of mesoaccumbens neurons and a tonic excitatory regulation of mesoprefrontal DA neurons, and NMDA receptors mediated primarily phasic responses to behaviorally relevant stimuli (Takahata and Moghaddam, 2000). Loss of this descending glutamatergic input thus could produce subcortical dopaminergic hyperactivity and prefrontal hypoactivity, similar to what has been postulated to occur in schizophrenia.

Overall, therefore, dopamine–glutamate interactions remain an area of active research. To date, other than COMT linkages, little evidence implicates intrinsic dopaminergic deficits in the pathophysiology of schizophrenia, although both D1 and D2 receptors are part of interactive cascades that may potently modulate glutamatergic function. D1 agonists, in particular, have been suggested as potential psychotherapeutic agents based on their ability to potentiate NMDA responses (Goldman-Rakic *et al.*, 2004). However, clinical data supporting this hypothesis to date remain lacking. Further, based on patterns of cognitive change in COMT genotypes and patterns of D1 alteration in schizophrenia, D1 agonist effects may be limited to prefrontal-type deficits, and may fail to affect more distributed aspects of neurocognitive dysfunction in schizophrenia.

B. LINKAGE-ASSOCIATION STUDIES IN SCHIZOPHRENIA

Adoption studies of schizophrenia suggest that ~50% of the risk for schizophrenia is genetic, with the other 50% being attributable to environmental factors. Candidate genes for schizophrenia have only recently been identified, however, and considerable controversy continues to surround many of the targets. Nevertheless, a consistent and somewhat surprising finding (to the uninitiated) from genetic studies in schizophrenia is that several of the identified genes interact closely with glutamatergic mechanisms in general, and NMDA receptors in particular. As such, these studies provide additional support for glutamatergic theories of the disorder.

One of the best established candidate genes for schizophrenia is neuregulin, a brain transmitter that mediates its effects primarily through ErbB3 and ErbB4 receptors. A positive linkage to the 8p locus encoding the neuregulin 1 (NRG1) gene was first reported in 2002 (Stefansson *et al.*, 2002). A meta-analysis of 13 studies published through November, 2005 confirmed the association, suggesting an odd ratio of 1.22 and a *p* value of $<10^{-9}$. Linkages to polymorphisms in the gene encoding the ErbB4 receptor, particularly in Ashkenazi Jews, have also been reported (Silberberg *et al.*, 2006). In initial studies, it was suggested that NRG1 might mediate its risk-enhancing effects based on interaction with NMDA receptors, based on the observation that NRG1 hypomorphs had fewer functional NMDA receptors than wild-type mice (Stefansson *et al.*, 2002). In a recent functional postmortem study, NRG1 stimulation was found to suppress NMDA receptor activation in prefrontal cortex tissue from schizophrenia patients to a greater extent than it did in tissue from matched comparison subjects (Hahn *et al.*, 2006). Thus, increased expression of NRG1 may increase risk for schizophrenia primarily by downregulating cortical NMDA receptor-mediated neurotransmission.

Two other genes with strong relationship to schizophrenia, D-amino acid oxidase (DAAO) and G72 (aka DAOA), have also been linked to schizophrenia in at least some studies. DAAO is the primary enzyme responsible for degradation of D-serine in brain. G72 is a modulatory subunit for DAAO that appears to have arisen during late primate evolution. An initial linkage of both genes to schizophrenia was first reported in 2002 in a Russian cohort (Chumakov *et al.*, 2002), with the most active combination of DAAO and G72 (i.e., the forms that would produce lowest D-serine levels) producing the greatest risk for developing schizophrenia. These genetic findings resonate with independent neurochemical findings of decreased CSF D-serine levels in schizophrenia (Hashimoto *et al.*, 2005). The DAAO and G72 findings were subsequently confirmed in an independent German sample (Schumacher *et al.*, 2004), and the G72 finding in an Ashkenazi cohort (Korostishevsky *et al.*, 2004). In contrast, no association with either gene was found in a recently studied Taiwanese cohort (Liu *et al.*, 2006). A study by Goldberg *et al.* (2006) also did not find significant associations of

DAAO or G72 and schizophrenia, although G72 was strongly associated with cognitive dysfunction and reduced hippocampal activation during an episodic working memory task. Thus, whether polymorphisms of DAAO and G72 explain increased risk for schizophrenia, they may be associated with accompanying neurocognitive dysfunction.

To date, there is also limited evidence implicating NMDA receptors directly in the genetics of schizophrenia. One positive linkage study was reported in an African Bantu population but has not been replicated (Riley *et al.*, 1997). Studies have also reported some linkages between NR2 subunits and either schizophrenia itself (Di Maria *et al.*, 2004) or with specific clinical features of the disorder (Chiu *et al.*, 2003; Itokawa *et al.*, 2003). Other potential risk genes for schizophrenia such as dysbindin (DTNBP1), disrupted in schizophrenia-1 (DISC-1), RGS4, and metabotropic glutamate receptor 3 (GRM-3) may also converge on glutamatergic systems (Harrison and Weinberger, 2005; Moghaddam, 2003; Weinberger, 2005), although further clarification is needed concerning functional consequences of risk haplotypes for brain function.

C. ENVIRONMENTAL AND NEUROCHEMICAL FACTORS

Finally, environmental factors that contribute to development of schizophrenia may also converge on NMDA receptors. For example, it has been hypothesized that perinatal hypoxia, an important risk factor for schizophrenia, leads to neurotoxic degeneration of NMDA-bearing cells, an effect that may only produce behavioral symptoms later in development (Olney *et al.*, 1999). Similarly, schizophrenia has been associated with decreased plasma levels of the NMDA agonists glycine (Sumiyoshi *et al.*, 2004) and D-serine (Hashimoto *et al.*, 2003), and increased levels of homocysteine (Levine *et al.*, 2002; Susser *et al.*, 1998), an agent that may act as a functional NMDA antagonist. Levels of kynurenic acid, an endogenous NMDA and nicotine receptor antagonist, may also be high in schizophrenia (Erhardt *et al.*, 2001; Schwarcz *et al.*, 2001) and lead to inhibition of glutamatergic/NMDA function.

A final compound of potential etiological interest in schizophrenia is glutathione. Glutathione regulates NMDA receptors at the redox site. Low glutathione levels have been reported in CSF and prefrontal cortex in schizophrenia *in vivo* (Do *et al.*, 2000). In hippocampal slices, reduced glutathione levels are associated with reduced presynaptic glutamate release along with postsynaptic NMDA activity, consistent with the phenotype observed in schizophrenia (Steullet *et al.*, 2006). Although determinants of various neurochemical levels in brain are unknown at present, present findings suggest that alterations in metabolism or environmental exposure may explain significant variance in risk for developing schizophrenia, along with genetic factors.

VI. Future Research and Treatment Implications

Over the last 40 years, the dopamine model has been the leading neurochemical hypothesis of schizophrenia. This model has proven heuristically valuable, with all current medications for schizophrenia functioning primarily to block dopamine D2 receptors. Yet it remains unlikely that dopaminergic dysfunction, on its own, can fully account for the wide range of symptoms and neurocognitive deficits seen in schizophrenia. Glutamatergic models provide an alternate approach for conceptualizing the brain abnormalities associated with schizophrenia. As opposed to dopaminergic agonists, NMDA antagonists produce negative and cognitive symptoms of schizophrenia, along with positive symptoms, and induce neuropsychological deficits that are extremely similar to those observed in schizophrenia. At present, there are no approved medications for treatment of either negative symptoms or neurocognitive dysfunction. New treatment approaches aimed at potentiating glutamatergic neurotransmission particularly at NMDA receptors, however, offer some new hope for future clinical development.

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References

- Aalto, S., Ihalaenen, J., Hirvonen, J., Kajander, J., Scheinin, H., Tanila, H., Nagren, K., Vilkinan, H., Gustafsson, L. L., Syvalahti, E., and Hietala, J. (2005). Cortical glutamate-dopamine interaction and ketamine-induced psychotic symptoms in man. *Psychopharmacology (Berl.)* **182**, 375–383.
- Abdul-Monim, Z., Neill, J. C., and Reynolds, G. P. (2006). Sub-chronic psychotomimetic phencyclidine induces deficits in reversal learning and alterations in parvalbumin-immunoreactive expression in the rat. *J. Psychopharmacol.*
- Abi-Dargham, A. (2003). Probing cortical dopamine function in schizophrenia: What can D1 receptors tell us? *World Psychiatry* **2**, 166–171.
- Abi-Dargham, A., Mawlawi, O., Lombardo, I., Gil, R., Martinez, D., Huang, Y., Hwang, D. R., Keilp, J., Kochan, L., Van Heertum, R., Gorman, J. M., and Laruelle, M. (2002). Prefrontal dopamine D1 receptors and working memory in schizophrenia. *J. Neurosci.* **22**, 3708–3719.
- Abi-Saab, W. M., D'Souza, D. C., Moghaddam, B., and Krystal, J. H. (1998). The NMDA antagonist model for schizophrenia: Promise and pitfalls. *Pharmacopsychiatry* **31**(Suppl. 2), 104–109.

- Adler, C. M., Malhotra, A. K., Elman, I., Goldberg, T., Egan, M., Pickar, D., and Breier, A. (1999). Comparison of ketamine-induced thought disorder in healthy volunteers and thought disorder in schizophrenia. *Am. J. Psychiatry* **156**, 1646–1649.
- Anand, A., Charney, D. S., Oren, D. A., Berman, R. M., Hu, X. S., Cappiello, A., and Krystal, J. H. (2000). Attenuation of the neuropsychiatric effects of ketamine with lamotrigine: Support for hyperglutamatergic effects of N-methyl-D-aspartate receptor antagonists. *Arch. Gen. Psychiatry* **57**, 270–276.
- Aubrey, K. R., and Vandenberg, R. J. (2001). N[3-(4'-fluorophenyl)-3-(4'-phenylphenoxy)propyl] sarcosine (NFPS) is a selective persistent inhibitor of glycine transport. *Br. J. Pharmacol.* **134**, 1429–1436.
- Balla, A., Koneru, R., Smiley, J., Serksen, H., and Javitt, D. C. (2001). Continuous phencyclidine treatment induces schizophrenia-like hyperreactivity of striatal dopamine release. *Neuropsychopharmacology* **25**, 157–164.
- Balla, A., Serksen, H., Serra, M., Koneru, R., and Javitt, D. C. (2003). Subchronic continuous phencyclidine administration potentiates amphetamine-induced frontal cortex dopamine release. *Neuropsychopharmacology* **28**, 34–44.
- Barch, D. M., and Carter, C. S. (2005). Amphetamine improves cognitive function in medicated individuals with schizophrenia and in healthy volunteers. *Schizophr. Res.* **77**, 43–58.
- Bennett, S., and Gronier, B. (2005). Modulation of striatal dopamine release *in vitro* by agonists of the glycineB site of NMDA receptors; interaction with antipsychotics. *Eur. J. Pharmacol.* **527**, 52–59.
- Benquet, P., Gee, C. E., and Gerber, U. (2002). Two distinct signaling pathways upregulate NMDA receptor responses via two distinct metabotropic glutamate receptor subtypes. *J. Neurosci.* **22**, 9679–9686.
- Bergeron, R., Meyer, T. M., Coyle, J. T., and Greene, R. W. (1998). Modulation of N-methyl-D-aspartate receptor function by glycine transport. *Proc. Natl. Acad. Sci. USA* **95**, 15730–15734.
- Bilder, R. M., Lipschutz-Broch, L., Reiter, G., Geisler, S., Mayerhoff, D., and Lieberman, J. A. (1991). Neuropsychological deficits in the early course of first episode schizophrenia. *Schizophr. Res.* **5**, 198–199.
- Bilder, R. M., Goldman, R. S., Robinson, D., Reiter, G., Bell, L., Bates, J. A., Pappadopulos, E., Willson, D. F., Alvir, J. M., Woerner, M. G., Geisler, S., Kane, J. M., *et al.* (2000). Neuropsychology of first-episode schizophrenia: Initial characterization and clinical correlates. *Am. J. Psychiatry* **157**, 549–559.
- Breier, A., Adler, C. M., Weisenfeld, N., Su, T. P., Elman, I., Picken, L., Malhotra, A. K., and Pickar, D. (1998). Effects of NMDA antagonism on striatal dopamine release in healthy subjects: Application of a novel PET approach. *Synapse* **29**, 142–147.
- Brody, S. A., Conquet, F., and Geyer, M. A. (2003a). Disruption of prepulse inhibition in mice lacking mGluR1. *Eur. J. Neurosci.* **18**, 3361–3366.
- Brody, S. A., Conquet, F., and Geyer, M. A. (2004). Effect of antipsychotic treatment on the prepulse inhibition deficit of mGluR5 knockout mice. *Psychopharmacology (Berl.)* **172**:187–195.
- Brown, A., Carlyle, I., Clark, J., Hamilton, W., Gibson, S., McGarry, G., McEachen, S., Rae, D., Thorn, S., and Walker, G. (2001). Discovery and SAR of org 24598-a selective glycine uptake inhibitor. *Bioorg. Med. Chem. Lett.* **11**, 2007–2009.
- Buchanan, R. W., Javitt, D. C., Marder, S. R., Schooler, N. R., Heresco-Levy, U., Gold, J. M., McMahon, R. P., and Carpenter, W. T. (2004). Is Glutamatergic Therapy Efficacious in Schizophrenia? Results from the CONSIST Study. Presented at the 2004 ACNP Meeting, San Juan, PR.
- Butler, P. D., Zemon, V., Schechter, I., Saperstein, A. M., Hoptman, M. J., Lim, K. O., Revheim, N., Silipo, G., and Javitt, D. C. (2005). Early-stage visual processing and cortical amplification deficits in schizophrenia. *Arch. Gen. Psychiatry* **62**, 495–504.

- Carlsson, A. (1988). The current status of the dopamine hypothesis of schizophrenia. *Neuropsychopharmacology* **1**, 179–186.
- Carlsson, A. (2006). The neurochemical circuitry of schizophrenia. *Pharmacopsychiatry* **39**(Suppl. 1), S10–S14.
- Carpenter, W. T., Heinrichs, D. W., and Wagman, A. M. (1988). Deficit and non-deficit forms of schizophrenia: The concept. *Am. J. Psychiatry* **145**, 578–583.
- Cartmell, J., Monn, J. A., and Schoepp, D. D. (2000). Attenuation of specific PCP-evoked behaviors by the potent mGlu2/3 receptor agonist, LY379268 and comparison with the atypical antipsychotic, clozapine. *Psychopharmacology (Berl.)* **148**, 423–429.
- Cepeda, C., and Levine, M. S. (1998). Dopamine and N-methyl-D-aspartate receptor interactions in the neostriatum. *Dev. Neurosci.* **20**, 1–18.
- Cepeda, C., and Levine, M. S. (2006). Where do you think you are going? The NMDA-D1 receptor trap. *Sci. STKE* **2006**, pe20.
- Cepeda, C., Hurst, R. S., Altemus, K. L., Flores-Hernandez, J., Calvert, C. R., Jokel, E. S., Grandy, D. K., Low, M. J., Rubinstein, M., Ariano, M. A., and Levine, M. S. (2001). Facilitated glutamatergic transmission in the striatum of D2 dopamine receptor-deficient mice. *J. Neurophysiol.* **85**, 659–670.
- Chartoff, E. H., Heusner, C. L., and Palmiter, R. D. (2005). Dopamine is not required for the hyperlocomotor response to NMDA receptor antagonists. *Neuropsychopharmacology* **30**, 1324–1333.
- Chatterton, J. E., Awobuluyi, M., Premkumar, L. S., Takahashi, H., Talantova, M., Shin, Y., Cui, J., Tu, S., Sevarino, K. A., Nakanishi, N., Tong, G., Lipton, S. A., *et al.* (2002). Excitatory glycine receptors containing the NR3 family of NMDA receptor subunits. *Nature* **415**, 793–798.
- Chen, L., Muhlhauser, M., and Yang, C. R. (2003). Glycine transporter-1 blockade potentiates NMDA-mediated responses in rat prefrontal cortical neurons *in vitro* and *in vivo*. *J. Neurophysiol.* **89**, 691–703.
- Chiu, H. J., Wang, Y. C., Liou, Y. J., Lai, I. C., and Chen, J. Y. (2003). Association analysis of the genetic variants of the N-methyl-D-aspartate receptor subunit 2b (NR2b) and treatment-refractory schizophrenia in the Chinese. *Neuropsychobiology* **47**, 178–181.
- Chojnacka-Wojcik, E., Klodzinska, A., and Pilc, A. (2001). Glutamate receptor ligands as anxiolytics. *Curr. Opin. Invest. Drugs* **2**, 1112–1119.
- Chumakov, I., Blumenfeld, M., Guerassimenko, O., Cavarec, L., Palicio, M., Abderrahim, H., Bougueleret, L., Barry, C., Tanaka, H., La Rosa, P., Puech, A., Tahri, N., *et al.* (2002). Genetic and physiological data implicating the new human gene G72 and the gene for D-amino acid oxidase in schizophrenia. *Proc. Natl. Acad. Sci. USA* **99**, 13675–13680.
- Clark, M., Johnson, B. G., Wright, R. A., Monn, J. A., and Schoepp, D. D. (2002). Effects of the mGlu2/3 receptor agonist LY379268 on motor activity in phencyclidine-sensitized rats. *Pharmacol. Biochem. Behav.* **73**, 339–346.
- Cosford, N. D., Tehrani, L., Roppe, J., Schweiger, E., Smith, N. D., Anderson, J., Bristow, L., Brodtkin, J., Jiang, X., McDonald, I., Rao, S., Washburn, M., *et al.* (2003). 3-[(2-Methyl-1, 3-thiazol-4-yl)ethynyl]-pyridine: A potent and highly selective metabotropic glutamate subtype 5 receptor antagonist with anxiolytic activity. *J. Med. Chem.* **46**, 204–206.
- Coyle, J. T. (1996). The glutamatergic dysfunction hypothesis for schizophrenia. *Harv. Rev. Psychiatry* **3**, 241–253.
- Davis, K. L., Kahn, R. S., Ko, G., and Davidson, M. (1991). Dopamine in schizophrenia: A review and reconceptualization. *Am. J. Psychiatry* **148**, 1474–1486.
- Depoortere, R., Dargazanli, G., Estenne-Bouhtou, G., Coste, A., Lanneau, C., Desvignes, C., Poncelet, M., Heaulme, M., Santucci, V., Decobert, M., Cudennec, A., Voltz, C., *et al.* (2005). Neurochemical, electrophysiological and pharmacological profiles of the selective inhibitor of

- the glycine transporter-1 SSR504734, a potential new type of antipsychotic. *Neuropsychopharmacology* **30**, 1963–1985.
- DeRosse, P., Funke, B., Burdick, K. E., Lencz, T., Goldberg, T. E., Kane, J. M., Kucherlapati, R., and Malhotra, A. K. (2006). COMT genotype and manic symptoms in schizophrenia. *Schizophr. Res.* **87**(1–3), 28–31.
- Di Maria, E., Gulli, R., Begni, S., De Luca, S., Bignotti, A., Pasini, A., Bellone, E., Pizzuti, A., Dallapiccola, B., Novelli, G., Ajmar, F., Gennarelli, M., *et al.* (2004). Variations in the NMDA receptor subunit 2B gene (GRIN2B) and schizophrenia: A case-control study. *Am. J. Med. Genet.* **128B**, 27–29.
- Dickinson, D., Ragland, J. D., Calkins, M. E., Gold, J. M., and Gur, R. C. (2006). A comparison of cognitive structure in schizophrenia patients and healthy controls using confirmatory factor analysis. *Schizophr. Res.* **85**, 20–29.
- Do, K. Q., Trabesinger, A. H., Kirsten-Kruger, M., Lauer, C. J., Dydak, U., Hell, D., Holsboer, F., Boesiger, P., and Cuenod, M. (2000). Schizophrenia: Glutathione deficit in cerebrospinal fluid and prefrontal cortex *in vivo*. *Eur. J. Neurosci.* **12**, 3721–3728.
- Dursun, S. M., and Deakin, J. F. (2001). Augmenting antipsychotic treatment with lamotrigine or topiramate in patients with treatment-resistant schizophrenia: A naturalistic case-series outcome study. *J. Psychopharmacol.* **15**, 297–301.
- Dursun, S. M., McIntosh, D., and Milliken, H. (1999). Clozapine plus lamotrigine in treatment-resistant schizophrenia. *Arch. Gen. Psychiatry* **56**, 950.
- Ehlis, A. C., Reif, A., Herrmann, M. J., Lesch, K. P., and Fallgatter, A. J. (2006). Impact of catechol-O-methyltransferase on prefrontal brain functioning in schizophrenia spectrum disorders. *Neuropsychopharmacology*.
- Erhardt, S., Blennow, K., Nordin, C., Skogh, E., Lindstrom, L. H., and Engberg, G. (2001). Kynurenic acid levels are elevated in the cerebrospinal fluid of patients with schizophrenia. *Neurosci. Lett.* **313**, 96–98.
- Evins, A. E., Amico, E. T., Shih, V., and Goff, D. C. (1997). Clozapine treatment increases serum glutamate and aspartate compared to conventional neuroleptics. *J. Neural. Transm.* **104**, 761–766.
- Evins, A. E., Fitzgerald, S. M., Wine, L., Rosselli, R., and Goff, D. C. (2000). Placebo-controlled trial of glycine added to clozapine in schizophrenia. *Am. J. Psychiatry* **157**, 826–828.
- Flores, C., and Coyle, J. T. (2003). Regulation of glutamate carboxypeptidase II function in corticolimbic regions of rat brain by phencyclidine, haloperidol, and clozapine. *Neuropsychopharmacology* **28**, 1227–1234.
- Gabernet, L., Pauly-Evers, M., Schwerdel, C., Lentz, M., Bluethmann, H., Vogt, K., Alberati, D., Mohler, H., and Boison, D. (2005). Enhancement of the NMDA receptor function by reduction of glycine transporter-1 expression. *Neurosci. Lett.* **373**, 79–84.
- Ghose, S., Weickert, C. S., Colvin, S. M., Coyle, J. T., Herman, M. M., Hyde, T. M., and Kleinman, J. E. (2004). Glutamate carboxypeptidase II gene expression in the human frontal and temporal lobe in schizophrenia. *Neuropsychopharmacology* **29**, 117–125.
- Goff, D. C., Tsai, G., Manoach, D. S., Flood, J., Darby, D., and Coyle, J. T. (1996). D-cycloserine added to clozapine for patients with schizophrenia. *Am. J. Psychiatry* **153**, 1628–1630.
- Goff, D. C., Henderson, D. C., Evins, A. E., and Amico, E. (1999a). A placebo-controlled crossover trial of D-cycloserine added to clozapine in patients with schizophrenia. *Biol. Psychiatry* **45**, 512–514.
- Goff, D. C., Tsai, G., Levitt, J., Amico, E., Manoach, D., Schoenfeld, D. A., Hayden, D. L., McCarley, R., and Coyle, J. T. (1999b). A placebo-controlled trial of D-cycloserine added to conventional neuroleptics in patients with schizophrenia. *Arch. Gen. Psychiatry* **56**, 21–27.

- Goff, D. C., Leahy, L., Berman, I., Posever, T., Herz, L., Leon, A. C., Johnson, S. A., and Lynch, G. (2001). A placebo-controlled pilot study of the ampakine CX516 added to clozapine in schizophrenia. *J. Clin. Psychopharmacol.* **21**, 484–487.
- Goff, D. C., Lambert, J. S., Miller, A., Patel, J. K., Manschreck, T., Freudenreich, O., Green, M. F., Leon, A., and Johnson, S. (2005). Placebo-controlled, add-on trial of CX516 (Ampakine) in schizophrenia. *Neuropsychopharmacology* **30**, S127.
- Gold, S., Arndt, S., Nopoulos, P., O'Leary, D. S., and Andreasen, N. C. (1999). Longitudinal study of cognitive function in first-episode and recent-onset schizophrenia. *Am. J. Psychiatry* **156**, 1342–1348.
- Goldberg, T. E., Straub, R. E., Callicott, J. H., Hariri, A., Mattay, V. S., Bigelow, L., Coppola, R., Egan, M. F., and Weinberger, D. R. (2006). The G72/G30 gene complex and cognitive abnormalities in schizophrenia. *Neuropsychopharmacology* **31**, 2022–2032.
- Goldman-Rakic, P. S., Castner, S. A., Svensson, T. H., Siever, L. J., and Williams, G. V. (2004). Targeting the dopamine D1 receptor in schizophrenia: Insights for cognitive dysfunction. *Psychopharmacology (Berl.)* **174**, 3–16.
- Hahn, C. G., Wang, H. Y., Cho, D. S., Talbot, K., Gur, R. E., Berrettini, W. H., Bakshi, K., Kamins, J., Borgmann-Winter, K. E., Siegel, S. J., Gallop, R. J., and Arnold, S. E. (2006). Altered neuregulin 1-erbB4 signaling contributes to NMDA receptor hypofunction in schizophrenia. *Nat. Med.* **12**, 824–828.
- Hampson, R. E., Rogers, G., Lynch, G., and Deadwyler, S. A. (1998a). Facilitative effects of the ampakine CX516 on short-term memory in rats: Correlations with hippocampal neuronal activity. *J. Neurosci.* **18**, 2748–2763.
- Hampson, R. E., Rogers, G., Lynch, G., and Deadwyler, S. A. (1998b). Facilitative effects of the ampakine CX516 on short-term memory in rats: Enhancement of delayed-nonmatch-to-sample performance. *J. Neurosci.* **18**, 2740–2747.
- Harrison, P. J., and Weinberger, D. R. (2005). Schizophrenia genes, gene expression, and neuro-pathology: On the matter of their convergence. *Mol. Psychiatry* **10**, 40–68; image 5.
- Harsing, L. G., Jr., Solyom, S., and Salamon, C. (2001). The role of glycineB binding site and glycine transporter (GlyT1) in the regulation of [3H]GABA and [3H]glycine release in the rat brain. *Neurochem. Res.* **26**, 915–923.
- Harsing, L. G., Jr., Gacsalyi, I., Szabo, G., Schmidt, E., Sziray, N., Sebban, C., Tesolin-Decros, B., Matyus, P., Egyed, A., Spedding, M., and Levay, G. (2003). The glycine transporter-1 inhibitors NFPS and Org 24461: A pharmacological study. *Pharmacol. Biochem. Behav.* **74**, 811–825.
- Hartvig, P., Valtysson, J., Lindner, K. J., Kristensen, J., Karlsten, R., Gustafsson, L. L., Persson, J., Svensson, J. O., Oye, I., Antoni, G., *et al.* (1995). Central nervous system effects of subdissociative doses of (S)-ketamine are related to plasma and brain concentrations measured with positron emission tomography in healthy volunteers. *Clin. Pharmacol. Ther.* **58**, 165–173.
- Hashimoto, K., Fukushima, T., Shimizu, E., Komatsu, N., Watanabe, H., Shinoda, N., Nakazato, M., Kumakiri, C., Okada, S., Hasegawa, H., Imai, K., and Iyo, M. (2003). Decreased serum levels of D-serine in patients with schizophrenia: Evidence in support of the N-methyl-D-aspartate receptor hypofunction hypothesis of schizophrenia. *Arch. Gen. Psychiatry* **60**, 572–576.
- Hashimoto, K., Engberg, G., Shimizu, E., Nordin, C., Lindstrom, L. H., and Iyo, M. (2005). Reduced D-serine to total serine ratio in the cerebrospinal fluid of drug naive schizophrenic patients. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **29**, 767–769.
- Heidinger, V., Manzerra, P., Wang, X. Q., Strasser, U., Yu, S. P., Choi, D. W., and Behrens, M. M. (2002). Metabotropic glutamate receptor 1-induced upregulation of NMDA receptor current: Mediation through the Pyk2/Src-family kinase pathway in cortical neurons. *J. Neurosci.* **22**, 5452–5461.

- Henry, S. A., Lehmann-Masten, V., Gasparini, F., Geyer, M. A., and Markou, A. (2002). The mGluR5 antagonist MPEP, but not the mGluR2/3 agonist LY314582, augments PCP effects on prepulse inhibition and locomotor activity. *Neuropharmacology* **43**, 1199–1209.
- Heresco-Levy, U., Javitt, D. C., Ermilov, M., Silipo, G., and Shimoni, J. (1998). Double-blind, placebo-controlled, crossover trial of D-cycloserine adjuvant therapy for treatment-resistant schizophrenia. *Int. J. Neuropsychopharmacol.* **1**, 131–136.
- Heresco-Levy, U., Javitt, D. C., Ermilov, M., Mordel, C., Silipo, G., and Lichtenstein, M. (1999). Efficacy of high-dose glycine in the treatment of enduring negative symptoms of schizophrenia. *Arch. Gen. Psychiatry* **56**, 29–36.
- Heresco-Levy, U., Ermilov, M., Shimoni, J., Shapira, B., Silipo, G., and Javitt, D. C. (2002). Placebo-controlled trial of D-cycloserine added to conventional neuroleptics, olanzapine, or risperidone in schizophrenia. *Am. J. Psychiatry* **159**, 480–482.
- Heresco-Levy, U., Ermilov, M., Lichtenberg, P., Bar, G., and Javitt, D. C. (2004). High-dose glycine added to olanzapine and risperidone for the treatment of schizophrenia. *Biol. Psychiatry* **55**, 165–171.
- Heresco-Levy, U., Javitt, D. C., Ebstein, R., Vass, A., Lichtenberg, P., Bar, G., Catinari, S., and Ermilov, M. (2005). D-serine efficacy as add-on pharmacotherapy to risperidone and olanzapine for treatment-refractory schizophrenia. *Biol. Psychiatry* **57**, 577–585.
- Honey, R. A., Turner, D. C., Honey, G. D., Sharar, S. R., Kumaran, D., Pomarol-Clotet, E., McKenna, P., Sahakian, B. J., Robbins, T. W., and Fletcher, P. C. (2003). Subdissociative dose ketamine produces a deficit in manipulation but not maintenance of the contents of working memory. *Neuropsychopharmacology* **28**, 2037–2044.
- Hood, W. F., Compton, R. P., and Monahan, J. B. (1989). D-cycloserine: A ligand for the N-methyl-D-aspartate coupled glycine receptor has partial agonist characteristics. *Neurosci. Lett.* **98**, 91–95.
- Ingvar, M., Ambros-Ingerson, J., Davis, M., Granger, R., Kessler, M., Rogers, G. A., Schehr, R. S., and Lynch, G. (1997). Enhancement by an ampakine of memory encoding in humans. *Exp. Neurol.* **146**, 553–559.
- Isaac, J. T., Nicoll, R. A., and Malenka, R. C. (1999). Silent glutamatergic synapses in the mammalian brain. *Can. J. Physiol. Pharmacol.* **77**, 735–737.
- Itokawa, M., Yamada, K., Yoshitsugu, K., Toyota, T., Suga, T., Ohba, H., Watanabe, A., Hattori, E., Shimizu, H., Kumakura, T., Ebihara, M., Meerabux, J. M., et al. (2003). A microsatellite repeat in the promoter of the N-methyl-D-aspartate receptor 2A subunit (GRIN2A) gene suppresses transcriptional activity and correlates with chronic outcome in schizophrenia. *Pharmacogenetics* **13**, 271–278.
- Javitt, D. C. (1987). Negative schizophrenic symptomatology and the PCP (phencyclidine) model of schizophrenia. *Hillside J. Clin. Psychiatry* **9**, 12–35.
- Javitt, D. C., and Frusciant, M. (1997). Glycylododecylamide, a phencyclidine behavioral antagonist, blocks cortical glycine uptake: Implications for schizophrenia and substance abuse. *Psychopharmacology (Berl.)* **129**, 96–98.
- Javitt, D. C., and Zukin, S. R. (1991). Recent advances in the phencyclidine model of schizophrenia. *Am. J. Psychiatry* **148**, 1301–1308.
- Javitt, D. C., Sershen, H., Hashim, A., and Lajtha, A. (1997). Reversal of phencyclidine-induced hyperactivity by glycine and the glycine uptake inhibitor glycylododecylamide. *Neuropsychopharmacology* **17**, 202–204.
- Javitt, D. C., Balla, A., Sershen, H., and Lajtha, A. (1999). A.E. Bennett Research Award. Reversal of phencyclidine-induced effects by glycine and glycine transport inhibitors. *Biol. Psychiatry* **45**, 668–679.
- Javitt, D. C., Sershen, H., Hashim, A., and Lajtha, A. (2000). Inhibition of striatal dopamine release by glycine and glycylododecylamide. *Brain Res. Bull.* **52**, 213–216.

- Javitt, D. C., Silipo, G., Cienfuegos, A., Shelley, A. M., Bark, N., Park, M., Lindenmayer, J. P., Suckow, R., and Zukin, S. R. (2001). Adjunctive high-dose glycine in the treatment of schizophrenia. *Int. J. Neuropsychopharmacol.* **4**, 385–392.
- Javitt, D. C., Balla, A., Burch, S., Suckow, R., Xie, S., and Sershen, H. (2004). Reversal of phencyclidine-induced dopaminergic dysregulation by N-methyl-D-aspartate receptor/glycine-site agonists. *Neuropsychopharmacology* **29**, 300–307.
- Javitt, D. C., Duncan, L., Balla, A., and Sershen, H. (2005a). Inhibition of system A-mediated glycine transport in cortical synaptosomes by therapeutic concentrations of clozapine: Implications for mechanisms of action. *Mol. Psychiatry* **10**, 275–287.
- Javitt, D. C., Hashim, A., and Sershen, H. (2005b). Modulation of striatal dopamine release by glycine transport inhibitors. *Neuropsychopharmacology* **30**, 649–656.
- Jentsch, J. D., and Roth, R. H. (1999). The neuropsychopharmacology of phencyclidine: From NMDA receptor hypofunction to the dopamine hypothesis of schizophrenia. *Neuropsychopharmacology* **20**, 201–225.
- Jentsch, J. D., Redmond, D. E., Jr., Elsworth, J. D., Taylor, J. R., Youngren, K. D., and Roth, R. H. (1997). Enduring cognitive deficits and cortical dopamine dysfunction in monkeys after long-term administration of phencyclidine. *Science* **277**, 953–955.
- Jentsch, J. D., Elsworth, J. D., Taylor, J. R., Redmond, D. E., Jr., and Roth, R. H. (1998). Dysregulation of mesoprefrontal dopamine neurons induced by acute and repeated phencyclidine administration in the nonhuman primate: Implications for schizophrenia. *Adv. Pharmacol.* **42**, 810–814.
- Johnson, S. A., Luu, N. T., Herbst, T. A., Knapp, R., Lutz, D., Arai, A., Rogers, G. A., and Lynch, G. (1999). Synergistic interactions between ampakines and antipsychotic drugs. *J. Pharmacol. Exp. Ther.* **289**, 392–397.
- Jourdi, H., Lu, X., Yanagihara, T., Lauterborn, J. C., Bi, X., Gall, C. M., and Baudry, M. (2005). Prolonged positive modulation of alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors induces calpain-mediated PSD-95/Dlg/ZO-1 protein degradation and AMPA receptor down-regulation in cultured hippocampal slices. *J. Pharmacol. Exp. Ther.* **314**, 16–26.
- Kapur, S., and Remington, G. (2001). Dopamine D(2) receptors and their role in atypical antipsychotic action: Still necessary and may even be sufficient. *Biol. Psychiatry* **50**, 873–883.
- Karlsson, P., Farde, L., Halldin, C., and Sedvall, G. (2002). PET study of D(1) dopamine receptor binding in neuroleptic-naïve patients with schizophrenia. *Am. J. Psychiatry* **159**, 761–767.
- Kay, S. R., Fiszbein, A., and Opler, L. A. (1987). The positive and negative syndrome scale (PANSS) for schizophrenia. *Schizophr. Bull.* **13**, 261–276.
- Keefe, R. S., Bilder, R. M., Harvey, P. D., Davis, S. M., Palmer, B. W., Gold, J. M., Meltzer, H. Y., Green, M. F., Miller del, D., Canive, J. M., Adler, L. W., Manschreck, T. C., et al. (2006). Baseline neurocognitive deficits in the CATIE schizophrenia trial. *Neuropsychopharmacology* **31**, 2033–2046.
- Kegeles, L. S., Abi-Dargham, A., Zea-Ponce, Y., Rodenhiser-Hill, J., Mann, J. J., Van Heertum, R. L., Cooper, T. B., Carlsson, A., and Laruelle, M. (2000). Modulation of amphetamine-induced striatal dopamine release by ketamine in humans: Implications for schizophrenia. *Biol. Psychiatry* **48**, 627–640.
- Kegeles, L. S., Martinez, D., Kochan, L. D., Hwang, D. R., Huang, Y., Mawlawi, O., Suckow, R. F., Van Heertum, R. L., and Laruelle, M. (2002). NMDA antagonist effects on striatal dopamine release: Positron emission tomography studies in humans. *Synapse* **43**, 19–29.
- Kinney, G. G., Burno, M., Campbell, U. C., Hernandez, L. M., Rodriguez, D., Bristow, L. J., and Conn, P. J. (2003a). Metabotropic glutamate subtype 5 receptors modulate locomotor activity and sensorimotor gating in rodents. *J. Pharmacol. Exp. Ther.* **306**, 116–123.
- Kinney, G. G., Sur, C., Burno, M., Mallorga, P. J., Williams, J. B., Figueroa, D. J., Wittmann, M., Lemaire, W., and Conn, P. J. (2003b). The glycine transporter type 1 inhibitor

- N-[3-(4'-fluorophenyl)-3-(4'-phenylphenoxy)propyl]sarcosine potentiates NMDA receptor-mediated responses *in vivo* and produces an antipsychotic profile in rodent behavior. *J. Neurosci.* **23**, 7586–7591.
- Kinney, J. W., Davis, C. N., Tabarean, I., Conti, B., Bartfai, T., and Behrens, M. M. (2006). A specific role for NR2A-containing NMDA receptors in the maintenance of parvalbumin and GAD67 immunoreactivity in cultured interneurons. *J. Neurosci.* **26**, 1604–1615.
- Korostishevsky, M., Kaganovich, M., Cholostoy, A., Ashkenazi, M., Ratner, Y., Dahary, D., Bernstein, J., Bening-Abu-Shach, U., Ben-Asher, E., Lancet, D., Ritsner, M., and Navon, R. (2004). Is the G72/G30 locus associated with schizophrenia? Single nucleotide polymorphisms, haplotypes, and gene expression analysis. *Biol. Psychiatry* **56**, 169–176.
- Kotter, R. (1994). Postsynaptic integration of glutamatergic and dopaminergic signals in the striatum. *Prog. Neurobiol.* **44**, 163–196.
- Kremer, I., Vass, A., Gorelik, I., Bar, G., Blararu, M., Javitt, D. C., and Heresco-Levy, U. (2004). Placebo-controlled trial of lamotrigine added to conventional and atypical antipsychotics in schizophrenia. *Biol. Psychiatry* **56**, 441–446.
- Krystal, J. H., Karper, L. P., Seibyl, J. P., Freeman, G. K., Delaney, R., Bremner, J. D., Heninger, G. R., Bowers, M. B., Jr., and Charney, D. S. (1994). Subanesthetic effects of the noncompetitive NMDA antagonist, ketamine, in humans. Psychotomimetic, perceptual, cognitive, and neuroendocrine responses. *Arch. Gen. Psychiatry* **51**, 199–214.
- Krystal, J. H., Abi-Saab, W., Perry, E., D'Souza, D. C., Liu, N., Gueorguieva, R., McDougall, L., Hunsberger, T., Belger, A., Levine, L., and Breier, A. (2004). Preliminary evidence of attenuation of the disruptive effects of the NMDA glutamate receptor antagonist, ketamine, on working memory by pretreatment with the group II metabotropic glutamate receptor agonist, LY354740, in healthy human subjects. *Psychopharmacology (Berl.)* **179**(1), 303–309.
- Krystal, J. H., Perry, E. B., Jr., Gueorguieva, R., Belger, A., Madonick, S. H., Abi-Dargham, A., Cooper, T. B., Macdougall, L., Abi-Saab, W., and D'Souza, D. C. (2005). Comparative and interactive human psychopharmacologic effects of ketamine and amphetamine: Implications for glutamatergic and dopaminergic model psychoses and cognitive function. *Arch. Gen. Psychiatry* **62**, 985–994.
- Kulagina, N. V., Zigmond, M. J., and Michael, A. C. (2001). Glutamate regulates the spontaneous and evoked release of dopamine in the rat striatum. *Neuroscience* **102**, 121–128.
- Lachman, H. M., Nolan, K. A., Mohr, P., Saito, T., and Volavka, J. (1998). Association between catechol O-methyltransferase genotype and violence in schizophrenia and schizoaffective disorder. *Am. J. Psychiatry* **155**, 835–837.
- Lahti, A. C., Weiler, M. A., Tamara Michaelidis, B. A., Parwani, A., and Tamminga, C. A. (2001). Effects of ketamine in normal and schizophrenic volunteers. *Neuropsychopharmacology* **25**, 455–467.
- Lane, H. Y., Huang, C. L., Wu, P. L., Liu, Y. C., Chang, Y. C., Lin, P. Y., Chen, P. W., and Tsai, G. (2006). Glycine transporter I inhibitor, N-methylglycine (sarcosine), added to clozapine for the treatment of schizophrenia. *Biol. Psychiatry* **60**(6), 645–649.
- Laruelle, M. (1998). Imaging dopamine transmission in schizophrenia. A review and meta-analysis. *Q. J. Nucl. Med.* **42**, 211–221.
- Laruelle, M., Abi-Dargham, A., van Dyck, C. H., Rosenblatt, W., Zea-Ponce, Y., Zoghbi, S. S., Baldwin, R. M., Charney, D. S., Hoffer, P. B., Kung, H. F., and Innis, R. B. (1995). SPECT imaging of striatal dopamine release after amphetamine challenge. *J. Nucl. Med.* **36**, 1182–1190.
- Laruelle, M., Abi-Dargham, A., van Dyck, C. H., Gil, R., D'Souza, C. D., Erds, J., McCance, E., Rosenblatt, W., Fingado, C., Zoghbi, S. S., Baldwin, R. M., Seibyl, J. P., *et al.* (1996). Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug-free schizophrenic subjects. *Proc. Natl. Acad. Sci. USA* **93**, 9235–9240.
- Laruelle, M., Abi-Dargham, A., Gil, R., Kegeles, L., and Innis, R. (1999). Increased dopamine transmission in schizophrenia: Relationship to illness phases. *Biol. Psychiatry* **46**, 56–72.

- Le Pen, G., Kew, J., Alberati, D., Borroni, E., Heitz, M. P., and Moreau, J. L. (2003). Prepulse inhibition deficits of the startle reflex in neonatal ventral hippocampal-lesioned rats: Reversal by glycine and a glycine transporter inhibitor. *Biol. Psychiatry* **54**, 1162–1170.
- Leveque, J. C., Macias, W., Rajadhyaksha, A., Carlson, R. R., Barczak, A., Kang, S., Li, X. M., Coyle, J. T., Haganir, R. L., Heckers, S., and Konradi, C. (2000). Intracellular modulation of NMDA receptor function by antipsychotic drugs. *J. Neurosci.* **20**, 4011–4020.
- Levine, J., Stahl, Z., Sela, B. A., Gavendo, S., Ruderman, V., and Belmaker, R. H. (2002). Elevated homocysteine levels in young male patients with schizophrenia. *Am. J. Psychiatry* **159**, 1790–1792.
- Lieberman, J. A., Perkins, D., Belger, A., Chakos, M., Jarskog, F., Boteva, K., and Gilmore, J. (2001). The early stages of schizophrenia: Speculations on pathogenesis, pathophysiology, and therapeutic approaches. *Biol. Psychiatry* **50**, 884–897.
- Linn, G. S., O'Keefe, R. T., Schroeder, C. E., Lifshitz, K., and Javitt, D. C. (1999). Behavioral effects of chronic phencyclidine in monkeys. *Neuroreport* **10**, 2789–2793.
- Lipina, T., Labrie, V., Weiner, I., and Roder, J. (2005). Modulators of the glycine site on NMDA receptors, D-serine and ALX 5407, display similar beneficial effects to clozapine in mouse models of schizophrenia. *Psychopharmacology (Berl.)* **179**, 54–67.
- Liu, Y. L., Fann, C. S., Liu, C. M., Chang, C. C., Wu, J. Y., Hung, S. I., Liu, S. K., Hsieh, M. H., Hwang, T. J., Chan, H. Y., Chen, J. J., Faraone, S. V., *et al.* (2006). No association of G72 and D-amino acid oxidase genes with schizophrenia. *Schizophr. Res.* **87**(1–3), 15–20.
- Lorrain, D. S., Bacceti, C. S., Bristow, L. J., Anderson, J. J., and Varney, M. A. (2003). Effects of ketamine and N-methyl-D-aspartate on glutamate and dopamine release in the rat prefrontal cortex: Modulation by a group II selective metabotropic glutamate receptor agonist LY379268. *Neuroscience* **117**, 697–706.
- Luby, E. D., Gottlieb, J. S., Cohen, B. D., Rosenbaum, G., and Domino, E. F. (1962). Model psychoses and schizophrenia. *Am. J. Psychiatry* **119**, 61–67.
- MacDonald, A. W., III, Carter, C. S., Kerns, J. G., Ursu, S., Barch, D. M., Holmes, A. J., Stenger, V. A., and Cohen, J. D. (2005). Specificity of prefrontal dysfunction and context processing deficits to schizophrenia in never-medicated patients with first-episode psychosis. *Am. J. Psychiatry* **162**, 475–484.
- Maeda, J., Suhara, T., Okauchi, T., and Semba, J. (2003). Different roles of group I and group II metabotropic glutamate receptors on phencyclidine-induced dopamine release in the rat prefrontal cortex. *Neurosci. Lett.* **336**, 171–174.
- Makino, C., Fujii, Y., Kikuta, R., Hirata, N., Tani, A., Shibata, A., Ninomiya, H., Tashiro, N., Shibata, H., and Fukumaki, Y. (2003). Positive association of the AMPA receptor subunit GluR4 gene (GRIA4) haplotype with schizophrenia: Linkage disequilibrium mapping using SNPs evenly distributed across the gene region. *Am. J. Med. Genet.* **116B**, 17–22.
- Malhotra, A. K., Pinals, D. A., Weingartner, H., Sirocco, K., Missar, C. D., Pickar, D., and Breier, A. (1996). NMDA receptor function and human cognition: The effects of ketamine in healthy volunteers. *Neuropsychopharmacology* **14**, 301–307.
- Malhotra, A. K., Adler, C. M., Kennison, S. D., Elman, I., Pickar, D., and Breier, A. (1997a). Clozapine blunts N-methyl-D-aspartate antagonist-induced psychosis: A study with ketamine. *Biol. Psychiatry* **42**, 664–668.
- Malhotra, A. K., Pinals, D. A., Adler, C. M., Elman, I., Clifton, A., Pickar, D., and Breier, A. (1997b). Ketamine-induced exacerbation of psychotic symptoms and cognitive impairment in neuroleptic-free schizophrenics. *Neuropsychopharmacology* **17**, 141–150.
- Marengo, S., Egan, M. F., Goldberg, T. E., Knable, M. B., McClure, R. K., Winterer, G., and Weinberger, D. R. (2002). Preliminary experience with an ampakine (CX516) as a single agent for the treatment of schizophrenia: A case series. *Schizophr. Res.* **57**, 221–226.
- Martina, M., Gorfinkel, Y., Halman, S., Lowe, J. A., Periyalwar, P., Schmidt, C. J., and Bergeron, R. (2004). Glycine transporter type 1 blockade changes NMDA receptor-mediated responses and

- LTP in hippocampal CA1 pyramidal cells by altering extracellular glycine levels. *J. Physiol.* **557**, 489–500.
- Martina, M., B-Turcotte, M., Tsai, G., Tiberi, M., Coyle, J. T., and Bergeron, R. (2005). Reduced glycine transporter type 1 expression leads to major changes in glutamatergic neurotransmission of CA1 hippocampal neurones in mice. *J. Physiol.* **563**, 777–793.
- McGhie, A., and Chapman, J. (1961). Disorders of attention and perception in early schizophrenia. *Br. J. Med. Psychol.* **34**, 103–116.
- Melone, M., Bragina, L., and Conti, F. (2003). Clozapine-induced reduction of glutamate transport in the frontal cortex is not mediated by GLAST and EAAC1. *Mol. Psychiatry* **8**, 12–13.
- Meyer-Lindenberg, A., Nichols, T., Callicott, J. H., Ding, J., Kolachana, B., Buckholtz, J., Mattay, V. S., Egan, M., and Weinberger, D. R. (2006). Impact of complex genetic variation in COMT on human brain function. *Mol. Psychiatry* **11**, 867–877.
- Miller, D. W., and Abercrombie, E. D. (1996). Effects of MK-801 on spontaneous and amphetamine-stimulated dopamine release in striatum measured with *in vivo* microdialysis in awake rats. *Brain Res. Bull.* **40**, 57–62.
- Missale, C., Fiorentini, C., Busi, C., Collo, G., and Spano, P. F. (2006). The NMDA/D1 receptor complex as a new target in drug development. *Curr. Top. Med. Chem.* **6**, 801–808.
- Miyamoto, E. (2006). Molecular mechanism of neuronal plasticity: Induction and maintenance of long-term potentiation in the hippocampus. *J. Pharmacol. Sci.* **100**, 433–442.
- Moghaddam, B. (2003). Bringing order to the glutamate chaos in schizophrenia. *Neuron* **40**, 881–884.
- Moghaddam, B., and Adams, B. W. (1998). Reversal of phencyclidine effects by a group II metabotropic glutamate receptor agonist in rats. *Science* **281**, 1349–1352.
- Morgan, C. J., Mofeez, A., Brandner, B., Bromley, L., and Curran, H. V. (2004a). Acute effects of ketamine on memory systems and psychotic symptoms in healthy volunteers. *Neuropsychopharmacology* **29**, 208–218.
- Morgan, C. J., Mofeez, A., Brandner, B., Bromley, L., and Curran, H. V. (2004b). Ketamine impairs response inhibition and is positively reinforcing in healthy volunteers: A dose-response study. *Psychopharmacology (Berl.)* **172**, 298–308.
- Muly, E. C., Maddox, M., and Smith, Y. (2003). Distribution of mGluR1alpha and mGluR5 immunolabeling in primate prefrontal cortex. *J. Comp. Neurol.* **467**, 521–535.
- Narendran, R., Frankle, W. G., Keefe, R., Gil, R., Martinez, D., Slistein, M., Kegeles, L. S., Talbot, P. S., Huang, Y., Hwang, D. R., Khenissi, L., Cooper, T. B., *et al.* (2005). Altered prefrontal dopaminergic function in chronic recreational ketamine users. *Am. J. Psychiatry* **162**, 2352–2359.
- Newcomer, J. W., Farber, N. B., Jevtic-Todorovic, V., Selke, G., Melson, A. K., Hershey, T., Craft, S., and Olney, J. W. (1999). Ketamine-induced NMDA receptor hypofunction as a model of memory impairment and psychosis. *Neuropsychopharmacology* **20**, 106–118.
- Nicola, S. M., Surmeier, J., and Malenka, R. C. (2000). Dopaminergic modulation of neuronal excitability in the striatum and nucleus accumbens. *Annu. Rev. Neurosci.* **23**, 185–215.
- Okubo, Y., Suhara, T., Suzuki, K., Kobayashi, K., Inoue, O., Terasaki, O., Someya, Y., Sassa, T., Sudo, Y., Matsushima, E., Iyo, M., Tateno, Y., *et al.* (1997). Decreased prefrontal dopamine D1 receptors in schizophrenia revealed by PET. *Nature* **385**, 634–636.
- Olney, J. W., Newcomer, J. W., and Farber, N. B. (1999). NMDA receptor hypofunction model of schizophrenia. *J. Psychiatr. Res.* **33**, 523–533.
- Onn, S. P., West, A. R., and Grace, A. A. (2000). Dopamine-mediated regulation of striatal neuronal and network interactions. *Trends Neurosci.* **23**, S48–S56.
- Overall, J. E., and Gorham, D. E. (1961). The brief psychiatric rating scale. *Psychol. Rep.* **10**, 799–812.
- Oye, I., Paulsen, O., and Mørset, A. (1992). Effects of ketamine on sensory perception: Evidence for a role of N-methyl-D-aspartate receptors. *J. Pharmacol. Exp. Ther.* **260**, 1209–1213.
- Paquet, M., and Smith, Y. (2003). Group I metabotropic glutamate receptors in the monkey striatum: Subsynaptic association with glutamatergic and dopaminergic afferents. *J. Neurosci.* **23**, 7659–7669.

- Parwani, A., Weiler, M. A., Blaxton, T. A., Warfel, D., Hardin, M., Frey, K., and Lahti, A. C. (2005). The effects of a subanesthetic dose of ketamine on verbal memory in normal volunteers. *Psychopharmacology (Berl.)* **183**, 265–274.
- Peris, J., Dwoskin, L. P., and Zahniser, N. R. (1988). Biphasic modulation of evoked [3H]D-aspartate release by D-2 dopamine receptors in rat striatal slices. *Synapse* **2**, 450–456.
- Pietraszek, M., Golembiowska, K., Bijak, M., Ossowska, K., and Wolfarth, S. (2002). Differential effects of chronic haloperidol and clozapine administration on glutamatergic transmission in the fronto-parietal cortex in rats: Microdialysis and electrophysiological studies. *Naunyn-Schmiedeberg Arch. Pharmacol.* **366**, 417–424.
- Pilc, A., Klodzinska, A., Branski, P., Nowak, G., Palucha, A., Szewczyk, B., Tatarczynska, E., Chojnacka-Wojcik, E., and Wieronska, J. M. (2002). Multiple MPEP administrations evoke anxiolytic- and antidepressant-like effects in rats. *Neuropharmacology* **43**, 181–187.
- Pin, J. P., and Acher, F. (2002). The metabotropic glutamate receptors: Structure, activation mechanism and pharmacology. *Curr. Drug Target CNS Neurol. Disord.* **1**, 297–317.
- Rabinowicz, E. F., Silipo, G., Goldman, R., and Javitt, D. C. (2000). Auditory sensory dysfunction in schizophrenia: Imprecision or distractibility? *Arch. Gen. Psychiatry* **57**, 1149–1155.
- Radant, A. D., Bowdle, T. A., Cowley, D. S., Kharasch, E. D., and Roy-Byrne, P. P. (1998). Does ketamine-mediated N-methyl-D-aspartate receptor antagonism cause schizophrenia-like oculomotor abnormalities? *Neuropsychopharmacology* **19**, 434–444.
- Reynolds, G. P., Abdul-Monim, Z., Neill, J. C., and Zhang, Z. J. (2004). Calcium binding protein markers of GABA deficits in schizophrenia: Postmortem studies and animal models. *Neurotox. Res.* **6**, 57–61.
- Reynolds, L. M., Cochran, S. M., Morris, B. J., Pratt, J. A., and Reynolds, G. P. (2005). Chronic phencyclidine administration induces schizophrenia-like changes in N-acetylaspartate and N-acetylaspartylglutamate in rat brain. *Schizophr. Res.* **73**, 147–152.
- Riley, B. P., Tahir, E., Rajagopalan, S., Mogudi-Carter, M., Faure, S., Weissenbach, J., Jenkins, T., and Williamson, R. (1997). A linkage study of the N-methyl-D-aspartate receptor subunit gene loci and schizophrenia in southern African Bantu-speaking families. *Psychiatr. Genet.* **7**, 57–74.
- Rowland, L. M., Astur, R. S., Jung, R. E., Bustillo, J. R., Lauriello, J., and Yeo, R. A. (2005). Selective cognitive impairments associated with NMDA receptor blockade in humans. *Neuropsychopharmacology* **30**, 633–639.
- Saykin, A. J., Gur, R. C., Gur, R. E., Mozley, P. D., Mozley, L. H., Resnick, S. M., Kester, D. B., and Stafiniak, P. (1991). Neuropsychological function in schizophrenia: Selective impairment in memory and learning. *Arch. Gen. Psychiatry* **48**, 618–624.
- Schoepp, D. D., and Marek, G. J. (2002). Preclinical pharmacology of mGlu2/3 receptor agonists: Novel agents for schizophrenia? *Curr. Drug Target CNS Neurol. Disord.* **1**, 215–225.
- Schumacher, J., Jamra, R. A., Freudenberger, J., Becker, T., Ohlraun, S., Otte, A. C., Tullius, M., Kovalenko, S., Bogaert, A. V., Maier, W., Rietschel, M., Propping, P., et al. (2004). Examination of G72 and D-amino-acid oxidase as genetic risk factors for schizophrenia and bipolar affective disorder. *Mol. Psychiatry* **9**, 203–207.
- Schwarcz, R., Rassoulpour, A., Wu, H. Q., Medoff, D., Tamminga, C. A., and Roberts, R. C. (2001). Increased cortical kynurenate content in schizophrenia. *Biol. Psychiatry* **50**, 521–530.
- Shurman, B., Horan, W. P., and Nuechterlein, K. H. (2005). Schizophrenia patients demonstrate a distinctive pattern of decision-making impairment on the Iowa Gambling Task. *Schizophr. Res.* **72**, 215–224.
- Silberberg, G., Darvasi, A., Pinkas-Kramarski, R., and Navon, R. (2006). The involvement of ErbB4 with schizophrenia: Association and expression studies. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **141**, 142–148.
- Smith, A. D., and Bolam, J. P. (1990). The neural network of the basal ganglia as revealed by the study of synaptic connections of identified neurones. *Trends Neurosci.* **13**, 259–265.

- Smith, G. S., Schloesser, R., Brodie, J. D., Dewey, S. L., Logan, J., Vitkun, S. A., Simkowitz, P., Hurley, A., Cooper, T., Volkow, N. D., and Cancro, R. (1998). Glutamate modulation of dopamine measured *in vivo* with positron emission tomography (PET) and 11C-raclopride in normal human subjects. *Neuropsychopharmacology* **18**, 18–25.
- Starr, M. S. (1995). Glutamate/dopamine D1/D2 balance in the basal ganglia and its relevance to Parkinson's disease. *Synapse* **19**, 264–293.
- Stefansson, H., Sigurdsson, E., Steinthorsdottir, V., Bjornsdottir, S., Sigmundsson, T., Ghosh, S., Brynjolfsson, J., Gunnarsdottir, S., Ivarsson, O., Chou, T. T., Hjaltason, O., Birgisdottir, B., *et al.* (2002). Neuregulin 1 and susceptibility to schizophrenia. *Am. J. Hum. Genet.* **71**, 877–892.
- Steullet, P., Neijt, H. C., Cuenod, M., and Do, K. Q. (2006). Synaptic plasticity impairment and hypofunction of NMDA receptors induced by glutathione deficit: Relevance to schizophrenia. *Neuroscience* **137**, 807–819.
- Strous, R. D., Cowan, N., Ritter, W., and Javitt, D. C. (1995). Auditory sensory (“echoic”) memory dysfunction in schizophrenia. *Am. J. Psychiatry* **152**, 1517–1519.
- Sumiyoshi, T., Anil, A. E., Jin, D., Jayathilake, K., Lee, M., and Meltzer, H. Y. (2004). Plasma glycine and serine levels in schizophrenia compared to normal controls and major depression: Relation to negative symptoms. *Int. J. Neuropsychopharmacol.* **7**, 1–8.
- Susser, E., Brown, A. S., Klonowski, E., Allen, R. H., and Lindenbaum, J. (1998). Schizophrenia and impaired homocysteine metabolism: A possible association. *Biol. Psychiatry* **44**, 141–143.
- Swanson, C. J., and Schoepp, D. D. (2002). The group II metabotropic glutamate receptor agonist (–)-2-oxa-4-aminobicyclo[3.1.0]hexane-4,6-dicarboxylate (LY379268) and clozapine reverse phencyclidine-induced behaviors in monoamine-depleted rats. *J. Pharmacol. Exp. Ther.* **303**, 919–927.
- Takahata, R., and Moghaddam, B. (2000). Target-specific glutamatergic regulation of dopamine neurons in the ventral tegmental area. *J. Neurochem.* **75**, 1775–1778.
- Tamminga, C. A., Holcomb, H. H., Gao, X., and Lahti, A. C. (1995). Glutamate pharmacology and the treatment of schizophrenia: Current status and future directions. *Int. Clin. Psychopharmacol.* **10**(Suppl. 3), 29–37.
- Tanaka, H., Grooms, S. Y., Bennett, M. V., and Zukin, R. S. (2000). The AMPAR subunit GluR2: Still front and center-stage. *Brain Res.* **886**, 190–207.
- Tiihonen, J., Hallikainen, T., Ryyanen, O. P., Repo-Tiihonen, E., Kotilainen, I., Eronen, M., Toivonen, P., Wahlbeck, K., and Putkonen, A. (2003). Lamotrigine in treatment-resistant schizophrenia: A randomized placebo-controlled crossover trial. *Biol. Psychiatry* **54**, 1241–1248.
- Torrey, E. F., Barci, B. M., Webster, M. J., Bartko, J. J., Meador-Woodruff, J. H., and Knable, M. B. (2005). Neurochemical markers for schizophrenia, bipolar disorder, and major depression in postmortem brains. *Biol. Psychiatry* **57**, 252–260.
- Toth, E., Weiss, B., Banay-Schwartz, M., and Lajtha, A. (1986). Effect of glycine derivatives on behavioral changes induced by 3-mercaptopropionic acid or phencyclidine in mice. *Res. Comm. Psychol. Psychiat. Behav.* **11**, 1–9.
- Tsai, G. E., Passani, L. A., Slusher, B. S., Carter, R., Baer, L., Kleinman, J. E., and Coyle, J. T. (1995). Abnormal excitatory neurotransmitter metabolism in schizophrenic brains. *Arch. Gen. Psychiatry* **52**, 829–836.
- Tsai, G. E., Yang, P., Chung, L. C., Lange, N., and Coyle, J. T. (1998). D-serine added to antipsychotics for the treatment of schizophrenia. *Biol. Psychiatry* **44**, 1081–1089.
- Tsai, G. E., Yang, P., Chung, L. C., Tsai, I. C., Tsai, C. W., and Coyle, J. T. (1999). D-serine added to clozapine for the treatment of schizophrenia. *Am. J. Psychiatry* **156**, 1822–1825.
- Tsai, G. E., Lane, H. Y., Yang, P., Chong, M. Y., and Lange, N. (2004a). Glycine transporter I inhibitor, N-methylglycine (sarcosine), added to antipsychotics for the treatment of schizophrenia. *Biol. Psychiatry* **55**, 452–456.

- Tsai, G. E., Ralph-Williams, R. J., Martina, M., Bergeron, R., Berger-Sweeney, J., Dunham, K. S., Jiang, Z., Caine, S. B., and Coyle, J. T. (2004b). Gene knockout of glycine transporter 1: Characterization of the behavioral phenotype. *Proc. Natl. Acad. Sci. USA* **101**, 8485–8490.
- Tsai, G. E., Yang, P., Chang, Y. C., and Chong, M. Y. (2005). D-alanine added to antipsychotics for the treatment of schizophrenia. *Biol. Psychiatry* **59**(3), 230–234.
- Tseng, K. Y., and O'Donnell, P. (2004). Dopamine-glutamate interactions controlling prefrontal cortical pyramidal cell excitability involve multiple signaling mechanisms. *J. Neurosci.* **24**, 5131–5139.
- Tsukada, H., Nishiyama, S., Fukumoto, D., Sato, K., Kakiuchi, T., and Domino, E. F. (2005). Chronic NMDA antagonism impairs working memory, decreases extracellular dopamine, and increases D1 receptor binding in prefrontal cortex of conscious monkeys. *Neuropsychopharmacology* **30**, 1861–1869.
- Tuominen, H. J., Tiihonen, J., and Wahlbeck, K. (2005). Glutamatergic drugs for schizophrenia: A systematic review and meta-analysis. *Schizophr. Res.* **72**, 225–234.
- Umbricht, D., Schmid, L., Koller, R., Vollenweider, F. X., Hell, D., and Javitt, D. C. (2000). Ketamine-induced deficits in auditory and visual context-dependent processing in healthy volunteers: Implications for models of cognitive deficits in schizophrenia. *Arch. Gen. Psychiatry* **57**, 1139–1147.
- van Berckel, B. N., Evenblij, C. N., van Loon, B. J., Maas, M. F., van der Geld, M. A., Wynne, H. J., van Ree, J. M., and Kahn, R. S. (1999). D-cycloserine increases positive symptoms in chronic schizophrenic patients when administered in addition to antipsychotics: A double-blind, parallel, placebo-controlled study. *Neuropsychopharmacology* **21**, 203–210.
- van Berckel, B. N., Kegeles, L. S., Waterhouse, R., Guo, N., Hwang, D. R., Huang, Y., Narendran, R., Van Heertum, R., and Laruelle, M. (2006). Modulation of amphetamine-induced dopamine release by group II metabotropic glutamate receptor agonist LY354740 in non-human primates studied with positron emission tomography. *Neuropsychopharmacology* **31**, 967–977.
- Vollenweider, F. X., Vontobel, P., Oye, I., Hell, D., and Leenders, K. L. (2000). Effects of (S)-ketamine on striatal dopamine: A [¹¹C]raclopride PET study of a model psychosis in humans. *J. Psychiatr. Res.* **34**, 35–43.
- Weinberger, D. R. (2005). Genetic mechanisms of psychosis: *In vivo* and postmortem genomics. *Clin. Ther.* **27**(Suppl. A), S8–S15.
- Weinberger, D. R., Egan, M. F., Bertolino, A., Callicott, J. H., Mattay, V. S., Lipska, B. K., Berman, K. F., and Goldberg, T. E. (2001). Prefrontal neurons and the genetics of schizophrenia. *Biol. Psychiatry* **50**, 825–844.
- West, A. R., and Grace, A. A. (2002). Opposite influences of endogenous dopamine D1 and D2 receptor activation on activity states and electrophysiological properties of striatal neurons: Studies combining *in vivo* intracellular recordings and reverse microdialysis. *J. Neurosci.* **22**, 294–304.
- Wexler, B. E., Stevens, A. A., Bowers, A. A., Sernyak, M. J., and Goldman-Rakic, P. S. (1998). Word and tone working memory deficits in schizophrenia. *Arch. Gen. Psychiatry* **55**, 1093–1096.
- Woods, S. W., Thomas, L., Tully, E., Hawkins, K. A., Miller, T. J., Rosen, J. L., Pearson, G., and McGlashan, T. H. (2004). Effects of oral glycine in the schizophrenia prodrome. *Schiz. Res.* **70**, 79.
- Yee, B. K., Balic, E., Singer, P., Schwerdel, C., Grampp, T., Gabernet, L., Knuesel, I., Benke, D., Feldon, J., Mohler, H., and Boison, D. (2006). Disruption of glycine transporter 1 restricted to forebrain neurons is associated with a procognitive and antipsychotic phenotypic profile. *J. Neurosci.* **26**, 3169–3181.
- Yurgelun-Todd, D. A., Coyle, J. T., Gruber, S. A., Renshaw, P. F., Silveri, M. M., Amico, E., Cohen, B., and Goff, D. C. (2005). Functional magnetic resonance imaging studies of schizophrenic patients during word production: Effects of D-cycloserine. *Psychiatry Res.* **138**, 23–31.

DECIPHERING THE DISEASE PROCESS OF SCHIZOPHRENIA: THE CONTRIBUTION OF CORTICAL GABA NEURONS

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Schizophrenia is a devastating illness that is manifest through a variety of clinical signs and symptoms. Among these, impairments in certain critical cognitive functions, such as working memory, appear to represent the core features of the disorder. In this chapter, we review the evidence indicating that disturbances in neurotransmission by a subset of GABA neurons in the dorsolateral prefrontal cortex are commonly present in schizophrenia. Despite both pre- and postsynaptic compensatory responses, the resulting pathophysiological process, alterations in the perisomatic inhibitory regulation of pyramidal neurons, underlies a reduced capacity for the synchronization of neuronal activity at gamma frequencies that is required for working memory function. We also discuss several pathogenetic mechanisms that could rise to the alterations in GABA neurotransmission and consider the implication of these findings for therapeutic interventions to improve cognitive function in individuals with schizophrenia.

I. Working Memory Impairments: A Core Feature of Schizophrenia

Of the multiple clinical features of schizophrenia, disturbances in certain cognitive processes, such as impairments in attention, some types of memory, and executive function, appear to represent core features of the illness (Elvevåg

and Goldberg, 2000). Cognitive abnormalities have been observed during the premorbid and prodromal phases of the illness (Davidson *et al.*, 1999), at the initial onset of psychosis (Saykin *et al.*, 1994), and throughout the later stages of the illness (Heaton *et al.*, 1994). Perhaps most important, the degree of cognitive impairment is the best predictor of long-term outcome in individuals with schizophrenia (Green, 1996).

At least some of the critical cognitive deficits in schizophrenia reflect alterations in working memory (the ability to transiently maintain and manipulate a limited amount of information in order to guide thought or behavior) that is mediated by the circuitry of the dorsolateral prefrontal cortex (DLPFC) (Miller and Cohen, 2001). Many individuals with schizophrenia perform poorly on working memory tasks and exhibit altered activation of the DLPFC when attempting to perform such tasks (Callicott *et al.*, 2003; Perlstein *et al.*, 2001; Weinberger *et al.*, 1986). In contrast, these abnormalities have not been found in individuals with other psychotic disorders (MacDonald *et al.*, 2005) or major depression (Barch *et al.*, 2003). The altered activation of the DLPFC during working memory tasks predicts the severity of cognitive disorganization symptoms in subjects with schizophrenia (Perlstein *et al.*, 2001), and reduced working memory capacity has been suggested to be rate limiting in the performance of other cognitive tasks in schizophrenia (Silver *et al.*, 2003).

II. Working Memory Impairments and Altered GABA Neurotransmission in the DLPFC

Working memory depends on the coordinated and sustained firing of subsets of DLPFC pyramidal neurons between the temporary presentation of a stimulus cue and the later initiation of a behavioral response (Goldman-Rakic, 1995). Although other neurotransmitter systems are also involved, inhibitory signaling via γ -aminobutyric acid (GABA) appears to be critical for this pattern of activity in DLPFC pyramidal neurons during working memory. Fast-spiking GABA neurons in monkey DLPFC are active during the delay period of working memory tasks (Wilson *et al.*, 1994) and are necessary for task-related firing and the spatial tuning of pyramidal neurons during working memory (Rao *et al.*, 2000). In addition, the injection of GABA antagonists in the DLPFC disrupts working memory performance (Sawaguchi *et al.*, 1989). Thus, these findings suggest that disturbances in GABA neurotransmission in the DLPFC could contribute to the working memory impairments in schizophrenia.

Consistent with this hypothesis, markers of GABA neurotransmission are altered in the DLPFC of subjects with schizophrenia. For example, reduced expression of the mRNA for the 67-kDa isoform of glutamic acid decarboxylase (GAD₆₇), an enzyme that synthesizes GABA, (Akbarian *et al.*, 1995; Guidotti

et al., 2000; Hashimoto *et al.*, 2005b; Mirnics *et al.*, 2000; Vawter *et al.*, 2002; Volk *et al.*, 2000) is one of the most consistent findings in postmortem studies of individuals with schizophrenia (Torrey *et al.*, 2005). The only exception to this finding was reported in one cohort of elderly, chronically hospitalized individuals with schizophrenia (Dracheva *et al.*, 2004). Although less extensively studied, the deficit in GAD₆₇ mRNA appears to be accompanied by a corresponding decrease in the cognate protein (Guidotti *et al.*, 2000). In contrast, both the overall protein and mRNA expression levels (Guidotti *et al.*, 2000) of another synthesizing enzyme for GABA, GAD₆₅, and the density of GAD₆₅-immunoreactive axon terminals (Benes *et al.*, 2000) were reported to be unchanged in the DLPFC of subjects with schizophrenia. Interestingly, elimination of the GAD₆₅ gene in mice has a limited effect on cortical levels of GABA, whereas genetically engineered reductions in GAD₆₇ mRNA expression are associated with profound decreases in cortical GAD activity and GABA content (Asada *et al.*, 1997).

In the DLPFC of subjects with schizophrenia, GAD₆₇ mRNA expression is undetectable in a subpopulation (about 25–30%) of GABA neurons (Akbarian *et al.*, 1995; Volk *et al.*, 2000), whereas the majority of GABA neurons have expression levels of GAD₆₇ mRNA that do not differ from normal comparison subjects (Volk *et al.*, 2000). Furthermore, in the same individuals, the mRNA expression for the GABA membrane transporter (GAT1), a protein responsible for reuptake of released GABA into nerve terminals, is similarly decreased in a subpopulation of GABA neurons (Volk *et al.*, 2001). The affected GABA neurons appear to be principally located in cortical layers 2–5; neither GAD₆₇ nor GAT1 mRNA expression is altered in layer 6. Together, these findings suggest that both the synthesis and reuptake of GABA are greatly reduced in a subset of DLPFC inhibitory neurons in schizophrenia.

The affected GABA neurons include those that contain the calcium-binding protein parvalbumin (PV), which is present in ~25% of GABA neurons in the primate DLPFC (Condé *et al.*, 1994), as demonstrated by the decreased expression of PV mRNA in layers 3 and 4, but not in layers 2, 5, or 6, of the DLPFC in subjects with schizophrenia (Hashimoto *et al.*, 2003). However, in contrast to the findings for GAD₆₇ and GAT1 mRNAs, the density of neurons with detectable levels of PV mRNA was not changed in subjects with schizophrenia, but the expression level of PV mRNA per neuron was significantly decreased. In addition, within the same subjects, the expression level of PV mRNA per neuron was strongly correlated with the change in density of GAD₆₇ mRNA-positive neurons. Furthermore, dual label *in situ* hybridization studies demonstrated that approximately half of PV mRNA-positive neurons in subjects with schizophrenia lacked detectable levels of GAD₆₇ mRNA (Hashimoto *et al.*, 2003). Finally, these findings were consistent with the results of immunocytochemical studies that reported similar densities of PV-immunoreactive neurons in the DLPFC of normal comparison and schizophrenia subjects (Beasley *et al.*, 2002;

Woo *et al.*, 1997). Thus, PV-containing GABA neurons are not reduced in number in the DLPFC of subjects with schizophrenia, but they do exhibit reduced expression of several critical genes, indicating that they are present but functionally impaired. In contrast, the expression of the mRNA for calretinin, a calcium-binding protein present in ~50% of GABA neurons in the primate DLPFC (Condé *et al.*, 1994), is not altered in schizophrenia (Hashimoto *et al.*, 2003), nor is the density of calretinin-immunoreactive neurons (Daviss and Lewis, 1995) or axon terminals (Woo *et al.*, 1997).

In addition to their calcium-binding protein content, PV-containing neurons are distinguishable from other cortical GABA neurons (Fig. 1) by their firing patterns, preferred synaptic targets, and morphological features (Kawaguchi, 1995). For example, in macaque monkey DLPFC, cluster analysis of multiple physiological features revealed that PV-containing neurons are distinctly different from all other types of GABA neurons (Krimmer *et al.*, 2005; Zaitsev *et al.*, 2005). Furthermore, PV-positive neurons in the primate DLPFC are composed of two morphologically distinct subtypes (Condé *et al.*, 1994). Chandelier (or axoaxonic) neurons furnish a linear array of axon terminals (termed cartridges) that synapse exclusively on the axon initial segment of pyramidal neurons (Somogyi, 1977), whereas the axons of wide-arbor (basket) neurons have a much larger spread than those of chandelier cells and their axon terminals principally target the cell body and proximal dendrites of pyramidal neurons (Lewis and Lund, 1990). Both of these types of PV-containing neurons have indistinguishable fast-spiking, nonadapting patterns of firing (González-Burgos *et al.*, 2005). The proximity of the perisomatic inhibitory synapses formed by PV-containing chandelier and wide-arbor neurons to the site of action potential generation in pyramidal neurons suggests that these GABA neurons are specialized to powerfully regulate the output of pyramidal neurons. For example, during hippocampal oscillations *in vivo* chandelier cells exhibit maximal firing probability 180° out of phase with pyramidal neurons, indicating the ability of inhibitory inputs from chandelier neurons to facilitate the rhythmic entrainment of pyramidal cell discharge, time locking the activity of local populations of pyramidal cells to fire together (Klausberger *et al.*, 2003). In contrast, an *in vitro* study indicated that GABA inputs to the axon initial segment of pyramidal neurons could actually sufficiently depolarize pyramidal cells to fire action potentials under certain conditions (Szabadics *et al.*, 2006); however, whether this observation holds under *in vivo* conditions remains to be determined.

PV-containing GABA neurons also undergo marked and distinctive refinements in the monkey DLPFC during adolescence (Fig. 2) (Cruz *et al.*, 2003; Erickson and Lewis, 2002). These developmental trajectories suggest that PV-containing neurons contribute to the increased engagement of DLPFC circuitry in (Lewis, 1997), and improved performance of (Diamond, 2002; Luna *et al.*, 2004), working memory during adolescence, providing rationale for the hypothesis that alterations in PV-positive neurons contribute to working memory dysfunction in schizophrenia.

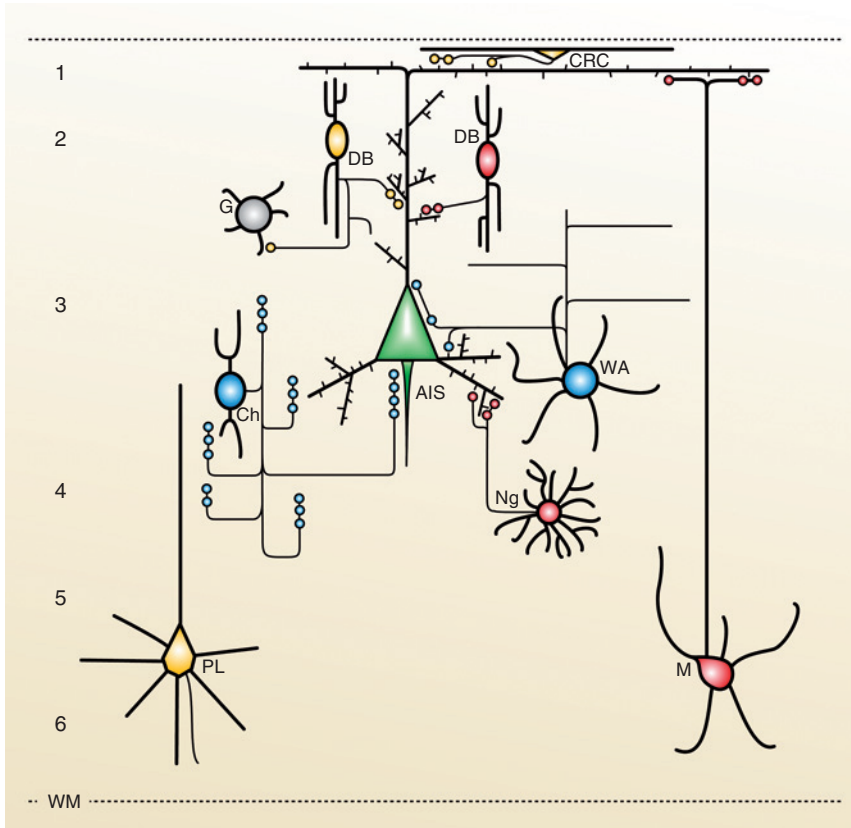


FIG. 1. Morphological and biochemical features of subclasses of cortical GABA neurons. This schematic diagram illustrates the calcium-binding protein content (blue, parvalbumin; red, calbindin; yellow, calretinin) and location of inhibitory synaptic inputs to a pyramidal neuron (green) of different morphological classes of cortical GABA neurons. The chandelier (Ch) and wide-arbor (WA) or basket neurons provide inhibitory input to the axon initial segment (AIS) or cell body and proximal dendrites, respectively, of pyramidal neurons. In contrast, the calbindin-containing double bouquet (DB), neurogliaform (Ng), and Martinotti (M) neurons tend to provide inhibitory inputs to the distal dendrites of pyramidal neurons. Finally, calretinin-containing neurons appear to target both pyramidal cell distal dendrites and other GABA (G) neurons. CRC, Cajal-Retzius cell; PL, pyramidal-like neuron. Reprinted from Lewis *et al.* (2005).

Furthermore, these developmental changes during adolescence could contribute to unmasking the consequences of inherited abnormalities in the regulation of GABA neurotransmission in schizophrenia and may help explain why certain life experiences during adolescence (e.g., stress or cannabis exposure) appear to increase the risk of the illness (Lewis and Levitt, 2002). Consistent with this

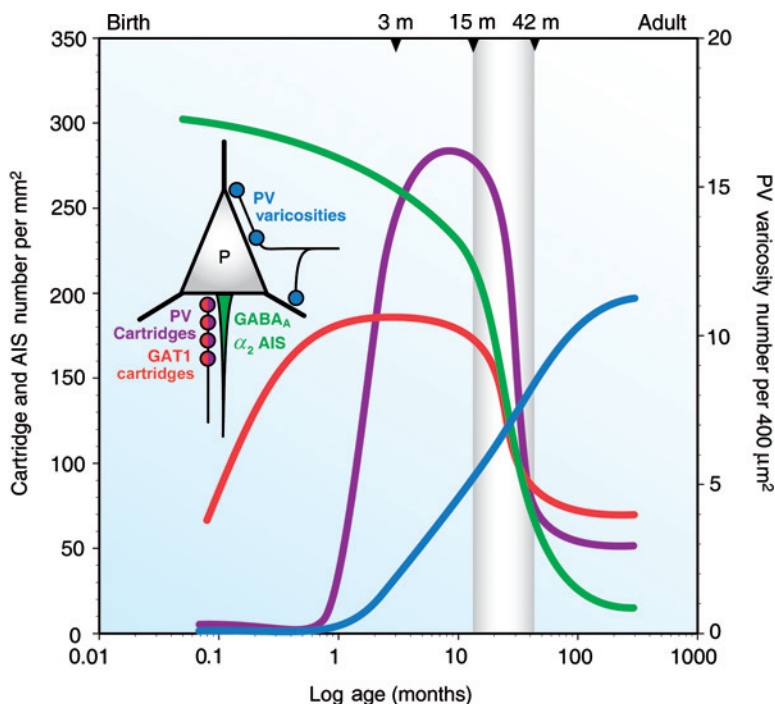


FIG. 2. Postnatal development of inputs from PV-containing GABA neurons to pyramidal neurons in monkey DLPFC. The axon terminals of chandelier neurons form vertical arrays of boutons (cartridges) that are immunoreactive for PV or GAT1 and that outline the axon initial segment of pyramidal neurons. Although the developmental time course differs somewhat for these two markers, the density of labeled cartridges is low in the DLPFC of newborn monkeys, increases to reach a peak prior to the onset of puberty, and then declines markedly during adolescence (shaded area between 15 and 42 months of age) to adult levels. These density changes in PV- and GAT1-IR cartridges appear to reflect developmental shifts in the concentration of these proteins. Interestingly, the peak and subsequent decline in the density of labeled cartridges occur prior to the age when the peak density of PV-immunoreactive varicosities, putative axon terminals from the wide-arbor (basket) class of PV-containing GABA neurons is achieved. Postsynaptically, the detectability of the α_2 subunit of the GABA_A receptor in pyramidal neuron axon initial segment (AIS) is high at birth, and then markedly declines during adolescence before stable adult levels are achieved. Reprinted from Lewis *et al.* (2005).

hypothesis, the density of chandelier neuron axon cartridges immunoreactive for GAT1 was significantly reduced in the DLPFC of subjects with schizophrenia (Woo *et al.*, 1998), with the effect most significant in the middle cortical layers (Pierri *et al.*, 1999). In contrast, measures of GAT1 immunoreactivity in other populations of axon terminals were unchanged (Woo *et al.*, 1998). Thus, in concert

with the observations of cell type-selective alterations in gene expression, these findings suggest that chandelier neurons in the DLPFC of subjects with schizophrenia express decreased levels of PV mRNA and undetectable levels of GAD₆₇ and GAT1 mRNAs, with the latter resulting in reduced GAT1 protein in chandelier neuron axon cartridges. The potential relevance of these findings for working memory dysfunction in schizophrenia is strengthened by the failure to find such disturbances in subjects with other psychiatric disorders or in monkeys exposed chronically to antipsychotic medications in a fashion that mimics the clinical treatment of schizophrenia (Hashimoto *et al.*, 2003; Pierri *et al.*, 1999; Volk *et al.*, 2000, 2001, 2002). Furthermore, GABA levels are not altered in the prefrontal cortex of unmedicated subjects with remitted major depressive disorder, indicating that if such changes are present in symptomatic depression (as suggested from studies of the visual cortex (Sanacora *et al.*, 1999)), they are not a persistent characteristic of this illness (Hasler *et al.*, 2005).

However, the pathophysiological significance of these changes depends on how they affect GABA neurotransmission at the synapse between the chandelier neuron and the pyramidal cell axon initial segment. Specifically, do these findings reflect deficient inhibition, resulting from a primary reduction in GABA synthesis, or excessive inhibition, secondary to a reduction in GABA reuptake? Interestingly, receptors containing GABA_A α_2 subunits are found principally at inhibitory synapses onto pyramidal neuron axon initial segments (Nusser *et al.*, 1996). In the DLPFC of subjects with schizophrenia, the density of pyramidal neuron axon initial segments immunoreactive for the GABA_A α_2 subunit is more than double that of control subjects (Volk *et al.*, 2002), apparently reflecting higher levels of α_2 subunits in the axon initial segment, since neither the density of pyramidal neurons (Pierri *et al.*, 2003) nor of their axon initial segments (Cruz *et al.*, 2004) is increased in these same subjects. Thus, in the DLPFC of subjects with schizophrenia, GABA_A receptors seem to be upregulated at pyramidal neuron axon initial segments in response to deficient GABA release from chandelier neuron axon terminals (Fig. 3).

Consistent with this interpretation, the reduced presynaptic levels of PV and GAT1 in chandelier cells appear to represent compensatory responses to a deficit in GABA release. For example, a reduction in PV would be expected to be associated with increased GABA release since, by buffering presynaptic Ca²⁺ transients, PV reduces the Ca²⁺-dependent facilitation of GABA release during periods of repetitive firing (Vreugdenhil *et al.*, 2003). Similarly, reduced levels of GAT1 would be expected to prolong the duration of inhibitory postsynaptic currents (IPSCs) when neighboring synapses are activated synchronously (Overstreet and Westbrook, 2003). Thus, the combination of reduced presynaptic levels of PV and GAT1 proteins in chandelier axon cartridges and the postsynaptic upregulation of GABA_A receptors at the axon initial segment of pyramidal neurons in the DLPFC of subjects with schizophrenia could act synergistically to increase

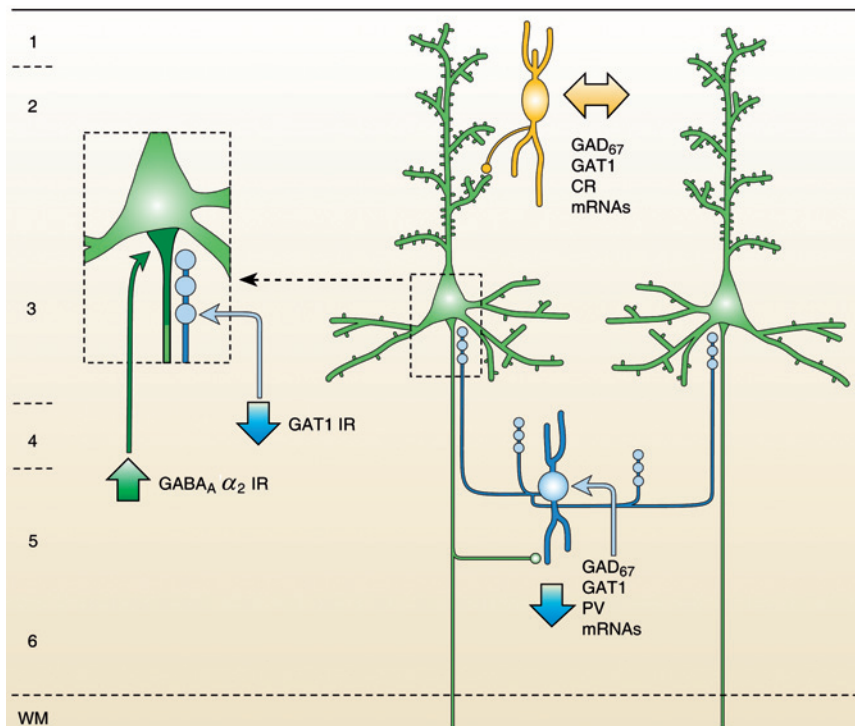


FIG. 3. Schematic summary of alterations in GABA circuitry in the DLPFC of subjects with schizophrenia. Reduced levels of gene expression in chandelier neurons (blue) are associated with a decrease in immunoreactivity (IR) for GAT1 in the axon cartridges of these neurons and an upregulation of GABA_A receptor α_2 subunit immunoreactivity in the postsynaptic axon initial segment of pyramidal neurons (green). In contrast, gene expression does not appear to be altered in the calretinin-containing subclass of GABA neurons (yellow). Reprinted from Lewis *et al.* (2005).

the efficacy of GABA neurotransmission at pyramidal cell axon initial segments during the types of repetitive activity that are associated with working memory. However, it appears that in schizophrenia these compensatory mechanisms are not adequate to overcome the effects of decreased GABA synthesis in chandelier neurons.

Similar pre- and postsynaptic alterations might also be present in the inputs of PV-containing wide-arbor neurons to the perisomatic region of pyramidal neurons. For example, the density of PV-immunoreactive puncta, possibly the axon terminals of wide-arbor neurons (Erickson and Lewis, 2002), is reduced in the middle layers, but not in the superficial layers, of the DLPFC of subjects with schizophrenia (Lewis *et al.*, 2001), paralleling the laminar pattern of decreased PV

mRNA expression in schizophrenia (Hashimoto *et al.*, 2003). Furthermore, the increased density of GABA_A receptors in the DLPFC of subjects with schizophrenia found in ligand-binding studies (Benes *et al.*, 1996; Hanada *et al.*, 1987) was most prominent at pyramidal neuron cell bodies (Benes *et al.*, 1996). Together, these data suggest that GABA_A receptors located at the soma and axon initial segments of pyramidal neurons are locally upregulated in schizophrenia in response to a reduction in perisomatic inhibitory input from chandelier and wide-arbor neurons.

However, abnormalities in PV neurons alone may not completely account for the deficits in expression of GAD₆₇ and GAT1 mRNAs since such changes were also observed in cortical layers 1 and 2, where relatively few PV-containing GABA neurons are located (Condé *et al.*, 1994) and where no changes in PV mRNA expression were found (Hashimoto *et al.*, 2003). Thus, other subpopulations of GABA neurons present in these layers, such as those that express the calcium-binding protein calbindin and/or the neuropeptides somatostatin or cholecystokinin, may also be altered in schizophrenia (Gabriel *et al.*, 1996; Virgo *et al.*, 1995). Indeed, we reported a marked decrease in mRNA levels for the somatostatin precursor protein in the DLPFC of subjects with schizophrenia (Hashimoto *et al.*, 2005a).

Are these abnormalities in GABA neurotransmission restricted to the DLPFC or representative of a disturbance distributed across other cortical regions that may contribute to other aspects of the clinical syndrome of schizophrenia? The anterior cingulate cortex, superior temporal gyrus, and hippocampal formation also appear to be sites of dysfunction in schizophrenia (Harrison and Lewis, 2003). Initial studies of the hippocampus reported a reduction in the density of nonpyramidal, putative GABA neurons (Benes *et al.*, 1998) and an increase in GABA_A receptor binding (Benes *et al.*, 1998) in schizophrenia. However, a study from the same research group did not detect a difference in the expression of either GAD₆₇ or GAD₆₅ mRNAs in the hippocampus of subjects with schizophrenia, although both transcripts were found to decrease in subjects with bipolar disorder (Heckers *et al.*, 2002). In the anterior cingulate cortex of subjects with schizophrenia, the densities of nonpyramidal neurons (Benes *et al.*, 1998) and of neurons immunoreactive for the calcium-binding protein calbindin (Cotter *et al.*, 2002) were reported to be reduced in layer 2, as was the density of GAD₆₇ mRNA-positive neurons (Woo *et al.*, 2004). Within the superior temporal gyrus, GAD₆₇ mRNA levels were found to be reduced (Impagnatiello *et al.*, 1998). In addition, the density of GAT1-immunoreactive axon cartridges was reduced in this region, although to a much lower extent than in the DLPFC of the same subjects with schizophrenia (Konopaske *et al.*, 2006). Thus, the available data suggest that alterations in GABA neurotransmission in schizophrenia may be a common feature across regions of the neocortex, but not of the hippocampus (Heckers *et al.*, 2002).

III. Potential Pathogenetic Mechanisms for Cell Type-Specific Alterations in GABA Neurons

Several different mechanisms have been suggested as the proximal cause of the alterations in PV-positive neurons in schizophrenia on the basis of correlated changes in individuals with the illness and evidence from animal models that these correlations represent cause and effect (Lewis *et al.*, 2005). Of these possibilities, alterations in N-methyl-D-aspartate (NMDA) receptor-mediated excitatory neurotransmission and deficits in neurotrophin signaling appear to have the strongest empirical basis.

A. REDUCED EXCITATORY DRIVE VIA NMDA RECEPTORS

The deficit in GAD₆₇ mRNA expression has been suggested to represent an activity-dependent change in response to reduced activity of excitatory circuits in the DLPFC (Akbarian *et al.*, 1995; Jones, 1997). One source of such excitatory inputs is the mediodorsal nucleus of the thalamus, the principal source of thalamic projections to the DLPFC. Interestingly, initial studies reported that the number of neurons in this nucleus was reduced in subjects with schizophrenia (Byne *et al.*, 2002; Pakkenberg, 1990, 1992; Popken *et al.*, 2000; Young *et al.*, 2000); however, studies have failed to confirm these observations (Cullen *et al.*, 2003; Danos *et al.*, 2003; Dorph-Petersen *et al.*, 2004; Nielsen *et al.*, 2004; Young *et al.*, 2000). Furthermore, experimental reductions in neuron number in the mediodorsal thalamus of rodents did not produce alterations in the expression of GAD₆₇ mRNA in the prefrontal cortex (Volk and Lewis, 2003). However, a range of other alterations, such as a decreased density of dendritic spines (a marker of excitatory synaptic inputs to pyramidal neurons) in the DLPFC of subjects with schizophrenia (Garey *et al.*, 1998; Glantz and Lewis, 2000), are consistent with reduced excitatory drive in DLPFC circuits, although the source of these altered inputs has yet to be determined.

One potential source of such altered inputs is the hippocampus. A variety of structural and functional abnormalities in the hippocampus have been observed in subjects with schizophrenia, and some of these appear to be correlated with alterations in the DLPFC (Bertolino *et al.*, 1996). To explore this association, investigators have employed a rodent model in which lesions of the ventral hippocampus are created neonatally (Lipska and Weinberger, 2000). In adulthood, these animals, in addition to mimicking a number of other phenotypic features of schizophrenia, show deficits in GAD₆₇ expression in the prefrontal cortex (Lipska *et al.*, 2003). However, whether the deficits in GAD₆₇ mRNA expression exhibit the cell type specificity, and are accompanied by the other changes in GABA markers present in schizophrenia, has not yet been examined.

These alterations in excitatory neurotransmission might differentially affect PV-, and not calretinin (CR)-containing GABA neurons because PV-containing

cells receive a larger complement of excitatory inputs (Lewis and Moghaddam, 2006). For example, in the rodent hippocampus, the total number of excitatory synapses onto PV-positive neurons is nearly an order of magnitude greater than the number onto CR-positive neurons (Gulyás *et al.*, 1999). Similarly, the density of asymmetric excitatory synapses on PV-positive dendrites in monkey DLPFC is significantly greater than on CR-positive dendrites (Melchitzky and Lewis, 2003). In addition, cell type differences have been reported for the NMDA receptor. For example, immunoreactivity for the NMDAR1 subunit was detected in the majority (50–90%) of PV-positive neurons, but in <10% of CR-positive neurons in monkey neocortex (Huntley *et al.*, 1994, 1997).

Convergent lines of evidence also indicate that PV-containing neurons are particularly sensitive to manipulation of excitatory signaling via NMDA receptors. First, administration of ketamine, an NMDA receptor antagonist, was associated with a decrease in the density of PV-immunoreactive neurons in the rodent hippocampus (Keilhoff *et al.*, 2004). Similarly, chronic exposure to PCP, another NMDA receptor antagonist, also resulted in decreased PV mRNA expression in the prefrontal cortex (Cochran *et al.*, 2003). Interestingly, in the latter study, the density of PV mRNA-positive neurons was unchanged following PCP, but the expression level of PV mRNA per neuron was decreased by 25%. These findings are strikingly similar to the pattern of PV mRNA expression changes observed in the DLPFC cortex of subjects with schizophrenia (Hashimoto *et al.*, 2003). Second, in cultures of mouse cortical neurons, ketamine induced a decrease in both PV and GAD₆₇ immunoreactivity specifically in PV interneurons, an effect that appeared to be mediated by NR2A-, and not NR2B-containing NMDA receptors (Kinney *et al.*, 2006). Consistent with the cell-type selectivity of the effect, the ratio of NR2A/NR2B was observed to be fivefold higher in PV-positive neurons than in pyramidal cells. Third, in living slice preparations from mouse entorhinal cortex, both the genetically engineered reduction in lysophosphatidic acid-1 (LPA-1) receptor and the acute blockade of NMDA receptors produced a laminar-specific decrease in induced gamma oscillations (see below), and in the LPA-1-deficient animals, these physiological changes were associated with an ~40% laminar-specific reduction in the number of GABA- and PV-containing neurons, without a change in the number of CR-positive neurons (Cunningham *et al.*, 2006). Together, these findings suggest that the alterations in GABA neurotransmission selective for PV-containing neurons in schizophrenia might be a downstream consequence of impaired NMDA receptor-mediated glutamatergic inputs.

B. REDUCED NEUROTROPHIN SIGNALING

Signaling by the neurotrophin, brain-derived neurotrophic factor (BDNF), through its receptor tyrosine kinase (TrkB) promotes the development of GABA neurons and induces the expression of GABA-related proteins, including GAD₆₇

and GAT1 (Marty *et al.*, 2000; Yamada *et al.*, 2002). In addition, TrkB is predominantly expressed by PV-containing, and not by calretinin-containing, GABA neurons, suggesting cell type-specificity of these effects (Cellerino *et al.*, 1996). Indeed, in mice, genetically engineered to overexpress BDNF, the development of cortical GABA neurons was accelerated and accompanied by a precocious increase in the number of neurons containing PV (Huang *et al.*, 1999). Thus, reduced BDNF-TrkB signaling might be an “upstream” event contributing to the altered expression of GABA-related genes in the DLPFC of subjects with schizophrenia. Consistent with this hypothesis, the mRNA and protein levels for both BDNF and TrkB reduced in the DLPFC of subjects with schizophrenia (Hashimoto *et al.*, 2005b; Weickert *et al.*, 2003, 2005). In contrast, levels of the mRNA encoding the receptor tyrosine kinase for neurotrophin-3, TrkC, were unchanged (Hashimoto *et al.*, 2005b).

Comparisons with the results of other studies indicate that these reduced mRNA levels represent alterations in gene expression and not a loss of DLPFC neurons in schizophrenia. For example, total neuron number is not altered in the prefrontal cortex of subjects with schizophrenia (Thune *et al.*, 2001). In addition, the density of DLPFC pyramidal neurons has been reported to be modestly increased across cortical layers (Selemon *et al.*, 1995) or to be unchanged in layer 3 (Pierri *et al.*, 2003) in schizophrenia. Similarly, the density of all nonpyramidal neurons has been reported to be slightly increased (Selemon *et al.*, 1995) or unchanged (Akbarian *et al.*, 1995), and as noted above, the densities of PV-immunoreactive (Beasley *et al.*, 2002; Woo *et al.*, 1997) and PV mRNA-positive neurons (Hashimoto *et al.*, 2003) were not altered in the DLPFC of subjects with schizophrenia. Because BDNF mRNA is expressed by pyramidal neurons, and because TrkB mRNA is expressed in both pyramidal and PV-containing GABA neurons, the absence of a reduction in neuron number in both of these neuronal populations in schizophrenia indicates that the decrease in BDNF and TrkB mRNAs is due to a downregulation in the expression of the transcripts.

Consistent with the hypothesis that altered GABA-related gene expression is driven by reduced BDNF-TrkB signaling in schizophrenia subjects, the changes in TrkB and GAD₆₇ mRNA expression levels were strongly correlated ($r = 0.74$, $p < 0.001$) in the same subjects, and a positive correlation between the changes in BDNF and GAD₆₇ mRNA expression levels ($r = 0.52$, $p = 0.007$) was also observed. Interestingly, the correlation was significantly ($p = 0.043$) stronger between TrkB and GAD₆₇ mRNAs than between BDNF and GAD₆₇ mRNAs, suggesting that altered TrkB might be a pathogenetic mechanism driving the reduced GABA-related gene expression in schizophrenia.

Of course, such correlations in human studies do not demonstrate a cause and effect relationship. However, the idea that reduced signaling through TrkB receptors could be a primary determinant of cortical GABA-related gene expression changes in schizophrenia was supported by studies in TrkB hypomorphic

mice in which the insertion of floxed TrkB cDNA (*fBZ*) resulted in decreased TrkB expression (Xu *et al.*, 2000a). Compared to wild-type mice, TrkB mRNA expression levels in the prefrontal cortex were significantly decreased by 42% and 75% in mice with *fBZ/+* and *fBZ/fBZ* genotypes, respectively, and in *fBZ/fBZ* mice, expression levels of GAD₆₇ and PV mRNAs in the prefrontal cortex were significantly decreased by 25% and 40%, respectively (Hashimoto *et al.*, 2005b). In addition, in the *fBZ/+* mice, the expression levels of GAD₆₇ and PV mRNAs were intermediate between the wild-type control and *fBZ/fBZ* mice. Furthermore, the cellular pattern of reduced GAD₆₇ mRNA expression in these mice precisely paralleled that seen in schizophrenia (Volk *et al.*, 2000). That is, the density of neurons with detectable levels of GAD₆₇ mRNA was significantly reduced, but the level of GAD₆₇ mRNA expression per neuron was unchanged (Hashimoto *et al.*, 2005b). Furthermore, consistent with the selective vulnerability of a GABA neuron subpopulation in schizophrenia, TrkB genotype had no effect on the expression of calretinin mRNA. Thus, the alterations in GABA-related gene expression in TrkB hypomorphic mice replicate those found in subjects with schizophrenia at both the tissue and cellular levels.

In contrast, although BDNF mRNA expression was decreased by 80% in the prefrontal cortex of mice with a neuron-specific inducible knockout of *bdnf* (Monteggia *et al.*, 2004), no alterations in the expression levels of GAD₆₇ or PV mRNAs were present in these animals, whether the *bdnf* knockout was induced during embryogenesis or in adulthood (Hashimoto *et al.*, 2005b). Together, these findings suggest that changes in TrkB expression, and not in BDNF expression, regulate GABA-related gene expression in the prefrontal cortex.

Thus, these observations support the hypothesis that the deficit in expression of GAD₆₇ mRNA in schizophrenia is the direct result of reduced TrkB expression in GABA neurons. However, this expression deficit may also be an indirect consequence of alterations in pyramidal neurons. For example, BDNF-TrkB signaling appears to promote somatodendritic development (McAllister *et al.*, 1995; Xu *et al.*, 2000b) and spine formation (Horch *et al.*, 1999) in pyramidal neurons, although spine density was not reduced in the prefrontal cortex of mice with the inducible knockout of *bdnf* (Hill *et al.*, 2005). Interestingly, in the DLPFC of subjects with schizophrenia, pyramidal neurons exhibit decreased somal size, dendritic length, and spine density (Garey *et al.*, 1998; Glantz and Lewis, 2000; Pierri *et al.*, 2001; Rajkowska *et al.*, 1998), consistent with an effect of reduced BDNF-TrkB signaling on pyramidal neurons in the illness. These findings, in concert with evidence that BDNF-TrkB signaling directly affects the efficacy of excitatory neurotransmission among pyramidal neurons (Kang and Schuman, 1995; Xu *et al.*, 2000a), suggest that reduced expression of TrkB in pyramidal neurons may both alter their morphology and cause a decrease in their activity. The resulting reduction in pyramidal neuron activity may lead to reduced gene expression in GABA neurons, especially those containing PV, which (in contrast

to calretinin-containing GABA neurons) receive direct excitatory inputs from neighboring pyramidal neurons (Melchitzky and Lewis, 2003).

IV. Connecting Alterations in PV-Positive Neurons to Working Memory Impairments: Decreased Gamma Band Synchrony in Schizophrenia

If reduced signaling via the TrkB receptor results in deficient chandelier cell-mediated inhibition of pyramidal neurons, how do such changes in GABA neurotransmission give rise to altered working memory? Interestingly, PV-containing inhibitory neurons are involved in the induction and maintenance of gamma oscillations in pyramidal neurons. In particular, networks of PV-containing, fast-spiking GABA neurons in the middle cortical layers, formed via both chemical and electrical synapses, give rise to oscillatory activity in the gamma band (30–80 Hz) range (Tamas *et al.*, 2000). The coordinated oscillatory context provided by these networks of inhibitory neurons is thought to create the discrete temporal structure necessary for ensembles of pyramidal neurons to perform specific functions, such as those involved in working memory.

Consistent with this interpretation, gamma band oscillations are induced and sustained in the DLPFC during the delay period of working memory tasks (Tallon-Baudry *et al.*, 1998). In addition, the amplitude, or power, of gamma oscillations in the DLPFC appears to increase in proportion to working memory load (Howard *et al.*, 2003). In the frontal cortex of subjects with schizophrenia, phase locking of gamma activity to the stimulus onset is impaired (Spencer *et al.*, 2003), and gamma band power in the DLPFC is reduced during the delay period of a working memory task (Cho *et al.*, 2004).

Thus, a deficit in pyramidal cell synchronization, resulting from impaired perisomatic inhibition via PV-containing GABA neurons, may contribute to the reported deficits in gamma oscillations, and consequently working memory dysfunction, in schizophrenia. Several features of PV-containing neurons, and of their alterations in schizophrenia, may explain how this occurs. First, the axonal arborizations of individual chandelier and wide-arbor neurons are highly divergent and target the axon initial segments and cell bodies, respectively, of a large number of pyramidal neurons (Peters, 1984), enabling them to regulate the firing of local groups of pyramidal neurons (Cobb *et al.*, 1995). Second, in the monkey DLPFC, PV-containing GABA neurons and pyramidal cells share certain excitatory inputs in common, including projections from neighboring pyramidal neurons and from the mediodorsal thalamus (Melchitzky and Lewis, 2003; Melchitzky *et al.*, 1999, 2001). Thus, excitatory input from these sources stimulates both PV-containing and pyramidal neurons simultaneously, resulting in a secondary, temporally delayed

perisomatic inhibitory input to pyramidal neurons. This disynaptic inhibitory input appears to limit the window of time, and thereby increases the temporal precision, for the summation of excitatory inputs needed to evoke pyramidal neuron firing (Pouille and Scanziani, 2001). Consequently, a deficiency in chandelier cell and/or wide-arbor neuron perisomatic input to pyramidal neurons would be expected to reduce the magnitude of pyramidal cell synchrony, and thus of gamma band power, in the DLPFC.

V. Treatment Implications

How do these findings inform our understanding of novel targets for pharmacological intervention in schizophrenia? Drugs with selective agonist activity at GABA_A receptors containing the α_2 subunit may provide an effective approach to enhance chandelier neuron inhibition of DLPFC pyramidal neurons in schizophrenia by increasing the synchronization of pyramidal cell firing at gamma frequencies and consequently improving working memory function (Lewis *et al.*, 2004; Volk and Lewis, 2005). The α_2 subunit of the GABA_A receptor represents a highly selective target for enhancing inhibition at the axon initial segment of pyramidal neurons because it is predominantly restricted to this location (Nusser *et al.*, 1996).

Drugs that directly activate α_2 -containing GABA_A receptors independent of the presence of GABA, or that generally increase the firing rate of chandelier cells, may actually disrupt the timing of disynaptic inhibition of pyramidal neurons. Furthermore, agents that nonselectively inhibit GABA reuptake may act too broadly at other GABA synaptic sites that are not altered in schizophrenia. In contrast, drugs that enhance the postsynaptic response to the release of GABA from chandelier cell axons, such as a GABA_A α_2 -selective benzodiazepine, would increase the frequency of opening of chloride ion channels in the presence of GABA. Currently available benzodiazepines are not selective for GABA_A receptors containing the α_2 subunit, and might cause a generalized and nonspecific increase in cortical inhibition. Thus, treatment with an α_2 -selective benzodiazepine would be predicted to augment the postsynaptic inhibitory response at pyramidal neuron axon initial segments in a manner that incorporates the critical timing of chandelier neuron firing essential for synchronizing pyramidal neuron activity. Furthermore, since the anxiolytic effects of benzodiazepines appear to be mediated by GABA_A receptors that contain the α_2 subunit (Löw *et al.*, 2000), α_2 -specific agents may both improve cognitive function and reduce the stress responses that have been linked to the exacerbations of psychotic symptoms in schizophrenia (Carpenter *et al.*, 1999).

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References

- Akbarian, S., Kim, J. J., Potkin, S. G., Hagman, J. O., Tafazzoli, A., Bunney, W. E., Jr., and Jones, E. G. (1995). Gene expression for glutamic acid decarboxylase is reduced without loss of neurons in prefrontal cortex of schizophrenics. *Arch. Gen. Psychiatry* **52**, 258–266.
- Asada, H., Kawamura, Y., Maruyama, K., Kume, H., Ding, R., Kanbara, N., Kuzume, H., Sanbo, M., Yagi, T., and Obata, K. (1997). Cleft palate and decreased brain γ -aminobutyric acid in mice lacking the 67-kDa isoform of glutamic acid decarboxylase. *Proc. Natl. Acad. Sci. USA* **94**, 6496–6499.
- Barch, D. M., Sheline, Y. I., Csernansky, J. G., and Snyder, A. Z. (2003). Working memory and prefrontal cortex dysfunction: Specificity to schizophrenia compared with major depression. *Biol. Psychiatry* **53**, 376–384.
- Beasley, C. L., Zhang, Z. J., Patten, I., and Reynolds, G. P. (2002). Selective deficits in prefrontal cortical GABAergic neurons in schizophrenia defined by the presence of calcium-binding proteins. *Biol. Psychiatry* **52**, 708–715.
- Benes, F. M., Vincent, S. L., Marie, A., and Khan, Y. (1996). Up-regulation of GABA-A receptor binding on neurons of the prefrontal cortex in schizophrenic subjects. *Neuroscience* **75**, 1021–1031.
- Benes, F. M., Kwok, E. W., Vincent, S. L., and Todtenkopf, M. S. (1998). Reduction of nonpyramidal cells in section CA2 of schizophrenics and manic depressives. *Biol. Psychiatry* **44**, 88–97.
- Benes, F. M., Todtenkopf, M. S., Logiotatos, P., and Williams, M. (2000). Glutamate decarboxylase (65)-immunoreactive terminals in cingulate and prefrontal cortices of schizophrenic and bipolar brain. *J. Chem. Neuroanat.* **20**, 259–269.
- Bertolino, A., Nawroz, S., Mattay, V. S., Barnett, A. S., Duyn, J. H., Moonen, C. T. W., Frank, J. A., Tedeschi, G., and Weinberger, D. R. (1996). Regionally specific pattern of neurochemical pathology in schizophrenia as assessed by multislice proton magnetic resonance spectroscopic imaging. *Am. J. Psychiatry* **153**, 1554–1563.
- Byne, W., Buchsbaum, M. S., Mattiace, L. A., Hazlett, E. A., Kemether, E., Elhakem, S. L., Purohit, D. P., Haroutunian, V., and Jones, L. (2002). Postmortem assessment of thalamic nuclear volumes in subjects with schizophrenia. *Am. J. Psychiatry* **159**, 59–65.
- Callicott, J. H., Mattay, V. S., Verchinski, B. A., Marenco, S., Egan, M. F., and Weinberger, D. R. (2003). Complexity of prefrontal cortical dysfunction in schizophrenia: More than up or down. *Am. J. Psychiatry* **160**, 2209–2215.
- Carpenter, W. T., Jr., Buchanan, R. W., Kirkpatrick, B., and Breier, A. F. (1999). Diazepam treatment of early signs of exacerbation in schizophrenia. *Am. J. Psychiatry* **156**, 299–303.
- Cellerino, A., Maffei, L., and Domenici, L. (1996). The distribution of brain-derived neurotrophic factor and its receptor trkB in parvalbumin-containing neurons of the rat visual cortex. *Eur. J. Neurosci.* **8**, 1190–1197.
- Cho, R. Y., Konecky, R. O., and Carter, C. S. (2004). Impaired task-set maintenance and frontal cortical gamma-band synchrony in schizophrenia. *Soc. Neurosci. Abstr.* **34** (348), 13.

- Cobb, S. R., Buhl, E. H., Halasy, K., Paulsen, O., and Somogyi, P. (1995). Synchronization of neuronal activity in hippocampus by individual GABAergic interneurons. *Nature* **378**, 75–78.
- Cochran, S. M., Kennedy, M., McKerchar, C. E., Steward, L. J., Pratt, J. A., and Morris, B. J. (2003). Induction of metabolic hypofunction and neurochemical deficits after chronic intermittent exposure to phencyclidine: Differential modulation by antipsychotic drugs. *Neuropsychopharmacology* **28**, 265–275.
- Condé, F., Lund, J. S., Jacobowitz, D. M., Baimbridge, K. G., and Lewis, D. A. (1994). Local circuit neurons immunoreactive for calretinin, calbindin D-28k, or parvalbumin in monkey prefrontal cortex: Distribution and morphology. *J. Comp. Neurol.* **341**, 95–116.
- Cotter, D., Landau, S., Beasley, C., Stevenson, R., Chana, G., MacMillan, L., and Everall, I. (2002). The density and spatial distribution of GABAergic neurons, labelled using calcium binding proteins, in the anterior cingulate cortex in major depressive disorder, bipolar disorder, and schizophrenia. *Biol. Psychiatry* **51**, 377–386.
- Cruz, D. A., Eggan, S. M., and Lewis, D. A. (2003). Postnatal development of pre- and post-synaptic GABA markers at chandelier cell inputs to pyramidal neurons in monkey prefrontal cortex. *J. Comp. Neurol.* **465**, 385–400.
- Cruz, D. A., Eggan, S. M., Azmitia, E. C., and Lewis, D. A. (2004). Serotonin1A receptors at the axon initial segment of prefrontal pyramidal neurons in schizophrenia. *Am. J. Psychiatry* **161**, 739–742.
- Cullen, T. J., Walker, M. A., Parkinson, N., Craven, R., Crow, T. J., Esiri, M. M., and Harrison, P. J. (2003). A postmortem study of the mediodorsal nucleus of the thalamus in schizophrenia. *Schizophr. Res.* **60**, 157–166.
- Cunningham, M. O., Hunt, J., Middleton, S., LeBeau, F. E., Gillies, M. J., Davies, C. H., Maycox, P. R., Whittington, M. A., and Racca, C. (2006). Region-specific reduction in entorhinal gamma oscillations and parvalbumin-immunoreactive neurons in animal models of psychiatric illness. *J. Neurosci.* **26**, 2767–2776.
- Danos, P., Baumann, B., Krämer, A. V., Bernstein, H.-G., Stauch, R., Krell, D., Falkai, P., and Bogerts, B. (2003). Volumes of association thalamic nuclei in schizophrenia: A postmortem study. *Schizophr. Res.* **60**, 141–155.
- Davidson, M., Reichenberg, A., Rabinowitz, J., Weiser, M., Kaplan, Z., and Mark, M. (1999). Behavioral and intellectual markers for schizophrenia in apparently healthy male adolescents. *Am. J. Psychiatry* **156**, 1328–1335.
- Daviss, S. R., and Lewis, D. A. (1995). Local circuit neurons of the prefrontal cortex in schizophrenia: Selective increase in the density of calbindin-immunoreactive neurons. *Psychiatry Res.* **59**, 81–96.
- Diamond, A. (2002). Normal development of prefrontal cortex from birth to young adulthood: Cognitive functions, anatomy and biochemistry. In “Principles of Frontal Lobe Function” (D. T. Stuss and R. T. Knight, Eds.), pp. 466–503. Oxford University Press, London.
- Dorph-Petersen, K. -A., Pierri, J. N., Sun, Z., Sampson, A. R., and Lewis, D. A. (2004). Stereological analysis of the mediodorsal thalamic nucleus in schizophrenia: Volume, neuron number, and cell types. *J. Comp. Neurol.* **472**, 449–462.
- Dracheva, S., Elhakem, S. L., McGurk, S. R., Davis, K. L., and Haroutunian, V. (2004). GAD67 and GAD65 mRNA and protein expression in cerebrocortical regions of elderly patients with schizophrenia. *J. Neurosci. Res.* **76**, 581–592.
- Elvevåg, B., and Goldberg, T. E. (2000). Cognitive impairment in schizophrenia is the core of the disorder. *Crit. Rev. Neurobiol.* **14**, 1–21.
- Erickson, S. L., and Lewis, D. A. (2002). Postnatal development of parvalbumin- and GABA transporter-immunoreactive axon terminals in monkey prefrontal cortex. *J. Comp. Neurol.* **448**, 186–202.
- Gabriel, S. M., Davidson, M., Haroutunian, V., Powchik, P., Bierer, L. M., Purohit, D. P., Perl, D. P., and Davis, K. L. (1996). Neuropeptide deficits in schizophrenia vs. Alzheimer's disease cerebral cortex. *Biol. Psychiatry* **39**, 82–91.

- Garey, L. J., Ong, W. Y., Patel, T. S., Kanani, M., Davis, A., Mortimer, A. M., Barnes, T. R. E., and Hirsch, S. R. (1998). Reduced dendritic spine density on cerebral cortical pyramidal neurons in schizophrenia. *J. Neurol. Neurosurg. Psychiatry* **65**, 446–453.
- Glantz, L. A., and Lewis, D. A. (2000). Decreased dendritic spine density on prefrontal cortical pyramidal neurons in schizophrenia. *Arch. Gen. Psychiatry* **57**, 65–73.
- Goldman-Rakic, P. S. (1995). Cellular basis of working memory. *Neuron* **14**, 477–485.
- González-Burgos, G., Krimer, L. S., Povysheva, N. V., Barrionuevo, G., and Lewis, D. A. (2005). Functional properties of fast spiking interneurons and their synaptic connections with pyramidal cells in primate dorsolateral prefrontal cortex. *J. Neurophysiol.* **93**, 942–953.
- Green, M. F. (1996). What are the functional consequences of neurocognitive deficits in schizophrenia? *Am. J. Psychiatry* **153**, 321–330.
- Guidotti, A., Auta, J., Davis, J. M., Gerevini, V. D., Dwivedi, Y., Grayson, D. R., Impagnatiello, F., Pandey, G., Pesold, C., Sharma, R., Uzunov, D., and Costa, E. (2000). Decrease in reelin and glutamic acid decarboxylase₆₇ (GAD₆₇) expression in schizophrenia and bipolar disorder. *Arch. Gen. Psychiatry* **57**, 1061–1069.
- Gulyás, A. I., Megias, M., Emri, Z., and Freund, T. F. (1999). Total number and ratio of excitatory and inhibitory synapses converging onto single interneurons of different types in the CA1 area of the rat hippocampus. *J. Neurosci.* **19**, 10082–10097.
- Hanada, S., Mita, T., Nishino, N., and Tanaka, C. (1987). [³H]Muscimol binding sites increased in autopsied brains of chronic schizophrenics. *Life Sci.* **40**, 239–266.
- Harrison, P. J., and Lewis, D. A. (2003). Neuropathology of schizophrenia. In “Schizophrenia” (S. Hirsch and D. R. Weinberger, Eds.), 2nd ed., pp. 310–325. Blackwell Science Ltd., Oxford.
- Hashimoto, T., Volk, D. W., Eggan, S. M., Mirnics, K., Pierri, J. N., Sun, Z., Sampson, A. R., and Lewis, D. A. (2003). Gene expression deficits in a subclass of GABA neurons in the prefrontal cortex of subjects with schizophrenia. *J. Neurosci.* **23**, 6315–6326.
- Hashimoto, T., Arion, G., Unger, T., Volk, D. W., Mirnics, K., and Lewis, D. A. (2005a). Analysis of gaba-specific transcriptome in the prefrontal cortex of subjects with schizophrenia. *Soc. Neurosci. Abstr.* **35**, 675.4.
- Hashimoto, T., Bergen, S. E., Nguyen, Q. L., Xu, B., Monteggia, L. M., Pierri, J. N., Sun, Z., Sampson, A. R., and Lewis, D. A. (2005b). Relationship of brain-derived neurotrophic factor and its receptor TrkB to altered inhibitory prefrontal circuitry in schizophrenia. *J. Neurosci.* **25**, 372–383.
- Hasler, G., Neumeister, A., van der Veen, J. W., Tuminis, T., Bain, E. E., Shen, J., Drevets, W. C., and Charney, D. S. (2005). Normal prefrontal gamma-aminobutyric acid levels in remitted depressed subjects determined by proton magnetic resonance spectroscopy. *Biol. Psychiatry* **58**, 969–973.
- Heaton, R., Paulsen, J. S., McAdams, L. A., Kuck, J., Zisook, S., Braff, D., Harris, J., and Jeste, D. V. (1994). Neuropsychological deficits in schizophrenics: Relationship to age, chronicity, and dementia. *Arch. Gen. Psychiatry* **51**, 469–476.
- Heckers, S., Stone, D., Walsh, J., Schick, J., Koul, P., and Benes, F. M. (2002). Differential hippocampal expression of glutamic acid decarboxylase 65 and 67 messenger RNA in bipolar disorder and schizophrenia. *Arch. Gen. Psychiatry* **59**, 521–529.
- Hill, J. J., Kolluri, N., Hashimoto, T., Wu, Q., Sampson, A. R., Monteggia, L. M., and Lewis, D. A. (2005). Analysis of pyramidal neuron morphology in an inducible knockout of brain-derived neurotrophic factor. *Biol. Psychiatry* **57**, 932–934.
- Horch, H. W., Krüttgen, A., Portbury, S. D., and Katz, L. C. (1999). Destabilization of cortical dendrites and spines by BDNF. *Neuron* **23**, 353–364.
- Howard, M. W., Rizzuto, D. S., Caplan, J. B., Madsen, J. R., Lisman, J., Aschenbrenner-Scheibe, R., Schulze-Bonhage, A., and Kahana, M. J. (2003). Gamma oscillations correlate with working memory load in humans. *Cereb. Cortex* **13**, 1369–1374.

- Huang, Z. J., Kirkwood, A., Pizzorusso, T., Porciatti, V., Morales, B., Bear, M. F., Maffei, L., and Tonegawa, S. (1999). BDNF regulates the maturation of inhibition and the critical period of plasticity in mouse visual cortex. *Cell* **98**, 739–755.
- Huntley, G. W., Vickers, J. C., Janssen, W., Brose, N., Heinemann, S. F., and Morrison, J. H. (1994). Distribution and synaptic localization of immunocytochemically identified NMDA receptor subunit proteins in sensory-motor and visual cortices of monkey and human. *J. Neurosci.* **14**, 3603–3619.
- Huntley, G. W., Vickers, J. C., and Morrison, J. H. (1997). Quantitative localization of NMDAR1 receptor subunit immunoreactivity in inferotemporal and prefrontal association cortices of monkey and human. *Brain Res.* **749**, 245–262.
- Impagnatiello, F., Guidotti, A. R., Pesold, C., Dwivedi, Y., Caruncho, H., Pisu, M. G., Uzunov, D. P., Smalheiser, N. R., Davis, J. M., Pandey, G. N., Pappas, G. D., Teuting, P., *et al.* (1998). A decrease of reelin expression as a putative vulnerability factor in schizophrenia. *Proc. Natl. Acad. Sci. USA* **95**, 15718–15723.
- Jones, E. G. (1997). Cortical development and thalamic pathology in schizophrenia. *Schizophr. Bull.* **23**, 483–501.
- Kang, H., and Schuman, E. M. (1995). Long-lasting neurotrophin-induced enhancement of synaptic transmission in the adult hippocampus. *Science* **267**, 1658–1662.
- Kawaguchi, Y. (1995). Physiological subgroups of nonpyramidal cells with specific morphological characteristics in layer II/III of rat frontal cortex. *J. Neurosci.* **15**, 2638–2655.
- Keilhoff, G., Becker, A., Grecksch, G., Wolf, G., and Bernstein, H. G. (2004). Repeated application of ketamine to rats induces changes in the hippocampal expression of parvalbumin, neuronal nitric oxide synthase and cFOS similar to those found in human schizophrenia. *Neuroscience* **126**, 591–598.
- Kinney, J. W., Davis, C. N., Tabarean, I., Conti, B., Bartfai, T., and Behrens, M. M. (2006). A specific role for NR2A-containing NMDA receptors in the maintenance of parvalbumin and GAD67 immunoreactivity in cultured interneurons. *J. Neurosci.* **26**, 1604–1615.
- Klausberger, T., Magill, P. J., Marton, L. F., Roberts, J. D. B., Cobden, P. M., Buzsaki, G., and Somogyi, P. (2003). Brain-state- and cell-type specific firing of hippocampal interneurons *in vivo*. *Nature* **421**, 844–848.
- Konopaske, G. T., Sweet, R. A., Wu, Q., Sampson, A., and Lewis, D. A. (2006). Regional specificity of chandelier neuron axon terminal alterations in schizophrenia. *Neuroscience* **138**, 189–196.
- Krimer, L. S., Zaitsev, A. V., Czanner, G., Kroner, S., Gonzalez-Burgos, G., Povysheva, N. V., Iyengar, S., Barrionuevo, G., and Lewis, D. A. (2005). Cluster analysis-based physiological classification and morphological properties of inhibitory neurons in layers 2–3 of monkey dorsolateral prefrontal cortex. *J. Neurophysiol.* **94**, 3009–3022.
- Lewis, D. A. (1997). Development of the primate prefrontal cortex. In “Neurodevelopment & Adult Psychopathology” (M. S. Keshavan and R. M. Murray, Eds.), pp. 12–30. Cambridge University Press, Cambridge.
- Lewis, D. A., and Levitt, P. (2002). Schizophrenia as a disorder of neurodevelopment. *Ann. Rev. Neurosci.* **25**, 409–432.
- Lewis, D. A., and Lund, J. S. (1990). Heterogeneity of chandelier neurons in monkey neocortex: Corticotropin-releasing factor and parvalbumin immunoreactive populations. *J. Comp. Neurol.* **293**, 599–615.
- Lewis, D. A., and Moghaddam, B. (2006). Cognitive dysfunction in schizophrenia: Convergence of GABA and glutamate alterations. *Arch. Neurol.* **63**, 1372–1376.
- Lewis, D. A., Cruz, D. A., Melchitzky, D. S., and Pierri, J. N. (2001). Lamina-specific deficits in parvalbumin-immunoreactive varicosities in the prefrontal cortex of subjects with schizophrenia: Evidence for fewer projections from the thalamus. *Am. J. Psychiatry* **158**, 1411–1422.
- Lewis, D. A., Volk, D. W., and Hashimoto, T. (2004). Selective alterations in prefrontal cortical GABA neurotransmission in schizophrenia: A novel target for the treatment of working memory dysfunction. *Psychopharmacology* **174**, 143–150.

- Lewis, D. A., Hashimoto, T., and Volk, D. W. (2005). Cortical inhibitory neurons and schizophrenia. *Nat. Rev. Neurosci.* **6**, 312–324.
- Lipska, B. K., and Weinberger, D. R. (2000). To model a psychiatric disorder in animals: Schizophrenia as a reality test. *Neuropsychopharmacology* **23**, 223–239.
- Lipska, B. K., Lerman, D. N., Khaing, Z. Z., Weickert, C. S., and Weinberger, D. R. (2003). Gene expression in dopamine and GABA systems in an animal model of schizophrenia: Effects of antipsychotic drugs. *Eur. J. Neurosci.* **18**, 391–402.
- Löw, K., Crestani, F., Keist, R., Benke, D., Brünig, I., Benson, J. A., Fritschy, J.-M., Rülcke, T., Bluethmann, H., Möhler, H., and Rudolph, U. (2000). Molecular and neuronal substrate for the selective attenuation of anxiety. *Science* **290**, 131–134.
- Luna, B., Garver, K. E., Urban, T. A., Lazar, N. A., and Sweeney, J. A. (2004). Maturation of cognitive processes from late childhood to adulthood. *Child Dev.* **75**, 1357–1372.
- MacDonald, A. W., III, Carter, C. S., Kerns, J. G., Ursu, S., Barch, D. M., Holmes, A. J., Stenger, V. A., and Cohen, J. D. (2005). Specificity of prefrontal dysfunction and context processing deficits to schizophrenia in never-medicated patients with first-episode psychosis. *Am. J. Psychiatry* **162**, 475–484.
- Marty, S., Wehrle, R., and Sotelo, C. (2000). Neuronal activity and brain-derived neurotrophic factor regulate the density of inhibitory synapses in organotypic slice cultures of postnatal hippocampus. *J. Neurosci.* **20**, 8087–8095.
- McAllister, A. K., Lo, D. C., and Katz, L. C. (1995). Neurotrophins regulate dendritic growth and developing visual cortex. *Neuron* **15**, 791–803.
- Melchitzky, D. S., and Lewis, D. A. (2003). Pyramidal neuron local axon terminals in monkey prefrontal cortex: Differential targeting of subclasses of GABA neurons. *Cereb. Cortex* **13**, 452–460.
- Melchitzky, D. S., Sesack, S. R., and Lewis, D. A. (1999). Parvalbumin-immunoreactive axon terminals in macaque monkey and human prefrontal cortex: Laminar, regional and target specificity of Type I and Type II synapses. *J. Comp. Neurol.* **408**, 11–22.
- Melchitzky, D. S., González-Burgos, G., Barrionuevo, G., and Lewis, D. A. (2001). Synaptic targets of the intrinsic axon collaterals of supragranular pyramidal neurons in monkey prefrontal cortex. *J. Comp. Neurol.* **430**, 209–221.
- Miller, E. K., and Cohen, J. D. (2001). An integrative theory of prefrontal cortex function. *Annu. Rev. Neurosci.* **24**, 167–202.
- Mirnics, K., Middleton, F. A., Marquez, A., Lewis, D. A., and Levitt, P. (2000). Molecular characterization of schizophrenia viewed by microarray analysis of gene expression in prefrontal cortex. *Neuron* **28**, 53–67.
- Monteggia, L. M., Barrot, M., Powell, C. M., Berton, O., Galanis, V., Gemelli, T., Meuth, S., Nagy, A., Greene, R. W., and Nestler, E. J. (2004). Essential role of brain-derived neurotrophic factor in adult hippocampal function. *Proc. Natl. Acad. Sci. USA* **101**, 10827–10832.
- Nielsen, R. D., Abitz, M., Andersen, B. B., and Pakkenberg, B. (2004). Neuron and glial cell numbers in subdivisions of the mediodorsal (MD) nucleus of the thalamus in schizophrenic subjects and controls. *Soc. Neurosci. Abstr.* **34**, 110.7.
- Nusser, Z., Sieghart, W., Benke, D., Fritschy, J.-M., and Somogyi, P. (1996). Differential synaptic localization of two major γ -aminobutyric acid type A receptor α subunits on hippocampal pyramidal cells. *Proc. Natl. Acad. Sci. USA* **93**, 11939–11944.
- Overstreet, L. S., and Westbrook, G. L. (2003). Synapse density regulates independence at unitary inhibitory synapses. *J. Neurosci.* **23**, 2618–2626.
- Pakkenberg, B. (1990). Pronounced reduction of total neuron number in mediodorsal thalamic nucleus and nucleus accumbens in schizophrenics. *Arch. Gen. Psychiatry* **47**, 1023–1028.
- Pakkenberg, B. (1992). The volume of the mediodorsal thalamic nucleus in treated and untreated schizophrenics. *Schizophr. Res.* **7**, 95–100.

- Perlstein, W. M., Carter, C. S., Noll, D. C., and Cohen, J. D. (2001). Relation of prefrontal cortex dysfunction to working memory and symptoms in schizophrenia. *Am. J. Psychiatry* **158**, 1105–1113.
- Peters, A. (1984). Chandelier cells. In "Cerebral Cortex" (E. G. Jones and A. Peters, Eds.), Vol. 1, pp. 361–380. Plenum Press, New York.
- Pierri, J. N., Chaudry, A. S., Woo, T.-U., and Lewis, D. A. (1999). Alterations in chandelier neuron axon terminals in the prefrontal cortex of schizophrenic subjects. *Am. J. Psychiatry* **156**, 1709–1719.
- Pierri, J. N., Volk, C. L. E., Auh, S., Sampson, A., and Lewis, D. A. (2001). Decreased somal size of deep layer 3 pyramidal neurons in the prefrontal cortex of subjects with schizophrenia. *Arch. Gen. Psychiatry* **58**, 466–473.
- Pierri, J. N., Volk, C. L., Auh, S., Sampson, A., and Lewis, D. A. (2003). Somal size of prefrontal cortical pyramidal neurons in schizophrenia: Differential effects across neuronal subpopulations. *Biol. Psychiatry* **54**, 111–120.
- Popken, G. J., Bunney, W. E., Jr., Potkin, S. G., and Jones, E. G. (2000). Subnucleus-specific loss of neurons in medial thalamus of schizophrenics. *Proc. Natl. Acad. Sci. USA* **97**, 9276–9280.
- Pouille, F., and Scanziani, M. (2001). Enforcement of temporal fidelity in pyramidal cells by somatic feed-forward inhibition. *Science* **293**, 1159–1163.
- Rajkowska, G., Selemon, L. D., and Goldman-Rakic, P. S. (1998). Neuronal and glial somal size in the prefrontal cortex: A postmortem morphometric study of schizophrenia and Huntington disease. *Arch. Gen. Psychiatry* **55**, 215–224.
- Rao, S. G., Williams, G. V., and Goldman-Rakic, P. S. (2000). Destruction and creation of spatial tuning by disinhibition: GABA_A blockade of prefrontal cortical neurons engaged by working memory. *J. Neurosci.* **20**, 485–494.
- Sanacora, G., Mason, G. F., Rothman, D. L., Behar, K. L., Hyder, F., Petroff, O. A. C., Berman, R. M., Charney, D. S., and Krystal, J. H. (1999). Reduced cortical γ -aminobutyric acid levels in depressed patients determined by proton magnetic resonance spectroscopy. *Arch. Gen. Psychiatry* **56**, 1043–1047.
- Sawaguchi, T., Matsumura, M., and Kubota, K. (1989). Delayed response deficits produced by local injection of bicuculline into the dorsolateral prefrontal cortex in Japanese macaque monkeys. *Exp. Brain Res.* **75**, 457–469.
- Saykin, A. J., Shtasel, D. L., Gur, R. E., Kester, D. B., Mozley, L. H., Stafiniak, P., and Gur, R. C. (1994). Neuropsychological deficits in neuroleptic naive patients with first-episode schizophrenia. *Arch. Gen. Psychiatry* **51**, 124–131.
- Selemon, L. D., Rajkowska, G., and Goldman-Rakic, P. S. (1995). Abnormally high neuronal density in the schizophrenic cortex: A morphometric analysis of prefrontal area 9 and occipital area 17. *Arch. Gen. Psychiatry* **52**, 805–818.
- Silver, H., Feldman, P., Bilker, W., and Gur, R. C. (2003). Working memory deficit as a core neuropsychological dysfunction in schizophrenia. *Am. J. Psychiatry* **160**, 1809–1816.
- Somogyi, P. (1977). A specific axo-axonal interneuron in the visual cortex of the rat. *Brain Res.* **136**, 345–350.
- Spencer, K. M., Nestor, P. G., Salisbury, D. F., Shenton, M. E., and McCarley, R. W. (2003). Abnormal neural synchrony in schizophrenia. *J. Neurosci.* **23**, 7407–7411.
- Szabadics, J., Varga, C., Molnar, G., Olah, S., Barzo, P., and Tamas, G. (2006). Excitatory effect of GABAergic axo-axonic cells in cortical microcircuits. *Science* **311**, 233–235.
- Tallon-Baudry, C., Bertrand, O., Peronnet, F., and Pernier, J. (1998). Induced gamma-band activity during the delay of a visual short-term memory task in humans. *J. Neurosci.* **18**, 4244–4254.
- Tamas, G., Buhl, E. H., Lorincz, A., and Somogyi, P. (2000). Proximally targeted GABAergic synapses and gap junctions synchronize cortical interneurons. *Nat. Neurosci.* **3**, 366–371.
- Thune, J. J., Uytings, H. B. M., and Pakkenberg, B. (2001). No deficit in total number of neurons in the prefrontal cortex in schizophrenics. *J. Psychiatr. Res.* **35**, 15–21.

- Torrey, E. F., Barci, B. M., Webster, M. J., Bartko, J. J., Meador-Woodruff, J. H., and Knable, M. B. (2005). Neurochemical markers for schizophrenia, bipolar disorder, and major depression in postmortem brains. *Biol. Psychiatry* **57**, 252–260.
- Vawter, M. P., Crook, J. M., Hyde, T. M., Kleinman, J. E., Weinberger, D. R., Becker, K. G., and Freed, W. J. (2002). Microarray analysis of gene expression in the prefrontal cortex in schizophrenia: A preliminary study. *Schizophr. Res.* **58**, 11–20.
- Virgo, L., Humphries, C., Mortimer, A., Barnes, T., Hirsch, S. R., and de Belleruche, J. (1995). Cholecystokinin messenger RNA deficit in frontal and temporal cerebral cortex in schizophrenia. *Biol. Psychiatry* **37**, 694–701.
- Volk, D. W., and Lewis, D. A. (2003). Effects of a mediodorsal thalamus lesion on prefrontal inhibitory circuitry: Implications for schizophrenia. *Biol. Psychiatry* **53**, 385–389.
- Volk, D. W., and Lewis, D. A. (2005). GABA targets for the treatment of cognitive dysfunction in schizophrenia. *Curr. Neuropharmacol.* **3**, 45–62.
- Volk, D. W., Austin, M. C., Pierri, J. N., Sampson, A. R., and Lewis, D. A. (2000). Decreased glutamic acid decarboxylase67 messenger RNA expression in a subset of prefrontal cortical gamma-aminobutyric acid neurons in subjects with schizophrenia. *Arch. Gen. Psychiatry* **57**, 237–245.
- Volk, D. W., Austin, M. C., Pierri, J. N., Sampson, A. R., and Lewis, D. A. (2001). GABA transporter-1 mRNA in the prefrontal cortex in schizophrenia: Decreased expression in a subset of neurons. *Am. J. Psychiatry* **158**, 256–265.
- Volk, D. W., Pierri, J. N., Fritschy, J.-M., Auh, S., Sampson, A. R., and Lewis, D. A. (2002). Reciprocal alterations in pre- and postsynaptic inhibitory markers at chandelier cell inputs to pyramidal neurons in schizophrenia. *Cereb. Cortex* **12**, 1063–1070.
- Vreugdenhil, M., Jefferys, J. G., Celio, M. R., and Schwaller, B. (2003). Parvalbumin-deficiency facilitates repetitive IPSCs and gamma oscillations in the hippocampus. *J. Neurophysiol.* **89**, 1414–1422.
- Weickert, C. S., Hyde, T. M., Lipska, B. K., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2003). Reduced brain-derived neurotrophic factor in prefrontal cortex of patients with schizophrenia. *Mol. Psychiatry* **8**, 592–610.
- Weickert, C. S., Ligons, D. L., Romanczyk, T., Ungaro, G., Hyde, T. M., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2005). Reductions in neurotrophin receptor mRNAs in the prefrontal cortex of patients with schizophrenia. *Mol. Psychiatry* **10**, 637–650.
- Weinberger, D. R., Berman, K. F., and Zec, R. F. (1986). Physiologic dysfunction of dorsolateral prefrontal cortex in schizophrenia. I. Regional cerebral blood flow evidence. *Arch. Gen. Psychiatry* **43**, 114–124.
- Wilson, F. A., O'Scalaidhe, S. P., and Goldman-Rakic, P. S. (1994). Functional synergism between putative gamma-aminobutyrate-containing neurons and pyramidal neurons in prefrontal cortex. *Proc. Natl. Acad. Sci. USA* **91**, 4009–4013.
- Woo, T.-U., Miller, J. L., and Lewis, D. A. (1997). Schizophrenia and the parvalbumin-containing class of cortical local circuit neurons. *Am. J. Psychiatry* **154**, 1013–1015.
- Woo, T.-U., Whitehead, R. E., Melchitzky, D. S., and Lewis, D. A. (1998). A subclass of prefrontal gamma-aminobutyric acid axon terminals are selectively altered in schizophrenia. *Proc. Natl. Acad. Sci. USA* **95**, 5341–5346.
- Woo, T.-U., Walsh, J. P., and Benes, F. M. (2004). Density of glutamic acid decarboxylase 67 messenger RNA-containing neurons that express the N-methyl-D-aspartate receptor subunit NR2A in the anterior cingulate cortex in schizophrenia and bipolar disorder. *Arch. Gen. Psychiatry* **61**, 649–657.
- Xu, B., Gottschalk, W., Chow, A., Wilson, R. I., Schnell, E., Zang, K., Wang, D., Nicoll, R. A., Lu, B., and Reichardt, L. F. (2000a). The role of brain-derived neurotrophic factor receptors in the mature hippocampus: Modulation of long-term potentiation through a presynaptic mechanism involving TrkB. *J. Neurosci.* **20**, 6888–6897.

- Xu, B., Ruff, N. L., Zang, Y. A., McConnell, S. K., Strycker, M. P., and Reichardt, L. F. (2000b). Cortical degeneration in the absence of neurotrophin signaling: Dendritic retraction and neuronal loss after removal of the receptor TrkB. *Neuron* **26**, 233–245.
- Yamada, M. K., Nakanishi, K., Ohba, S., Nakamura, T., Ikegaya, Y., Nishiyama, N., and Matsuki, N. (2002). Brain-derived neurotrophic factor promotes the maturation of GABAergic mechanisms in cultured hippocampal neurons. *J. Neurosci.* **22**, 7580–7585.
- Young, K. A., Manaye, K. F., Liang, C.-L., Hicks, P. B., and German, D. C. (2000). Reduced number of mediodorsal and anterior thalamic neurons in schizophrenia. *Biol. Psychiatry* **47**, 944–953.
- Zaitsev, A. V., Gonzalez-Burgos, G., Povysheva, N. V., Kroner, S., Lewis, D. A., and Krimer, L. S. (2005). Localization of calcium-binding proteins in physiologically and morphologically characterized interneurons of monkey dorsolateral prefrontal cortex. *Cereb. Cortex* **15**, 1178–1186.

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ALTERATIONS OF SEROTONIN TRANSMISSION IN SCHIZOPHRENIA

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A role for serotonin alterations in the pathophysiology of schizophrenia has long been suspected because of the psychotogenic effects of serotonergic agonists and the therapeutic effects of 5-HT₂ antagonism. This chapter is a review of the evidence derived from pharmacological studies, postmortem, and imaging studies that have assessed the role of serotonin transmission in schizophrenia. While a clear picture of specific serotonergic alterations in schizophrenia has not emerged despite much research, this review reinforces a modulatory role of serotonergic agents on dopamine transmission in schizophrenia, which may contribute to the therapeutic effects of atypical antipsychotics.

I. Introduction

A dysfunction of 5-hydroxytryptamine (5-HT) function in schizophrenia was first postulated because of the structural similarity between 5-HT and the hallucinogenic drug lysergic acid diethylamide (LSD; Gaddum, 1954;

Wooley and Shaw, 1954) and renewed after the introduction of clozapine in the United States, a drug with negligible liability for extrapyramidal side effects (EPS) and superior antipsychotic properties compared to typical antipsychotics (Kane *et al.*, 1988). The superior efficacy of clozapine has been attributed to its relatively potent 5-HT₂ receptor antagonism (Meltzer, 1991), prompting the development of pure 5-HT₂ antagonists or “balanced” 5-HT₂-D2 antagonists as potential antipsychotics. So far, available data indicates acceptable antipsychotic efficacy for combined 5-HT₂-D2 antagonists, but not for pure 5-HT₂ antagonists. At the same time, we have learned more about the potential role of other serotonergic receptors in the action of atypical antipsychotics. In addition, postmortem studies, cerebrospinal fluid (CSF), clinical challenge, and imaging studies of serotonergic receptors and transporters performed over the last decade have suggested a serotonergic dysfunction in the brains of patients with schizophrenia. In this chapter, we will first review the evidence for alterations of serotonin transmission in schizophrenia and its implications for the therapeutic effects of antipsychotics. The putative role of 5-HT transmission in schizophrenia will then be discussed in the context of all the data reviewed, the relevant dopaminergic and serotonergic interactions, and the recent advances in the conceptualization of the dopamine (DA) hypothesis of schizophrenia.

II. Alteration of 5-HT Receptors in Schizophrenia

The involvement of alteration of 5-HT transmission in the pathophysiology of schizophrenia is supported by numerous postmortem studies, which have been reviewed elsewhere (Abi-Dargham *et al.*, 1997; Breier, 1995; Lieberman *et al.*, 1998; Meltzer *et al.*, 1999). The most consistent abnormalities of 5-HT markers in schizophrenia are a reduction in cortical 5-HT transporters density and an increase in cortical 5-HT_{1A} receptor binding. A decrease in 5-HT_{2A} density has also been frequently noted, but this observation might be secondary to previous neuroleptic exposure (Table I).

A. 5-HT TRANSPORTERS

5-HT transporters are located on presynaptic serotonergic terminals and are believed to provide an index of serotonergic innervation. Three studies reported decreased density of 5-HT transporters in the frontal cortex of patients with schizophrenia. Laruelle *et al.* (1993) reported decreased density of 5-HT transporters, labeled with [³H]paroxetine, in the frontal cortex of schizophrenic patients, as compared to controls, while no changes were observed in the

TABLE I
ALTERATIONS IN 5-HT RECEPTORS IN SCHIZOPHRENIA: POSTMORTEM STUDIES

References	Site	Ligand	B_{max}	K_D	Brain region
Ohuaha <i>et al.</i> (1993)	5-HT uptake site	[3H]citalopram	Decrease	No change	Frnt Ctx
Laruelle <i>et al.</i> (1993)		[3H]citalopram	Decrease	No change	Frnt Ctx
Joyce <i>et al.</i> (1993)		[3H]cyanoimipramine	Decrease	No change	Frnt Ctx
Dean <i>et al.</i> (1995)		[3H]citalopram	No change	Increase	Hippocampus
Hashimoto <i>et al.</i> (1991)		[3H]8-OH-DPAT	Increase		Frnt Ctx, BA 10
					Temp Ctx
Joyce <i>et al.</i> (1993)		[3H]8-OH-DPAT	Nonsignificant increase		Frnt Ctx BA 9, Cing Ctx Motor Ctx, Hippoc.
Simpson <i>et al.</i> (1996)		[3H]8-OH-DPAT	Increase		Frnt Ctx BA 12 and 11
Sumiyoshi <i>et al.</i> (1996)		[3H]8-OH-DPAT	Increase		PFC
Burnet <i>et al.</i> (1996b)		[3H]8-OH-DPAT	Increase		Frnt Ctx BA 46
Burnet <i>et al.</i> (1997)	5-HT _{1A}	[3H]8-OH-DPAT	Increase		Frnt Ctx BA 46
Gurevich <i>et al.</i> (1997)		[3H]WAY-100635	Increase		Frnt Ctx BA 9, 44, 6, Cing Ctx
Dean <i>et al.</i> (1999)		[3H]8-OH-DPAT	No change		Frnt Ctx BA 9, 8, 10
Bennett <i>et al.</i> (1979)		[3H]LSD	Decrease	No change	Frnt Ctx
Whitaker <i>et al.</i> (1981)		[3H]LSD	No change	No change	Frnt Ctx
Mita <i>et al.</i> (1986)		[3H]ketanserin	Decrease	No change	Frnt Ctx
Arora and Meltzer (1991)		[3H]spiperone	Decrease	No change	Frnt Ctx
Reynolds <i>et al.</i> (1983)		[3H]ketanserin	No change	No change	Frnt Ctx
Laruelle <i>et al.</i> (1993)		[3H]ketanserin	No change	No change	Frnt Ctx
Joyce <i>et al.</i> (1993)		[3H]ketanserin	No change	No change	Frnt Ctx
Dean <i>et al.</i> (1999)	5-HT ₂	[3H]ketanserin	Decrease		Frnt Ctx BA 9
Abi-Dargham <i>et al.</i> (1993)		[3H]LY278584	No change	No change	Amygdala
Dean <i>et al.</i> (1999)		[3H]GR113808	No change		Frnt Ctx BA 9, 8, 10

occipital cortex of the same subjects (Laruelle *et al.*, 1993). Joyce *et al.* (1993) reported decreased 5-HT transporter density, labeled with [^3H]cyanoimipramine, in the frontal and cingulate cortices in patients with schizophrenia, while no changes were observed in the motor cortex, temporal cortex, and hippocampus. In the same schizophrenic patients, increased 5-HT transporter density was observed in the striatum. Ohuoha *et al.* (1993) reported decreased density of 5-HT transporters, labeled with [^3H]citalopram in the frontal cortex of schizophrenic patients as compared to controls. Dean *et al.* (1995) reported decreased affinity of the transporters in the hippocampus but did not replicate the findings of decreased 5-HT transporters in the frontal cortex in schizophrenia, possibly due to methodological differences, as they examined a different area of the frontal cortex than did the other investigators. Similarly negative findings were reported by Gurevich *et al.* (1997).

Brain-imaging studies provide the opportunity to study well-characterized and medication free patients. However, the development of radiotracers for *in vivo* imaging of serotonin transporter (SERT) has been difficult. [^{11}C] 3-Amino-4-(2-dimethylaminomethyl-phenylsulfanyl)-benzonitrile (DASB) was successfully produced and evaluated in humans (Ginovart *et al.*, 2001; Houle *et al.*, 2000; Meyer *et al.*, 2001). [^{11}C]DASB provides higher specific to nonspecific binding ratios than previous tracers such as [^{11}C]McN 5652, and thus an enhanced reliability in the assessment of SERT density (Frankle *et al.*, 2004; Huang *et al.*, 2002; Szabo *et al.*, 2002).

We used [^{11}C]DASB to compare SERT availability between medication-free subjects with schizophrenia and matched controls. Brain regions included in this analysis were those where the concentration of SERT is high enough for [^{11}C]DASB to provide reliable quantification of SERT availability. There were no significant group differences in SERT availability in any brain region. This study failed to support the postmortem findings of decreased SERT affinity in the hippocampus in schizophrenia (Dean *et al.*, 1995; Naylor *et al.*, 1996) as well as the decrease in SERT in the cingulate cortex observed in one postmortem study (Joyce *et al.*, 1993). In the midbrain, this study agreed with the negative results of the previous imaging study with [^{123}I] β -CIT (Laruelle *et al.*, 2000).

In addition, the level of SERT binding was not significantly related to the severity of positive, negative, or depressive symptoms.

Studies in larger samples with radiotracers which allow quantification of cortical 5-HT transporters may be needed to provide a more definitive picture of serotonin innervation in brains of patients with schizophrenia.

B. 5-HT_{1A} RECEPTORS

As noted by Joyce, "Postmortem studies of the concentration of 5-HT_{1A} receptors in schizophrenic patients represent a rare consensus in schizophrenia: all studies published to date found an elevation of this receptor subtype in

schizophrenic patients" (Gurevich and Joyce, 1997). With the exception of one negative study (Dean *et al.*, 1999), seven out of eight studies reported elevations of 5-HT_{1A} receptors in frontal cortex of patients with schizophrenia (Burnet *et al.*, 1996b, 1997; Gurevich and Joyce, 1997; Hashimoto *et al.*, 1991; Joyce *et al.*, 1993; Simpson *et al.*, 1996; Sumiyoshi *et al.*, 1996). All studies were performed with the 5-HT_{1A} receptor agonist [³H]8-hydroxy-2-[di-n-propyl-amino]tetralin ([³H]8-OH-DPAT), except for the study of Burnet *et al.* (1997), which used [3H]WAY-100635. Two studies were performed with homogenate binding and five with autoradiography. All studies included samples from the Prefrontal Cortex (PFC): a significant increase has been reported in all prefrontal Brodmann areas (BA) studied, including the dorso-lateral-prefrontal cortex (DLPFC) (areas 8, 9, 45, and 46), the frontal pole (BA 10), the orbitofrontal cortex (OFC) (BA 11 and 12), and some premotor areas (BA 44, 6). The effect size (difference in the means divided by SD) varied considerably, from small (0.24) to large (1.13), with an average effect size of 0.85 ± 0.34 . A similar effect size was obtained when the average was weighted by the number of cases included per study. Combined, these studies reached a significant level of $p = 0.0006$. Other regions were evaluated: in the cingulate, two studies reported an increase and two found no change. An increase was also reported in the temporal cortex, motor cortex, and hippocampus, but none of these findings were confirmed by the other studies.

The finding of increased 5-HT_{1A} receptors was reported in patients on and off drugs at time of death. Moreover, subchronic (21 days) treatment with haloperidol or clozapine does not affect the density of prefrontal 5-HT_{1A} receptors (Burnet *et al.*, 1996a; Shapiro *et al.*, 1995; Stockmeier *et al.*, 1996). However, as most patients received antipsychotics and other psychotropic medications for years, the possibility that this increase may be a long-term effect of treatment cannot be excluded. An increase in 5-HT_{1A} receptors in the OFC (but not the DLPFC) has also been reported in suicide victims (Arango *et al.*, 1995), raising questions about the disease specificity of this alteration in patients with schizophrenia. However, four studies failed to detect abnormalities in [3H]8-OH-DPAT binding in the frontal cortex of suicide victims (Arranz *et al.*, 1994; Dillon *et al.*, 1991; Lowther *et al.*, 1997; Matsubara *et al.*, 1991). In the PFC, 5-HT_{1A} receptors are concentrated in layers I-II, with lower densities in layers III-IV (Dillon *et al.*, 1991; Hoyer *et al.*, 1986a; Joyce *et al.*, 1993; Lidow *et al.*, 1989). At the ultrastructural level, prefrontal neuronal 5-HT_{1A} receptors are mostly present on the axon hillock of pyramidal cells (Azmitia *et al.*, 1996; Francis *et al.*, 1992). Striatal lesions which induce a degeneration of corticostriatal projections decrease 5-HT_{1A} receptors in the deep cortical layers, indicating that 5-HT_{1A} receptors are located on pyramidal cells projecting to the striatum (Francis *et al.*, 1992). In primate pyramidal cortical neurons, 5-HT_{1A} receptors are observed in high levels in the initial segment of the axons (axon hillock; Azmitia *et al.*, 1996). This localization is consistent with inhibition of action potential by 5-HT_{1A}

agonists (Azmitia *et al.*, 1996). Given the postmortem and clinical suggestion of reduced 5-HT innervation in the PFC in schizophrenia, one could propose that the increase in 5-HT_{1A} receptors observed in this region may be due to a functional upregulation of these receptors. This hypothesis is not supported by a majority of studies in rodents, which failed to observe an upregulation of 5-HT_{1A} receptors following 5,7-dihydroxytryptamine lesions of the 5-HT system (Hensler *et al.*, 1991; Kia *et al.*, 1996; Lawrence *et al.*, 1993; Pranzatelli *et al.*, 1994). Nevertheless, if the alteration of the 5-HT system in the PFC in schizophrenia is neurodevelopmental, the relevance of lesions in adult rodents is limited. A neurodevelopmental alteration is suggested by the study of Slater *et al.* (1998) showing a lack of regression of vermal 5-HT_{1A} receptors in cerebellum of patients with schizophrenia compared to controls.

Unfortunately this consistent data from postmortem studies received little support from the imaging studies undertaken to better explore the clinical relevance of these findings.

Three studies examined 5-HT_{1A} receptor levels in schizophrenia relative to controls *in vivo* using [11C]WAY 100635 positron emission tomography (PET). In the first study, Tauscher *et al.* (2002) reported an overall increase in [11C]WAY 100635 binding across nine regions in subjects with schizophrenia compared to controls, with post hoc analysis revealing significant difference in the medial temporal cortex. The second PET study to explore this issue found a reduction in 5-HT_{1A}-binding parameters in the amygdala (Yasuno *et al.*, 2004).

In the third study, we used [11C]WAY 100635 to compare 5-HT_{1A} availability between medication free subjects with schizophrenia and matched controls. We investigated the same brain regions where differences in the density of 5-HT_{1A} receptors in schizophrenia have been described as well as additional regions in which the concentration of 5-HT_{1A} is high enough for [11C]WAY 100635 to provide reliable quantification of 5-HT_{1A} availability. These regions included four prefrontal cortical regions (dorsolateral, medial, orbital, and subgenual), the parietal, temporal, and occipital cortices, the anterior cingulate cortex, insular cortex, the medial temporal lobe (amygdala, entorhinal cortex, hippocampus, and parahippocampal gyrus), and the dorsal raphe nucleus. We found no significant group differences in 5-HT_{1A} availability in any brain region. Within the patient group, the level of 5-HT_{1A} binding was not significantly related to the severity of positive, negative, or depressive symptoms. The explanation for the difference in findings among all three studies, reporting increase, decrease, and no change, in our case, is not clear. The sample size in the present study was slightly larger than either of the two prior studies. There were fewer drug-naïve subjects in our sample compared to the study of Tauscher *et al.* (2002). However, we did not detect a difference in [11C]WAY 100635 binding when the data from these individuals was analyzed separately. Slight differences in methodology exist across the studies,

including a longer scan time in the current study (110 min vs 60 min; Tauscher *et al.*, 2002, and 90 min; Yasuno *et al.*, 2004) and different data modeling strategies (kinetic modeling with arterial input in the current study compared with a reference region approach in the other two studies). These technical differences would be expected to affect the schizophrenic and control groups equally within each study, and therefore one would expect similar results across studies. In fact, when we analyzed our data using a reference region approach we found no differences between the groups.

Although differences exist between these three PET studies, none of the studies confirmed the findings reported in the postmortem literature exploring 5-HT_{1A} density in schizophrenic subjects. The postmortem studies report an increase in 5-HT_{1A} density in subjects with schizophrenia when compared to controls. This increase is observed in the dorsolateral prefrontal cortex in most studies (Burnet *et al.*, 1996b, 1997; Gurevich and Joyce, 1997; Hashimoto *et al.*, 1991; Simpson *et al.*, 1996; Sumiyoshi *et al.*, 1996), as well as in the anterior cingulate (Burnet *et al.*, 1996b; Gurevich and Joyce, 1997; Joyce *et al.*, 1993) and motor cortex (Gurevich and Joyce, 1997; Joyce *et al.*, 1993) in others. The reason for this lack of consistency in findings may relate to the resolution of these very different techniques. Some postmortem studies show more pronounced increase in 5-HT_{1A} density within superficial cortical layers (Gurevich and Joyce, 1997), whereas others show no difference while exploring specific cellular locations within the prefrontal cortex such as the axon initial segment (Cruz *et al.*, 2004). Using PET, with the currently available technology, it is not possible to explore layer specific differences in receptor density within the cortex, limiting the resolution of these types of studies to relatively large cortical regions.

C. 5-HT₂ RECEPTORS

Studies of 5-HT₂ receptors in the frontal cortex of patients with schizophrenia have generated conflicting results. Early studies were performed with [³H]LSD, a ligand which labels both 5-HT₁ and 5-HT₂ families (Peroutka and Snyder, 1979). Bennett *et al.* (1979) reported decreased [³H]LSD binding in the frontal cortex of patients with schizophrenia as compared to controls. Because [³H]5-HT binding was not reduced in these samples, the decreased [³H]LSD binding was attributed to the 5-HT_{2A} sites, which display relatively low affinity for 5-HT. A second study performed with [³H]LSD showed no differences in the frontal cortex between schizophrenic and control samples (Whitaker *et al.*, 1981).

The more selective 5-HT₂ ligands [³H]ketanserin and [³H]spiperone, were used in five studies to evaluate frontal density of 5-HT₂ receptors. Since both ligands are relatively more selective for 5-HT_{2A} than 5-HT_{2C} (Choudhary *et al.*, 1992; Leysen, 1990), these studies can be viewed as measuring the 5-HT_{2A} rather

than the 5-HT_{2C}. Three studies demonstrated a significant decrease in 5-HT₂ density in the frontal cortex of schizophrenic patients (Arora and Meltzer, 1991; Dean *et al.*, 1999; Mita *et al.*, 1986) while no changes were reported in the other three studies (Joyce *et al.*, 1993; Laruelle *et al.*, 1993; Reynolds *et al.*, 1983). Given that 5-HT₂ antagonists downregulate 5-HT₂ receptors (Andree *et al.*, 1986; Helmeste and Tang, 1983; Leysen *et al.*, 1987) and that most antipsychotic drugs display 5-HT₂ antagonism (Leysen *et al.*, 1978, 1982; Wander *et al.*, 1987), these differences may reflect differences in the antemortem medication. Supporting this interpretation, a PET study with [¹⁸F]setoperone failed to detect any significant changes in 5-HT₂ density in drug-naïve patients with schizophrenia (Lewis *et al.*, 1997). Alternatively, these differences in the density of 5-HT₂ receptors in the frontal cortex in schizophrenia may be related to the heterogeneity of the disease. Laruelle *et al.* (1993) observed that, while schizophrenic patients who committed suicide had 5-HT₂ levels comparable to controls, schizophrenic patients who died from natural causes had significantly lower 5-HT₂ levels than controls. Interestingly, none of the patients in the series of Mita *et al.* (1986) and Arora *et al.* (1991) committed suicide. Thus, three studies suggest decreased frontal 5-HT₂ density in nonsuicide schizophrenic patients. The cause of death was not reported by Reynolds *et al.* and the series of Joyce *et al.* included both suicide and nonsuicide victims. The significance of this finding is unclear. Suicide per se has been associated with increased frontal 5-HT₂ receptor density in some (Arango *et al.*, 1990; Hrdina *et al.*, 1993; Mann *et al.*, 1986; Stanley and Mann, 1983) but not all (Cheetham *et al.*, 1988; Gross-Isseroff *et al.*, 1990; Laruelle *et al.*, 1993; Lowther *et al.*, 1994; Owen *et al.*, 1983, 1986) postmortem studies. Therefore, this factor has to be controlled for in studies of 5-HT₂ density in schizophrenia as it may account for some of the discrepancies in the findings. Decreased 5-HT₂ density may be associated with predominantly negative symptoms and a lower incidence of suicide.

Three PET studies in drug-naïve or drug-free patients with schizophrenia reported normal cortical 5-HT_{2A} receptor binding (Lewis *et al.*, 1999; Okubo *et al.*, 2000; Trichard *et al.*, 1998), while one study reported a significant decrease in PFC 5-HT_{2A} binding in a small group ($n = 6$) of drug-naïve schizophrenic patients (Ngan *et al.*, 2000).

D. OTHER RECEPTORS

No change in the density of 5-HT₃ receptors was observed in the amygdala of patients with schizophrenia (Abi-Dargham *et al.*, 1993). A study found no change in the density of 5-HT₄ receptors in frontal cortex of patients with schizophrenia (Dean *et al.*, 1999).

III. Pharmacological Manipulation of 5-HT Transmission in Schizophrenia

In addition to the direct evidence reviewed above that 5-HT transmission might be affected in schizophrenia, pharmacological interventions modifying 5-HT transmission provided data implicating 5-HT transmission in the mediation of schizophrenia symptomatology. This evidence is specially compelling regarding interventions modifying 5-HT_{2A} receptor function.

A. 5-HT PRECURSORS

The amino acid L-tryptophan is the dietary precursor of 5-HT. Administration of large doses of L-tryptophan increases the synthesis of 5-HT in the brain (Wurtman *et al.*, 1981). During the 1960s and 1970s, numerous studies examined the effects of 5-HT precursors on the clinical symptoms of schizophrenia. Lauer *et al.* (1958) and Pollin *et al.* (1961) administered tryptophan with iproniazid to schizophrenic patients and reported mood elevation, increased involvement, and motor activity. Given the concomitant use of Monoamine oxidase inhibitors (MAOI), these data are difficult to interpret. Bowers (1970) reported mild improvement in Brief Psychiatric Rating Scale (BPRS) in schizophrenic patients treated with L-tryptophan at doses of 2–4 g/day in combination with vitamin B6. Gillin *et al.* (1976) observed that tryptophan administration (20 g/day) had no effect in schizophrenia. Chouinard *et al.* (1978) tested the clinical efficacy of tryptophan (2–6 g/day) and benserazide coadministration against chlorpromazine, and concluded that the antipsychotic action of the tryptophan-benserazide combination was inferior to that of chlorpromazine. Morand *et al.* (1983) described a decrease in aggressivity in schizophrenic patients treated by tryptophan (4 g/day). In summary, tryptophan administration may produce a limited improvement in negative symptoms. Of interest is the fact that worsening of psychosis was not reported. Similar results were reported during the administration of the immediate 5-HT precursor, L-5-hydroxytryptophan, although some patients presented exacerbation of psychotic symptoms, maybe related to the fact that L-5-hydroxytryptamine administration increases also catecholaminergic function (Bigelow *et al.*, 1979; van Praag *et al.*, 1987; Wyatt *et al.*, 1972).

B. 5-HT DEPLETING AGENTS

Two studies with limited number of patients investigated the effects of the tryptophan hydroxylase inhibitor, *p*-chlorophenylalanine (pCPA). Casachia *et al.* (1975) reported improvement in three out of four acute schizophrenics during pCPA treatment (1250 mg/day), while DeLisi *et al.* (1982) reported no significant

changes in chronic schizophrenic patients ($n = 7$, 3000 mg/day). Fenfluramine, a halogenated amphetamine derivative, is believed to cause depletion of the serotonergic system when administered chronically. Shore *et al.* (1985) and Stahl *et al.* (1985) failed to show significant changes during fenfluramine treatment in placebo-controlled studies (8–12 weeks). Soper *et al.* (1990) demonstrated that fenfluramine treatment produced worsening of communication competence and thought disorder in treatment-resistant schizophrenic patients. In summary, these studies suggest that 5-HT depleting agents are not useful in the treatment of schizophrenia, and may even further impair cognitive functioning.

C. 5-HT_{2A} AGONISM: LSD AND “MODEL” PSYCHOSIS

The observation of an LSD-induced psychosis in healthy subjects was the first indication of a potential relationship between serotonin function and schizophrenia. The early reports in the 1950s emphasized the clinical similarities between LSD-induced psychosis and schizophrenia (Rinkel *et al.*, 1955). These were followed by numerous studies which examined carefully the differences such as the prevalence of visual as opposed to auditory hallucinations, the absence of thought disorder, the preservation of affect and insight (Hollister, 1962). However, these differences were lessened when the comparison involved early as opposed to chronic schizophrenics (Freedman and Chapman, 1973) and when cross-cultural differences in schizophrenic symptomatology were examined (Murphy *et al.*, 1963). One study (Langs and Barr, 1968) attempted to compare LSD effects to the different subtypes of schizophrenia and found similarities between the drug group and the paranoid but not the undifferentiated patients. Interestingly enough, the authors described a higher rate of overlap of symptoms for those drug subjects with poorly integrated premorbid personality. Overall, LSD-induced psychosis seemed to be a potential model for some (i.e., hallucinations and paranoid delusions), but not all aspects of schizophrenia (such as disorganization and negative symptoms; Fishman, 1983). Administration of mescaline (3,4,5-trimethoxyphenethylamine), a phenethylamine hallucinogen, to healthy volunteers resulted, similarly, in symptoms of dissolution of ego boundaries, visual hallucinations, “oceanic boundlessness,” and passivity experiences (Hermle *et al.*, 1992). Similar findings were described in humans with psilocybin (Vollenweider *et al.*, 1998). Disturbances in performance on neuropsychological tasks and alterations of cerebral blood flow measured with single photon emission tomography and ⁹⁹Tm-HMPAO have also been described.

1. *Neuropharmacological Effects of Hallucinogens*

The first observation of an effect of LSD on serotonergic transmission was made in 1961 by Freedman (1961). Since, with the discovery of more than

15 subtypes of 5-HT receptors, much more is known about the effects of LSD on central serotonergic receptors. LSD inhibits serotonergic cells in raphe nuclei through a direct agonism on the presynaptic 5-HT_{1A} site, thus reducing the firing of these neurons and the release of serotonin. It also acts as a weak agonist on the postsynaptic 5-HT_{1A} site. LSD has high affinity for all other 5-HT₁ subtypes and for 5-HT_{5A,5B} (Matthes *et al.*, 1993), 5-HT₆ (Shen *et al.*, 1993), and 5-HT₇ receptors. However, the hallucinogenic effect of LSD has been linked to its affinity for the 5-HT₂ receptor, as this property is shared by substituted phenethylamine hallucinogens, such as mescaline, DOI (1-(2,5-dimethoxy-4-iodophenyl)-2-amino-propane hydrochloride), DOB (4-bromo-2,5-dimethoxyphenylisopropylamine), and DOM (2,5-dimethoxy-4-methylamphetamine) (Aghajanian, 1994), and other indoleamine hallucinogens such as DMT (*N,N*-dimethyltryptamine) and psilocybin. Phenethylamine hallucinogens are in general more selective for the 5-HT₂ receptor than LSD. A strong correlation was described between effective doses of indoleamines (LSD) and phenethylamine hallucinogens and their respective potency at the 5-HT₂ receptor (Glennon and Titeler, 1984; Titeler *et al.*, 1988), suggesting that 5-HT₂ receptors mediate the hallucinogenic effects of these drugs. Most data indicates a specific 5-HT_{2A} mechanism, although a 5-HT_{2C} effect cannot be ruled out. LSD has been reported to be an antagonist at the 5-HT₂ site by some investigators, and a full or partial agonist (Glennon, 1990) by others. However, data demonstrates clearly a partial agonist effect of LSD and DOI on the 5-HT_{2A} receptors in the piriform cortex in the rat (Marek and Aghajanian, 1996). This partial agonism effect may explain why LSD can appear as an antagonist, since it can decrease the effect of the agonist, when coadministered at high doses. Its dual effect on 5-HT₂ (stimulatory) and 5-HT_{1A} (inhibitory) can also explain how it may appear as an antagonist, since it can modulate its own effect. Psilocybin's psychotomimetic effects in humans were blocked by ketanserin, a pure 5-HT_{2A} antagonist, and risperidone (Vollenweider, 1998). As psilocybin is a potent 5-HT_{2A} and weak 5-HT_{1A} agonist, this study demonstrates, in humans, that 5-HT_{2A} activation is psychogenic. Another study by the same group showed that this psychosis may be, at least partly, mediated by an increased release of DA, as evidenced by a 20% decrease of [¹¹C]raclopride binding after psilocybin administration in the striatum of human subjects (Vollenweider *et al.*, 1999).

2. Anatomical Substrates of Hallucinogens

5-HT_{2A} receptors are present in high concentration in the olfactory bulb, hippocampus, frontal cortex, piriform, and entorhinal cortices, while 5-HT_{2C} receptors are present in highest density in choroid plexus, anterior olfactory nucleus, piriform and entorhinal cortices, striatum, amygdala, and Substantia Nigra (SN) (Hoyer *et al.*, 1986a,b; Martin and Humphrey, 1994). Hallucinogens have been shown to interact with 5-HT₂ receptors in the locus coeruleus (LC) and the cortex in rats. In the

LC, their effects were reversed by 5-HT₂ antagonists (Rasmussen and Aghajanian, 1986) and by various antipsychotics (Rasmussen and Aghajanian, 1988). The reversal of effect was correlated with the 5-HT₂ binding affinity of the antipsychotic medications. In the piriform cortex in the rat it has been shown that serotonin induces activation of GABAergic interneurons through the 5-HT_{2A} receptor resulting in an enhancement of spontaneous inhibitory postsynaptic potentials in the pyramidal cells (IPSPs) (Gellman and Aghajanian, 1993; Marek and Aghajanian, 1994; Sheldon and Aghajanian, 1991). Aghajanian and Marek (1999) have shown that 5-HT, through 5-HT_{2A} receptors, enhances spontaneous excitatory postsynaptic potentials (EPSPs) in pyramidal cells of layer V of the neocortex, through a focal action on apical dendrites, the main targets for excitatory corticocortical and thalamocortical inputs. This activation leads to an increase in asynchronous release of glutamate by pyramidal cells. This suggests a facilitation of glutamatergic transmission in the cortex via 5-HT_{2A} agonism, and may be consistent with the data of Farber *et al.* (1998) showing that 5-HT_{2A} agonism can prevent the vacuolization related to NMDA neurotoxicity in rodent brain. However, this increase in glutamate release can lead to an alteration in corticocortical and corticosubcortical transmission.

In summary, these studies overall suggest a strong relationship between 5-HT_{2A} stimulation and hallucinogen-induced psychosis. More similarities have been described between hallucinogen-induced psychosis and positive symptoms of schizophrenia as opposed to negative symptoms. Thus, one can conclude that alterations in 5-HT_{2A} function may mediate positive symptoms of schizophrenia, possibly by affecting directly or indirectly other transmitter systems such as DA and glutamate. This is consistent with the therapeutic efficacy of the new atypical neuroleptics known to have a strong 5-HT₂ antagonism.

D. 5-HT_{2A} ANTAGONISM, CLOZAPINE, AND ATYPICALITY

Almost all antipsychotic drugs have appreciable affinity for the 5-HT₂ receptors. 5-HT₂ receptors were initially termed "serotonergic component of neuroleptic receptors" and it was proposed as early as 1978 that this component may play an important role in the antipsychotic properties of neuroleptics (Leysen *et al.*, 1978). Nevertheless, as average clinical doses correlated better with D2 affinity rather than 5-HT₂ affinity, D2 receptor blockade was proposed to be the principal mechanism of action of neuroleptic drugs (Creese *et al.*, 1976; Peroutka and Snyder, 1980; Seeman and Lee, 1975). Drugs such as clozapine, chlorpromazine, thioridazine, and pipamperone had significantly higher affinity for 5-HT₂ than for D2 receptors. However, they were usually prescribed at high doses which induced D2 blockade, suggesting that 5-HT₂ blockade was not the principal mechanism of their antipsychotic action. The inverse was true for

haloperidol and fluphenazine, which did not significantly block 5-HT₂ receptors at average clinical doses (Peroutka and Snyder, 1980).

More recently, the demonstration of the superior efficacy of clozapine for treatment of schizophrenia and of its low incidence of EPS has promoted a renewed interest in the role of 5-HT₂ antagonism in schizophrenia. Given the lack of pharmacological specificity of clozapine, many theories have been proposed to account for its particular clinical profile and have been extensively reviewed elsewhere (Canton *et al.*, 1990; Deutch *et al.*, 1991; Lieberman, 1993; Meltzer, 1991; Seeman, 1992). Most prominent hypotheses include a higher *in vivo* selectivity of clozapine as compared to typical neuroleptics for (1) "corticolimbic" D2 receptors, as compared to "striatal" D2 receptors (Altar *et al.*, 1986; Deutch *et al.*, 1992), possibly as an effect of lower competition with endogenous DA in corticolimbic regions than in striatal regions (Seeman, 1990); (2) D4 receptors (Seeman, 1992; Van Tol *et al.*, 1991); (3) 5-HT₂ receptors (Altar *et al.*, 1986; Meltzer, 1989; Meltzer *et al.*, 1989; Rasmussen and Aghajanian, 1988). While a host of preclinical and clinical data support each of these assumptions, the introduction of new compounds with a more narrow pharmacological profile is indispensable to identify which of these putative mechanisms are critical for a clozapine-like atypical profile. For example, supporting the first hypothesis is the relative corticolimbic selectivity of benzamides with atypical profiles such as sulpiride or remoxipride, compounds which are otherwise devoid of D4 and 5-HT₂ selectivity as compared to D2. Compounds with high D4/D2 selectivity other than clozapine or pure D4 receptor antagonists have not been shown to be effective antipsychotics (Bristow *et al.*, 1997; Kramer *et al.*, 1997; Sanyal and VanTol, 1997). Many new compounds support the hypothesis that a relative 5-HT₂ to D2 selectivity provide "atypical" properties, the first one extensively tested being risperidone.

Following an extensive study of the *in vitro* receptor affinity profile of typical and putative atypical compounds, Meltzer *et al.* (1989) proposed that a ratio of 5-HT₂ pK_i to D2 pK_i > 1.19 (corresponding to a 25-fold selectivity for 5-HT₂ as compared to D2) was desirable to achieve an atypical profile. Risperidone, a compound not included in the original study of Meltzer *et al.* (1989) provides a 19-fold selectivity, slightly lower than the cutoff point originally proposed, but still more selective than the typical chlorpromazine (sevenfold selectivity). Placebo-controlled studies have demonstrated the antipsychotic efficacy of risperidone (Meco *et al.*, 1989; Mesotten *et al.*, 1989), and comparison studies with haloperidol or perphenazine have shown superior antipsychotic properties of risperidone (Claus *et al.*, 1992; Hoyberg *et al.*, 1993). The US-Canadian collaboration study included 388 schizophrenic patients divided in six groups: placebo, 2, 6, 10, or 16 mg daily risperidone and haloperidol 20 mg daily for 8 weeks (Marder and Meibach, 1994). Positive symptoms were significantly reduced as compared to placebo in the 6-, 10-, 16-mg risperidone groups and in the 20-mg

haloperidol group. Negative symptoms were significantly reduced only in the 6- and 16-mg risperidone group, but not in the 20-mg haloperidol group. EPS were significantly higher than placebo in the 16-mg risperidone group and in the 20-mg haloperidol group.

Because haloperidol is significantly more selective toward D2 than 5-HT₂ and because neither haloperidol nor risperidone have prominent antimuscarinic properties, this study provided the best data to date to evaluate the impact of the addition of a preferential 5-HT₂ blockade to D2 blockade in the treatment of schizophrenia. This study supports the hypothesis that a "balanced" 5-HT₂/D2 blockade has superior efficacy in the treatment of negative symptoms and a lower EPS liability. However, EPS and negative symptoms can be correlated and difficult to distinguish clinically. The observation of a significant improvement in negative symptoms despite a similar incidence of EPS in the 16-mg risperidone group as compared to the 20-mg haloperidol group suggests that improvement in negative symptoms is unrelated to decreased EPS. This study also indicates that 5-HT₂ blockade does not affect the incidence of EPS in the presence of complete or near complete D2 blockade. Other atypical agents have been introduced: olanzapine (Tollefson *et al.*, 1997), quetiapine (Arvanitis and Miller, 1997), and ziprasidone (Daniel *et al.*, 1999; Keck *et al.*, 1998; Seeger *et al.*, 1995). These have been demonstrated to be effective in the treatment of positive and negative symptoms in schizophrenia, with fewer side effects than typical neuroleptics. Most studies have shown a preferential response of negative symptoms to atypical antipsychotics versus typicals (Kane *et al.*, 1988; Tollefson *et al.*, 1997). This, however, was attributed by some investigators to an improvement in secondary negative symptoms, that is those related to positive symptoms, depression, EPS, or environmental deprivation, versus the primary negative symptoms otherwise characterized by the deficit syndrome, resulting in controversial debates (Meltzer, 1995). Meta-analyses of available studies have been published showing generally slight advantages of atypical antipsychotics in the treatment of negative symptoms (Leucht *et al.*, 1999). Despite the traditional resistance to treatment of cognitive impairments in schizophrenia, data related to atypical neuroleptics suggest that these drugs may have relatively greater efficacy than typical neuroleptics for treating these deficits (Meltzer and McGurk, 1999; Weinberger and Gallhofer, 1997). The improvement in negative symptoms and cognition is generally attributed to increased dopaminergic tone in the frontal cortex induced by 5-HT_{2A} antagonism, and possibly 5-HT_{1A} agonism for some compounds (see below).

While these treatment studies clarify the therapeutic effect of a combined 5-HT₂ and D2 antagonism, they do not inform us about the potential benefit of "pure" 5-HT₂ antagonists in the treatment of schizophrenia. Ritanserin, a potent 5-HT_{2A/2C} antagonist, is not devoid of activity at the D2 receptor, but its 5-HT₂/D2 selectivity is three times higher than risperidone. In predominantly type II

schizophrenic patients, ritanserin augmentation of classical antipsychotics, as compared to placebo augmentation, was found to induce a significant reduction in BPRS mainly due to a decrease in negative symptoms such as anergia, anxiety/depression (Gelders, 1989). In this trial, ritanserin was more potent than placebo in reducing EPS, a finding which has been replicated (Bersani *et al.*, 1990; Miller *et al.*, 1990). However, therapeutic effects of ritanserin administered alone remain controversial and need further study (Bleekers *et al.*, 1990; Wiesel *et al.*, 1994). Pipamperone, a highly selective 5-HT₂/D2 drug, has been characterized by low EPS profile and antiautistic, disinhibiting, and resocializing effects (Gelders, 1989; Leysen *et al.*, 1978). MDL 100907, a compound with high affinity for 5-HT_{2A} receptors and negligible affinity for D2 receptors, has shown promising properties in preclinical studies predictive of atypical antipsychotic properties (Sorensen *et al.*, 1993). However, a therapeutic effect of MDL 100907 in phase II trials in patients with schizophrenia has not been demonstrated yet, despite many years of development so far, suggesting that pure 5-HT_{2A} antagonism alone may not be sufficient to have a clinically effective antipsychotic agent. D2 blockade remains so far a necessary component for a therapeutic antipsychotic effect, as demonstrated by the fact that no known effective antipsychotic lacks D2 antagonism. Clinical trials with fananserin, an antagonist at D4 and 5-HT_{2A} receptors, show lack of efficacy, illustrating the notion that neither D4 or 5-HT_{2A} antagonism, in the absence of D2 antagonism, seem to be associated with clinical improvement (Truffinet *et al.*, 1999). In conclusion, antagonism at 5-HT_{2A} when added to D2 antagonism may contribute to the atypical profile of an antipsychotic, that is, to better tolerability, fewer motor side effects, and better efficacy on negative symptoms, but alone, may not confer antipsychotic properties.

E. ACTION OF ANTIPSYCHOTIC DRUGS AT OTHER SEROTONERGIC RECEPTORS

Most atypical antipsychotics have affinities for multiple serotonergic receptors, as summarized in Table II, the 5-HT_{1A}, 5-HT_{2C}, 5-HT₆, and 5-HT₇ deserve further discussion, as 5-HT₃ antagonists have been shown to lack antipsychotic property (Newcomer *et al.*, 1992). The actions of 17 antipsychotic agents at the 5-HT_{1A} were explored by (Newman-Tancredi *et al.*, 1998). Clozapine, ziprasidone, and quetiapine exhibited partial agonist activity and marked affinity at the human 5-HT_{1A} receptors, similar to their affinity at D2 receptors. In contrast, risperidone and sertindole displayed low affinity at 5-HT_{1A} receptors and behaved as "neutral" antagonists. Likewise the "typical" neuroleptics, haloperidol, pimozide, raclopride, and chlorpromazine exhibited relatively low affinity and "neutral" antagonist activity. This study suggests that agonist activity at 5-HT_{1A} receptors may be beneficial, although it is clear from clinical experience with drugs such as buspirone that 5-HT_{1A} agonism without D2 blockade does not

TABLE II
AFFINITIES OF SELECTED ANTIPSYCHOTIC DRUGS FOR 5-HT RECEPTORS

Drug	5-HT _{1A}	5-HT _{1B}	5-HT _{1D}	5-HT _{2A}	5-HT _{2C}	5-HT ₃	5-HT ₆	5-HT ₇
Clozapine	132 ^{a,*}	1200 ^b	980 ^b	4 ^c	5 ^c	69 ^b	9.5 ^d	6.3 ^e
Olanzapine	1637 ^{a,*}	1355 ^b	800 ^b	1.9 ^c	2.8 ^c	57 ^b	10 ^d	104 ^e
Risperidone	292 ^a	1325 ^b	100 ^b	0.39 ^c	6.4 ^c		2400 ^d	1.39 ^e
Quetiapine	250 ^{a,*}	5400 ^b	6220 ^b	82 ^c	1500 ^c	170 ^b	33 ^d	
Sertindole	433 ^a			0.2 ^c	0.51 ^c			
Ziprasidone	1.24 ^{a,*}			0.25 ^c	0.55 ^c			
Amperozide	3100 ^c			20 ^c	440 ^c		1600 ^d	549 ^e
Remoxipride	11000 ^c			23000 ^c	5500 ^c			5000 ^e
Haloperidol	1910 ^a		6950 ^b	28 ^c	1500 ^c	>1000 ^b	6600 ^d	263 ^e
Aripiprazola	5.6 ^f	830 ^f	68 ^f	22 ^f	76 ^f	630 ^f	570 ^f	10.3 ^f

^aValues taken from Newman-Tancredi *et al.* (1998), measured in cloned human receptors.

^bBymaster *et al.* (1996), in rat brain (Bymaster *et al.*, 1999).

^cValues reported in Arnt *et al.* (1997), available from Lunbeck Pharmacological screening system (Arnt, 1998).

^dKohen *et al.* (1996), values obtained in human cloned receptors.

^eRoth *et al.* (1994), in cloned rat receptors.

^fShapiro *et al.* (2003).

*Indicates an agonist effect at this receptor.

confer antipsychotic properties. Data has indicated that DA release in the frontal cortex induced by clozapine and other atypical antipsychotics is mediated by 5-HT_{1A} agonism (Rollema *et al.*, 1997). This provides a mechanism for a potential role of this receptor in alleviating negative symptoms and cognitive impairment, an important property of atypical antipsychotics. Currently, new compounds with strong affinity for this receptor are under development and may shed further light on its contribution to the treatment of schizophrenia.

Clozapine and olanzapine have high affinities for the newly discovered 5-HT₆ receptor ($K_{is} < 20$ nM), while clozapine and risperidone but not olanzapine displayed affinities for the 5-HT₇ receptor lower than 15 nM in cloned rat cells (Roth *et al.*, 1994). In addition, this study showed that several typical antipsychotic agents (chlorpromazine and fluphenazine) had high affinities for both the 5-HT₆ and 5-HT₇ receptors with pimozide displaying the highest affinity of all the typical antipsychotic agents tested for the 5-HT₇ receptor ($K_i = 0.5$ nM). This study seems to indicate that a high affinity for 5-HT₆ or 5-HT₇ does not relate to the atypical properties of antipsychotics, as it is shared by some of the typical antipsychotics, and is present in a minority of atypical antipsychotics, unlike 5-HT_{2A} antagonism.

Studies have shown that serotonin has opposing effects via different serotonergic receptor subtypes. A study by Martin *et al.* (1997) showed that ritanserin, a mixed 5-HT_{2A/2C} antagonist, counteracts the inhibitory effect of MDL

100907 on the hyperlocomotion induced by MK-801 in a mouse model of schizophrenia. In another elegant series of experiments (Martin *et al.*, 1998), the same group showed that the effect of MDL 100907 was abolished by serotonin depletion achieved by pCPA treatment, and restored by restitution of endogenous serotonin. This finding suggests that activation of 5-HT_{2A} receptors is stimulatory while activation of 5-HT_{2C} receptors is inhibitory. A first conclusion from this study is that agonism at 5-HT_{2C} receptor may be antipsychotic. Another conclusion is that 5-HT_{2C} antagonist effects may depend on increased serotonergic tone. This would suggest that a response to atypical antipsychotics may be observed in patients with high serotonergic transmission. This conclusion is in agreement with clinical studies showing that a high serotonin tone, reflected by a low pretreatment CSF HVA/5HIAA ratio, predicts preferential response to clozapine (see references in Martin *et al.*, 1998). According to this line of thinking, agonism at the presynaptic 5-HT_{1A} receptor would result in a decreased serotonergic tone and would diminish the beneficial effects of 5-HT_{2A} antagonists, while therapeutic strategies aiming at increasing serotonergic tone, would be expected to enhance the efficacy of 5-HT_{2A} antagonists. An alternative interpretation of this data is that atypical antipsychotics may benefit most those patients without serotonergic deficits. Severe deficits in serotonergic function may underlie treatment resistance.

IV. 5-HT-DA Interactions Relevant to Schizophrenia

A. VTA DA NEURONS ACTIVITY

Activity of DA neurons is inhibited by raphe stimulation. This effect is mediated by serotonin (Dray *et al.*, 1978; Fibiger and Miller, 1977) and by local dendritic release of DA (Nedergaard *et al.*, 1988; Williams and Davies, 1983) promoting D2 autoreceptor activation. In addition, this inhibitory effect of serotonin on Ventral Tegmental Area (VTA) and SN neurons may be mediated by 5-HT₂ receptors: acute systemic administration of the 5-HT₂ antagonist ritanserin increased the burst firing and firing rate of VTA and SN neurons (Ugedo *et al.*, 1989). This effect required the presence of intact endogenous 5-HT, as it was not observed after pCPA treatment. Thus, VTA and SN are under tonic inhibition by 5-HT neurons, possibly via DA dendritic release mediated by 5-HT₂ receptors. Acute administration of ritanserin increased DA concentration in the extracellular fluid in the accumbens measured with microdialysis (Devaud and Hollingsworth, 1991). Since extracellular DA concentration is thought to depend more on the tonic than the phasic release of DA (Grace, 1991), this observation suggests that 5-HT₂ antagonism increases the tonic release of DA in the terminal fields. Furthermore, VTA neurons are more sensitive than SN neurons to the disinhibiting effect of 5-HT₂ antagonists.

Low doses of ritanserin or ICS 169,369, a highly selective and potent 5-HT₂ antagonist, increase A10 firing rate but do not affect A9 cells. At higher doses, this selective effect of 5-HT₂ antagonists on VTA neurons is lost (Goldstein *et al.*, 1989).

In addition to interactions at the level of the VTA, 5-HT₂ receptors antagonism promotes DA release in the terminal fields of the VTA projection. In the prefrontal cortex, local administration of the 5-HT_{2A} antagonists clozapine, ritanserin, ICS 205,930, amperozide, and MDL 100907 increase DA efflux measured with microdialysis (Hertel *et al.*, 1996). Together, these observations suggest that 5-HT₂ blockade might enhance DA activity at the level of the VTA and the DA terminals.

Activation of VTA DA systems by 5-HT₂ blockade also prevents dysregulation of VTA DA functions observed following reductions in prefrontal cortex glutamatergic input. Local and reversible cooling of the prefrontal cortex alters the pattern of activity of VTA cells, from their normal, burst firing activity to a regular "pacemaker" pattern (Svensson and Tung, 1989). This alteration of firing activity has been proposed to mediate some of the negative symptoms associated with hypofrontality in schizophrenia such as poor drive and reward. Administration of the NMDA antagonist phencyclidine, known to induce both positive and negative symptoms in humans (Allen and Young, 1978; Snyder, 1980), produces the same effects as hypofrontality on VTA neurons, that is a reduction in burst activity (Svensson, 1993). Ritanserin and amperozide, both potent 5-HT₂ blockers, protect VTA DA cells from deactivation induced by cooling of prefrontal cortex or phencyclidine administration (Grenhoff *et al.*, 1990; Svensson, 1993; Svensson *et al.*, 1989, 1995). These observations are also compatible with a stimulating effect of 5-HT₂ antagonists on DA neuronal activity.

In summary, 5-HT₂ antagonists might reduce negative symptoms in schizophrenia through activation of midbrain DA projections to the limbic system and cerebral cortex. Since VTA DA neurons projecting to the accumbens are involved in drive and reward (Bozarth, 1986), it has been proposed that activation of VTA neurons by 5-HT₂ antagonists might provide a basis for their thymosthenic action and the improvement in negative symptoms (Ugedo *et al.*, 1989).

Behavioral studies performed with raphe lesions or pCPA treatment have consistently demonstrated an *enhancement* of amphetamine effects after 5-HT depletion. However, more recently, electrophysiological and microdialysis studies have demonstrated that selective 5-HT₂ blockade *decreases* amphetamine effects on locomotion. MDL 100907, a selective 5-HT_{2A} antagonist, blocks amphetamine stimulated locomotion at a dose that does not affect spontaneous locomotion (Sorensen *et al.*, 1993). While having no effect on basal DA extracellular concentration, 5-HT₂ antagonists such as MDL 100907, amperozide, and ketanserin decrease MDMA-mediated DA release measured by microdialysis

(Nash, 1990; Schmidt *et al.*, 1994). Electrophysiological data support the fact that 5-HT₂ blockade decreases DA mediated amphetamine effects by interfering with regulation of DA synthesis.

D-Amphetamine produces a marked inhibition of DA activity as recorded by single-cell recording. This inhibition is due to neural feedback loops in the SN and to the stimulation of somatodendritic receptors following DA release in VTA (Bunney and Aghajanian, 1976; Wang, 1981). α -Methyl-paratyrosine (α MPT), an inhibitor of tyrosine hydroxylase, the rate-limiting step in DA synthesis, attenuates amphetamine-induced DA release (Butcher *et al.*, 1988) and blocks amphetamine-induced slowing of cell firing (Bunney and Aghajanian, 1976). Thus, DA synthesis plays a major role in the effect of amphetamine on DA neurons. Interestingly, the selective 5-HT₂ antagonists ritanserin and MDL 28,133A also significantly suppress the effect of amphetamine on VTA neurons. This effect was, however, restored when L-dopa was coadministered with a 5-HT₂ antagonist. Since L-dopa enters the DA synthetic pathway beyond the point of synthesis regulation (tyrosine hydroxylase), it was proposed that 5-HT regulates tyrosine hydroxylase activity via 5-HT₂ receptors. While the 5-HT₂ agonist DOI hydrochloride does not appear to increase DA synthesis when given alone, it greatly potentiates amphetamine-induced increase in DA synthesis (Huang and Nichols, 1993) and amphetamine-induced DA release (Ichikawa and Meltzer, 1995). These data suggest that 5-HT₂ stimulation may be needed to maintain the increase in phasic DA neuronal activity, as observed after administration of stimulants or during stress. Thus, in this model, 5-HT₂ blockade would decrease DA phasic activity (a tyrosine hydroxylase dependent process) without affecting tonic basal DA activity (Schmidt *et al.*, 1993). Since positive symptoms are associated with increased DA release in schizophrenia (Laruelle *et al.*, 1996), these preclinical observations suggest that 5-HT₂ antagonism might have a beneficial effect on positive symptoms.

V. Discussions

The hypothesis that schizophrenia may be associated with decreased tonic DA activity and increased phasic activity (Grace, 1993) provides a framework in which the effects of a balanced 5-HT₂/D2 antagonism could be conceptualized. The tonic mode, or baseline mode, plays a role in motivation and drive, and appears to be regulated by a corticostriatal and cortico-VTA glutamatergic input. In contrast, the phasic mode is responsible for a rapid increase of DA in the synapse, in response to emotions or stress. One function of the tonic baseline activity is to regulate the sensitivity of the system to the phasic release of DA. A decrease in the tonic activity would result in an increased sensitivity of the

phasic activity. In schizophrenia, cortical lesions may induce a hypoactivity of the corticostriatal and corticolimbic glutamatergic projections, leading to a decrease in the tonic release of DA, which may be associated with negative symptoms such as lack of drive and motivation. This decreased tonic activity in turn induces a state of hypersensitivity of the DA system to the phasic release, which can be the substrate of positive symptoms (Grace, 1993).

Examination of 5-HT-DA interactions mediated by the 5-HT_{2A} receptors lead to the following suggestion regarding the mechanism of action of "balanced" 5-HT_{2A}/D2 agents. It is suggested that 5-HT_{2A} blockade acts as a buffer to narrow the range of DA activity in the VTA projection territories, elevating the baseline activity (thus reducing negative symptoms) and decreasing the amplitude of the phasic reactivity (thus reducing positive symptoms). Decreased tonic activity of VTA is observed after inactivation of the frontal cortex or administration of NMDA antagonists. 5-HT_{2A} antagonists restore normal VTA DA activity after inactivation of the frontal cortex, and this might account for improvement in negative symptoms. Obviously, if stimulation of mesocorticolimbic DA baseline activity is the mechanism mediating the improvement in negative symptoms attributed to 5-HT_{2A} antagonism, this effect would be lost in the presence of a complete blockade of D2 receptors (Kapur and Remington, 1996). Thus, a moderate rather than a complete D2 receptors occupancy might be desirable to permit 5-HT_{2A} antagonism to exert its therapeutic action on the negative symptoms.

On the other hand, positive symptoms could be reduced by attenuation of DA phasic activity via blockade of 5-HT_{2A}-stimulated tyrosine hydroxylase activity. If increased DA phasic activity is at least partially dependent on 5-HT_{2A}-stimulated tyrosine hydroxylase activity, 5-HT₂ blockade might reduce the tyrosine hydroxylase dependent DA phasic release responsible for the positive symptoms of schizophrenia. One study seems to contradict this hypothesis so far, where clozapine and risperidone treatment did not normalize the amphetamine-induced DA release in patients with schizophrenia compared to controls (Breier *et al.*, 1999). However, the effect of D2 blockade may have confounded the interpretation of the effect of the 5-HT₂ antagonism in this study. Revisiting this issue after treatment with a pure 5-HT₂ antagonist such as MDL 100907 is warranted.

So far, 5-HT_{2A} antagonists show modest efficacy as stand-alone treatments for schizophrenia. However, they do not appear to be as effective in treating schizophrenia as haloperidol. In addition, it is now clear that 5-HT_{2A} antagonists do not increase the D2 receptor blocking threshold associated with emergence of EPS since the threshold of D2 receptor occupancy associated with EPS is not markedly different between these drugs and drugs devoid of 5-HT_{2A} antagonism (Kapur *et al.*, 1995, 1998; Knable *et al.*, 1997; Nyberg *et al.*, 1993). The benefit derived from the combination of 5-HT_{2A} antagonism with a partial D2 blockade may relate to the following two factors: (1) 5-HT_{2A} antagonism increases DA release in the

cortex, this effect might lead to an improvement in negative symptoms and cognition, possibly through an increased stimulation of the D1 receptor. This effect may be potentiated by agonism at the 5-HT_{1A} receptor. (2) 5-HT_{2A} antagonists may decrease the tyrosine hydroxylase dependent, or phasic, DA release in subcortical areas and improve positive symptoms, 5-HT_{2A} antagonists may be even more beneficial in presence of high serotonergic tone, since increased stimulation of the 5-HT_{2A} receptors, because of their location on apical dendrites of most pyramidal cells in the cortex, may lead to a dysregulation of glutamatergic transmission and corticocortical as well as corticosubcortical transmission. As we understand better the role of the 5-HT_{2A} receptors in the treatment of schizophrenia, much remains to be learned about the other receptors, although the evidence so far does not suggest they play a role as prominent as the 5-HT_{2A}. More research is needed to further clarify the role of these receptors.

In conclusion, results of postmortem studies indicate possible alteration of 5-HT transmission in the prefrontal cortex of patients with schizophrenia. Decreased 5-HT transporter density, increased 5-HT_{1A} receptors, and decreased 5-HT_{2A} receptors have all been suggested by postmortem studies in schizophrenia, but these findings have not been consistently replicated and have not yielded conclusive evidence so far. On the other hand, stimulation of 5-HT_{2A} receptors is psychogenic, although LSD-induced psychosis is an imperfect model of the illness. 5-HT_{2A} receptor blockade is a useful augmentation or modulation of D2 receptor blockade, but 5-HT_{2A} antagonism alone has not yet demonstrated incisive antipsychotic properties. If 5-HT_{2A} blockade per se does not produce antipsychotic effects in patients with schizophrenia, it would support the argument that these symptoms are not primarily due to hyperstimulation of 5-HT_{2A} receptors. The potential of pharmacological interventions targeted at other 5-HT receptors (such as 5-HT_{1A} agonism) remains to be clarified. Overall, a comprehensive model of alterations of 5-HT transmission in schizophrenia has not yet emerged and additional research is needed, not only to clarify possible alterations of 5-HT systems in schizophrenia but also to establish their significance in terms of modulation of other transmitters systems (DA and glutamate), role in symptomatology, and treatment opportunity.

References

- Abi-Dargham, A., Laruelle, M., Lipska, B., Jaskiw, G. E., Wong, D. T., Robertson, D. W., Weinberger, D. R., and Kleinman, J. E. (1993). Serotonin 5-HT₃ receptors in schizophrenia: A postmortem study of the amygdala. *Brain Res.* **616**, 53–57.
- Abi-Dargham, A., Laruelle, M., Aghajanian, G. K., Charney, D., and Krystal, J. (1997). The role of serotonin in the pathophysiology and treatment of schizophrenia. *J. Neuropsychiatry Clin. Neurosci.* **9**, 1–17.

- Aghajanian, G. K. (1994). Serotonin and the action of LSD in the brain. *Psychiatr. Ann.* **24**, 137–141.
- Aghajanian, G. K., and Marek, G. J. (1999). Serotonin, via 5-HT_{2A} receptors, increases EPSCs in layer V pyramidal cells of prefrontal cortex by an asynchronous mode of glutamate release. *Brain Res.* **825**, 161–171.
- Allen, R. M., and Young, S. J. (1978). Phencyclidine-induced psychosis. *Am. J. Psychiatry* **135**, 1081–1084.
- Altar, C. A., Wasley, A. M., Neale, R. F., and Stone, G. A. (1986). Typical and atypical antipsychotic occupancy of D₂ and S₂ receptors: An autoradiographic analysis in rat brain. *Brain Res. Bull.* **16**, 517–525.
- Andree, T. H., Mikini, M., Yong, C. Y., Koenig, J. I., and Meltzer, H. Y. (1986). Differential effects of subchronic treatment with various neuroleptic agents on serotonin₂ receptors in rat cerebral cortex. *J. Neurochem.* **46**, 191–197.
- Arango, V., Ernsberg, P., Mazuk, P. M., Chen, J. S., Tierney, H., Stanley, M., Reis, D. J., and Mann, J. J. (1990). Autoradiographic demonstration of increased 5-HT₂ and β -adrenergic receptors in the brains of suicide victims. *Arch. Gen. Psychiatry* **47**, 1038–1047.
- Arango, V., Underwood, M. D., Gubbi, A. V., and Mann, J. J. (1995). Localized alterations in pre- and postsynaptic serotonin binding sites in the ventrolateral prefrontal cortex of suicide victims. *Brain Res.* **688**, 121–133.
- Arnt, J. (1998). Pharmacological differentiation of classical and novel antipsychotics. *Int. Clin. Psychopharmacol.* **13**(Suppl. 3), S7–S14.
- Arora, R. C., and Meltzer, H. Y. (1991). Serotonin₂ (5-HT₂) receptor binding in frontal cortex of schizophrenic patients. *J. Neural. Transm.* **85**, 19–29.
- Arranz, B., Eriksson, A., Mellerup, E., Plenge, P., and Marcusson, J. (1994). Brain 5-HT_{1A}, 5-HT_{1D}, and 5-HT₂ receptors in suicide victims. *Biol. Psychiatry* **35**, 457–463.
- Arvanitis, L. A., and Miller, B. G. (1997). Multiple fixed doses of “Seroquel” (quetiapine) in patients with acute exacerbation of schizophrenia: A comparison with haloperidol and placebo. The Seroquel Trial 13 Study Group. *Biol. Psychiatry* **42**, 233–246.
- Azmitia, E. C., Gannon, P. J., Kheck, N. M., and Whitaker-Azmitia, P. M. (1996). Cellular localization of the 5-HT_{1A} receptor in primate brain neurons and glial cells. *Neuropsychopharmacology* **14**, 35–46.
- Bennett, J. P., Enna, S. J., Bylund, D. B., Gillin, J. C., Wyatt, R. J., and Snyder, S. H. (1979). Neurotransmitter receptors in frontal cortex of schizophrenics. *Arch. Gen. Psychiatry* **36**, 927–934.
- Bersani, G., Grispini, A., Marini, S., Valducci, M., and Ciani, N. (1990). 5-HT₂ antagonist ritanserin in neuroleptic induced parkinsonism: A double blind comparison with orphenadrine and placebo. *Clin. Neuropharmacol.* **13**, 500–506.
- Bigelow, L., Walls, P., Gillin, J., and Wyatt, R. (1979). Clinical effects of L-5-hydroxytryptophan administration in chronic schizophrenic patients. *Biol. Psychiatry* **14**, 53–67.
- Bleekers, E., Goldfarb, J., and Green, J. P. (1990). Ritanserin in the treatment of negative symptoms in chronic schizophrenic patients. In “Collegium Internationale Neuro-psychopharmacologicum (CINP).” Kyoto, Japan, p. 221.
- Bowers, M. B. J. (1970). Cerebrospinal fluid 5-hydroxyindoles and behavior after L-tryptophan and pyridoxine administration to psychiatric patients. *Neuropharmacology* **9**, 599–604.
- Bozarth, M. A. (1986). Neural basis of psychomotor stimulant and opiate reward: Evidence suggesting an involvement of a common dopaminergic system. *Behav. Brain Res.* **22**, 107–116.
- Breier, A. (1995). Serotonin, schizophrenia and antipsychotic drug action. *Schizophr. Res.* **14**, 187–202.
- Breier, A., Su, T. P., Malhotra, A. K., Elman, L., Adler, C. M., Weisenfeld, N. I., and Pickar, D. (1999). Effects of atypical antipsychotic drug treatment on amphetamine-induced striatal dopamine release in patients with psychotic disorders. *Neuropsychopharmacology* **20**, 340–345.

- Bristow, L. J., Kramer, M. S., Kulagowski, J., Patel, S., Ragan, C. I., and Seabrook, G. R. (1997). Schizophrenia and L-745,870, a novel dopamine D4 receptor antagonist. *Trends Pharmacol. Sci.* **18**, 186–188.
- Bunney, B. S., and Aghajanian, G. K. (1976). *d*-Amphetamine-induced inhibition of central dopaminergic neurons: Mediation by a striato-nigral feedback pathway. *Science* **192**, 391–393.
- Burnet, P. W., Chen, C. P., McGowan, S., Franklin, M., and Harrison, P. J. (1996a). The effects of clozapine and haloperidol on serotonin-1A, -2A and -2C receptor gene expression and serotonin metabolism in the rat forebrain. *Neuroscience* **73**, 531–540.
- Burnet, P. W., Eastwood, S. L., and Harrison, P. J. (1996b). 5-HT_{1A} and 5-HT_{2A} receptor mRNAs and binding site densities are differentially altered in schizophrenia. *Neuropsychopharmacology* **15**, 442–455.
- Burnet, P. W. J., Eastwood, S. L., and Harrison, P. J. (1997). [H-3]WAY-100635 for 5-HT_{1A} receptor autoradiography in human brain: A comparison with [H-3]8-OH-DPAT and demonstration of increased binding in the frontal cortex in schizophrenia. *Neurochem. Int.* **30**, 565–574.
- Butcher, S. P., Fairbrother, I. S., Kelly, J. S., and Arbutnot, G. W. (1988). Amphetamine-induced dopamine release in rat striatum: An *in vivo* microdialysis study. *J. Neurochem.* **50**, 346–355.
- Bymaster, F. P., Nelson, D. L., DeLapp, N. W., Falcone, J. F., Eckols, K., Truex, L. L., Foreman, M. M., Lucaites, V. L., and Calligaro, D. O. (1999). Antagonism by olanzapine of dopamine D₁, serotonin₂, muscarinic, histamine H₁ and alpha 1-adrenergic receptors *in vitro*. *Schizophr. Res.* **37**, 107–122.
- Canton, H., Verrielle, L., and Colpaert, F. (1990). Binding of typical and atypical antipsychotics to 5-HT_{1c} and 5-HT₂ sites: Clozapine potently interact with 5-HT_{1c} sites. *Eur. J. Pharmacol.* **191**, 93–96.
- Cheetham, S. C., Crompton, M. R., Katona, C. L. E., and Horton, R. W. (1988). Brain 5-HT₂ receptor binding sites in depressed suicide victims. *Brain Res.* **443**, 272–280.
- Choudhary, M. S., Craig, S., and Roth, B. L. (1992). Identification of receptor domains that modify ligand binding to 5-hydroxytryptamine₂ and 5-hydroxytryptamine_{1c} serotonin receptors. *Mol. Pharmacol.* **42**, 627–633.
- Chouinard, G., Annable, L., Young, S., and Sourkes, T. L. (1978). A controlled study of tryptophan-benserazide in schizophrenia. *Commun. Psychopharmacol.* **2**, 21–31.
- Claus, A., Bollen, J., DeCuyper, H., Eneman, M., Malfroid, M., Peuskens, J., and Heylen, S. (1992). Risperidone versus haloperidol in the treatment of chronic schizophrenic inpatients: A multicentre double-blind comparative study. *Acta Psychiatr. Scand.* **85**, 295–305.
- Creese, I., Burt, D. R., and Snyder, S. H. (1976). Dopamine receptor binding predicts clinical and pharmacological potencies of antischizophrenic drugs. *Science* **19**, 481–483.
- Cruz, D. A., Eggen, S. M., Azmitia, E. C., and Lewis, D. A. (2004). Serotonin_{1A} receptors at the axon initial segment of prefrontal pyramidal neurons in schizophrenia. *Am. J. Psychiatry* **161**, 739–742.
- Daniel, D. G., Zimbroff, D. L., Potkin, S. G., Reeves, K. R., Harrigan, E. P., and Lakshminarayanan, M. (1999). Ziprasidone 80 mg/day and 160 mg/day in the acute exacerbation of schizophrenia and schizoaffective disorder: A 6-week placebo-controlled trial. Ziprasidone Study Group. *Neuropsychopharmacology* **20**, 491–505.
- Dean, B., Opeskin, K., Pavey, G., Naylor, L., Hill, C., Keks, N., and Copolov, D. L. (1995). [3H] paroxetine binding is altered in the hippocampus but not the frontal cortex or caudate nucleus from subjects with schizophrenia. *J. Neurochem.* **64**, 1197–1202.
- Dean, B., Tomaskovic-Crook, E., Opeskin, K., Keks, N., and Copolov, D. (1999). No change in the density of the serotonin_{1A} receptor, the serotonin₄ receptor or the serotonin transporter in the dorsolateral prefrontal cortex from subjects with schizophrenia. *Neurochem. Int.* **34**, 109–115.
- DeLisi, L., Freed, W., Gillin, J., Kleinman, J., Bigelow, L., and Wyatt, R. (1982). P-chlorophenylalanine trials in schizophrenic patients. *Biol. Psychiatry* **11**, 471–477.

- Deutch, A., Moghadam, B., Innis, R., Krystal, J., Aghajanian, G., Bunney, B., and Charney, D. (1991). Mechanisms of action of atypical antipsychotic drugs. Implication for novel therapeutic strategies for schizophrenia. *Schizophr. Res.* **4**, 121–156.
- Deutch, A. Y., Lee, M. C., and Iadarola, M. J. (1992). Regionally specific effects of atypical antipsychotic drugs on striatal fos expression: The nucleus accumbens shell as a locus of antipsychotic action. *Mol. Cell. Neurosci.* **3**, 332–341.
- Devaud, L. L., and Hollingsworth, E. B. (1991). Effect of the 5-HT₂ antagonist, ritanserin, on biogenic amines in the rat nucleus accumbens. *Eur. J. Pharmacol.* **192**, 427–429.
- Dillon, K. A., Gross-Isseroff, R., Israeli, M., and Biegon, A. (1991). Autoradiographic analysis of serotonin 5-HT_{1A} receptor binding in the human brain postmortem: Effects of age and alcohol. *Brain Res.* **554**, 56–64.
- Dray, A., Davies, J., Oakley, N. R., Tongroach, P., and Vellucci, S. (1978). The dorsal and medial raphe projections to the substantia nigra in the rat: Electrophysiological, biochemical and behavioural observations. *Brain Res.* **151**, 431–442.
- Farber, N. B., Newcomer, J. W., and Olney, J. W. (1998). The glutamate synapse in neuropsychiatric disorders. Focus on schizophrenia and Alzheimer's disease. *Prog. Brain Res.* **116**, 421–437.
- Fibiger, H. C., and Miller, J. J. (1977). An anatomical and electrophysiological investigation of the serotonergic projection from the dorsal raphe nucleus to the substantia nigra in the rat. *Neuroscience* **2**, 975–987.
- Fishman, L. (1983). Dreams, hallucinogenic drug states and schizophrenia: A psychological and biological comparison. *Schizophr. Bull.* **9**, 73–94.
- Francis, P. T., Pangalos, M. N., Pearson, R. C., Middlemiss, D. N., Stratmann, G. C., and Bowen, D. M. (1992). 5-Hydroxytryptamine_{1A} but not 5-hydroxytryptamine₂ receptors are enriched on neocortical pyramidal neurones destroyed by intrastriatal volkensin. *J. Pharmacol. Exp. Ther.* **261**, 1273–1281.
- Frankle, W. G., Huang, Y., Hwang, D. R., Talbot, P. S., Slifstein, M., Van Heertum, R., Abi-Dargham, A., and Laruelle, M. (2004). Comparative evaluation of serotonin transporter radioligands ¹¹C-DASB and ¹¹C-McN 5652 in healthy humans. *J. Nucl. Med.* **45**, 682–694.
- Freedman, B., and Chapman, L. (1973). Early subjective experiences in schizophrenic episodes. *J. Abnormal Psychol.* **82**, 46–54.
- Freedman, D. (1961). Effects of LSD-25 on brain serotonin. *J. Pharmacol. Exp. Ther.* **134**, 160–166.
- Gaddum, J. H. (1954). Drug antagonistic to 5-hydroxytryptamine. In "Ciba Foundation Symposium on Hypertension" (G. W. Wolstenholme, Ed.), pp. 75–77. Little Brown and Company, Boston.
- Gelders, Y. G. (1989). Thymosthenic agents, a novel approach in the treatment of schizophrenia. *Br. J. Psychiatry* **155**, 33–36.
- Gellman, R. L., and Aghajanian, G. K. (1993). Pyramidal cells in piriform cortex receive a convergence of inputs from monoamine activated GABAergic interneurons. *Brain Res.* **600**, 63–73.
- Gillin, J., Kaplan, J., and Wyatt, R. (1976). Clinical effects of tryptophan in chronic schizophrenic patients. *Biol. Psychiatry* **11**, 635–639.
- Genovart, N., Wilson, A. A., Meyer, J. H., Hussey, D., and Houle, S. (2001). Positron emission tomography quantification of [¹¹C]-DASB binding to the human serotonin transporter: Modeling strategies. *J. Cereb. Blood Flow Metab.* **21**, 1342–1353.
- Glennon, R. (1990). Do classical hallucinogens act as 5-HT₂ agonists or antagonists? *Neuropsychopharmacology* **3**, 509–517.
- Glennon, R., and Titeler, M. (1984). Evidence for 5-HT₂ involvement in the mechanism of action of hallucinogenic agents. *Life Sci.* **35**, 2505–2511.
- Goldstein, J. M., Litwin, L. C., Sutton, E. B., and Malick, J. B. (1989). Effects of ICI 169,369, a selective serotonin₂ antagonist, in electrophysiological tests predictive of antipsychotic activity. *J. Pharmacol. Exp. Ther.* **249**, 673–680.

- Grace, A. A. (1991). Phasic versus tonic dopamine release and the modulation of dopamine system responsivity: A hypothesis for the etiology of schizophrenia. *Neuroscience* **41**, 1–24.
- Grace, A. A. (1993). Cortical regulation of subcortical systems and its possible relevance to schizophrenia. *J. Neural. Transm.* **91**, 111–134.
- Grenhoff, J., Tung, C. S., Ugedo, L., and Svensson, T. H. (1990). Effects of amperozide, a putative antipsychotic drug, on rat midbrain dopamine neurons recorded *in vivo*. *Pharmacol. Toxicol.* **66**(Suppl. 1), 29–33.
- Gross-Isseroff, R., Salama, D., Israeli, M., and Biegon, A. (1990). Autoradiographic analysis of [3 H] ketanserin binding in the human brain postmortem: Effect of suicide. *Brain Res.* **507**, 208–215.
- Gurevich, E. V., and Joyce, J. N. (1997). Alterations in the cortical serotonergic system in schizophrenia: A postmortem study. *Biol. Psychiatry* **42**, 529–545.
- Hashimoto, T., Nishino, N., Nakai, H., and Tanaka, C. (1991). Increase in serotonin 5-HT_{1A} receptors in prefrontal and temporal cortices of brains from patients with chronic schizophrenia. *Life Sci.* **48**, 355–363.
- Helmeste, D. M., and Tang, S. W. (1983). Unusual acute effects of antidepressants and neuroleptics on 5-HT₂-serotonergic receptors. *Life Sci.* **33**, 2527–2533.
- Hensler, J. G., Kovachich, G. B., and Frazer, A. (1991). A quantitative autoradiographic study of serotonin_{1A} receptor regulation. Effect of 5,7-dihydroxytryptamine and antidepressant treatments. *Neuropsychopharmacology* **4**, 131–144.
- Hermle, L., Funfgeld, M., Oepen, G., Botsch, H., Borchardt, D., Gouzoulis, E., Fehrenbach, R., and Spitzer, M. (1992). Mescaline-induced psychopathological, neuropsychological and neurometabolic effects in normal subjects: Experimental psychosis as a tool for psychiatric research. *Biol. Psychiatry* **32**, 976–991.
- Hertel, P., Nomikos, G. G., Iurlo, M., and Svensson, T. H. (1996). Risperidone: Regional effects *in vivo* on release and metabolism of dopamine and serotonin in the rat brain. *Psychopharmacology (Berl.)* **124**, 74–86.
- Hollister, L. E. (1962). Drug-induced psychoses and schizophrenic reactions: A critical comparison. *Ann. N. Y. Acad. Sci.* **96**, 80–88.
- Houle, S., Ginovart, N., Hussey, D., Meyer, J. H., and Wilson, A. A. (2000). Imaging the serotonin transporter with positron emission tomography: Initial human studies with [11 C]DAPP and [11 C]DASB. *Eur. J. Nucl. Med.* **27**, 1719–1722.
- Hoyberg, O. J., Fensbo, C., Remvig, J., Lingjaerde, O., Sloth-Nielsen, M., and Salvesen, I. (1993). Risperidone versus perphenazine in the treatment of chronic schizophrenic patients with acute exacerbations. *Acta Psychiatr. Scand.* **88**, 395–402.
- Hoyer, D., Pazos, A., Probst, A., and Palacios, J. M. (1986a). Serotonin receptors in the human brain. I. Characterization and autoradiographic localization of 5-HT_{1A} recognition sites. Apparent absence of 5-HT_{1B} recognition sites. *Brain Res.* **376**, 85–96.
- Hoyer, D., Pazos, A., Probst, A., and Palacios, J. M. (1986b). Serotonin receptors in the human brain. II. Characterization and autoradiographic localization of 5-HT_{1C} and 5-HT₂ recognition sites. *Brain Res.* **376**, 97–107.
- Hrdina, P. D., Demeter, E., Vu, T. B., Sótónyi, P., and Palkovits, M. (1993). 5-HT uptake sites and 5-HT₂ receptors in brain of antidepressant-free suicide victims/depressives: Increase in 5-HT₂ sites in cortex and amygdala. *Brain Res.* **614**, 37–44.
- Huang, X., and Nichols, D. E. (1993). 5HT₂ mediated potentiation of dopamine synthesis and central serotonergic deficits. *Eur. J. Pharmacol.* **238**, 291–296.
- Huang, Y., Hwang, D. R., Narendran, R., Sudo, Y., Chatterjee, R., Bae, S. A., Mawlawi, O., Kegeles, L. S., Wilson, A. A., Kung, H. F., and Laruelle, M. (2002). Comparative evaluation in nonhuman primates of five PET radiotracers for imaging the serotonin transporters: [11 C]McN 5652, [11 C]ADAM, [11 C]DASB, [11 C]DAPA, and [11 C]AFM. *J. Cereb. Blood Flow Metab.* **22**, 1377–1398.

- Ichikawa, J., and Meltzer, H. Y. (1995). D. O. L., a 5-HT_{2A/2C} receptor agonist, potentiates amphetamine-induced dopamine release in rat striatum. *Brain Res.* **698**, 204–208.
- Joyce, J. N., Shane, A., Lexow, N., Winokur, A., Casanova, M. F., and Kleinman, J. E. (1993). Serotonin uptake sites and serotonin receptors are altered in the limbic system of schizophrenics. *Neuropsychopharmacology* **8**, 315–336.
- Kane, J., Honigfeld, G., Singer, J., Meltzer, H. Y., and the Clozaril Collaborative Study Group (1988). Clozapine for the treatment-resistant schizophrenic. *Arch. Gen. Psychiatry* **45**, 789–796.
- Kapur, S., and Remington, G. (1996). Serotonin-dopamine interaction and its relevance to schizophrenia. *Am. J. Psychiatry* **153**, 466–476[Review].
- Kapur, S., Remington, G., Zipursky, R. B., Wilson, A. A., and Houle, S. (1995). The D2 dopamine receptor occupancy of risperidone and its relationship to extrapyramidal symptoms: A PET study. *Life Sci.* **57**, L103–L107.
- Kapur, S., Zipursky, R. B., Remington, G., Jones, C., DaSilva, J., Wilson, A. A., and Houle, S. (1998). 5-HT₂ and D2 receptor occupancy of olanzapine in schizophrenia: A PET investigation. *Am. J. Psychiatry* **155**, 921–928.
- Keck, P., Jr., Buffenstein, A., Ferguson, J., Feighner, J., Jaffe, W., Harrigan, E. P., and Morrissey, M. R. (1998). Ziprasidone 40 and 120 mg/day in the acute exacerbation of schizophrenia and schizoaffective disorder: A 4-week placebo-controlled trial. *Psychopharmacology (Berl.)* **140**, 173–184.
- Kia, H. K., Miquel, M. C., Brisorgueil, M. J., Daval, G., Riad, M., El Mestikawy, S., Hamon, M., and Verge, D. (1996). Immunocytochemical localization of serotonin_{1A} receptors in the rat central nervous system. *J. Comp. Neurol.* **365**, 289–305.
- Knable, M. B., Heinz, A., Raedler, T., and Weinberger, D. R. (1997). Extrapyramidal side effects with risperidone and haloperidol at comparable D2 receptor occupancy levels. *Psychiatr. Res. Neuroimag.* **75**, 91–101.
- Kohen, R., Metcalf, M. A., Khan, N., Druck, T., Huebner, K., Lachowicz, J. E., Meltzer, H. Y., Sibley, D. R., Roth, B. L., and Hamblin, M. W. (1996). Cloning, characterization, and chromosomal localization of a human 5-HT₆ serotonin receptor. *J. Neurochem.* **66**, 47–56.
- Kramer, M. S., Last, B., Getson, A., and Reines, S. A. (1997). The effects of a selective D4 dopamine receptor antagonist (L-745,870) in acutely psychotic inpatients with schizophrenia. D4 Dopamine Antagonist Group. *Arch. Gen. Psychiatry* **54**, 567–572.
- Langs, R. J., and Barr, H. L. (1968). Lysergic acid diethylamide (LSD-25) and schizophrenic reactions. *J. Nerv. Ment. Dis.* **147**, 163–172.
- Laruelle, M., Abi-Dargham, A., Casanova, M. F., Toti, R., Weinberger, D. R., and Kleinman, J. E. (1993). Selective abnormalities of prefrontal serotonergic receptors in schizophrenia: A postmortem study. *Arch. Gen. Psychiatry* **50**, 810–818.
- Laruelle, M., Abi-Dargham, A., van Dyck, C. H., Gil, R., D'Souza, C. D., Erdos, J., McCance, E., Rosenblatt, W., Fingado, C., Zoghbi, S. S., Baldwin, R. M., Seibyl, J. P., et al. (1996). Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug-free schizophrenic subjects. *Proc. Natl. Acad. Sci. USA* **93**, 9235–9240.
- Laruelle, M., Abi-Dargham, A., van Dyck, C., Gil, R., D'Souza, D. C., Krystal, J., Seibyl, J., Baldwin, R., and Innis, R. (2000). Dopamine and serotonin transporters in patients with schizophrenia: An imaging study with [¹²³I]beta-CIT. *Biol. Psychiatry* **47**, 371–379.
- Lauer, J., Inskip, W., Bernshon, J., and Zeller, E. (1958). Observations on schizophrenic patients after iproniazid and tryptophan. *Arch. Neurol. Psychiatr.* **80**, 122.
- Lawrence, J. A., Olverman, H. J., Shirakawa, K., Kelly, J. S., and Butcher, S. P. (1993). Binding of 5-HT_{1A} receptor and 5-HT transporter ligands in rat cortex and hippocampus following cholinergic and serotonergic lesions. *Brain Res.* **612**, 326–329.
- Leucht, S., Pitschel-Walz, G., Abraham, D., and Kissling, W. (1999). Efficacy and extrapyramidal side-effects of the new antipsychotics olanzapine, quetiapine, risperidone, and sertindole compared

- to conventional antipsychotics and placebo. A meta-analysis of randomized controlled trials. *Schizophr. Res.* **35**, 51–68.
- Lewis, R., Kapur, S., Jones, C., DaSilva, J., Poy, P., Brown, G., Wilson, A., Houle, S., and Zipursky, R. (1997). PET study of 5-HT₂ receptors density in schizophrenia. *Biol. Psychiatry* **41**, 63S.
- Lewis, R., Kapur, S., Jones, C., DaSilva, J., Brown, G. M., Wilson, A. A., Houle, S., and Zipursky, R. B. (1999). Serotonin 5-HT₂ receptors in schizophrenia: A PET study using [18F]setoperone in neuroleptic-naïve patients and normal subjects. *Am. J. Psychiatry* **156**, 72–78.
- Leysen, J., Niemegeers, C., Tollenaere, J., and Laduron, P. (1978). Serotonergic component of neuroleptic receptors. *Nature* **272**, 168–171.
- Leysen, J. E. (1990). Gaps and peculiarities in 5-HT₂ receptor studies. *Neuropsychopharmacology* **3**, 361–369.
- Leysen, J. E., Niemegeers, J. E., Van Nueten, J. M., and Laduron, P. M. (1982). [³H]Ketanserin (R 41 468), a selective ³H-ligand for serotonin-2 receptor binding sites: Binding properties, brain distribution and functional role. *Mol. Pharmacol.* **21**, 301–314.
- Leysen, J. E., Van Gompel, P., de Chaffoy de Courcelles, D., and Niemegeers, C. J. E. (1987). Opposite regulation of serotonin S₂ and dopamine D₂ receptors in rat brain following chronic receptor blockade. *J. Recep. Res.* **7**, 223–239.
- Lidow, M. S., Goldman-Rakic, P. S., Gallager, D. W., and Rakic, P. (1989). Quantitative autoradiographic mapping of serotonin 5-HT₁ and 5-HT₂ receptors and uptake sites in the neocortex of the rhesus monkey. *J. Comp. Neurol.* **280**, 27–42.
- Lieberman, J. A. (1993). Understanding the mechanism of action of atypical antipsychotic drugs. *Br. J. Psychiatry* **163**, 7–18.
- Lieberman, J. A., Mailman, R. B., Duncan, G., Sikich, L., Chakos, M., Nichols, D. E., and Kraus, J. E. (1998). Serotonergic basis of antipsychotic drug effects in schizophrenia. *Biol. Psychiatry* **44**, 1099–1117.
- Lowther, S., De Paermentier, F., Crompton, R., Katona, C. L. E., and Horton, R. W. (1994). Brain 5-HT₂ receptors in suicide victims: Violence of death, depression and effects of antidepressant treatment. *Brain Res.* **642**, 281–289.
- Lowther, S., DePaermentier, F., Cheetham, S. C., Crompton, M. R., Katona, C. L. E., and Horton, R. W. (1997). 5-HT_{1A} receptor binding sites in post-mortem brain samples from depressed suicides and controls. *J. Affect. Disord.* **42**, 199–207.
- Mann, J. J., Stanley, M., Mc Bride, P. A., and Mc Ewen, B. S. (1986). Increased serotonin₂ and beta receptor in the frontal cortices of suicide victims. *Arch. Gen. Psychiatry* **43**, 954–959.
- Marder, S. R., and Meibach, R. C. (1994). Risperidone in the treatment of schizophrenia. *Am. J. Psychiatry* **151**, 825–835.
- Marek, G. J., and Aghajanian, G. K. (1994). Excitation of interneurons in piriform cortex by 5-hydroxytryptamine: Blockade by MDL 100,907, a highly selective 5-HT_{2A} receptor antagonist. *Eur. J. Pharmacol.* **259**, 137–141.
- Marek, G. J., and Aghajanian, G. K. (1996). LSD and the phenethylamine hallucinogen DOI are potent partial agonists at 5-HT_{2A} receptors on interneurons in rat piriform cortex. *J. Pharmacol. Exp. Ther.* **278**, 1373–1382.
- Martin, G., and Humphrey, P. (1994). Classification review: Receptors for 5-hydroxytryptamine: Current perspectives on classification and nomenclature. *Neuropharmacology* **33**, 261–273.
- Martin, P., Waters, N., Carlsson, A., and Carlsson, M. L. (1997). The apparent antipsychotic action of the 5-HT_{2a} receptor antagonist M100907 in a mouse model of schizophrenia is counteracted by ritanserin—Rapid communication. *J. Neural. Transm.* **104**, 561–564.
- Martin, P., Waters, N., Schmidt, C. J., Carlsson, A., and Carlsson, M. L. (1998). Rodent data and general hypothesis: Antipsychotic action exerted through 5-HT_{2A} receptor antagonism is dependent on increased serotonergic tone. *J. Neural. Transm.* **105**, 365–396.

- Matsubara, S., Arora, R. C., and Meltzer, H. Y. (1991). Serotonergic measures in suicide brain: 5-HT_{1A} binding sites in frontal cortex of suicide victims. *J. Neural Transm. Gen. Sect.* **85**, 181–194.
- Matthes, H., Boschert, U., Amlaiky, N., Grailhe, R., Plassat, J., Muscatelli, F., Mattei, M., and Hen, R. (1993). Mouse 5-hydroxytryptamine_{5A} and 5-hydroxytryptamine_{5B} receptors define a new family of serotonin receptors: Cloning, functional expression and chromosomal localization. *Mol. Pharmacol.* **43**, 313–319.
- Meco, G., Bedini, L., Bonifati, V., and Sonsini, U. (1989). Risperidone in the treatment of chronic schizophrenia with tardive dyskinesia. *Curr. Ther. Res.* **46**, 876–883.
- Meltzer, C. C., Price, J. C., Mathis, C. A., Greer, P. J., Cantwell, M. N., Houck, P. R., Mulsant, B. H., Ben-Eliezer, D., Lopresti, B., DeKosky, S. T., and Reynolds, C. F., III (1999). PET imaging of serotonin type 2A receptors in late-life neuropsychiatric disorders. *Am. J. Psychiatry* **156**, 1871–1878.
- Meltzer, H. (1989). Clinical studies on the mechanism of action of clozapine: The dopamine-serotonin hypothesis of schizophrenia. *Psychopharmacology* **99**, S18–S27.
- Meltzer, H., Matsubara, S., and Lee, J. (1989). Classification of typical and atypical antipsychotic drugs on the basis of dopamine D-1, D-2, and serotonin₂ pki values. *J. Pharmacol. Exp. Ther.* **251**, 238–246.
- Meltzer, H. Y. (1991). The mechanism of action of novel antipsychotic drugs. *Schizophr. Bull.* **17**, 263–286.
- Meltzer, H. Y. (1995). The role of serotonin in schizophrenia and the place of serotonin-dopamine antagonist antipsychotics. *J. Clin. Psychopharmacol.* **15**, S2–S3.
- Meltzer, H. Y., and McGurk, S. R. (1999). The effects of clozapine, risperidone, and olanzapine on cognitive function in schizophrenia [In Process Citation]. *Schizophr. Bull.* **25**, 233–255.
- Mesotten, F., Suy, E., Pietquin, M., Burton, P., Heylen, S., and Gelders, S. (1989). Therapeutic effect and safety of increasing doses of risperidone (R 64766) in psychotic patients. *Psychopharmacology* **99**, 445–449.
- Meyer, J. H., Wilson, A. A., Ginovart, N., Goulding, V., Hussey, D., Hood, K., and Houle, S. (2001). Occupancy of serotonin transporters by paroxetine and citalopram during treatment of depression: A [(11)C]DASB PET imaging study. *Am. J. Psychiatry* **158**, 1843–1849.
- Miller, C. H., Fleischhacker, W. W., Ehrman, H., and Kane, J. M. (1990). Treatment of neuroleptic induced akathisia with the 5-HT₂ antagonist ritanserin. *Psychopharmacol. Bull.* **26**, 373–376.
- Mita, T., Hanada, S., Nishino, N., Kuno, T., Nakai, H., Yamadori, T., Mizoi, Y., and Tanaka, C. (1986). Decreased serotonin S₂ and increased dopamine D₂ receptors in chronic schizophrenics. *Biol. Psychiatry* **21**, 1407–1414.
- Morand, C., Young, S., and Ervin, F. (1983). Clinical response of aggressive schizophrenics to oral tryptophan. *Biol. Psychiatry* **18**, 575–578.
- Murphy, H., Wittkower, E., Fried, J., and Ellenberger, H. (1963). Cross-cultural survey of schizophrenic symptomatology. *Int. J. Soc. Psychiatry* **9**, 237–249.
- Nash, J. F. (1990). Ketanserin pretreatment attenuates MDMA-induced dopamine release in the striatum as measured by *in vivo* microdialysis. *Life Sci.* **47**, 2401–2408.
- Naylor, L., Dean, B., Opeskin, K., Pavey, G., Hill, C., Keks, N., and Copolov, D. (1996). Changes in the serotonin transporter in the hippocampus of subjects with schizophrenia identified using [3H]paroxetine. *J. Neural. Transm. Gen. Sect.* **103**, 749–757.
- Nedergaard, S., Bolam, J. P., and Greenfield, S. A. (1988). Facilitation of a dendritic calcium conductance by 5-hydroxytryptamine in the substantia nigra. *Nature* **333**, 174–176.
- Newcomer, J. W., Faustman, W. O., Zipursky, R. B., and Csernansky, J. G. (1992). Zacopride in schizophrenia: A single-blind serotonin type 3 antagonist trial. *Arch. Gen. Psychiatry* **49**, 751.
- Newman-Tancredi, A., Gavaudan, S., Conte, C., Chaput, C., Touzard, M., Verrielle, L., Audinot, V., and Millan, M. J. (1998). Agonist and antagonist actions of antipsychotic agents at 5-HT_{1A} receptors: A [35S]GTPgammaS binding study. *Eur. J. Pharmacol.* **355**, 245–256.

- Ngan, E. T., Yatham, L. N., Ruth, T. J., and Liddle, P. F. (2000). Decreased serotonin 2A receptor densities in neuroleptic-naïve patients with schizophrenia: A PET study using [(18)F]setoperone. *Am. J. Psychiatry* **157**, 1016–1018.
- Nyberg, S., Farde, L., Eriksson, L., Halldin, C., and Eriksson, B. (1993). 5-HT₂ and D₂ dopamine receptor occupancy in the living human brain. A PET study with risperidone. *Psychopharmacology* **110**, 265–672.
- Ohuoha, D. C., Hyde, T. M., and Kleinman, J. E. (1993). The role of serotonin in schizophrenia: An overview of the nomenclature, distribution and alterations of serotonin receptors in the central nervous system. *Psychopharmacology* **112**, S5–S15[Review].
- Okubo, Y., Suhara, T., Suzuki, K., Kobayashi, K., Inoue, O., Terasaki, O., Someya, Y., Sassa, T., Sudo, Y., Matsushima, E., Iyo, M., Tateno, Y., *et al.* (2000). Serotonin 5-HT₂ receptors in schizophrenic patients studied by positron emission tomography. *Life Sci.* **66**, 2455–2464.
- Owen, F., Cross, A., Crow, T., Deakin, J., Ferrier, I., Lofthouse, R., and Poulter, M. (1983). Brain 5-HT₂ receptors and suicide. *Lancet* **2**, 1256.
- Owen, F., Chambers, D. R., Cooper, S. J., Crow, T. J., Johnson, J. A., Lofthouse, R., and Poulter, M. (1986). Serotonergic mechanisms in brain of suicide victims. *Brain Res.* **362**, 185–188.
- Peroutka, S. J., and Snyder, S. H. (1979). Multiple serotonin receptors: Differential binding of [³H] 5-hydroxytryptamine, [³H]lysergic acid diethylamide and [³H]spiperidol. *Mol. Pharmacol.* **16**, 687–699.
- Peroutka, S. J., and Snyder, S. S. (1980). Relationship of neuroleptics drugs effects at brain dopamine, serotonin, *̢*-adrenergic and histamine receptors to clinical potency. *Am. J. Psychiatry* **137**, 1518–1522.
- Pollin, W., Cardon, P. V., and Kety, S. S. (1961). Effects of amino acid feedings in schizophrenic patients treated with iproniazid. *Science* **133**, 104–105.
- Pranzatelli, M. R., Durkin, M. M., and Barkai, A. I. (1994). Quantitative autoradiography of 5-hydroxytryptamine_{1A} binding sites in rats with chronic neonatal 5,7-dihydroxytryptamine lesions. *Dev. Brain Res.* **80**, 1–6.
- Rasmussen, K., and Aghajanian, G. (1986). Effects of hallucinogens on spontaneous and sensory-evoked locus coeruleus unit activity in the rat: Reversal by selective 5-HT₂ antagonists. *Brain Res.* **385**, 395–400.
- Rasmussen, K., and Aghajanian, G. K. (1988). Potency of antipsychotics. *Neuropsychopharmacology* **1**, 101–107.
- Reynolds, G. P., Rossor, M. N., and Ivesen, L. L. (1983). Preliminary studies of human cortical 5-HT₂ receptors and their involvement in schizophrenia and neuroleptic drug action. *J. Neural. Transm. Suppl.* **18**, 273–277.
- Rinkel, M., Hyde, R., Solomon, H., and Hoagland, H. (1955). Experimental psychiatry II: Clinical and physio-chemical observations in experimental psychosis. *Am. J. Psychiatry* **111**, 881–895.
- Rollema, H., Lu, Y., Schmidt, A. W., and Zorn, S. H. (1997). Clozapine increases dopamine release in prefrontal cortex by 5-HT_{1A} receptor activation. *Eur. J. Pharmacol.* **338**, R3–R5.
- Roth, B. L., Craig, S. C., Choudhary, M. S., Uluer, A., Monsma, F., Jr., Shen, Y., Meltzer, H. Y., and Sibley, D. R. (1994). Binding of typical and atypical antipsychotic agents to 5-hydroxytryptamine-6 and 5-hydroxytryptamine-7 receptors. *J. Pharmacol. Exp. Ther.* **268**, 1403–1410.
- Sanyal, S., and VanTol, H. H. M. (1997). Review the role of dopamine D₄ receptors in schizophrenia and antipsychotic action. *J. Psychiatr. Res.* **31**, 219–232.
- Schmidt, C. J., Kehne, J. H., Carr, A. A., Fadayel, G. M., Humphreys, T. M., Kettler, H. J., McCloskey, T. C., Padich, R. A., Taylor, V. L., and Sorensen, S. M. (1993). Contribution of serotonin neurotoxins to understanding psychiatric disorders: The role of 5-HT₂ receptors in schizophrenia and antipsychotic activity. *Int. Clin. Psychopharmacol.* **8**, 25–32.

- Schmidt, C. J., Sullivan, C. K., and Fadayel, G. M. (1994). Blockade of striatal 5-hydroxytryptamine (2) receptors reduces the increase in extracellular concentrations of dopamine produced by the amphetamine analogue 3,4-methylenedioxymethamphetamine. *J. Neurochem.* **62**, 1382–1389.
- Seeger, T. F., Seymour, P. A., Schmidt, A. W., Zorn, S. H., Schulz, D. W., Lebel, L. A., McLean, S., Guanowsky, V., Howard, H. R., Lowe, J. A., III, *et al.* (1995). Ziprasidone (CP-88,059): A new antipsychotic with combined dopamine and serotonin receptor antagonist activity. *J. Pharmacol. Exp. Ther.* **275**, 101–113.
- Seeman, P. (1990). Atypical neuroleptics: Role of multiple receptors, endogenous dopamine, and receptor linkage. *Acta Psychiatr. Scand.* **82**, 14–20.
- Seeman, P. (1992). Dopamine receptor sequences. Therapeutic levels of neuroleptics occupy D2 receptors, clozapine occupies D4. *Neuropsychopharmacology* **7**, 261–284.
- Seeman, P., and Lee, T. (1975). Antipsychotic drugs: Direct correlation between clinical potency and presynaptic action on dopamine neurons. *Science* **188**, 1217–1219.
- Shapiro, L. A., Offord, S. J., and Ordway, G. A. (1995). The effect of chronic treatment with a novel aryl-piperazine antipsychotic on monoamine receptors in rat brain. *Brain Res.* **677**, 250–256.
- Shapiro, D. A., Renock, S., Arrington, E., Chiodo, L. A., Liu, L. X., Sibley, D. R., Roth, B. L., and Mailman, R. (2003). Aripiprazole, a novel antipsychotic drug with a unique and robust pharmacology. *Neuropsychopharmacology* **28**, 1400–1411.
- Sheldon, P., and Aghajanian, G. K. (1991). Excitatory responses to serotonin (5-HT) in neurons of the rat piriform cortex: Evidence for mediation by 5-HT_{1C} receptors in pyramidal cells and 5-HT₂ receptors in interneurons. *Synapse* **9**, 208–218.
- Shen, Y., Monsma, F., Jr., Metcalf, M. A., Jose, P. A., Hamblin, M. W., and Sibley, D. R. (1993). Molecular cloning and expression of a 5-hydroxytryptamine₇ serotonin receptor subtype. *J. Biol. Chem.* **268**, 18200–18204.
- Shore, D., Korpi, E. R., Bigelow, L. B., Zec, R. F., and Wyatt, R. J. (1985). Fenfluramine and chronic schizophrenia. *Biol. Psychiatry* **20**, 329–352.
- Simpson, M. D., Lubman, D. I., Slater, P., and Deakin, J. F. (1996). Autoradiography with [3H]8-OH-DPAT reveals increases in 5-HT_{1A} receptors in ventral prefrontal cortex in schizophrenia. *Biol. Psychiatry* **39**, 919–928.
- Slater, P., Doyle, C. A., and Deakin, J. F. (1998). Abnormal persistence of cerebellar serotonin-1A receptors in schizophrenia suggests failure to regress in neonates. *J. Neural. Transm.* **105**, 305–315.
- Snyder, S. (1980). Phencyclidine. *Nature* **285**, 355–356.
- Soper, H. V., Elliott, R. O., Rejzner, A. A., and Marshall, B. D. (1990). Effects of fenfluramine on neuropsychological and communicative functioning in treatment-refractory schizophrenia patients. *J. Clin. Psychopharmacol.* **10**, 168–175.
- Sorensen, S. M., Kehne, J. H., Fadayel, G. M., Humphreys, T. M., Ketteler, H. J., Sullivan, C. K., Taylor, V. L., and Schmidt, C. J. (1993). Characterization of the 5-HT₂ receptor antagonist MDL 100907 as a putative atypical antipsychotic: Behavioral, electrophysiological and neurochemical studies. *J. Pharmacol. Exp. Ther.* **266**, 684–691.
- Stahl, S. M., Uhr, S. B., and Berger, P. A. (1985). Pilot study on the effects of fenfluramine on negative symptoms in twelve schizophrenic patients. *Biol. Psychiatry* **20**, 1098–1102.
- Stanley, M., and Mann, J. J. (1983). Increased serotonin-2 binding sites in frontal cortex of suicide victims. *Lancet* **2**, 214–216.
- Stockmeier, C. A., Shapiro, L. A., Haycock, J. W., Thompson, P. A., and Lowy, M. T. (1996). Quantitative subregional distribution of serotonin-1A receptors and serotonin transporters in the human dorsal raphe. *Brain Res.* **727**, 1–12.
- Sumiyoshi, T., Stockmeier, C. A., Overholser, J. C., Dilley, G. E., and Meltzer, H. Y. (1996). Serotonin-1A receptors are increased in postmortem prefrontal cortex in schizophrenia. *Brain Res.* **708**, 209–214.

- Svensson, T. H. (1993). Mode of action of atypical neuroleptic drug: Role of 5HT₂ receptor antagonism. *Schizophr. Res.* **11**, 115P.
- Svensson, T. H., and Tung, C. S. (1989). Local cooling of pre-frontal cortex induces pace-maker-like firing of dopamine neurons in rat ventral tegumental area *in vivo*. *Acta Physiol. Scand.* **136**, 135–136.
- Svensson, T. H., Tung, C. S., and Grenhoff, J. (1989). The 5-HT₂ antagonist ritanserin blocks the effect of prefrontal cortex inactivation on rat A10 dopamine neurons *in vivo*. *Acta Physiol. Scand.* **136**, 497–498.
- Svensson, T. H., Mathe, J. M., Andersson, J. L., Nomikos, G. G., Hildebrand, B. E., and Marcus, M. (1995). Mode of action of atypical neuroleptics in relation to the phencyclidine model of schizophrenia: Role of 5-HT₂ receptor and alpha 1-adrenoceptor antagonism. *J. Clin. Psychopharmacol.* **15**, 11S–18S.
- Szabo, Z., McCann, U. D., Wilson, A. A., Scheffel, U., Owonikoko, T., Mathews, W. B., Ravert, H. T., Hilton, J., Dannals, R. F., and Ricaurte, G. A. (2002). Comparison of (+)-(11)C-McN5652 and (11)C-DASB as serotonin transporter radioligands under various experimental conditions. *J. Nucl. Med.* **43**, 678–692.
- Tauscher, J., Kapur, S., Verhoeff, N. P., Hussey, D. F., Daskalakis, Z. J., Tauscher-Wisniewski, S., Wilson, A. A., Houle, S., Kasper, S., and Zipursky, R. B. (2002). Brain serotonin 5-HT_{1A} receptor binding in schizophrenia measured by positron emission tomography and [11C]WAY-100635. *Arch. Gen. Psychiatry* **59**, 514–520.
- Titeler, M., Lyon, R. A., and Glennon, R. A. (1988). Radioligand binding evidence implicates the brain 5HT₂ receptor as a site of action for LSD and phenylisopropylamine hallucinogens. *Psychopharmacology* **94**, 231–316.
- Tollefson, G. D., Beasley, C., Jr., Tran, P. V., Street, J. S., Krueger, J. A., Tamura, R. N., Graffeo, K. A., and Thieme, M. E. (1997). Olanzapine versus haloperidol in the treatment of schizophrenia and schizoaffective and schizophreniform disorders: Results of an international collaborative trial. *Am. J. Psychiatry* **154**, 457–465.
- Trichard, C., Paillere-Martinot, M. L., Attar-Levy, D., Blin, J., Feline, A., and Martinot, J. L. (1998). No serotonin 5-HT_{2A} receptor density abnormality in the cortex of schizophrenic patients studied with PET. *Schizophr. Res.* **31**, 13–17.
- Truffinet, P., Tammaing, C. A., Fabre, L. F., Meltzer, H. Y., Riviere, M. E., and Papillon-Downey, C. (1999). Placebo-controlled study of the D₄/5-HT_{2A} antagonist fananserin in the treatment of schizophrenia. *Am. J. Psychiatry* **156**, 419–425.
- Ugedo, L., Genhoff, J., and Svensson, T. H. (1989). Ritanserin, a 5-HT₂ receptor antagonist, activates midbrain dopamine neurons by blocking serotonergic inhibition. *Psychopharmacology* **98**, 45–50.
- van Praag, H. M., Lemus, C., and Kahn, R. (1987). Hormonal probes of central serotonergic activity: Do they really exist? *Biol. Psychiatry* **22**, 86–98.
- Van Tol, H. H. M., Bunzow, J. R., Guan, H.-C., Sunahara, R. K., Seeman, P., Niznik, H. B., and Civelli, O. (1991). Cloning of the gene for a human dopamine D₄ receptor with high affinity for the antipsychotic clozapine. *Nature* **350**, 610–614.
- Vollenweider, F. X. (1998). Advances and pathophysiological models of hallucinogenic drug actions in humans: A preamble to schizophrenia research. *Pharmacopsychiatry* **31**(Suppl. 2), 92–103.
- Vollenweider, F. X., Vollenweider-Scherpenhuyzen, M. F., Babler, A., Vogel, H., and Hell, D. (1998). Psilocybin induces schizophrenia-like psychosis in humans via a serotonin-2 agonist action. *Neuroreport* **9**, 3897–3902.
- Vollenweider, F. X., Vontobel, P., Hell, D., and Leenders, K. (1999). 5-HT modulation of dopamine release in basal ganglia in psilocybin-induced psychosis in man—a PET study with [11C] raclopride. *Neuropsychopharmacology* **20**, 424–433.

- Wander, T. J., Nelson, A., Okazaki, H., and Richelson, E. (1987). Antagonism by neuroleptic of serotonin 5-HT_{1A} and 5-HT₂ receptors of normal human brain *in vitro*. *Eur. J. Pharmacol.* **143**, 279–282.
- Wang, R. Y. (1981). Dopaminergic neurons in the rat ventral tegmental area. III. Effects of D- and L-amphetamine. *Brain Res. Rev.* **3**, 153–165.
- Weinberger, D. R., and Gallhofer, B. (1997). Cognitive function in schizophrenia. *Int. Clin. Psychopharmacol.* **12**(Suppl. 4), S29–S36.
- Whitaker, P. M., Crow, T. J., and Ferrier, I. N. (1981). Tritiated LSD binding in frontal cortex in schizophrenia. *Arch. Gen. Psychiatry* **38**, 278–280.
- Wiesel, F.-A., Nordstrom, A.-L., Farde, L., and Eriksson, B. (1994). An open clinical and biochemical study of ritanserin in acute patients with schizophrenia. *Psychopharmacology* **114**, 31–38.
- Williams, J., and Davies, J. A. (1983). The involvement of 5-hydroxytryptamine in the release of dendritic dopamine from slices of substantia nigra. *J. Pharm. Pharmacol.* **35**, 734–737.
- Wooley, D. W., and Shaw, E. (1954). A biological and pharmacological suggestion about certain mental disorder. *Proc. Natl. Acad. Sci. USA* **40**, 228–231.
- Wurtman, R. J., Heft, F., and Melamed, E. (1981). Precursor control of neurotransmitter synthesis. *Pharmacol. Rev.* **32**, 315–335.
- Wyatt, R. J., Vaughan, T., Galanter, M., Kaplan, J., and Green, J. (1972). Behavioral changes of chronic schizophrenic patients given L-5-hydroxytryptophan. *Science* **177**, 1124–1126.
- Yasuno, F., Suhara, T., Ichimiya, T., Takano, A., Ando, T., and Okubo, Y. (2004). Decreased 5-HT_{1A} receptor binding in amygdala of schizophrenia. *Biol. Psychiatry* **55**, 439–444.

SEROTONIN AND DOPAMINE INTERACTIONS IN RODENTS AND PRIMATES: IMPLICATIONS FOR PSYCHOSIS AND ANTIPSYCHOTIC DRUG DEVELOPMENT

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Since the late 1950s, appreciation of dopamine receptor blockade has played a primary role in understanding the mechanism underlying the therapeutic effects of antipsychotic drugs in schizophrenic patients in treating the positive symptoms of schizophrenia (e.g., delusions and hallucinations). Development of the second generation of antipsychotic drugs, otherwise known as atypical antipsychotic drugs, has resulted in treatments with improved subjective tolerability but relatively modest improvements in the negative symptoms of schizophrenia such as avolition, flat affect, and anhedonia. The major current challenge is to develop medications which can further improve negative symptoms treatment and also tackle the intractable clinical problems of cognitive impairment associated with schizophrenia. Further advances along these lines with respect to the dopaminergic and serotonergic neurotransmitter systems will be aided by an appreciation of the interaction between dopamine and serotonin receptor subtypes in a range of key brain structures, such as the prefrontal cortex, thalamus, striatum, amygdala, hippocampus, and the brain stem nuclei, from which the cell bodies of monoaminergic-containing neurons originate. Increasing emphasis on the use of animal models which are homologous to critical aspects of the pathophysiology

in the brains of schizophrenic patients will also be required, especially as negative symptoms and cognitive impairment become an important focus for generating novel therapeutics.

I. Introduction

Serotonin and dopamine represent two of the three monoaminergic neurotransmitter systems that play prominent roles in the action of most psychotropic drugs used for the treatment of major neuropsychiatric syndromes. Dopamine, especially, has played a critical role in therapeutics for schizophrenia since the initial hypothesis by Carlsson that blockade of dopamine receptors plays a role in their therapeutic effects for psychotic patients (Carlsson and Lindquist, 1963). This initial hypothesis has evolved over the last several decades to emphasize a relative hypofunctional state in the prefrontal cortex and a hyperfunctional state in the striatum (Abi-Dargham, 2004; Laruelle *et al.*, 2005; Winterer and Weinberger, 2004). The emergence of human PET studies of receptor occupancy demonstrated that occupancy of ~65% of dopamine D2 receptors is associated with the therapeutic effects of antipsychotic drugs (Farde *et al.*, 1992; Kapur *et al.*, 1999) except for clozapine and aripiprazole. Antipsychotic drug development in the last 15 years has emphasized serotonin (5-hydroxytryptamine, 5-HT) and 5-HT_{2A} receptor blockade as a mechanism to either minimize extrapyramidal symptoms (EPS) or increase efficacy for psychotic symptoms, negative or depressive symptoms, and cognitive dysfunction (Meltzer, 1999; Meltzer *et al.*, 1989). This has resulted in a number of “atypical” antipsychotic drugs or second generation antipsychotics (SGA). Here we discuss the underlying neurobiology for the dopamine and serotonergic neurotransmitter systems, their interactions, and gaps in our knowledge toward developing better therapeutic agents for schizophrenia. A large literature has developed describing interactions of dopamine and 5-HT regarding psychomotor stimulant drug effects, so understanding these relationships will be emphasized. A number of cortical and subcortical structures have been implicated in schizophrenia. Since structural changes and/or dysfunction of the prefrontal cortex (Selemon and Goldman-Rakic, 1999; Selemon *et al.*, 1995), thalamus (Clinton and Meador-Woodruff, 2004), dorsal and ventral striatum (nucleus accumbens, n. accumbens), and the hippocampal formation (Harrison, 2004) play prominent roles from postmortem studies of schizophrenic patients and neuroimaging studies from first-break schizophrenic patients, we will focus attention on how dopamine and 5-HT interact to modulate the function of these macrocircuits, with a special emphasis on the prefrontal cortex. Other regions are implicated in the neurodevelopmental changes, and

pathophysiology of schizophrenia such as the amygdala or the cerebellum will not be discussed in this chapter.

II. Dopamine and 5-HT Receptors

Dopamine has a neuromodulatory influence on neurotransmission in the brain by acting on five different dopamine G-protein-coupled receptor subtypes (Neve *et al.*, 2004) that are defined by both a dopamine D1-like (dopamine D1 and D5 receptors) and dopamine D2-like (dopamine D2, D3, and D4 receptors). The D1-like receptors typically couple to $G_{\alpha s}$ and $G_{\alpha olf}$ which leads to sequential activation of adenylyl cyclase, cyclic AMP-dependent protein kinase, and the protein phosphatase-1 inhibitor DARPP-32. This leads to pleiotropic effects on receptors, enzymes, ion channels, and transcription factors. Dopamine D1 receptors may also couple to phospholipase C. The activation of the phosphoinositide pathway and cAMP-dependent mobilization of intracellular Ca^{2+} is responsible for other signaling properties. For example, activation of either dopamine D1 or D5 receptors, if coexpressed with calcyon, can stimulate calcium release from intracellular stores after the cell has been primed by activation of $G_{\alpha q}$ -coupled receptors (Lezcano *et al.*, 2000). This type of interaction appears to be region specific as it occurs in the neocortex and hippocampus but not the striatum (Lezcano and Bergson, 2002). The dopamine D2-like receptors (dopamine D2, D3, and D4) couple to $G_{\alpha i}$ and $G_{\alpha o}$ heterotrimeric G-proteins to decrease activity of adenylyl cyclase but also regulate other effectors such as ion channels, phospholipases, protein kinases, and receptor tyrosine kinases via $G_{\beta \gamma}$ subunit interactions. These differential effects on postreceptor transduction pathways for the D1-like and D2-like receptor families emphasize that the effect of either increasing or decreasing dopaminergic transmission in a given region of the brain is dependent on the receptors activated by tonic (volume transmission) or phasic dopamine input (synaptic transmission) and the type of neuron (glutamatergic projection cell or GABAergic interneuron or projection cell) being affected (Onn *et al.*, 2006). Further complexity in dopamine receptor signaling due to protein-protein interactions (e.g., receptor oligomerization or receptor interactions with scaffolding and signal-switching proteins) are being uncovered (Bergson *et al.*, 2003; Neve *et al.*, 2004).

Serotonin has a neuromodulatory influence on neurotransmission in the brain and targets in the periphery by acting on at least 15 different 5-HT receptor subtypes (Aghajanian and Sanders-Bush, 2002; Barnes and Sharp, 1999) that have been classified according to 7 different families according to coupling to ion channels (5-HT₃ receptors) or heterotrimeric G-proteins (5-HT_{1A/1B/1D/1E/1F}, 5-HT_{2A/2B/2C}, 5-HT₄, 5-HT_{5A/5B}, 5-HT₆, and 5-HT₇ receptors). The 5-HT₁

family of receptors is negatively coupled to adenylyl cyclase through $G_{\alpha i}$ - and $G_{\alpha o}$ -containing G-proteins. The 5-HT₅ family may also share this coupling. In contrast, the 5-HT₄, 5-HT₆, and 5-HT₇ receptors are positively coupled to adenylyl cyclase through $G_{\alpha s}$. The 5-HT₂ family of receptors are coupled to phosphoinositide turnover and the arachidonate pathway via $G_{\alpha q}/G_{\alpha 11}$ -containing G-proteins that are linked to phospholipases C and A2, respectively. The 5-HT₃ receptor is directly coupled to ion channels and is the only ionotropic monoaminergic receptor.

When considering dopamine and serotonin interactions at a cellular level, activation of dopamine D1-like or 5-HT_{2A/2B/2C}, 5-HT₄, 5-HT₆, and 5-HT₇ receptors could potentiate the action of the other transmitter where these receptors are colocalized in the same cells with similar transduction pathways. Such a convergence of transduction pathways leading to an integrated modulation of DARPP-32 has been demonstrated in the rodent forebrain (prefrontal cortex, n. accumbens, neostriatum) for activation of dopamine D1, 5-HT₂, 5-HT₄, and 5-HT₆ receptors (Nishi *et al.*, 2000; Svenningsson *et al.*, 2002). Similarly, the dopamine D2-like and the 5-HT₁ receptor family and 5-HT₅ receptors could also potentiate the action of either dopamine or serotonin when these receptors are colocalized in the same cells with similar transduction pathways. Conversely, opposing effects of these neurotransmitters might be present where these receptors are localized to a glutamatergic projection cell or GABAergic interneuron, respectively. The later sections discussing the monoamine nuclei, the thalamus, the prefrontal cortex, the striatum, and the hippocampus will discuss the known relationships of these neurotransmitters and will also point out critical gaps to understanding the monoaminergic transmitter interactions.

III. Psychomotor Stimulants: A Dopamine-Serotonin Interaction "Case Study"

Prior to the elucidation of multiple dopamine and 5-HT receptors, a substantial body of work had arisen to suggest that decreasing serotonergic neurotransmission increased the behavioral effects of amphetamine, the direct acting dopamine agonist apomorphine, and dopamine (Breese *et al.*, 1974; Campbell and Fibiger, 1971; Carter and Pycock, 1978, 1979; Fuxe *et al.*, 1975; Green and Harvey, 1974; Hollister *et al.*, 1974; Lucki and Harvey, 1979; Neill *et al.*, 1972; Segal, 1976). Conversely, increasing serotonergic neurotransmission decreased the behavioral effects of amphetamine, apomorphine, and dopamine (Breese *et al.*, 1974; Carter and Pycock, 1978; Warbritton *et al.*, 1978).

Paradoxically, *in vivo* microdialysis studies have found that acute administration of the selective serotonin reuptake inhibitor (SSRI) fluoxetine with the catecholamine-enhancing antidepressant bupropion appears to result in a

synergistic increase in extracellular dopamine in the prefrontal cortex and n. accumbens compared to either agent alone (Li *et al.*, 2002). Similar work suggesting at least an additive effect between dopamine and 5-HT were revealed by the locomotor stimulation observed for a combination of the SSRI fluvoxamine and the nonselective reuptake inhibitor mazindol despite both drugs alone having either had no effect (mazindol) or decreasing locomotor activity (fluvoxamine). The attenuation of fluvoxamine/mazindol-induced hyperactivity by the 5-HT_{2A} receptor antagonist M100907, and potentiation of fluvoxamine/mazindol hyperactivity by the 5-HT_{2B/2C} receptor antagonist is consistent with reciprocal interactions of 5-HT_{2A} and 5-HT_{2C} receptors (McMahon and Cunningham, 2001b) discussed below. Clearly, different 5-HT receptors may mediate differential effects on dopamine function. These conflicting sets of observations exploring interactions of 5-HT and dopamine at mediating the effects of psychomotor stimulants or monoamine reuptake inhibitors may need to take into account activation of different 5-HT receptor subtypes with respect to tonic 5-HT release (volume transmission) versus phasic 5-HT release (direct synaptic transmission).

While the pharmacological effects of cocaine are frequently attributed to effects on dopamine, the ability of cocaine to inhibit 5-HT reuptake (Andersen, 1989; Gatley *et al.*, 1996) and interactions between 5-HT and the dopaminergic system also appear to be relevant for understanding the psychopharmacology of cocaine. Reciprocal interactions between dopamine and 5-HT at a global level would appear to play a role in the ability of the monoamine reuptake inhibitor cocaine to decrease locomotor activity in dopamine (DAT) knockout mice (Gainetdinov *et al.*, 1999). In DAT knockout mice, a tonic elevation of extracellular dopamine is thought to mediate the increase in locomotor activity in comparison to wild-type mice. However, in naïve rats, blockade of 5-HT_{2A} receptors attenuates the psychomotor activation induced by either amphetamine or cocaine (McMahon and Cunningham, 2001a; O'Neill *et al.*, 1999). Similar to effects reported above with SSRIs and the uptake inhibitor mazindol, there also appear to be opposing effects of 5-HT_{2A} versus 5-HT_{2C} receptor activation or blockade on cocaine hyperactivity or self-administration (Filip *et al.*, 2004; Fletcher *et al.*, 2002b; McMahon *et al.*, 2001).

Positive modulation of cocaine-induced hyperactivity or self-administration also appears with activation of 5-HT_{1A/7} receptors by 8-OH-DPAT (De La Garza and Cunningham, 2000). Similarly, activation of 5-HT_{1B} receptors in the ventral tegmental area (VTA) also increases cocaine-induced extracellular dopamine in the ipsilateral n. accumbens (O'Dell and Parsons, 2004). Overexpression of 5-HT_{1B} receptors in the VTA–n. accumbens pathway increases cocaine-induced locomotion and results in a leftward shift for the cocaine-induced conditioned place preference (Neumaier *et al.*, 2002). These effects are consistent with the absence of cocaine-induced place preference in 5-HT_{1B} receptor knockout mice (Belzung *et al.*, 2000). In contrast to cocaine-induced

effects, 5-HT_{1B} receptor activation in the n. accumbens decreases amphetamine self-administration. This discrepancy between the effects of 5-HT_{1B} receptor activation may be dependent on the presence of 5-HT_{1B} receptors in different cellular compartments (Fletcher *et al.*, 2002a). Pharmacological *blockade* of 5-HT₄ receptors in the n. accumbens shell attenuates cocaine-induced hyperactivity (McMahon and Cunningham, 1999). While 5-HT₆ receptor antagonism does not alter cocaine effects, amphetamine-induced behavioral effects were potentiated (Frantz *et al.*, 2002). Thus, activation of a number of serotonin receptors (5-HT_{1A/7}, 5-HT_{1B}, 5-HT_{2A}, 5-HT₄) does appear to mediate, in part, the behavioral and neurochemical effects of psychomotor stimulants. However, there also appears to be particular 5-HT receptors (e.g., 5-HT_{2C} receptors) which oppose the stimulating effects of amphetamines or cocaine.

IV. Monoaminergic Nuclei Interactions

Dopamine-containing cell bodies are localized primarily in the substantia nigra pars compacta (SNpc) and the VTA of the midbrain (Moore and Bloom, 1978). The SNpc projects to the dorsal striatum (caudate and putamen) and makes up the nigrostriatal dopaminergic system. The VTA has relatively precise topographical relationships with the prefrontal cortex (mesocortical dopaminergic system), the ventral striatum (n. accumbens), and other areas (thalamus, hippocampus, lateral septum, amygdala). Together, the projections to the n. accumbens and the other limbic structures outside of the prefrontal cortex make up the mesolimbic dopaminergic system. These nuclei which contain dopaminergic cells are not homogenous with respect to cell type. At least 15–20% of the neurons within the VTA are known to be GABAergic interneurons, and at least a portion of these are projection GABAergic interneurons to the prefrontal cortex (Carr and Sesack, 2000) and the n. accumbens (Van Bockstaele and Pickel, 1995). While these general relationships hold, it has also been proposed that the dopaminergic afferents to the primate thalamus may represent an additional distinct system involving cell bodies of origin in the hypothalamus, periaqueductal gray (PAG) matter, ventral mesencephalon, and the lateral parabrachial nucleus (Sanchez-Gonzalez *et al.*, 2005).

Serotonin-containing cell bodies which project to the limbic forebrain are found primarily in the dorsal raphe nucleus (DR) and the median raphe nucleus (MR) in the pons (Hensler, 2006). The DR projects to the prefrontal cortex, the lateral septum, ventral hippocampus, and n. accumbens as well as providing a relatively strong innervation to the substantia nigra pars reticulata (a nucleus containing the dendrites of SNpc cells) and VTA. The MR projections include the dorsal hippocampus, medial septum, and hypothalamus. Some areas such as

the intralaminar and midline thalamic nuclei receive a projection from both the DR and the MR (Azmitia, 1978). The midbrain raphe nuclei, like the midbrain SNpc and VTA, contain GABAergic interneurons. There is some evidence that at least some of these GABAergic interneurons may also be projection GABAergic interneurons (O'Hearn and Molliver, 1984; Van Bockstaele *et al.*, 1993). An interesting feature about the raphe nuclei is that most forebrain projection areas send back reciprocal projections via the habenula nuclei. However, the prefrontal cortex appears to be an area with a privileged direct projection to the dorsal raphe (Aghajanian and Wang, 1977; Hajos *et al.*, 1998; Martin-Ruiz *et al.*, 2001a; Peyron *et al.*, 1998; Sesack *et al.*, 1989).

The dorsal raphe nucleus also sends a dense serotonergic projection to the pars reticulata of the substantia nigra (SNr; Fallon and Loughlin, 1995). Dorsal raphe stimulation mainly induces inhibitory effects in SNpc cells, though non-5-HT afferents may be involved (Gervais and Rouillard, 2000). Both the dorsal raphe and the median raphe send axons which arborize in the VTA (Azmitia, 1978; Vertes, 1991; Vertes *et al.*, 1999). In turn, the VTA and cells from the A11 hypothalamic cell group project back to the dorsal raphe (Kalen *et al.*, 1988; Peyron *et al.*, 1995). While activation of both dopamine D1-like and D2-like receptors increase the firing rate of serotonergic-containing DR neurons and increase 5-HT release in the DR and striatum, it appears that dopamine D2 receptors outside of the DR are involved in these effects (Martin-Ruiz *et al.*, 2001b). GABAergic neurons in the PAG-expressing dopamine D2 mRNA are among candidate regions mediating this effect.

Regarding dopamine-serotonin interactions, multiple 5-HT receptors have different, and apparent, opposing influences on the presynaptic side of the dopaminergic system. There is a well-documented effect of 5-HT_{2C} receptor stimulation activating GABAergic cells in the SN and VTA, and thereby inhibiting frontocortical and accumbal dopaminergic transmission. Conversely, blockade of 5-HT_{2C} receptors increases activity of dopamine-containing cells in the VTA, and thereby increases dopaminergic transmission in the mesolimbic and mesocortical dopamine pathways (De Deurwaerdere *et al.*, 2004; DiMattio *et al.*, 2001; Millan *et al.*, 2000). Furthermore, *in vivo* evidence suggests that 5-HT_{2C} receptors inhibit dopamine release in the rat striatum and n. accumbens via constitutive activity (De Deurwaerdere *et al.*, 2004).

While activation of the 5-HT_{2C} receptor appears to play an inhibitory role on dopaminergic neurotransmission, activation of another receptor from the same family, the 5-HT_{2A} receptor, appears to facilitate dopaminergic neurotransmission in the mesolimbic and mesocortical pathways. For example, 5-HT_{2A} receptor mRNA tends to colocalize with a fraction of the dopamine-containing cells in regions of the VTA that preferentially projects to terminal fields of the mesolimbic pathway (Nocjar *et al.*, 2002). An ultrastructural study has found that 5-HT_{2A} receptors are present in the dendrites and soma of VTA dopamine-containing

cells (Doherty and Pickel, 2000). There is also a significant amount of functional data (discussed above) suggesting that 5-HT_{2A} receptor activation plays a role in the psychomotor stimulant effects of cocaine and amphetamine by enhancing extracellular levels of dopamine. The increase seen in extracellular DA following local infusion into the prefrontal cortex with the highly selective 5-HT_{2A} receptor antagonist suggests a facilitatory role for this receptor in the mesocortical pathway (Pehek *et al.*, 2001). Activation of cortical 5-HT_{2A} receptors also appears to increase stress-induced dopamine release in the mPFC (Pehek *et al.*, 2006). This effect may be mediated by polysynaptic neuronal circuits involving cortical pyramidal cells with 5-HT_{2A} receptors in the apical dendritic field (Bortolozzi *et al.*, 2005). The immunohistochemical localization of 5-HT_{2A} receptors in presumed monoaminergic axons of the prefrontal cortex might be an additional or alternative substrate for these latter findings (Miner *et al.*, 2003). A previous electron microscopic study had observed the presence of 5-HT_{2A} receptor immunoreactivity in small unmyelinated axons of the VTA that were likely either dopaminergic axons or axons from nondopaminergic cells coursing through the VTA (Doherty and Pickel, 2000). These opposing effects of 5-HT_{2A} versus 5-HT_{2C} receptors on dopaminergic neurotransmission highlight the critical importance of understanding the localization of different 5-HT receptors, SERT, and DAT with respect to both microcircuits and macrocircuits involving corticostriato-thalamic pathways.

5-HT_{1A} receptor partial agonist action has been discussed for a number of years with respect to development of antidepressant drugs by understanding the relative role of somatodendritic 5-HT_{1A} autoreceptors and 5-HT_{1A} postsynaptic receptors. There are a number of compounds currently in development possessing partial agonist action at 5-HT_{1A} receptors in addition to effects at other monoaminergic neurotransmitters such as dopamine D2/D3 receptors.

V. Serotonin and Dopamine in the Thalamus

The serotonergic innervation of the thalamus from the dorsal and median raphe is mainly distributed to the midline and intralaminar thalamic nuclei, the so-called “nonspecific” thalamic nuclei of Lorente de No. While the afferent projection from the dorsomedial nucleus (MD n.) of the thalamus to layer III of the neocortex is much discussed as a defining feature of the prefrontal cortex, the much less studied midline and intralaminar thalamic nuclei have a number of neuroanatomical relationships that make them critical structures with respect to major neuropsychiatric syndromes and therapeutics (Van der Werf *et al.*, 2000, 2002). First of all, these midline and intralaminar thalamic nuclei project to layers I and Va throughout the prefrontal cortex providing discrete laminar inputs to

the distal dendritic tuft and proximal apical dendrites of the principal output cells, the layer V pyramidal cells. The importance of the thalamus in coordinating higher functions of the cortex may be suggested by the tenfold greater number of corticothalamic fibers than thalamocortical fibers; the layer V pyramidal cells project back to the intralaminar and midline thalamic nuclei. Second, the intralaminar and midline thalamic nuclei make up the only projection from the thalamus to the dorsal and ventral striatum. Third, the midline and intralaminar thalamic nuclei also project to the amygdala, subiculum and return a principal projection from the brain stem reticular activating system. While the intralaminar and midline thalamic nuclei express a rich distribution of 5-HT_{2C} and 5-HT_{1B} receptor mRNA in the adult rat, activation of 5-HT_{2A} receptors appear to play an important role in inducing glutamate release from these afferents in layer I and Va of the prefrontal cortex (Aghajanian and Marek, 1997; Marek *et al.*, 2001). Thus, these anatomical relationships define a critical feature for the intralaminar and midline thalamic nuclei consistent with their physiological importance in mediating arousal and vigilance (Kinomura *et al.*, 1996).

While the serotonergic and noradrenergic innervation of the thalamus is much more prominent than that of dopamine, there is also evidence for physiological effects of dopamine within the thalamus. There is evidence for dopamine D2/D3 receptor binding in the human midline and intralaminar thalamic nuclei which is most intense for the paraventricular thalamic (PVT) nucleus (Rieck *et al.*, 2004). In addition to dopaminergic afferents to the PVT, there also appears to be a role for dopamine D2 receptor stimulation in modulating the MD n. Dopamine D2 receptor activation appears to increase the excitability of rat MD n. cells recorded using an *in vitro* preparation (Lavin and Grace, 1998). This direct effect of dopamine in the MD n. could have even a greater effect on distinct thalamocortical pathways given the convergence of axon terminals from the MD n. and the VTA (Kuroda *et al.*, 1996). However, it should also be noted that most of the dopaminergic cells of origin which project to the rodent MD n. and the PVT appear to arise from the hypothalamus rather than the VTA. In the primate, there does appear to be a projection from the SNr to the MD n., although the neurotransmitter phenotype of these cells has not been identified. In the primate, DAT immunopositive labeling has clearly suggested that dopamine does innervate the MD n. (Melchitzky and Lewis, 2001). As mentioned earlier, the heterogeneity of dopamine innervation to different regions in the thalamus has prompted one group to suggest that the "thalamic dopaminergic system" may be a novel system with respect to the classical recognition of the nigrostriatal, mesocortical, and mesolimbic dopaminergic systems (Sanchez-Gonzalez *et al.*, 2005). With respect to the potential complementary roles played by 5-HT_{2A} receptor activation versus dopamine D2 receptor activation on thalamocortical pathways, it is interesting that dopamine receptor activation, unlike 5-HT_{2A} or α_1 adrenergic receptor activation, does not appear to play a role at inducing excitatory postsynaptic

currents (EPSCs) recorded from layer V pyramidal cells, which appear to arise in part from the midline and intralaminar thalamic nuclei (Marek and Aghajanian, 1999).

VI. Dopamine and Serotonin in the Striatum

The effects of dopamine and serotonin in the striatum cannot be addressed without a brief description of the basal ganglia microcircuitry and macrocircuitry (Wilson, 1998). The neostriatum consists of dorsal striatum (caudate/putamen) and ventral striatum (n. accumbens). Increased dopaminergic neurotransmission in circuits running through the dorsal striatum is associated with motor stereotypies such as those induced by high dose-amphetamine treatment. The caudate/putamen receives inputs from sensory cortex, motor cortex, and prefrontal cortex that converge with projections from the intralaminar thalamic nuclei, dopaminergic inputs from the SNpc, and serotonergic inputs from the (DR). The GABAergic, substance P-containing medium spiny cells of the caudate/putamen then project to two major regions: the globus pallidus [external (GPe) and internal (GPi) segments] and the pars reticulata of the SNr. The “direct pathway” leading from the GPi and the SNr projects to the ventral thalamic nuclei tier, lateral habenula nuclei, and the deep layers of the superior colliculus. The “indirect pathway” involves GABAergic enkephalin-containing neostriatal medium spiny cells of the caudate/putamen which project to the external segment of the globus pallidus, which in turn projects to the subthalamic nucleus. It should be noted that the neostriatum does not provide a direct reciprocal projection back to the neocortex and the thalamus.

Enhanced dopaminergic neurotransmission in circuits running through the ventral striatum (n. accumbens) is associated with stimulation of increased locomotor activity induced by relatively low-dose amphetamine treatment. The n. accumbens core, unlike the dorsal striatum, receives serotonergic input via the median raphe. There are important topographical relationships of different areas of the prefrontal cortex, amygdala, thalamic, and hippocampus projecting onto the accumbal patch and matrix areas (Groenewegen *et al.*, 1999). For example, the n. accumbal patches project to the SNpc while the matrix areas project to the SNr. These patch (striosome) and matrix regions also have substantial differences with respect to μ -opioid receptor localization, calcium-binding proteins, and acetylcholinesterase [patch neuropil high in μ -opioid receptors; low in calbindin; acetylcholinesterase rich (Herkenham and Pert, 1981)]. Enkephalin and substance P have a more complex distribution, though they are differentially expressed in the patch or matrix in different regions of the neostriatum. The patch versus matrix distinction has important relationships to macrocircuitry afferent

relationships. For example, the n. accumbens core matrix receives projections from layer III or superficial layer V pyramidal cells in the prefrontal cortex in addition to strong afferents from a number of midline and intralaminar thalamic nuclei. The n. accumbens core patches receive projections from deep layer V pyramidal cells in the PFC (Gerfen, 1989) and amygdala but are relatively sparse with respect to thalamic afferents. However, it is important to note that the topography of afferents to the striatum from different prefrontal cortical layers to the striosomes (patch) or matrix can switch in different regions of the caudate/putamen (Ragsdale and Graybiel, 1990). One difference between the patch (striosome) and matrix region in the human caudate and putamen is that 5-HT_{2A} receptor binding appears more intense in the patch regions (Lopez-Gimenez *et al.*, 1999; Waeber and Palacios, 1994). 5-HT_{1A} receptor binding in the primate striatum also is more intense in the striosome or patch neuropil (Frechilla *et al.*, 2001; Mengod *et al.*, 1996).

This mosaic organization of the neostriatum and accumbens was previously identified in primates. The patch or striosome is equivalent to the cell islands while the matrix is similarly named in the primate (Goldman-Rakic, 1982). In the rodent, patterns of fos immunopositive cells have been studied with respect to predicting extrapyramidal side effects consistent with typical versus atypical antipsychotic drugs. The ratio striosome/matrix cells expressing fos immunopositive cells is greater for atypical antipsychotic drugs than for typical antipsychotic drugs such as haloperidol (Bubser and Deutch, 2002). However, single serotonin receptor subtypes such as the 5-HT_{1A} or 5-HT_{2A} receptor did not appear, by themselves, to account for these effects. These striatal compartments are differentially modulated under other circumstances. For example, the matrix compartment of the rat is preferentially activated during free movement, gentle restraint, or focal tactile stimulation during gentle restraint (Brown *et al.*, 2002). Conversely, stereotypy induced by cocaine or amphetamine or psychostimulant-induced sensitization appears to be associated with preferential early gene expression in the striosome compartment of the striatum (Canales and Graybiel, 2000; Capper-Loup *et al.*, 2002).

Dopamine can differentially affect the direct and indirect pathways of the striatum. Dopamine D1 receptors are preferentially expressed in the substance P-containing striatonigral direct pathway whereas dopamine D2 receptors are preferentially expressed on the enkephalin-containing spiny neurons making up the striatopallidal indirect pathway (Gerfen *et al.*, 1990; Surmeier *et al.*, 1996). Thus, dopamine would tend to enhance neurotransmission through the direct pathway while attenuating transmission in the indirect pathway. However, there is also a subpopulation of striatal principal cells (medium spiny neurons) which coexpress both D1 and D2 receptors (Surmeier *et al.*, 1996). While the actions of dopamine in the neostriatum are varied depending on dopamine receptor subtype and cellular phenotype, an important action of dopamine D1 receptors is to

enhance the glutamatergic input arriving from the prefrontal cortex/neocortex (Nicola *et al.*, 2000).

The n. accumbens shell region appears to be part of the extended amygdala and has yet different input-output relationships compared to the n. accumbens core (Heimer, 2003). In fact, the shell region is considered to be a constituent of the extended amygdala involving the n. accumbens shell, the central n. of the amygdala, and the bed n. of the stria terminalis (BNST).

Studies involving the lesioning of dopaminergic terminals in the striatum have revealed interesting relationships between dopamine and serotonin. Namely, neonatal destruction of dopaminergic nerve terminals in the dorsal striatum results in a 5-HT hyperinnervation of the adult striatum as reviewed elsewhere (Kostrzewa *et al.*, 1998).

VII. Dopamine and Serotonin in the Hippocampal Formation

The hippocampal formation includes the dentate gyrus, the four subdivisions of Ammon's horn (CA1/2/3/4), the subiculum, the entorhinal cortex, and the parahippocampal gyrus. Both the rat and cynomolgus monkey have a rich dopaminergic innervation of the entorhinal cortex, subiculum, and CA1 and/or CA3 (Baulac *et al.*, 1986; Samson *et al.*, 1990). Dopaminergic-containing neurons of the VTA project to the ventral hippocampus (Gasbarri *et al.*, 1994; Verney *et al.*, 1985).

All five dopamine receptor subtypes are expressed in the hippocampal formation, though with a differential distribution (Meador-Woodruff *et al.*, 1994). There is also a rich serotonergic innervation of the hippocampal formation from both the dorsal and especially the median raphe (Hensler, 2006). Of the serotonin receptors, the 5-HT_{1A} receptor has been associated with playing important functional roles in the hippocampus (Andrade and Nicoll, 1987; Haddjeri *et al.*, 1998). Alterations in the oscillatory frequencies of the hippocampus have been associated with important behavioral state changes; 5-HT_{1A} receptors in the hippocampus have been implicated in modulating hippocampal oscillations (Gordon *et al.*, 2005). The 5-HT₄ receptor induces a slow excitatory response to 5-HT in the hippocampus which has been compared functionally to 5-HT_{2A} receptor activation in the neocortex (Andrade and Chaput, 1991). In this respect, the 5-HT₄ in addition to 5-HT_{1A} receptors has been implicated as a potential target for cognitive-enhancing therapies for schizophrenic patients (Roth *et al.*, 2004). The 5-HT₆ receptor, localized in the hippocampus, striatum, cerebral cortex, and modulates cholinergic neurotransmission, is another target for cognitive enhancement by selective antagonists (Roth *et al.*, 2004).

The subiculum is important as a site of routing incoming activity to the hippocampus, prior to processing in the dentate gyrus and Ammon's horn. For example, these regions receive afferents from the reuniens n. and the anterior tier nuclei of the thalamus (Berendse and Groenewegen, 1991; van Groen and Wyss, 1995; Van Groen *et al.*, 1999). The ventral hippocampus, which has important anatomical relationships to the prefrontal cortex and n. accumbens, is of special interest with respect to neurodevelopmental disturbances of cortico-limbic circuits in schizophrenia. The hippocampus provides a monosynaptic input to the prefrontal cortex via the subiculum and temporal aspect of CA1 (Jay *et al.*, 1989; Laroche *et al.*, 1990; Wyss *et al.*, 1980). There does not appear to be a specific relationship with hippocampal afferents to the medial prefrontal cortex similar to the thalamocortical afferents from the medial dorsal n. (Carr and Sesack, 1996; Kuroda *et al.*, 1996). The subiculum also provides a massive input to the n. accumbens that is necessary for accumbal neurons to enter a depolarized, "up" state (O'Donnell and Grace, 1995). Dopamine D1-like and probably D2 receptors play at least a permissive role in the locomotor activation and increase in n. accumbens extracellular dopamine induced by local infusion of NMDA into the ventral hippocampus (Zornoza *et al.*, 2005).

VIII. Dopamine and Serotonin in the Prefrontal Cortex/Neocortex

Dopamine and serotonin are two of the aminergic neurotransmitters (along with norepinephrine and histamine) that are constituents of the ascending arousal system from the brain stem which also includes the reticular activating system. As such, an assessment of the role for dopamine and serotonin for cortical function must include their role in the ascending arousal system and the modular nature of the prefrontal cortex/neocortex (Grillner *et al.*, 2005; Silberberg *et al.*, 2005). The thick, tufted layer V pyramidal cells are the principal output cell for the prefrontal cortex/neocortex with projections to the thalamus, striatum, amygdala, brain stem, and spinal cord (Deschenes *et al.*, 1994). These layer V pyramidal cells provide the "driving" input to the midline and intralaminar thalamic nuclei discussed above (Sherman and Guillery, 1998). Untufted layer V pyramidal cells that do not extend tufts of dendrites up into layer I of the prefrontal cortex/neocortex project to the contralateral hemispheres. A population of layer VI corticothalamic pyramidal cells project to the first-order thalamic nuclei as a "modulatory" input. Thus, understanding relative dopaminergic or serotonergic control of layer V versus layer VI pyramidal cells has important implications for controlling or modulating different types of thalamocortical pathways.

Different subpopulations of cortical pyramidal cells also project to the striatum (Groenewegen *et al.*, 1999). For the rat ventral prelimbic region of the prefrontal cortex, for example, deep layer V pyramidal cells project to the dorsomedial portion of the shell and “patches” (see below) of the n. accumbens core. In contrast, layer III and superficial layer VI pyramidal neurons send fibers to the “matrix” compartment of the n. accumbens core.

Most of the remaining 20% of PFC/neocortical neurons are GABAergic interneurons which form a variety of classes based on nature of principal efferent contact (interneuron vs pyramidal cell), portion of the axonal/soma/dendritic surface that is contacted by the interneuron, nature of calcium-binding protein expressed by the cell, nature of peptide neurotransmitters colocalized to the interneuron (e.g., substance P, neurokinin B, CCK), and the electrophysiological phenotype of the interneuron. The critical importance of interneurons in sculpting and modulating oscillations of the cortex is highlighted for schizophrenia given the importance of some oscillations such as those in the gamma range as a substrate for higher cortical function (Symond *et al.*, 2005). In addition to the role of interneurons in shaping oscillations, the observations that interneurons appear to be involved in the pathophysiology of schizophrenia makes understanding the role of dopamine and 5-HT for interneuron function a critical question (Levitt *et al.*, 2004).

Differences exist for the serotonergic and dopaminergic projections to the prefrontal cortex and neocortex. First, serotonergic terminals are more widely and evenly distributed in different regions of the prefrontal cortex and neocortex (Tork, 1990). This is in contrast to the dopaminergic system which is limited largely to the prefrontal cortex in the rodent. The primate, however, has an expanded regional dopamine distribution outside of the prefrontal cortex (Berger *et al.*, 1991). Second, while both serotonergic and dopaminergic projections do differ in having distinct laminar distributions, they may even differ from region to region or even in different primate species (Crino *et al.*, 1993). It is the deep projections of dopaminergic cells to layer VI of the prefrontal cortex which appear to be diminished in schizophrenic patients (Akil *et al.*, 1999).

One area of similarity between serotonergic and dopaminergic neurotransmission in the prefrontal cortex and neocortex is both the absence (volume transmission) and the presence (synaptic transmission) of synaptic specializations associated neurotransmitter-releasing monoaminergic varicosities. Estimates for the frequency of synaptic specializations opposite to serotonin varicosities ranges from 28% to 90% in the rat prefrontal cortex or neocortex (Papadopoulos *et al.*, 1987; Seguela *et al.*, 1989). In contrast, very few synaptic specializations were observed opposing serotonergic varicosities in the primate motor-sensory cortex (DeFelipe and Jones, 1988). These observations of synaptic versus volume transmission may be of particular importance with regard to activation of particular

monoaminergic neurotransmitters on different cellular compartments when neurotransmitter release, reuptake, or metabolism is altered.

Interneurons have emerged as a significant target of 5-HT-containing varicosities for direct synaptic neurotransmission in the primate (Smiley and Goldman-Rakic, 1996), where only ~8% of the postsynaptic shafts opposing serotonin axons were identified as pyramidal cell dendrites. 5-HT_{2A} receptors have been localized to large- and medium-sized parvalbumin and calbindin-containing interneurons that would likely target the perisomatic region of pyramidal cells (Jakab and Goldman-Rakic, 1998, 2000). This is striking since the evidence suggests that most, if not all, cortical pyramidal cells contain 5-HT_{2A} receptors. In contrast, 5-HT₃ receptors are present solely in classes of interneurons that target the dendritic field of pyramidal cells (Jakab and Goldman-Rakic, 2000; Morales *et al.*, 1996).

One effect of activating 5-HT_{2A} receptor-containing interneurons in the cortex is to increase spontaneous inhibitory postsynaptic currents (IPSCs) recorded from pyramidal cells through layers II–VI. In layer V pyramidal cells, activation of 5-HT_{2A} receptors results in depolarization by an apparent closure of potassium channels and may also increase a persistent sodium current (Aghajanian and Marek, 1997; Araneda and Andrade, 1991; Tanaka and North, 1993). Another effect that is largely restricted to the major output cell of the prefrontal cortex and neocortex is an enhancement of spontaneous EPSCs which appears to involve an induction of glutamate release from thalamocortical afferents (layers I and Va) onto the apical dendritic field of layer V pyramidal cells (Aghajanian and Marek, 1997; Lambe *et al.*, 2000; Marek *et al.*, 2001). In contrast to these direct excitatory effects on different cellular compartments by activation of 5-HT_{2A} receptor, it should be kept in mind that the predominant effects of 5-HT in the cortex during electrically evoked potentials is a suppression of activity that appears to washout quickly when probed using *in vitro* slice preparations. This inhibitory effect on glutamatergic transmission is due at least in part to activation of 5-HT_{1B} receptors (Aghajanian and Marek, 1999; Read *et al.*, 1994; Tanaka and North, 1993).

In contrast to the relationship of serotonergic varicosities to synaptic specializations in interneurons, the major relationship for dopaminergic varicosities to synaptic specializations appears to be on pyramidal cell dendrites (Goldman-Rakic *et al.*, 1989; Smiley and Goldman-Rakic, 1993). While some of the dopamine D1 receptors in the human and monkey prefrontal cortex appear to be in association with synaptic specializations on dendritic spines opposite of presumed glutamatergic terminals, many dopamine D1 receptors may also have an extrasynaptic localization in pyramidal cells (Smiley *et al.*, 1994). In contrast to dopamine D1 and D5 receptors, the dopamine D4 receptor has been localized to GABAergic interneurons (Mrzljak *et al.*, 1996). The electrophysiological effects of dopamine

in the prefrontal cortex have been reviewed elsewhere (Seamans and Yang, 2004).

Groundbreaking studies emphasizing the importance of prefrontal cortical dopamine in modulating spatial working memory was the finding that depletion of dopamine in the rat or primate mPFC resulted in deficits approximately as robust as prefrontal cortical lesions (Brozoski *et al.*, 1979; Simon *et al.*, 1980). A number of studies since that time especially by Arnsten, Goldman-Rakic, and colleagues demonstrated that activation of dopamine D1 receptors facilitated prefrontal cortical executive function in the primate and rodent. Similarly, activation of dopamine D1 receptors in the mPFC increases the accuracy of rats performing on a five choice serial reaction time test (5-CSRTT) which measures attention (Granon *et al.*, 2000). An inverted “U” relationship between D1 receptor stimulation and PFC function exists (Zahrt *et al.*, 1997). Furthermore, enhanced dopaminergic function also appears to play a role in the disruption of working memory in the dorsolateral prefrontal cortex induced by stress (Arnsten and Goldman-Rakic, 1998).

Evidence also supports the hypothesis that activation and blockade of 5-HT_{2A} receptors impair and enhance, respectively, working memory tasks for which the DLPFC is a critical component. The hallucinogen LSD impairs delayed response and delayed alternation tasks in the rhesus monkey in a fashion consistent with prefrontal cortical lesions (Frederick *et al.*, 1997; Jarvik and Chorover, 1960). 5-HT_{2A} receptor activation appears to improve the tuning of PFC pyramidal cells and interneurons in primates performing a well-learned delayed response task. It was suggested that activation of 5-HT_{2A} receptors would adversely impact cognition with more demanding tasks (Williams *et al.*, 2002). Accordingly, the selective 5-HT_{2A} receptor antagonist EMD 281014 enhanced the performance of both young and aged rhesus monkeys on a delayed response task (Terry *et al.*, 2005). However, the effects of LSD appear to be more prominent on time estimation and motivation, than on short-term memory and attention (Frederick *et al.*, 1997). Similarly, hallucinogens that selectively activate the 5-HT₂ family of receptors have also been found to have profound effects on social interaction, especially affiliative behavior (Schlemmer and Davis, 1986).

Serotonin appears to play a role in a number of aspects of impulsivity as known from rodent studies and a review of the primate and human literature where 5-HT levels were decreased either by global lesion of serotonergic neurons or dietary manipulation (Winstanley *et al.*, 2004a). A number of studies using systemic administration of phenethylamine hallucinogenic drugs which activate the 5-HT₂ family of receptors or 5-HT receptor antagonists with at a 20- to 80-fold selectivity for 5-HT_{2A} versus 5-HT_{2C} receptors suggest that activation of 5-HT_{2A} impairs impulsivity as reflected in an increased premature responding on the 5-CSRTT (Carli and Samanin, 1992; Koskinen *et al.*, 2000; Passetti *et al.*, 2003; Winstanley *et al.*, 2004b). A local effect in the mPFC for 5-HT_{2A} receptors

on modulating impulsivity in this task is supported both by the ability of systemic administration of the selective 5-HT_{2A} receptor antagonist M100907 to prevent attentional impairment induced by local NMDA receptor blockade or for local infusion of M100907 itself to decrease premature responding and omissions (Carli *et al.*, 2004; Winstanley *et al.*, 2003).

Opposing influences of different 5-HT receptor subtypes have also been found using the 5-CSRTT. For example, in contrast to the improvement in impulsivity with systemic or local administration of a 5-HT_{2A} receptor antagonist, systemic administration of a 5-HT_{2C} receptor antagonist SB 242084 increased impulsivity (Winstanley *et al.*, 2004b). Analogous data has been produced where a selective 5-HT_{2A} receptor antagonist attenuated the increase in premature responding induced by NMDA receptor blockade whereas the 5-HT_{2C} receptor antagonist SB 242084 was without an effect (Higgins *et al.*, 2003). However, a 5-HT_{2B/2C} receptor antagonist did not share this effect with SB 242084 (Talpos *et al.*, 2006). Within the prefrontal cortex, local activation of 5-HT_{1A} receptors also improves visuospatial attention and decreases impulsivity, though with some differences in comparison to local blockade of 5-HT_{2A} receptors (Winstanley *et al.*, 2003). This observation would be consistent with the hypothesis that activation of 5-HT_{1A} and blockade of 5-HT_{2A} receptors could have therapeutic effects for cognitive impairment of schizophrenic patients (Roth *et al.*, 2004). In contrast to the effects of 5-HT_{2A} receptor blockade, a 5-HT₆ receptor antagonist had no effect on impulsivity (Talpos *et al.*, 2006). While these results with 5-HT_{2A} receptor blockade show a consistent effect on impulsivity as measured by premature responding on the 5-CSRTT, it should be noted that 5-HT_{2A} receptor antagonists did not counter impulsivity observed for choice procedures between a small immediate reward versus a larger, delayed reward (Talpos *et al.*, 2006; Winstanley *et al.*, 2004a).

IX. Animal Models

Two principal unmet medical needs in treating schizophrenic patients is improving negative symptoms and treating the cognitive impairment associated with schizophrenia (CIAS). The probability that a novel treatment would treat both the positive symptoms and cognitive impairment/negative symptoms in the near future seems remote. The MATRICS and TURNS initiative involving academia, industry, and the FDA assumes that new treatments targeting cognitive deficits will be added to current antipsychotic drugs (Geyer and Tamminga, 2004). A critical feature in moving toward these new treatment goals is a deeper appreciation of the neural substrates involved with respect to macrocircuitry relevant for schizophrenia (thalamocortical-striatal-hippocampal-brain stem).

An important task of future research is understanding the degree in which different cognitive enhancement strategies will be compatible with current antipsychotic drugs (Floresco *et al.*, 2006).

The neonatal excitotoxic hippocampal lesion (ventral hippocampus and ventral subiculum) is an animal model with evidence of altered dopaminergic activity in addition to decreased social behavior, impaired working memory, and enhanced sensitivity to drugs of abuse (Lipska, 2004). A developmental link with this model is emphasized by the ability of excitotoxic lesions of the mPFC to block certain behaviors in this syndrome such as hyperlocomotion to novelty and amphetamine (Lipska *et al.*, 1998). Furthermore, even reversible inactivation during the neonatal period leads to some of the characteristic dopamine- and glutamate-mediated changes in pharmacological sensitivity (Lipska, 2004). An additional key feature with this model is an enhanced sensitivity to the effects of NMDA antagonists which imparts additional clinical face validity.

Another developmental model for schizophrenia involves administering the methylating agent methylazoxymethanol acetate (MAM) on embryonic day 17. When the rats are tested as adults, they have a number of neuroanatomical features consistent with the neuropathology of schizophrenia such as size reduction in the mediodorsal thalamus, hippocampus, and prefrontal cortex and decreased neuron density without neuron loss in the prefrontal cortex (Moore *et al.*, 2006). These rats also exhibit a disruption of synaptically driven bistable membrane potentials in both the prefrontal cortex and the ventral striatum (Lavin *et al.*, 2005). A remarkable cognitive change in these rats is a deficit in reversal learning. With respect to the dopaminergic system in the prefrontal cortex, these rats exhibit a hypofunctional response to topical dopamine while the deep pyramidal cells are more sensitive to VTA stimulation. At a behavioral level, adult (but not adolescent) rats exhibit greater sensitivity to the locomotor activating effects of amphetamine (Moore *et al.*, 2006).

Since increases in striatal dopamine D2 receptors have been implicated in the pathophysiology of first-break and medicated schizophrenic patients, mice have been generated with a reversible increase level of striatal D2 receptors (Kellendonk *et al.*, 2006). These mice have cognitive deficits in the sphere of attentional set-shifting and also have decreased dopamine turnover in the prefrontal cortex that are consistent with altered function in the prefrontal cortex.

Another recent animal model for schizophrenia is a mouse strain with a dramatically reduced level of the obligate NMDA NR1 receptor subunit. These mice display reduced locomotor habituation to a novel environment, increased stereotypic activity, deficits in social interaction, and reduced prepulse inhibition of acoustic startle (Duncan *et al.*, 2004; Mohn *et al.*, 1999). Another characteristic of these mice bearing some similarity with schizophrenic patients is an increased motor stereotypy and reduced fos expression in the medial prefrontal and cingulate cortex in response to amphetamine (Miyamoto *et al.*, 2004).

X. Conclusions

A fundamental question in understanding the place of dopamine and serotonin interactions in treating schizophrenic patients will be the extent and ease that cognitive impairment and negative symptoms/deficit state can be treated by layering new therapeutics on a base of atypical antipsychotic drugs. Understanding the interactions of dopamine and serotonin with respect to modulating amino acid neurotransmission in macrocircuits involving the prefrontal cortex, the striatum, the thalamus, the hippocampus, and the brain stem with the ascending reticular activating system is a critical feature. Cognitive enhancement by modulating dopaminergic D1 receptor neurotransmission remains a relatively untapped direction. Cognitive enhancement with 5-HT_{1A} partial agonists, 5-HT_{2A} antagonists, 5-HT₄ partial agonists, and 5-HT₆ antagonists would appear to address these remaining unmet medical needs in schizophrenia. One region of the brain which would appear to require a greater emphasis would be the thalamus given the chemical heterogeneity of this anatomical structure and the relatively understudied components such as the midline and intralaminar thalamic nuclei, which appear to provide a crucial anatomical node that complements other thalamic systems which are easier to study such as the mediodorsal nucleus.

References

- Abi-Dargham, A. (2004). Do we still believe in the dopamine hypothesis? New data bring new evidence. *Int. J. Neuropsychopharmacol.* **7**(Suppl. 1), S1–S5.
- Aghajanian, G. K., and Marek, G. J. (1997). Serotonin induces excitatory postsynaptic potentials in apical dendrites of neocortical pyramidal cells. *Neuropharmacology* **36**, 589–599.
- Aghajanian, G. K., and Marek, G. J. (1999). Serotonin, via 5-HT_{2A} receptors, increases EPSCs in layer V pyramidal cells of prefrontal cortex by an asynchronous mode of glutamate release. *Brain Res.* **825**, 161–171.
- Aghajanian, G. K., and Sanders-Bush, E. (2002). Serotonin. In “Neuropsychopharmacology: The Fifth Generation of Progress” (K. L. Davis, D. S. Charney, J. T. Coyle, and C. Nemeroff, Eds.), pp. 15–34. Lippincott Williams & Wilkins, Philadelphia.
- Aghajanian, G. K., and Wang, R. Y. (1977). Habenular and other midbrain raphe afferents demonstrated by a modified retrograde tracing technique. *Brain Res.* **122**, 229–242.
- Akil, M., Pierri, J. N., Whitehead, R. E., Edgar, C. L., Mohila, C., Sampson, A. R., and Lewis, D. A. (1999). Lamina-specific alterations in the dopamine innervation of the prefrontal cortex in schizophrenic patients. *Am. J. Psychiatry* **156**, 1580–1589.
- Andersen, P. H. (1989). The dopamine uptake inhibitor GBR 12909: Selectivity and molecular mechanism of action. *Eur. J. Pharmacol.* **1989**, 493–504.
- Andrade, R., and Chaput, Y. (1991). 5-HT₄-like receptors mediate the slow excitatory response to serotonin in the rat hippocampus. *J. Pharmacol. Exp. Ther.* **257**, 930–937.

- Andrade, R., and Nicoll, R. A. (1987). Pharmacologically distinct actions of serotonin on single pyramidal neurones of the rat hippocampus recorded *in vitro*. *J. Physiol. (Lond.)* **394**, 99–124.
- Araneda, R., and Andrade, R. (1991). 5-Hydroxytryptamine₂ and 5-hydroxytryptamine_{1A} receptors mediate opposing responses on membrane excitability in rat association cortex. *Neuroscience* **40**, 399–412.
- Arnsten, A. F., and Goldman-Rakic, P. S. (1998). Noise stress impairs prefrontal cortical cognitive function in monkeys: Evidence for a hyperdopaminergic mechanism. *Arch. Gen. Psychiatry* **55**, 362–368.
- Azmitia, E. C. (1978). The serotonin-producing neurons of the midbrain median and dorsal raphe nuclei. In “Handbook of Psychopharmacology: Chemical Pathways of the Brain” (L. L. Iversen, S. Iversen, and S. H. Snyder, Eds.), Vol. 9. Plenum, New York, London.
- Barnes, N. M., and Sharp, T. (1999). A review of central 5-HT receptors and their function. *Neuropharmacology* **38**, 1083–1152.
- Baulac, M., Verney, C., and Berger, B. (1986). Innervation dopaminergique des regions parahippocamiques et hippocampiques du rat. *Rev. Neurol. (Paris)* **142**, 895–905.
- Belzung, C., Searce-Levie, K., Barreau, S., and Hen, R. (2000). Absence of cocaine-induced place conditioning in serotonin 5-HT_{1B} receptor knock-out mice. *Pharmacol. Biochem. Behav.* **66**, 221–225.
- Berendse, H. W., and Groenewegen, H. J. (1991). Restricted cortical termination fields of the midline and intralaminar thalamic nuclei in the rat. *Neuroscience* **42**, 73–102.
- Berger, B., Gaspar, P., and Verney, C. (1991). Dopaminergic innervation of the cerebral cortex: Unexpected differences between rodents and primates. *Trends Neurosci.* **14**, 21–27.
- Bergson, C., Levenson, R., Goldman-Rakic, P. S., and Lidow, M. S. (2003). Dopamine receptor-interacting proteins: The Ca²⁺ connection in dopamine signalling. *Trends Pharmacol. Sci.* **24**, 486–492.
- Bortolozzi, A., Diaz-Mataix, L., Scorza, M. C., Celada, P., and Artigas, F. (2005). The activation of 5-HT receptors in prefrontal cortex enhances dopaminergic activity. *J. Neurochem.* **95**, 1597–1607.
- Breese, G. R., Cooper, B. R., and Mueller, R. A. (1974). Evidence for involvement of 5-hydroxytryptamine in the actions of amphetamine. *Br. J. Pharmacol.* **52**, 307–312.
- Brown, L. L., Feldman, S. M., Smith, D. M., Cavanaugh, J. R., Ackermann, R. F., and Graybiel, A. M. (2002). Differential metabolic activity in the striosome and matrix compartments of the rat striatum during natural behaviors. *J. Neurosci.* **22**, 305–314.
- Brozoski, T. J., Brown, R. M., Rosvold, H. E., and Goldman, P. S. (1979). Cognitive deficit caused by regional depletion of dopamine in prefrontal cortex of rhesus monkey. *Science* **205**, 929–932.
- Bubser, M., and Deutch, A. Y. (2002). Differential effects of typical and atypical antipsychotic drugs on striosome and matrix components of the striatum. *Eur. J. Neurosci.* **15**, 713–720.
- Campbell, B. A., and Fibiger, H. C. (1971). Potentiation of amphetamine-induced arousal by starvation. *Nature* **233**, 424–425.
- Canales, J. J., and Graybiel, A. M. (2000). A measure of striatal function predicts motor stereotypy. *Nat. Neurosci.* **3**, 377–383.
- Capper-Loup, C., Canales, J. J., Kadaba, N., and Graybiel, A. M. (2002). Concurrent activation of dopamine D1 and D2 receptors is required to evoke neural and behavioral phenotypes of cocaine sensitization. *J. Neurosci.* **22**, 6218–6227.
- Carli, M., and Samanin, R. (1992). Serotonin₂ receptor agonists and serotonergic anorectic drugs affect rats' performance differently in a five-choice serial reaction time task. *Psychopharmacology* **106**, 228–234.
- Carli, M., Baviera, M., Invernizzi, R. W., and Balducci, C. (2004). The serotonin 5-HT_{2A} receptors antagonist M100907 prevents impairment in attentional performance by NMDA receptor blockade in the rat prefrontal cortex. *Neuropsychopharmacology* **29**, 1637–1647.

- Carlsson, A., and Lindquist, M. (1963). Effect of chlorpromazine and haloperidol on formation of 3-methoxytyramine and normetanephrine in mouse brain. *Acta Pharmacol. Toxicol. (Copenh.)* **20**, 140–144.
- Carr, D. B., and Sesack, S. R. (1996). Hippocampal afferents to the rat prefrontal cortex: Synaptic targets and relation to dopamine terminals. *J. Comp. Neurol.* **369**, 1–15.
- Carr, D. B., and Sesack, S. R. (2000). GABA-containing neurons in the rat ventral tegmental area project to the prefrontal cortex. *Synapse* **38**, 114–123.
- Carter, C. J., and Pycock, C. J. (1978). Differential effects of central serotonin manipulation on hyperactive and stereotyped behavior. *Life Sci.* **23**, 953–960.
- Carter, C. J., and Pycock, C. J. (1979). The effects of 5,7-dihydroxytryptamine lesions of extrapyramidal and mesolimbic sites on spontaneous motor behavior and amphetamine-induced stereotypy. *Naunyn Schmiedeberg's Arch. Pharmacol.* **308**, 51–54.
- Clinton, S. M., and Meador-Woodruff, J. H. (2004). Thalamic dysfunction in schizophrenia: Neurochemical, neuropathological, and *in vivo* imaging abnormalities. *Schizophr. Res.* **69**, 237–253.
- Crino, P. B., Morrison, J. H., and Hof, P. R. (1993). Monoaminergic innervation of cingulate cortex. In “Neurobiology of Cingulate Cortex and Limbic Thalamus” (B. A. Vogt and M. Gabriel, Eds.), pp. 285–310. Birkhauser, Boston.
- De Deurwaerdere, P., Navailles, S., Berg, K. A., Clarke, W. P., and Spampinato, U. (2004). Constitutive activity of the serotonin_{2C} receptor inhibits *in vivo* dopamine release in the rat striatum and nucleus accumbens. *J. Neurosci.* **24**, 3235–3241.
- De La Garza, R. I., and Cunningham, K. A. (2000). The effects of the 5-hydroxytryptamine_{1A} agonist 8-hydroxy-2-(di-n-propylamino)tetralin on spontaneous activity, cocaine-induced hyperactivity and behavioral sensitization: A microanalysis of locomotor activity. *J. Pharmacol. Exp. Ther.* **292**, 610–617.
- DeFelipe, J., and Jones, E. G. (1988). A light and electron microscopic study of serotonin-immunoreactive fibers and terminals in the monkey sensory-motor cortex. *Exp. Brain Res.* **71**, 171–182.
- Deschenes, M., Bourassa, J., and Pinault, D. (1994). Corticothalamic projections from layer V cells in rat are collaterals of long-range corticofugal axons. *Brain Res.* **664**, 215–219.
- DiMattio, V., De Blasi, A., Di Giulio, C., and Esposito, E. (2001). Role of 5-HT_{2C} receptor in the control of central dopamine function. *Trends Pharmacol. Sci.* **22**, 229–232.
- Doherty, M. D., and Pickel, V. M. (2000). Ultrastructural localization of the serotonin 2A receptor in dopaminergic neurons in the ventral tegmental area. *Brain Res.* **864**, 176–185.
- Duncan, G. E., Moy, S. S., Perez, A., Eddy, D. M., Zincow, W. M., and Lieberman, J. A. (2004). Deficits in sensorimotor gating and tests of social behavior in a genetic model of reduced NMDA receptor function. *Behav. Brain Res.* **153**, 507–519.
- Fallon, J. H., and Loughlin, S. E. (1995). Substantia nigra. In “The Rat Nervous System” (G. Paxinos, Ed.), pp. 215–237. Academic Press, Sydney.
- Farde, L., Nordstrom, A.-L., Wiesel, F.-A., Pauli, S., and Sedvall, G. (1992). Positron emission tomographic analysis of central D1 and D2 dopamine receptor occupancy in patients treated with classical neuroleptics and clozapine. *Arch. Gen. Psychiatry* **49**, 538–544.
- Filip, M., Bubar, M. J., and Cunningham, K. A. (2004). Contributions of serotonin (5-hydroxytryptamine; 5-HT) 5-HT₂ receptor subtypes to the hyperlocomotor effects of cocaine: Acute and chronic pharmacological analyses. *J. Pharmacol. Exp. Ther.* **310**, 1246–1254.
- Fletcher, P. J., Azampanah, A., and Korth, K. M. (2002a). Activation of 5-HT_{1B} receptors in the nucleus accumbens reduces self-administration of amphetamine on a progressive ratio schedule. *Pharmacol. Biochem. Behav.* **71**, 717–725.
- Fletcher, P. J., Grottick, A. J., and Higgins, G. A. (2002b). Differential effects of the 5-HT_{2A} receptor antagonist M100,907 and the 5-HT_{2C} receptor antagonist SB242,084 on cocaine-induced locomotor activity, cocaine self-administration and cocaine-induced reinstatement of responding. *Neuropsychopharmacology* **27**, 576–586.

- Floresco, S. B., Magyar, O., Ghods-Sharifi, S., Vexelman, C., and Tse, M. T. L. (2006). Multiple dopamine receptor subtypes in the medial prefrontal cortex of the rat regulate set-shifting. *Neuropsychopharmacology* **31**, 297–309.
- Frantz, K. J., Hansson, K. J., Stouffer, D. G., and Parsons, L. H. (2002). 5-HT₆ receptor antagonism potentiates the behavioral and neurochemical effects of amphetamine but not cocaine. *Neuropharmacology* **42**, 170–180.
- Frechilla, D., Cobreros, A., Saldise, L., Moratalla, R., Insausti, R., Luquin, M.-R., and Del Rio, J. (2001). Serotonin 5-HT_{1A} receptor expression is selectively enhanced in the striosomal compartment of chronic parkinsonian monkeys. *Synapse* **39**, 288–296.
- Frederick, D. L., Gillam, M. P., Lensing, S., and Paule, M. G. (1997). Acute effects of LSD on rhesus monkey operant test battery performance. *Pharmacol. Biochem. Behav.* **57**, 633–641.
- Fuxe, K., Agnati, L., and Everitt, B. (1975). Effects of metergoline on central monoamine neurons: Evidence of a selective blockade of central 5-HT receptors. *Neurosci. Lett.* **1**, 283–290.
- Gainetdinov, R. R., Wetsel, W. C., Jones, S. R., Levin, E. D., Jaber, M., and Caron, M. G. (1999). Role of serotonin in the paradoxical calming effect of psychostimulants on hyperactivity. *Science* **283**, 397–401.
- Gasbarri, A., Verney, C., Campana, E., and Pacitti, C. (1994). Anterograde and retrograde tracing from the ventral tegmental area to the hippocampal formation in the rat. *Brain Res. Bull.* **33**, 445–452.
- Gateley, S. J., Pan, D., Chen, R., Chaturvedi, G., and Ding, Y.-S. (1996). Affinities of methylphenidate derivatives for dopamine, norepinephrine and serotonin transporters. *Life Sci.* **58**, PL231–PL239.
- Gerfen, C. R. (1989). The neostriatal mosaic: Striatal patch-matrix organization is related to cortical lamination. *Science* **246**, 385–388.
- Gerfen, C. R., Engber, T. M., Mahan, L. C., Susel, Z., Chase, T. N., Monsma, F. J., and Sibley, D. R. (1990). D1 and D2 dopamine receptor-regulated gene expression of striatonigral and striatopallidal neurons. *Science* **250**, 1429–1432.
- Gervais, J., and Rouillard, C. (2000). Dorsal raphe stimulation differentially modulates dopaminergic neurons in the ventral tegmental area and substantia nigra. *Synapse* **35**, 281–291.
- Geyer, M. A., and Tamminga, C. A. (2004). Measurement and treatment research to improve cognition in schizophrenia: Neuropharmacological aspects. *Psychopharmacology* **174**, 1–2.
- Goldman-Rakic, P. S. (1982). Cytoarchitectonic heterogeneity of the primate neostriatum-subdivision into island and matrix cellular compartments. *J. Comp. Neurol.* **205**, 398–413.
- Goldman-Rakic, P. S., Leranth, C., Williams, S. M., Mons, N., and Geffard, M. (1989). Dopamine synaptic complex on pyramidal neurons in primate cerebral cortex. *Proc. Natl. Acad. Sci. USA* **86**, 9015–9019.
- Gordon, J. A., Lacefield, C. O., Kentros, C. G., and Hen, R. (2005). State-dependent alterations in hippocampal oscillations in serotonin_{1A} receptor-deficient mice. *J. Neurosci.* **25**, 6509–6519.
- Granon, S., Passetti, F., Thomas, K. L., Dalley, J. W., Everitt, B. J., and Robbins, T. W. (2000). Enhanced and impaired attentional performance after infusion of D1 dopaminergic receptor agents into rat prefrontal cortex. *J. Neurosci.* **20**, 1208–1215.
- Green, T. K., and Harvey, J. A. (1974). Enhancement of amphetamine action after interruption of ascending serotonergic pathways. *J. Pharmacol. Exp. Ther.* **190**, 109–117.
- Grillner, S., Markham, H., De Schutter, E., Silberberg, G., and LeBeau, F. E. N. (2005). Microcircuits in action: From CPGs to neocortex. *Trends Neurosci.* **28**, 525–533.
- Groenewegen, H. J., Wright, C. I., Beijer, A. V. J., and Voorn, P. (1999). Convergence and Segregation of ventral striatal inputs and outputs. In “Advancing from the Ventral Striatum to the Extended Amygdala” (J. F. McGinty, Ed.), Vol. 877, pp. 49–63. Annals NY Acad. Sci., New York.
- Haddjeri, N., Blier, P., and deMontigny, C. (1998). Long-term antidepressant treatments result in a tonic activation of forebrain 5-HT_{1A} receptors. *J. Neurosci.* **18**, 10150–10156.

- Hajos, M., Richards, C. D., Szekely, A. D., and Sharp, T. (1998). An electrophysiological and neuroanatomical study of the medial prefrontal cortical projection to the midbrain raphe nuclei in the rat. *Neuroscience* **87**, 95–108.
- Harrison, P. J. (2004). The hippocampus in schizophrenia: A review of the neuropathological evidence and its pathophysiological implications. *Psychopharmacology* **174**, 151–162.
- Heimer, L. (2003). A new anatomical framework for neuropsychiatric disorders and drug abuse. *Am. J. Psychiatry* **160**, 1726–1739.
- Hensler, J. G. (2006). Serotonergic modulation of the limbic system. *Neurosci. Biobehav. Rev.* **30**, 203–214.
- Herkenham, M., and Pert, C. B. (1981). Mosaic distribution of opiate receptors, parafascicular projections and acetylcholinesterase in rat striatum. *Nature* **291**, 415–418.
- Higgins, G. A., Enderlin, M., Haman, M., and Fletcher, P. J. (2003). The 5-HT_{2A} receptor antagonist M100,907 attenuates motor and ‘impulsive-type’ behaviors produced by NMDA receptor antagonism. *Psychopharmacology* **170**, 309–319.
- Hollister, A. S., Breese, G. R., and Cooper, B. R. (1974). Comparison of tyrosine hydroxylase and dopamine-hydroxylase inhibition with the effects of various 6-hydroxydopamine treatments on d-amphetamine induced motor activity. *Psychopharmacologia* **36**, 1–16.
- Jakab, R. L., and Goldman-Rakic, P. S. (1998). 5-HT_{2A} serotonin receptors in the primate cerebral cortex: Possible site of action of hallucinogens in pyramidal cell apical dendrites. *Proc. Natl. Acad. Sci. USA* **95**, 735–740.
- Jakab, R. L., and Goldman-Rakic, P. S. (2000). Segregation of serotonin 5-HT_{2A} and 5-HT₃ receptors in inhibitory circuits of the primate cerebral cortex. *J. Comp. Neurol.* **417**, 337–348.
- Jarvik, M. E., and Chorover, S. (1960). Impairment by lysergic acid diethylamide of accuracy in performance of a delayed alternation test in monkeys. *Psychopharmacologia* **1**, 221–230.
- Jay, T. M., Glowinski, J., and Thierry, A.-M. (1989). Selectivity of the hippocampal projection to the prefrontal area of the prefrontal cortex in the rat. *Brain Res.* **505**, 337–340.
- Kalen, P., Skagerberg, G., and Lindvall, O. (1988). Projections from the ventral tegmental area and mesencephalic raphe to the dorsal raphe nucleus in the rat. Evidence for a minor dopaminergic component. *Exp. Brain Res.* **73**, 69–77.
- Kapur, S., Zipursky, R. B., and Remington, G. (1999). Clinical and theoretical implications of 5-HT₂ and D₂ receptor occupancy of clozapine, risperidone, and olanzapine in schizophrenia. *Am. J. Psychiatry* **156**, 286–293.
- Kellendonk, C., Simpson, E. H., Polan, H. J., Malleret, G., Vronskaya, S., Winiger, V., Moore, H., and Kandel, E. R. (2006). Transient and selective overexpression of dopamine D₂ receptors in the striatum causes persistent abnormalities in prefrontal cortex functioning. *Neuron* **49**, 603–615.
- Kinomura, S., Larsson, J., Gulyas, B., and Roland, P. E. (1996). Activation by attention of the human reticular formation and thalamic intralaminar nuclei. *Science* **271**, 512–515.
- Koskinen, T., Ruotsalainen, S., Puumala, T., Lappalainen, R., Koivisto, E., Mannisto, P. T., and Sirvio, J. (2000). Activation of 5-HT_{2A} receptors impairs response control of rats in a five-choice serial reaction time task. *Neuropharmacology* **39**, 471–481.
- Kostrzewa, R. M., Reader, T. A., and Descarries, L. (1998). Serotonin neural adaptations to ontogenetic loss of dopamine neurons in rat brain. *J. Neurochem.* **70**, 889–898.
- Kuroda, M., Murakami, K., Iganashi, H., and Okada, A. (1996). The convergence of axon terminals of the mediodorsal thalamic nucleus and ventral tegmental area on pyramidal cells in layer V of the rat prefrontal cortex. *Eur. J. Neurosci.* **8**, 1340–1349.
- Lambe, E. K., Goldman-Rakic, P. S., and Aghajanian, G. K. (2000). Serotonin induces EPSCs preferentially in layer V pyramidal neurons of the frontal cortex in the rat. *Cereb. Cortex* **10**, 974–980.
- Laroche, S., Jay, T. M., and Thierry, A.-M. (1990). Long-term potentiation in the prefrontal cortex following stimulation of the hippocampal CA1/subicular region. *Neurosci. Lett.* **114**, 184–190.

- Laruelle, M., Kegeles, L. S., Frankle, G. W., Narendran, R., Gil, R., Talbot, P. S., Hwang, D.-R., Huang, Y., Cooper, T., Mark, S., and Abi-Dargham, A. (2005). Schizophrenia is associated with increased synaptic dopamine in associative rather than limbic regions of the striatum. *Neuropsychopharmacology* **30**(Suppl. 1), S196.
- Lavin, A., and Grace, A. A. (1998). Dopamine modulates the responsiveness of mediodorsal thalamic cells recorded *in vitro*. *J. Neurosci.* **18**, 10566–10578.
- Lavin, A., Moore, H. M., and Grace, A. A. (2005). Prenatal disruption of neocortical development alters prefrontal cortical neuron responses to dopamine in adult rats. *Neuropharmacology* **30**, 1426–1435.
- Levitt, P., Eagleson, K. L., and Powell, E. M. (2004). Regulation of neocortical interneuron development and the implications for neurodevelopmental disorders. *Trends Neurosci.* **27**, 400–406.
- Lezcano, N., and Bergson, C. (2002). D1/D5 dopamine receptors stimulate intracellular calcium release in primary cultures of neocortical and hippocampal neurons. *J. Neurophysiol.* **87**, 2167–2175.
- Lezcano, N., Mrzljak, L., Eubanks, S., Levenson, R., Goldman-Rakic, P. S., and Bergson, C. (2000). Dual signaling regulated by calcyon, a D1 dopamine receptor interacting protein. *Science* **287**, 1660–1664.
- Li, S. X.-M., Perry, K. W., and Wong, D. T. (2002). Influence of fluoxetine on the ability of bupropion to modulate extracellular dopamine and norepinephrine concentrations in three mesocorticolimbic areas of rats. *Neuropharmacology* **42**, 181–190.
- Lipska, B. K. (2004). Using animal models to test a neurodevelopmental hypothesis of schizophrenia. *J. Psychiatry Neurosci.* **29**, 282–286.
- Lipska, B. K., Al-Amin, H. A., and Weinberger, D. R. (1998). Excitotoxic lesions of the rat medial prefrontal cortex: Effects on abnormal behaviors associated with neonatal hippocampal damage. *Neuropsychopharmacology* **19**, 451–464.
- Lopez-Gimenez, J. F., Mengod, G., Palacios, J. M., and Vilario, M. T. (1999). Human striosomes are enriched in 5-HT_{2A} receptors: Autoradiographical visualization with [³H]MDL100,907, [¹²⁵I](±)DOI and [³H]ketanserin. *Eur. J. Neurosci.* **11**, 3761–3765.
- Lucki, I., and Harvey, J. A. (1979). Increased sensitivity to d- and l-amphetamine action after midbrain raphe lesions as measured by locomotor activity. *Neuropharmacology* **18**, 243–249.
- Marek, G. J., and Aghajanian, G. K. (1999). 5-HT_{2A} or α_1 -adrenoceptor activation induces excitatory postsynaptic currents in layer V pyramidal cells of the medial prefrontal cortex. *Eur. J. Pharmacol.* **367**, 197–206.
- Marek, G. J., Wright, R. A., Gewirtz, J. C., and Schoepp, D. D. (2001). A major role for thalamocortical afferents in serotonergic hallucinogen receptor function in the rat neocortex. *Neuroscience* **105**, 379–392.
- Martin-Ruiz, R., Puig, M. V., Celada, P., Shapirio, D. A., Roth, B. L., Mengod, G., and Artigas, F. (2001a). Control of serotonergic function in medial prefrontal cortex by serotonin-2A receptors through a glutamate-dependent mechanism. *J. Neurosci.* **21**, 9856–9866.
- Martin-Ruiz, R., Ugedo, L., Honrubia, M. A., Mengod, G., and Artigas, F. (2001b). Control of serotonergic neurons in rat brain by dopaminergic receptors outside the dorsal raphe nucleus. *J. Neurochem.* **77**, 762–775.
- McMahon, L. R., and Cunningham, K. A. (1999). Antagonism of 5-hydroxytryptamine₄ receptors attenuates hyperactivity induced by cocaine: Putative role for 5-hydroxytryptamine₄ receptors in the nucleus accumbens shell. *J. Pharmacol. Exp. Ther.* **291**, 300–307.
- McMahon, L. R., and Cunningham, K. A. (2001a). Antagonism of 5-hydroxytryptamine_{2A} receptors attenuates the behavioural effects of cocaine in rats. *J. Pharmacol. Exp. Ther.* **297**, 357–363.

- McMahon, L. R., and Cunningham, K. A. (2001b). Role of 5-HT_{2A} and 5-HT_{2B/2C} receptors in the behavioral interactions between serotonin and catecholamine reuptake inhibitors. *Neuropsychopharmacology* **24**, 319–329.
- McMahon, L. R., Filip, M., and Cunningham, K. A. (2001). Differential regulation of the mesoaccumbens circuit by serotonin 5-hydroxytryptamine (5-HT)_{2A} and 5-HT_{2C} receptors. *J. Neurosci.* **21**, 7781–7787.
- Meador-Woodruff, J. H., Grandy, D. K., Van Tol, H. H. M., Damask, S. P., Little, K. Y., Civelli, O. I., and Watson, S. J. (1994). Dopamine receptor gene expression in the human medial temporal lobe. *Neuropsychopharmacology* **10**, 239–248.
- Melchitzky, D. S., and Lewis, D. A. (2001). Dopamine transporter immunoreactive axons in the mediodorsal thalamic nucleus of the macaque monkey. *Neuroscience* **103**, 1033–1042.
- Meltzer, H. Y. (1999). The role of serotonin in antipsychotic drug action. *Neuropsychopharmacology* **21**, 106S–115S.
- Meltzer, H. Y., Matsubara, S., and Lee, J.-C. (1989). Classification of typical and atypical antipsychotic drugs on the basis of dopamine D-1, D-2 and serotonin₂ pK_i values. *J. Pharmacol. Exp. Ther.* **251**, 238–246.
- Mengod, G., Vilaro, M. T., Raurich, A., Lopez-Gimenez, J. F., Cortes, R., and Palacios, J. M. (1996). 5-HT receptors in mammalian brain: Receptor autoradiography and *in situ* hybridization studies of new ligands and newly identified receptors. *Histochem. J.* **28**, 747–758.
- Millan, M. J., Lejeune, F., and Gobert, A. (2000). Reciprocal autoreceptor and heteroreceptor control of serotonergic, dopaminergic and noradrenergic transmission in the frontal cortex: Relevance to the actions of antidepressant agents. *J. Psychopharmacol.* **14**, 114–138.
- Miner, L. A. H., Backstrom, J. R., Sanders-Bush, E., and Sesack, S. R. (2003). Ultrastructural localization of serotonin_{2A} receptors in the middle layers of the rat prefrontal cortex. *Neuroscience* **116**, 107–117.
- Miyamoto, S., Snouwaert, J. N., Koller, B. H., Moy, S. S., Lieberman, J. A., and Duncan, G. E. (2004). Amphetamine-induced Fos is reduced in limbic cortical regions but not in the caudate or accumbens in a genetic model of NMDA receptor hypofunction. *Neuropsychopharmacology* **29**, 2180–2188.
- Mohn, A. R., Gainetdinov, R. R., Caron, M. G., and Koller, B. H. (1999). Mice with reduced NMDA receptor expression display behaviors related to schizophrenia. *Cell* **98**, 427–436.
- Moore, H., Jentsch, J. D., Ghajarnia, M., Geyer, M. A., and Grace, A. A. (2006). (March 31 Epub). A neurobehavioral systems analysis of adult rats exposed to methylazoxymethanol acetate on E17: Implications for the neuropathology of schizophrenia. *Biol. Psychiatry* **60**(3), 253–264.
- Moore, R. Y., and Bloom, F. E. (1978). Central catecholamine neuron systems: Anatomy and physiology of the dopamine systems. *Ann. Rev. Neurosci.* **1**, 129–169.
- Morales, M., Battenberg, E., de Lecea, L., and Bloom, F. E. (1996). The type 3 serotonin receptor is expressed in a subpopulation of GABAergic neurons in the rat neocortex and hippocampus. *Brain Res.* **731**, 199–202.
- Mrzljak, L., Bergson, C., Pappy, M., Huff, R., Levenson, R., and Goldman-Rakic, P. S. (1996). Localization of dopamine D4 receptors in GABAergic neurons of the primate brain. *Nature* **381**, 245.
- Neill, D. B., Grant, L. D., and Grossman, S. P. (1972). Selective potentiation of locomotor effects of amphetamine by midbrain raphe lesions. *Physiol. Behav.* **9**, 655–657.
- Neumaier, J. F., Vincow, E. S., Arvanitogiannis, A., Wise, R. A., and Carlezon, W. A. J. (2002). Elevated expression of 5-HT_{1B} receptors in nucleus accumbens efferents sensitizes animals to cocaine. *J. Neurosci.* **22**, 10856–10863.
- Neve, K. A., Seamans, J. K., and Trantham-Davidson, H. (2004). Dopamine receptor signalling. *J. Recept. Signal Transduct. Res.* **24**, 165–205.
- Nicola, S. M., Surmeier, D. J., and Malenka, R. C. (2000). Dopaminergic modulation of neuronal excitability in the striatum and nucleus accumbens. *Annu. Rev. Neurosci.* **23**, 185–215.

- Nishi, A., Bibb, J. A., Snyder, G. L., Higashi, H., Nairn, A. C., and Greengard, P. (2000). Amplification of dopaminergic signalling by a positive feedback loop. *Proc. Natl. Acad. Sci. USA* **97**, 12840–12845.
- Nocjar, C., Roth, B. L., and Pehek, E. A. (2002). Localization of 5-HT_{2A} receptor on dopamine cells in subnuclei of the midbrain A10 cell group. *Neuroscience* **111**, 163–176.
- O'Dell, L. E., and Parsons, L. H. (2004). Serotonin1B receptors in the ventral tegmental area modulate cocaine-induced increases in nucleus accumbens dopamine levels. *J. Pharmacol. Exp. Ther.* **311**, 711–719.
- O'Donnell, P., and Grace, A. A. (1995). Synaptic interactions among excitatory afferents to nucleus accumbens neurons: Hippocampal gating of prefrontal cortical input. *J. Neurosci.* **15**, 3622–3638.
- O'Hearn, E., and Molliver, M. E. (1984). Organization of the raphe-cortical projections in the rat: A quantitative retrograde study. *Brain Res. Bull.* **13**, 709–726.
- O'Neill, M. F., Heron-Maxwell, C. L., and Shaw, G. (1999). 5-HT₂ receptor antagonism reduces hyperactivity induced by amphetamine, cocaine, and MK-801 but not D1 agonist C-APB. *Pharmacol. Biochem. Behav.* **63**, 237–243.
- Onn, S.-P., Wang, X.-B., Lin, M., and Grace, A. A. (2006). Dopamine D1 and D4 receptor subtypes differentially modulate recurrent excitatory synapses in prefrontal cortical pyramidal neurons. *Neuropsychopharmacology* **31**, 318–338.
- Papadopoulos, G. C., Parnevelas, J. G., and Buijs, R. M. (1987). Light and electron microscopic immunocytochemical analysis of the serotonin innervation of the rat visual cortex. *J. Neurocytol.* **16**, 883–892.
- Passetti, F., Dalley, J. W., and Robbins, T. W. (2003). Double dissociation of serotonergic and dopaminergic mechanisms on attentional performance using rodenet five-choice reaction time test. *Psychopharmacology* **165**, 136–145.
- Pehek, E. A., McFarlane, H. G., Maguschak, K., Price, B., and Pluto, C. P. (2001). M100,907, a selective 5-HT_{2A} antagonist, attenuates dopamine release in the rat medial prefrontal cortex. *Brain Res.* **888**, 51–59.
- Pehek, E. A., Nocjar, C., Roth, B. L., Byrd, T. A., and Mabrouk, O. S. (2006). Evidence for the preferential involvement of 5-HT_{2A} serotonin receptors in stress- and drug-induced dopamine release in the rat medial prefrontal cortex. *Neuropsychopharmacology* **31**, 265–277.
- Peyron, C., Luppi, P. H., Kitahama, K., Fort, P., Hermann, D. M., and Jouvet, M. (1995). Origin of the dopaminergic innervation of the rat dorsal raphe nucleus. *NeuroReport* **6**, 2527–2531.
- Peyron, C., Petit, J. M., Rampon, C., Jouvet, M., and Luppi, P. H. (1998). Forebrain afferents to the dorsal raphe nucleus demonstrated by retrograde and anterograde tracing methods. *Neuroscience* **82**, 443–468.
- Ragsdale, C. W., and Graybiel, A. M. (1990). A simple ordering of neocortical areas established by the compartmental organization of their striatal projections. *Proc. Natl. Acad. Sci. USA* **87**, 6196–6199.
- Read, H. L., Beck, S. G., and Dun, N. J. (1994). Serotonergic suppression of interhemispheric cortical synaptic potentials. *Brain Res.* **643**, 17–28.
- Rieck, R. W., Ansari, M. S., Whetsell, W. O. J., Deutch, A. Y., and Kessler, R. M. (2004). Distribution of dopamine D2-like receptor in the human thalamus: Autoradiographic and PET studies. *Neuropsychopharmacology* **29**, 362–372.
- Roth, B. L., Hanizavareh, S. M., and Blum, A. E. (2004). Serotonin receptors represent highly favorable molecular targets for cognitive enhancement in schizophrenia and other disorders. *Psychopharmacology* **174**, 17–24.
- Samson, Y., Wu, J. J., Friedman, A. H., and Davis, J. N. (1990). Catecholaminergic innervation of the hippocampus in the cynomolgus monkey. *J. Comp. Neurol.* **298**, 250–263.
- Sanchez-Gonzalez, M. A., Garcia-Cabezas, M. A., Rico, B., and Cavada, C. (2005). The primate thalamus is a key target for brain dopamine. *J. Neurosci.* **25**, 6076–6083.

- Schlemmer, R. F., and Davis, J. M. (1986). A primate model for the study of hallucinogens. *Pharmacol. Biochem. Behav.* **24**, 381–392.
- Seamans, J. K., and Yang, C. R. (2004). The principle features and mechanisms of dopamine modulation in the prefrontal cortex. *Prog. Neurobiol.* **74**, 1–57.
- Segal, D. S. (1976). Differential effects of p-chlorophenylalanine on amphetamine-induced locomotion and stereotypy. *Brain Res.* **116**, 267–276.
- Seguela, P., Watkins, K. C., and Descarries, L. (1989). Ultrastructural relationships of serotonin axon terminals in the cerebral cortex of the adult rat. *J. Comp. Neurol.* **289**, 129–142.
- Selemon, L. D., and Goldman-Rakic, P. S. (1999). The reduced neuropil hypothesis: A circuit based model of schizophrenia. *Biol. Psychiatry* **45**, 17–25.
- Selemon, L. D., Rajkowska, G., and Goldman-Rakic, P. S. (1995). Abnormally high neuronal density in the schizophrenic cortex. *Arch. Gen. Psychiatry* **52**, 805–818.
- Sesack, S. R., Deutch, A. Y., Roth, R. H., and Bunney, B. S. (1989). Topographical organization of the efferent projections of the medial prefrontal cortex in the rat: An anterograde tract-tracing study with Phaseolus vulgaris leucoagglutinin. *J. Comp. Neurol.* **290**, 213–242.
- Sherman, S. M., and Guillery, R. W. (1998). On the actions that one nerve cell can have on another: Distinguishing “drivers” from “modulators.” *Proc. Natl. Acad. Sci. USA* **95**, 7121–7126.
- Silberberg, G., Grillner, S., LeBeau, F. E. N., Maex, R., and Markham, H. (2005). Synaptic pathways in neural microcircuits. *Trends Neurosci.* **28**, 541–551.
- Simon, H., Scatton, B., and Le Moal, M. (1980). Dopaminergic A10 neurons are involved in cognitive function. *Nature* **286**, 150–151.
- Smiley, J. F., and Goldman-Rakic, P. S. (1993). Heterogenous targets of dopamine synapses in monkey prefrontal cortex demonstrated by serial section electron microscopy. A laminar analysis using the silver enhanced diaminobenzidine-sulfide (SEDS) immunolabeling technique. *Cereb. Cortex* **3**, 223–238.
- Smiley, J. F., and Goldman-Rakic, P. S. (1996). Serotonergic axons in monkey prefrontal cerebral cortex synapse predominantly on interneurons as demonstrated by serial section electron microscopy. *J. Comp. Neurol.* **367**, 431–443.
- Smiley, J. F., Levey, A. I., Ciliax, B. J., and Goldman-Rakic, P. S. (1994). D1 dopamine receptor immunoreactivity in human and monkey cerebral cortex: Predominant and extrasynaptic localization in dendritic spines. *Proc. Natl. Acad. Sci. USA* **91**, 5720–5724.
- Surmeier, D. J., Song, W.-J., and Yan, Z. (1996). Coordinated expression of dopamine receptors in neostriatal medium spiny neurons. *J. Neurosci.* **16**, 6579–6591.
- Svenningsson, P., Tzavara, E. T., Liu, F., Fienberg, A. A., Nomikos, G. G., and Greengard, P. (2002). DARP-32 mediates serotonergic neurotransmission in the forebrain. *Proc. Natl. Acad. Sci. USA* **99**, 3188–3193.
- Symond, M. B., Harris, A. W. F., Gordon, E., and Williams, L. M. (2005). “Gamma synchrony” in first-episode schizophrenia: A disorder of temporal connectivity. *Am. J. Psychiatry* **162**, 459–465.
- Talpos, J. C., Wilkinson, L. S., and Robbins, T. W. (2006). A comparison of multiple 5-HT receptors in two tasks measuring impulsivity. *J. Psychopharmacol.* **20**, 47–58.
- Tanaka, E., and North, R. A. (1993). Actions of 5-hydroxytryptamine on neurons of the rat cingulate cortex. *J. Neurophysiol.* **69**, 1749–1757.
- Terry, A. V. J., Buccafusco, J. J., and Bartoszyk, G. D. (2005). Selective serotonin 5-HT_{2A} receptor antagonist EMD 281014 improves delayed matching performance in young and aged rhesus monkeys. *Psychopharmacology* **179**, 725–732.
- Tork, I. (1990). Anatomy of the serotonergic system. In “Ann. N.Y. Acad. Sci.” (P. M. Whitaker-Azmitia and S. J. Peroutka, Eds.), Vol. 600, pp. 9–35. The New York Academy of Sciences, New York.
- Van Bockstaele, E. J., and Pickel, V. M. (1995). GABA-containing neurons in the ventral tegmental area project to nucleus accumbens in rat brain. *Brain Res.* **682**, 215–221.

- Van Bockstaele, E. J., Biswas, A., and Pickel, V. M. (1993). Topography of serotonin neurons in the dorsal raphe nucleus that send axon collaterals to the rat prefrontal cortex and nucleus accumbens. *Brain Res.* **624**, 188–198.
- Van der Werf, Y. D., Witter, M. P., Uylings, H. B. M., and Jolles, J. (2000). Neuropsychology of infarctions in the thalamus: A review. *Neuropsychologia* **38**, 613–627.
- Van der Werf, Y. D., Witter, M. P., and Groenewegen, H. J. (2002). The intralaminar and midline nuclei of the thalamus. Anatomical and functional evidence for participation in processes of arousal and awareness. *Brain Res. Rev.* **39**, 107–140.
- Van Groen, T., and Wyss, J. M. (1995). Projections from the anterodorsal and anteroventral nucleus of the thalamus to the limbic cortex in the rat. *J. Comp. Neurol.* **358**, 584–604.
- Van Groen, T., Kadish, I., and Wyss, J. M. (1999). Efferent connections of the anteromedial nucleus of the thalamus of the rat. *Brain Res. Rev.* **30**, 1–26.
- Verney, C., Baulac, M., Berger, B., Alvarez, C., Vigny, A., and Helle, K. B. (1985). Morphological evidence for a dopaminergic terminal field in the hippocampal formation of young and adult rat. *Neuroscience* **14**, 1039–1052.
- Vertes, R. P. (1991). A PHA-L analysis of ascending projections of the dorsal raphe nucleus in the rat. *J. Comp. Neurol.* **313**, 643–668.
- Vertes, R. P., Fortin, W. J., and Crane, A. M. (1999). Projections of the median raphe nucleus in the rat. *J. Comp. Neurol.* **407**, 555–582.
- Waeber, C., and Palacios, J. M. (1994). Binding sites for 5-hydroxytryptamine-2 receptor agonists are predominantly located in striosomes in the human basal ganglia. *Mol. Brain Res.* **24**, 199–209.
- Warbritton, J. O., III, Steward, M., and Baldessarini, R. J. (1978). Decreased locomotor activity and attenuation of amphetamine hyperactivity with intraventricular infusion of serotonin in the rat. *Brain Res.* **143**, 373–382.
- Williams, G. V., Rao, S. G., and Goldman-Rakic, P. S. (2002). The physiological role of 5-HT_{2A} receptors in working memory. *J. Neurosci.* **22**, 2843–2854.
- Wilson, C. J. (1998). Basal ganglia. In “The Synaptic Organization of the Brain” (G. M. Shepherd, Ed.), pp. 329–375. Oxford University Press, New York.
- Winstanley, C. A., Chudasama, Y., Dalley, J. W., Theobald, D. E. H., Glennon, J. C., and Robbins, T. W. (2003). Intra-prefrontal 8-OH-DPAT and M100907 improve visuospatial attention and decrease impulsivity on the five-choice serial reaction time task in rats. *Psychopharmacology* **167**, 304–314.
- Winstanley, C. A., Dalley, J. W., Theobald, D. E. H., and Robbins, T. W. (2004a). Fractionating impulsivity: Contrasting effects of central 5-HT depletion on different measures of impulsive behavior. *Neuropsychopharmacology* **29**, 1331–1343.
- Winstanley, C. A., Theobald, D. E. H., Dalley, J. W., Glennon, J. C., and Robbins, T. W. (2004b). 5-HT_{2A} and 5-HT_{2C} receptor antagonists have opposing effects on a measure of impulsivity: Interactions with global 5-HT depletion. *Psychopharmacology* **176**, 376–385.
- Winterer, G., and Weinberger, D. R. (2004). Genes, dopamine and cortical signal-to-noise ratio in schizophrenia. *Trends Neurosci.* **27**, 683–690.
- Wyss, J. M., Swanson, L., and Cowan, W. M. (1980). The organization of the fimbria, dorsal fornix, and ventral hippocampal commissure in the rat. *Anat. Embryol.* **158**, 303–316.
- Zahrt, J., Taylor, J. R., Mathew, R. G., and Arnsten, A. F. (1997). Supranormal stimulation of D1 dopamine receptors in the rodent prefrontal cortex impairs spatial working memory performance. *J. Neurosci.* **17**, 8528–8535.
- Zornoza, T., Cano-Cebrian, M. J., Miquel, M., Aragon, C., Polache, A., and Granero, L. (2005). Hippocampal dopamine receptors modulate the motor activation and the increase in dopamine levels in the rat n. accumbens evoked by chemical stimulation of the ventral hippocampus. *Neuropsychopharmacology* **30**, 843–852.

CHOLINERGIC CIRCUITS AND SIGNALING IN THE PATHOPHYSIOLOGY OF SCHIZOPHRENIA

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Central cholinergic signaling has long been associated with aspects of memory, motivation, and mood, each affected functions in neuropsychiatric disorders such as schizophrenia. In this chapter, we review evidence related to the core hypothesis that dysregulation of central cholinergic signaling contributes to the pathophysiology of schizophrenia. Although central cholinergic circuits are resistant to simplification—particularly when one tries to parse the contributions of various classes of cholinergic receptors to disease related phenomena—the potential role of ACh signaling in Schizophrenia pathophysiology deserves careful consideration for prospective therapeutics. The established role of cholinergic circuits in attentional tuning is considered along with recent work on how the patterning of cholinergic activity may modulate corticostriatal circuits affected in schizophrenia.

I. Introduction

Cholinergic innervation of cortical and striatal brain areas is extensive and diffuse, as are both pre- and postsynaptic targets for acetylcholine (ACh) interaction. Receptors for ACh (AChRs) come in two broad classes—ionotropic (nicotinic) and metabotropic (muscarinic)—each class having multiple subtypes with both opposing and synergistic actions. Activation of these receptors regulates neuronal excitability by interaction with pre- and postsynaptically localized ACh-binding sites. ACh can act as a tonic, diffuse signal, modulating the release of ACh and other transmitters, including dopamine, glutamate, and GABA. Alternatively, ACh can exert its effects via highly localized and directed interactions with neuronal AChRs to increase or decrease neuronal firing.

The complexity of CNS cholinergic circuits and signaling mechanisms produces a system in which origins and end results may be easier to appreciate than intervening steps. It is clear that ACh, released from the cholinergic inputs of the basal forebrain, striatal, and the pontomesencephalic (PM) areas, plays an important role in supporting neurocognitive and motivational functions of the prefrontal cortical, hippocampal, and ventral tegmental projections to the striatum (for reviews see Cragg, 2006; Gotti and Clementi, 2004; chapter by Martin and Freedman, this volume; Mesulam, 2004; Sarter *et al.*, 2005; Smythies, 2005; Wonnacott *et al.*, 2005). In addition, there is considerable evidence that events which reduce the amount of ACh at cholinergic targets may contribute to functional deficits—including deficits related to schizophrenia (Hyde and Crook, 2001; Sarter *et al.*, 2005; chapter by Martin and Freedman, this volume). But considerable confusion sets in when one tries to extract exactly *how* the intervening steps, with activation of muscarinic and/or nicotinic receptors and consequent changes in downstream circuits, are integrated to elicit the broad spectrum of effects modulated by cholinergic signaling.

Some of the confusion arises from attempts to reconcile the varying “anti-cholinergic” properties of antipsychotic medications with data on the effects of muscarinic agonists *per se*. Further confusion arises from the fact that commonly used cholinergic ligands may be less specific in their binding properties than previously thought: indeed, some compounds traditionally considered selective muscarinic antagonists may function as partial agonists or antagonists of other ACh (nicotinic and muscarinic) receptor subtypes. Finally, schemes that overemphasize the role of a particular ACh-signaling pathway to the exclusion of others, rather than viewing the function of cholinergic circuits as the result of the summation of actions of ACh at all of its receptors, may do more to confuse than enlighten. Anatomical and functional data underscore the interaction of cholinergic circuits with other neurotransmitter systems (Smiley *et al.*, 1999). Indeed, interaction of ACh with its full panoply of receptor sites elicits substantive changes in the synaptic transmission of dopamine, glutamate, serotonin, and GABA in a variety of brain regions.

We will first provide a thumbnail sketch of cholinergic circuits and then examine how they are involved in functions relevant to schizophrenia with focus on interactions with dopamine- and glutamate-mediated signaling. We will then review developmental/genetic, pathological, and pharmacological evidence for potential cholinergic contributions to schizophrenia. Although cholinergic signaling may not be the major site of circuit dysregulation underlying the etiology of schizophrenia, more knowledgeable manipulation of cholinergic systems may provide an untapped reservoir of considerable therapeutic potential in the treatment of the positive and negative symptoms of this complex disease.

II. ACh in Brain Regions Implicated in Schizophrenia

Central cholinergic circuits participate in aspects of memory formation, motivational and volitional behaviors, and affect. Each of these functions is altered in neuropsychiatric disorders, including schizophrenia. Cholinergic neurons in the CNS make up for any apparent deficit in numbers by projecting to a broad swath of cerebral cortical mantle, select portions of the temporal lobe, and by their profuse axonal arborizations throughout the corpus striatum. The schematic diagram presented in Fig. 1 attempts to bring some order to the cholinergic chaos by corraling the diverse targets of ACh innervation into a manageable subset of brain regions strongly implicated in schizophrenia. Our focus is represented in primary colors: red for cholinergic neuronal groups and a subset of their projections, and yellow for the chosen cholinceptive targets—brain regions that have been examined in detail in recent circuit analyses and that will be the focus of this chapter. Obviously this degree of simplification endangers the generality of our

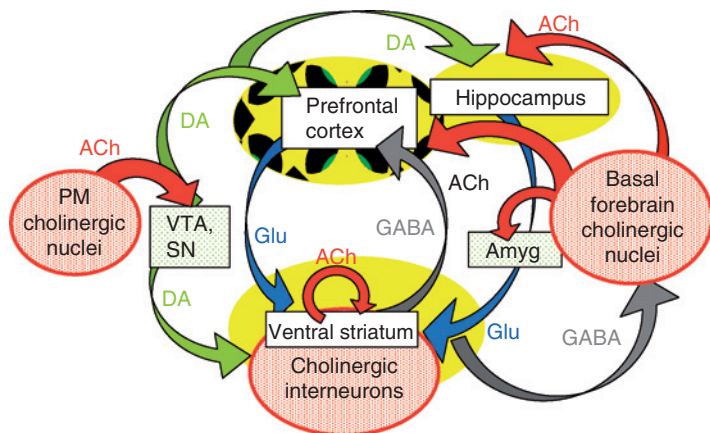


FIG. 1. Schematic diagram of cholinergic circuits (in red) and their projections within a subset of key brain regions affected in SZ. Cholinergic inputs to prefrontal cortex and hippocampus arise primarily from the basal forebrain group, including the septal cholinergic neurons, the nbM, the preoptic and diagonal band nuclei. Other contributors to the forebrain ACh group are neurons within the substantia innominata and ventral pallidum. The second major subgroup of ACh-containing neurons, the pontomesencephalic (PM) cholinergic neurons, provides input to brainstem aminergic nuclei (e.g., VTA, SN, and raphe). Cholinergic interneurons intrinsic to the basal ganglia are thought to modulate the relative impact of glutamatergic, dopaminergic, and GABAergic circuits within the ventral striatum. Potential mechanisms of cholinergic regulation of neuronal excitability in prefrontal cortex and hippocampus are also discussed in the text.

considerations—for example, there is little discussion of cholinergic signaling in amygdala, or in cingulate or somatosensory cortex—areas of study that have contributed important progress to our understanding of central cholinergic coding. Such omissions are not intended to infer relative impact on the field, but rather reflect the limits of time, space, and comprehension of the authors.

Overall, there are three major groups of cholinergic neurons and interneurons within the primate brain. Cholinergic inputs to prefrontal cortex and hippocampus arise primarily from the basal forebrain group, which includes the medial septal cholinergic neurons, the nucleus basalis of Meynert (nbM) and the preoptic and diagonal band nuclei (Fig. 1, right). Other contributors to the forebrain ACh groups are neurons within the substantia innominata and ventral pallidum. The relative contribution of cholinergic versus noncholinergic neurons to each of the basal forebrain nuclei ranges from <5% to 90% in human brain, with the nbM as the highest density forebrain cholinergic nucleus (Mesulam *et al.*, 1984; reviewed in Hyde and Crook, 2001). Other major targets of the basal forebrain groups include the amygdala, olfactory bulb, and hypothalamus (Woolf, 1991).

The second major subgroup of ACh-containing neurons (Fig. 1, left), the PM cholinergic neurons, provides input to brainstem aminergic nuclei (e.g., VTA,

SN, raphe) as well as to the cerebellum, thalamus, and hypothalamus (Woolf, 1991). In addition, the PM group of neurons project to the basal forebrain cholinergic neurons, thereby coordinating central cholinergic modulation of brainstem, midbrain, and forebrain circuits.

The final major group of cholinergic neurons consists of the ACh interneurons that are intrinsic to the basal ganglia. These intrastriatal neurons modulate the relative impact of multiple glutamatergic and dopaminergic circuits on the medium spiny GABAergic projection neurons of the striatum: our focus will be on the role of cholinergic circuits in the regulation of ventral striatal signaling.

A. CHOLINERGIC PATHWAYS WITHIN THE VENTRAL STRIATUM

Cholinergic neurons within the striatum are typically large, aspiny neurons that comprise 1–5% of the striatal interneurons, varying somewhat with species and consideration of dorsal versus ventral regions. The extensive arborizations of the striatal cholinergic interneurons throughout the corpus striatum provide a tonic level of ACh release, with the ultimate concentration of extracellular ACh being set (and reset) by the interplay of local ACh release and the activity of the omnipresent acetylcholinesterase (AChE).

Striatal cholinergic neurons are characterized by their tonic activation profile, and periods of relatively low activity (referred to as “pauses”) can be associated with salience and prediction of reward (Cragg, 2006; Graybiel *et al.*, 1994). The phasic lulls and peaks of the striatal cholinergic activity, which alters local ACh and choline concentrations, are influenced by corticostriatal projections from hippocampal subicular and prefrontal glutamatergic neurons (Fig. 1, in blue), as well as from amygdala, cingulate, and other cerebral and temporal cortices (not shown). In the rodent, the bulk of the hippocampal projections to the nucleus accumbens arise from ventral rather than dorsal hippocampal areas analogous to the more anterior portions of the primate hippocampus.

Dopaminergic inputs to the ventral striatum arise from the ventral tegmentum (VTA) and substantia nigra (SN), which themselves are recipients of cholinergic projections from PM neurons. Activation of a variety of pre- and postsynaptic dopamine receptors strongly regulates the release of ACh and the excitability of striatal cholinergic interneurons (Maurice *et al.*, 2004; Wang *et al.*, 2006). In addition, local circuits of opiate peptide and GABAergic neurons influence the net levels of striatal cholinergic tone.

Even this, admittedly limited, summary of key regulators of the spatial and temporal profile of ACh-mediated signaling in striatum reveals considerable complexity. Nevertheless, recent progress in dissecting the interaction of cholinergic circuits with dopaminergic and glutamatergic inputs to the striatum inspires considerable hope that we may be approaching a *bona fide* understanding

of how ACh works at least in *one* region that is known to effect in schizophrenia and that is particularly high in ACh tone (see below and Cragg, 2006; Calabresi *et al.*, 2000; Wilson, 2006; Wonnacott, 2005 for reviews; Wang *et al.*, 2006).

B. CHOLINERGIC PROJECTIONS IN PREFRONTAL CORTEX AND HIPPOCAMPUS

The principal source of cholinergic input to the PFC and hippocampus is from the basal forebrain nuclei, with particularly strong contributions from the medial septum in rodent and from the nbM in human brain. The primate prefrontal cortex receives a fairly homogeneous cholinergic input with the highest density of cholinergic marker-positive fibers in layers I, II, and V (Lewis, 1990; Smiley *et al.*, 1997). Cholinergic axons within the cerebral cortex of human brain are studded with numerous *en passant* swellings that serial EM reveals as primarily asymmetric type synapses. Close appositions of cholinergic synaptic profiles in cortex (Mesulam, 1999, 2004; Smiley *et al.*, 1997) as well as the prevalence of cholinergic marker-positive swellings in the vicinity of pyramidal and nonpyramidal neurons in PFC and hippocampus is consistent with proposed modulatory effects of ACh on both excitatory and inhibitory cortical circuits (Mansvelder *et al.*, 2006). Likewise, evidence has accrued that the release of ACh per se is likely subject to cholinergic, as well as dopaminergic, synaptic tuning in both PFC and hippocampus (DeBoer *et al.*, 1996; Moore *et al.*, 1999).

III. Physiology of ACh Circuits and Signaling in Brain Regions Implicated in Schizophrenia Pathology

A. ACh RECEPTORS IN THE CNS

When clinicians and patients contemplate the “anticholinergic” side effects of various drugs, they often focus on the diverse and distressing array of peripheral autonomic cholinergic actions, including alterations in gastrointestinal function, nausea, and changes in appetite. In fact, the binding sites with which cholinergic drugs interact in the CNS are just as diverse as those in the periphery and often more accessible than expected, despite the blood–brain barrier. Any careful deliberation on ACh-binding sites in the CNS must include ACh-degradative, synthetic, and transporter proteins, as well as the multimembered muscarinic and nicotinic receptor subtypes (for reviews see Calabresi *et al.*, 2000; Cobb and Davies, 2005; Gotti and Clementi, 2004; Mansvelder *et al.*, 2006; Newhouse *et al.*, 2004; Sarter *et al.*, 2005). Pharmacological agents originally identified for their

activity as AChE inhibitors (such as physostigmine and galantamine) are now known to act as partial agonists or antagonists of specific subtypes of CNS nicotinic AChRs (nAChRs). The oldest and most established “antimuscarinic” agent, atropine, blocks multiple classes of nAChRs at submicromolar concentrations—well within the clinically relevant and experimentally typical range of doses (Zwart *et al.*, 1999). Carbamylcholine is more than a muscarinic agonist—it gates deliciously long openings of nAChRs. Finally, the activation of different types of pre-synaptic nicotinic *and* muscarinic receptors can facilitate or depress the release of ACh itself (see below).

Awareness of the complexities in the number and pharmacodiversity of ACh-binding partners in the CNS is essential to evaluating the past and present literature on ACh circuits and signaling. Humbling though this may be, we are actually well positioned to do so: the last 20 years have yielded impressive advances in understanding the differential regulation, expression, targeting, and function of the many muscarinic (at least 5 genes, so far) and nicotinic (11 subunit genes) receptors (see below). With this knowledge in hand, we need to reassess the effects of the pharmaceuticals we have and work toward the development of agents that more selectively manipulate the synthesis, release, and binding(s) of ACh.

A quick primer then, on the most important of ACh-binding sites, from information largely extracted from the following reviews: Calabresi *et al.* (2000); Gotti and Clementi (2004); Laviolette and van der Kooy (2004); MacDermott *et al.* (1999); Mansvelder *et al.* (2006); Sarter and Parikh (2005); Smythies (2005); Wonnacot *et al.* (2005).

1. *Choline acetyltransferase (ChAT)*: This enzyme is responsible for ACh synthesis. The regulation of ChAT gene expression in the CNS is thought to be coordinated with that of vAChT, by virtue of a common “cholinergic locus” promoter. However, the distribution of these two proteins between somatodendritic and axonal domains may be regulated independently.

2. *ACh esterase (AChE)*: This binds ACh with micromolar affinity and is considered the principal degradative activity for ACh. AChE is one of the fastest turnover rate enzymes identified and is located primarily at intraneuronal and extracellular sites. Despite its preeminence as “the AChE,” recent work deleting AChE-encoding genes revealed that butyrylcholinesterase (BuChE) activity, which is associated with glial cells rather than neurons, can maintain grossly normal ACh balance. So BuChE is another ACh-binding partner to bear in mind.

3. *The vesicular ACh transporter (vAChT)*: This binds ACh with submicromolar affinity and translocates it into vesicular compartments within cholinergic neurons.

4. *Muscarinic (metabotropic) AChRs*: At least five genes are identified to date (M1–M5); M1, M2, and M4 subtypes predominate in the CNS. These ACh-binding proteins are coupled to a variety of G-proteins resulting in the

activation or inhibition of an even wider variety of enzymatic and ion channel targets. Note that in the CNS, only a subset of the muscarinic-binding sites are postsynaptic; other subtypes of muscarinic receptors are targeted to axonal/presynaptic sites where they modulate the release of glutamate, dopamine, and ACh, among other key players.

5. *Nicotinic (ionotropic) AChRs*: Twelve subunit genes ($\alpha 2$ – $\alpha 10$; $\beta 2$ – $\beta 4$) encode a group of proteins that are faintly related to—and pharmacologically very distinct from—the renowned muscle-type nicotinic receptors. Drugs that interact with subtypes of neuronal nicotinic receptors (e.g., nicotine, hexamethonium) barely touch the muscle receptor and vice versa. Also important to note is that in the CNS, nAChRs, just like muscarinic receptors, are targeted to *pre*- as well as postsynaptic locations. In fact, the role of presynaptic nicotinic receptors as modulators of dopamine, glutamate, GABA, serotonin, and ACh release is so prevalent in the CNS that their contribution as postsynaptic receptors is often overlooked (but see Frazier *et al.*, 1998; Jones and Yakel, 1997)!

In sum, a circumspect evaluation of how the dysregulation of cholinergic circuits may be involved in the pathophysiology requires recognition that ACh targets are many, perhaps not as pharmacologically distinct as previously considered and at pre-, post-, and perisynaptic locations. Viewing the function of cholinergic circuits as the result of the summation of actions of ACh at all of its receptors, although initially daunting, may resolve some apparent conflicts in the literature and guide the way to new therapeutic approaches (for reviews see Calabresi *et al.*, 2000; Gotti and Clementi, 2004; Laviolette and van der Kooy, 2004; MacDermott *et al.*, 1999; Mansvelder *et al.*, 2006; Sarter and Parikh, 2005; Sarter *et al.*, 2005; Smythies, 2005; Wonnacot *et al.*, 2005).

B. PHYSIOLOGY OF ACh CIRCUITS IN STRIATUM

The striatum is established stomping grounds for fans of central cholinergic circuits and ACh signaling. Although the numbers of cholinergic neurons in the striatum are small, they are the foremost, if not the exclusive, source of the high-pack cholinergic inputs in mammalian striatum. As discussed above, striatal cholinergic neurons are characterized by their large size, aspiny appearance, and tonic activation profile (hence the names ASpN and TANS neurons; Fig. 2). Changes in the activity profile of striatal TANS, referred to as “pauses,” are thought to arise in part from the slowing of autonomous pacemaker activity and in part to local changes in dopamine, glutamate, and GABA signaling (Cragg, 2006; Maurice *et al.*, 2004; Wang *et al.*, 2006). The association between changes in striatal cholinergic “tone” and salience/reward prediction has continued to stoke the fire of physiologists’ interests in the workings of striatal ACh circuits

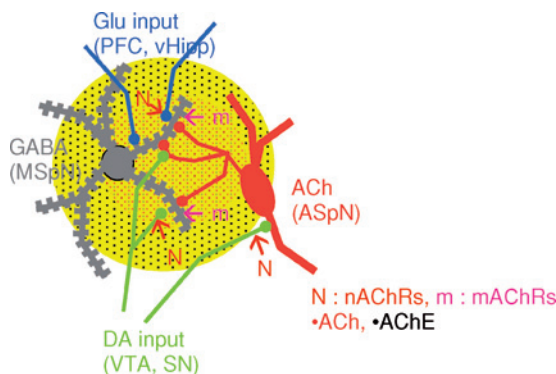


FIG. 2. Schematic diagram of an aspiny cholinergic neuron (ASpN) and its projections to convergent sites of glutamatergic and dopaminergic input on striatal GABAergic medium spiny projection neurons (MSpN). Changes in local [ACh], from pauses in the firing of these TANS, modulate the net output of the striatum by interactions with ACh receptors and binding sites in pre-, post-, and perisynaptic compartments.

(see Cragg, 2006 for review; Apicella, 2002; Maurice *et al.*, 2004; Wang *et al.*, 2006 for recent highlights).

Perhaps the best studied, although still mechanistically mysterious, role of cholinergic circuits in striatum is in their reciprocal interactions with dopaminergic inputs from the VTA and SN. Recent studies add new dimensions to prior evidence that ACh acts as a key regulator of striatal output by influencing the activity of GABAergic medium spiny neurons (MSpNs, Fig. 2). The newest twist is that ACh is likely to exert its modulatory control on striatal activity through interaction with *both* pre- and postsynaptic, nicotinic *and* muscarinic receptors (Fig. 2). Presynaptic nicotinic receptors have long been implicated in the regulation of striatal dopamine release, with reports identifying some of the nAChR subtypes involved in aspects of nicotine addiction in staggering detail (for review see Wonnacott *et al.*, 2005; Tapper *et al.*, 2004 for recent highlights). The vague term “regulation” was intentionally employed because one of the points of controversy has been whether dopamine release in striatum is enhanced or depressed by nicotine. It turns out that the answer may be both, depending on the frequency of firing of the dopamine neurons (see Cragg, 2006 for review; Partridge *et al.*, 2002; Rice and Cragg, 2004; Zhang and Sulzer, 2004; Zhou *et al.*, 2001). The effects of dopamine receptor agonists on modulating the release of ACh in striatum (as well as in PFC and in hippocampus, see below) are also well established (DeBoer *et al.*, 1996). But new results reveal that depending on the type and location of the dopaminergic and muscarinic receptors activated, the net effect may be to stably enhance or depress the activity of the GABAergic

MSPNs (Cragg, 2006; Wang *et al.*, 2006; Wilson, 2006;). Indeed, the potential for mutual tuning of TANS and DANS seems more than flexible enough to account for the differences in valence and timing of all of the synaptic changes observed in striatum (Calabresi *et al.*, 2000; Cragg, 2006; Maurice *et al.*, 2004; Wang *et al.*, 2006).

C. PHYSIOLOGY OF ACh CIRCUITS IN PFC AND HIPPOCAMPUS

The role of cholinergic signaling in aspects of memory and cognition are typically attributed to the broad spectrum of effects that ACh elicits in altering the excitability of prefrontal cortical and hippocampal circuits (for reviews see Albuquerque *et al.*, 1996; Buzsaki, 2002; Levin *et al.*, 2006; Mansvelder *et al.*, 2006; Newhouse *et al.*, 2004; Picciotto, 2003; Role and Berg, 1996; Sacco *et al.*, 2004; Sarter *et al.*, 2005; Smythies, 2005; Wonnacott *et al.*, 2005). Analysis of the pros and cons of the many theories on *how* ACh actually does what it does to regulate synaptic efficacy in these regions, as with striatum, is best served by considering the potential interaction of ACh with each of its five major sets of binding partners: ChAT, AChE, VAcHT, muscarinic, and nAChRs (see discussion above).

ACh transmission in cortex and hippocampus likely involves both localized release and tonic or “volume” transmission (Cobb and Davies, 2005; Vizi and Kiss, 1998). Activation of presynaptic ACh receptors modulates the release of glutamate, ACh, and dopamine in PFC and hippocampus (Colgin *et al.*, 2003; Laplante *et al.*, 2004; Lucas-Meunier *et al.*, 2003), enhancing or depressing transmission depending on the flavor(s) of AChRs expressed (Mansvelder *et al.*, 2006; Sarter and Parikh, 2005; Wonnacott *et al.*, 2005). Postsynaptic mAChRs and nAChRs have also been implicated in the modulation of PFC and hippocampal circuits (Cobb and Davies, 2005; Frazier *et al.*, 1998; Ji *et al.*, 2001; Jones and Yakel, 1997).

Perhaps the most important (albeit still controversial in detail) role of ACh circuits in cortex and in hippocampus is in the regulation of theta rhythm oscillatory activity (Buzsaki, 2002; Calabresi *et al.*, 2000; Cobb and Davies, 2005; Hasselmo, 2005; Lee *et al.*, 2005). Theta-frequency band oscillations constitute a prominent network pattern in all mammals, including humans. Theta activity has been proposed to underlie everything from temporal cooperativity of cortical and subcortical networks to coordinate modifications of synaptic connections within cortex and hippocampus per se (Buzsaki, 2002; Calabresi *et al.*, 2000; Cobb and Davies, 2005; Hasselmo, 2005). In any case, there is no doubt that cholinergic circuits, specifically the septal cholinergic projections, play an essential role in theta oscillations, as selective lesion of the ACh synthesizing neurons in the medial septal/diagonal band nuclei abolishes hippocampal theta.

Discrepancies arise in interpretation of studies that manipulate ACh by different pharmacological means—that is, M1-AChR versus AChE antagonists—which imply that other types of muscarinic and/or nicotinic AChRs may be involved (Buzsaki, 2002; Ji *et al.*, 2001).

IV. Developmental and Genetic Deficits in Schizophrenia That May Influence Function and Assembly of Cholinergic Systems

Schizophrenia is widely viewed as a neurodevelopmental disease, resulting from a combination of environmental challenges acting on susceptible genotypes. Significant progress has been made at identifying both relevant environmental and genetic risk factors (Bresnahan *et al.*, 2005; Harrison and Weinberger, 2005), although we have yet to make progress at understanding how these factors interact to dysregulate relevant circuits in the developing brain. The vertebrate forebrain contains relatively few cholinergic neurons, yet this population exerts widespread modulatory control over essentially all striatal–cortical networks.

Experimental studies have shown that during development, the cholinergic system is especially sensitive to environmental insults (e.g., ethanol, lead, organophosphates, tobacco smoke; Eriksson *et al.*, 2001; Reddy *et al.*, 2003; Robinson, 2002; Thomas *et al.*, 2000). These and other insults would certainly have the potential to interact with genetic vulnerabilities affecting brain development to produce deficits which could contribute to disease states.

A. DEVELOPMENT OF CHOLINERGIC SYSTEMS

Forebrain cholinergic neurons arise early in telencephalic development (~E10 in mouse which approximately corresponds to gestational day 40 in humans; Clancy *et al.*, 2001) in the medial ganglionic eminence, a ventricular/subventricular neurogenic zone that appears as a thickening along the ventral/medial wall of the ventricle (Brady *et al.*, 1989; Furusho, 2006; Marin *et al.*, 2000; Olsson *et al.*, 1998; Semba *et al.*, 1988). Presumptive forebrain projection cholinergic neurons migrate from the MGE (and possibly from the anterior entopeduncular/preoptic area) radially to take up locations in basal forebrain nuclei (medial septum, magnocellular nucleus, diagonal band of Broca), whereas the striatal cholinergic interneurons migrate tangentially from the MGE, occupying dispersed sites throughout the striatal plate. Subsequent to this early birth and migration from the MGE, 1–2 weeks pass before these neurons undergo maturation into cholinergic neurons (Aznavour *et al.*, 2005; Berger-Sweeney, 2003; Mechawar and Descarries, 2001). This delay allows time for other populations of predominantly GABAergic neurons to emerge from the MGE and LGE

and occupy their appropriate positions throughout the striatum and cortex for radial population of the neocortex with pyramidal cells and for the proper targeting of axons from the dorsal thalamus to project through the striatal region to innervate cortical structures (Flames *et al.*, 2004; Lopez-Bendito *et al.*, 2006; van Vulpén and van der Kooy, 1998). Some investigators propose that during this period the presumptive cholinergic neurons provide instructive signals that guide the targeting and differentiation of later born striatal populations (Berger-Sweeney, 2003; Hohmann, 2003; Hohmann and Berger-Sweeney, 1998).

During the first two postnatal weeks, the cholinergic interneurons elaborate robust networks of axons locally within the striatum, whereas the forebrain cholinergic neurons elaborate a wide array of axonal projections that target all neocortical regions (prefrontal, sensory, and motor) and the hippocampal formation. As a result these two relatively small populations of cholinergic neurons maximize their ability to interact with striatal-cortical networks. It is likely that the delay between the migration of newly formed cholinergic neurons from neurogenic zones and the elaboration of axonal projections plays a critical role in properly controlling the final wiring of the forebrain cholinergic system.

A number of recent studies using molecular genetic approaches in mice are beginning to clarify the developmental processes that determine the specification of forebrain cholinergic neurons. In particular, the basics of a cholinergic transcription factor code are emerging. A variety of experimental approaches has demonstrated that expression of several transcription factors is important (Mash 1, Olig2, Lmx7, Lmx8) or essential (Nkx2.1) for generating forebrain cholinergic neurons (Bachy and Retaux, 2006; Furusho *et al.*, 2006; Marin *et al.*, 2000; Mori *et al.*, 2004; Zhao *et al.*, 2003). Thus, the combined expression of these factors, and their target genes, probably accounts for much of the intrinsic identity of the cholinergic phenotype. However, these studies have not yet distinguished between factors that determine how a newborn cholinergic neuron migrates from the MGE (radially into basal septal regions or tangentially into the striatum), or whether an individual cholinergic neuron will elaborate a spatially restricted axonal network, as is the case of the striatal interneurons, or a broadly targeted set of cortical projections.

B. POTENTIAL ROLE OF NEUREGULIN 1

Neuregulin 1-ErbB signaling plays multiple critical roles in proper development of the neocortex, guiding both the tangential migration of MGE-derived GABAergic interneurons (Flames *et al.*, 2004) and proper navigation of axonal projections from the dorsal thalamus into the cortex (Lopez-Bendito *et al.*, 2006). Whether neuregulin also guides the migration and/or axon projections of forebrain cholinergic neurons is not known. We have seen apparent decreases

in the numbers of specific populations of forebrain interneurons in adult mice that are heterozygous for an isoform specific, targeted mutation in the neuregulin 1 gene (Wolpowitz *et al.*, 2000; Johnson, Talmage, and Role, Unpublished data).

Maturation and maintenance of cholinergic neurons depends on a number of extracellular signaling molecules. Of particular relevance to our discussion of cholinergic signaling and schizophrenia are nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), and cortical steroids. NGF, BDNF, and glucocorticoids regulate the expression of ChAT, enhance the connectivity of, and promote the survival of cholinergic neurons (Fagan *et al.*, 1997; Grosse *et al.*, 2005; Guijarro *et al.*, 2006; Johnston *et al.*, 1987; Mobley *et al.*, 1986; Phillips *et al.*, 2004; Sofroniew *et al.*, 2001; Takahashi, 1998; Takahashi and Goh, 1998; Ward and Hagg, 2000). Reports have demonstrated altered regulation of NGF (Parikh *et al.*, 2003), BDNF (Weickert *et al.*, 2003, 2005), and the HPA axis (Corcoran *et al.*, 2001, 2003) in schizophrenics. Whether these changes alter corticostriatal cholinergic tone remains to be seen.

Beyond general effects on synaptic structures, there is no clear evidence linking the products of identified schizophrenia susceptibility genes with the cholinergic system, with the notable exception of the neuregulin 1 gene. Neuregulin 1 has been linked to schizophrenia in multiple populations, and disease-associated changes in the relative expression of different neuregulin 1 isoforms is seen in the DFPLC and hippocampus. Neuregulin 1 isoforms play important roles in neurodevelopment, in particular in the patterning of the neocortex (Flames *et al.*, 2004; Lopez-Bendito *et al.*, 2006). At present, these latter roles for neuregulin 1 have focused on tangential migration of cortical interneurons and axonal projections from the dorsal thalamus to the neocortex. Given the relative spatial and temporal parallels between these events (GABAergic interneurons originate in the MGE during an overlapping time frame with the striatal cholinergic interneurons) and the reported decreases in the numbers of ventral striatal cholinergic interneurons in postmortem tissue from schizophrenics, it is important that studies of the role of neuregulin 1 in forebrain development be extended to the cholinergic system as well.

A more direct association between neuregulin 1 and cholinergic signaling exists at the level of the expression of the nAChRs. Two families of neuregulin 1 isoforms were identified originally by virtue of their ability to regulate the expression of nAChRs at peripheral synapses (Falls *et al.*, 1993; Yang *et al.*, 1998). Subsequently, a number of investigators have demonstrated that neuregulin also can increase the synaptic expression of $\alpha 7$ -containing nAChRs at central synapses (Kawai *et al.*, 2002; Liu *et al.*, 2001), and our laboratories have extended this story by demonstrating that neuregulin 1 signaling also regulates presynaptic expression and targeting of the $\alpha 7$ nAChRs (Role and Talmage, unpublished). These latter studies are particularly intriguing in light of the

well-documented deficits in $\alpha 7$ nAChRs in schizophrenics, the association of this deficit with defects in P50 measures of auditory gating in schizophrenics and their first degree relatives, and the association of $\alpha 7$ subunit gene promoter polymorphisms with these deficits (see chapter by Martin and Freedman, this volume; Leonard *et al.*, 1996, 2002).

V. Clinical and Preclinical Evidence for Deficits in Components of Brain Cholinergic Systems in Schizophrenia

There are numerous examples of deficits in components of brain cholinergic systems that have been linked with schizophrenia. They range from abnormal expression of receptors of various subtypes through decreases in cholinergic neurons in key areas.

A. DEFICITS IN COMPONENTS OF MUSCARINIC CHOLINERGIC TRANSMISSION

Several studies give evidence of alteration of muscarinic ACh receptors in the brains of schizophrenics. The majority of evidence in this realm points to decrements in binding suggestive of a decrease in available m1 muscarinic-binding sites in prefrontal cortex and hippocampus.

Using [(123)I]IQNB SPECT, one group found decreased muscarinic receptor availability in unmedicated patients with schizophrenia as compared to controls in a variety of cortical and subcortical brain regions (Raedler *et al.*, 2003). Another group used GTP- γ S binding to distinguish M2 and M3 muscarinic receptors and found no change in postmortem cortex from patients with schizophrenia as compared to controls, while finding a reduction in M1 (Scarr *et al.*, 2006). Other groups have found decreased pirenzapine binding in the hippocampus in brains of patients with schizophrenia (Crook *et al.*, 2000) and decreased M1 receptor mRNA in dorsolateral prefrontal cortex (Dean *et al.*, 2002) but not caudate nucleus (Dean *et al.*, 2000).

At the level of genetic findings, there is evidence for linkage of an M1 polymorphism to decreased performance on the Wisconsin Card Sort Test (Liao *et al.*, 2003). In addition, there is evidence that an M5 polymorphism confers susceptibility to schizophrenia. Interestingly, the M5 variant seems to confer risk only in combination with a nicotinic $\alpha 7$ polymorphism (De Luca *et al.*, 2004).

Finally, it has been reported that circulating antibodies to the M1 receptor can be identified in the serum of some schizophrenic patients. These antibodies

displace tritiated pirenzapine and have agonist-like properties at the M1 receptor in *in vitro* assays (Borda *et al.*, 2002).

B. DEFICITS IN COMPONENTS OF NICOTINIC CHOLINERGIC TRANSMISSION

There is also ample evidence of alterations of nAChRs in the brains of patients with schizophrenia. The most notable set of findings pertains to decrements in $\alpha 7$ -containing nAChRs and their function in sensory gating. These studies are dealt with in detail in the subsequent chapter by Martin and Freedman, this volume.

Apart from the well-defined case of $\alpha 7$ -containing nicotinic receptors, more global changes in nicotinic cholinergic receptors have been observed as well. One group observed decreased high-affinity nicotine and epibatidine binding in post-mortem brains from patients with schizophrenia as compared to controls (Breese *et al.*, 2000). The differences were seen in hippocampus, cortex, and caudate in the subgroup of patients versus controls who smoked. The same group reported increases in receptor sites in nicotine and haldol treated rats (Breese *et al.*, 2000). However, another group found elevation of nicotine binding in the striatum of patients with schizophrenia (Court *et al.*, 2000) and minimal changes in α -bungarotoxin binding in the thalamus (Court *et al.*, 1999).

C. DEFICITS IN CHOLINERGIC INNERVATION

Besides decrements in receptors for ACh, one group has observed a reduction in numbers of cholinergic interneurons in the ventral striatum (Holt *et al.*, 1999, 2005), but not in other striatal regions. However, cortical cholinesterase and ChAT activity is not reduced in the brains of patients with schizophrenia (Haroutunian *et al.*, 1994).

D. SUMMARY

There are numerous findings of abnormalities in the expression or distribution of many components of cholinergic systems in the brains of patients with schizophrenia, the bulk of which would be expected to lead to decrements in cholinergic neurotransmission. It is unclear whether all of these are primary deficits or in some cases downstream effects of other lesions. At least some cholinergic deficits may have to interact with deficits in other systems in order to confer disease vulnerability. However, despite these uncertainties, the preponderance of evidence points toward possible roles for abnormalities in cholinergic systems participating in the pathophysiology of schizophrenia.

VI. Evidence for Cholinergic Contributions to Schizophrenia Pathophysiology from Clinical and Preclinical Psychopharmacology

Correlation of the efficacy of medications used to treat target symptoms of schizophrenia with effects of those medications on cholinergic systems has been another way in which investigators have attempted to infer potential roles for cholinergic systems in the pathophysiology of schizophrenia. The complex receptor-binding properties of antipsychotic medications and complex relationships of target symptoms and medication side effects have made it difficult to make such inferences in a clear and convincing way. That said, there seems to be ample evidence that well-chosen cholinergic targets have potential to be therapeutic targets.

A. ACh RELEASE, MUSCARINIC BLOCKADE, PARTIAL AGONISTS, AND “ATYPICALITY”

Preclinical studies have demonstrated that many antipsychotic medications cause the release of ACh in cortical (Ichikawa *et al.*, 2002; Li *et al.*, 2005) and hippocampal regions (Johnson *et al.*, 2005; Shirazi-Southall *et al.*, 2002). While a broad range of antipsychotic medications have been shown to release cortical and hippocampal ACh, olanzapine and clozapine have been shown to be especially potent in this regard. Interestingly, these are the two antipsychotic medications which seem to have greater efficacy against a broader range of symptoms than other antipsychotic medications (Kane *et al.*, 1988, 2001; Lieberman *et al.*, 2003, 2005).

To some extent, antipsychotic-induced release of cortical ACh correlates with M2 binding affinity; antipsychotic medications with less M2 binding tend to be less potent inducers of cortical ACh release (Johnson *et al.*, 2005). Observations such as these combined with the observation that both clozapine and olanzapine bind muscarinic receptors with high affinity have led to the idea that anticholinergic properties of antipsychotic medications might be responsible for ACh release, and further might be correlated with “atypicality.” This scenario is plausible in that muscarinic receptors can serve as inhibitory presynaptic autoreceptors, providing a potential mechanism by which their blockade could augment ACh release.

However, olanzapine and clozapine may be only weakly antimuscarinic at the level of clinical symptoms, with fewer anticholinergic side effects than expected based on their potent *in vitro* displacement of muscarinic ligands (Bymaster *et al.*, 1996). This discrepancy is likely due both to subtype selectivity at muscarinic sites, and to partial agonist activity at M4 and M2 receptors (Bymaster *et al.*, 2003; Michal *et al.*, 1999).

Interestingly, clozapine has cognition impairing properties in mice, but the effect is complex, in that clozapine reduces scopolamine-induced impairment while its direct effects on cognition are reversed by cholinesterase inhibitors. This pattern of reduction of antagonist effects and reversal by treatments that increase the endogenous ligand is highly suggestive of partial agonist effects (Ninan and Kulkarni, 1996) or of interaction of ACh with other types of muscarinic and/or nicotinic receptors (see above). Of note, at least two other compounds, xanomeline (reviewed in Mirza *et al.*, 2003) and PTAC (Bymaster *et al.*, 1999), show antipsychotic-like activity in animal models and are also partial agonists at M4 and M2 receptors.

A further wrinkle in this data is introduced by the fact that both clozapine and olanzapine still release cortical and hippocampal ACh in mice lacking M2 and M4 receptors (Bymaster *et al.*, 2003). As such, neither blockade nor partial agonism at these sites appears to be required for ACh release. In view of recent studies implicating both presynaptic nicotinic and dopamine receptors in the modulation of ACh release, the potential contribution of these pathways to the effects of clozapine and olanzapine should, perhaps, be considered.

B. BOTH PROCHOLINERGIC AND ANTICHOLINERGIC COMPOUNDS MAY AMELIORATE OR WORSEN DIFFERENT SYMPTOM DOMAINS

Adding further to confusion about the role of cholinergic transmission in schizophrenia, muscarinic agonists and antagonists exert opposing effects on various schizophrenia symptom clusters. There has been extensive interest in this area because of the use of medications with “anticholinergic” properties as antipsychotics (e.g., chlorpromazine, perphenazine) and because of the use of medications such as benztropine and biperiden to counter the parkinsonian side effects of traditional neuroleptic antipsychotics. For some time, the prevailing opinion seems to have been that anticholinergic effects were fairly neutral in relation to symptoms of schizophrenia, but closer examination has revealed a more complicated picture.

Several studies showed some increase in positive symptoms (hallucinations and delusions) when anticholinergic medications were added to neuroleptics. In a placebo-controlled trial, procyclidine or placebo was added to flupenthixol in a group of 36 patients, with the subsequent finding that those receiving procyclidine had more positive symptoms than those receiving placebo (Johnstone *et al.*, 1983). Another study compared symptoms in 47 patients receiving neuroleptics during periods with and without treatment with benztropine or trihexyphenidyl. Again, the anticholinergic medication was associated with an increase in positive symptoms (Singh *et al.*, 1987). A study showed that biperiden increased positive symptoms in a small group of schizophrenic patients when added during a medication free

period, strengthening the case that the symptomatic worsening is a direct anticholinergic effect rather than one that depends on interaction with the effects of other medications (Tandon *et al.*, 1991).

Anticholinergic compounds such as benztropine have been shown to cause memory impairment (Brebion *et al.*, 2004; Tune *et al.*, 1982). In one small study, higher serum anticholinergic levels correlated with worsened recall but improved reaction time (Strauss *et al.*, 1990). It is possible that improvements in reaction time represent an anti-parkinsonian effect as all patients in the study were taking antipsychotic medications. In another study, single injections of benztropine or glycopyrrolate impaired free recall (McEvoy and Freter, 1989). Overall, it appears likely that anticholinergic medications could worsen some cognitive deficits in schizophrenia.

At the same time, there is a small body of evidence showing that anticholinergic may decrease some negative symptoms in schizophrenia. This provides an instance in which the domain of cognitive symptoms and the domain of negative symptoms can be separated from one another in part based on how they are affected by pharmacological treatment.

In one study, treatment of a small number of patients with trihexyphenidyl resulted in decreases in affective flattening, avolition, and anhedonia/asociality (Tandon *et al.*, 1992). In another study by the same group, biperiden also reduced negative symptoms (while increasing positive symptoms; Tandon *et al.*, 1991). Abuse of trihexyphenidyl and benztropine has at times been cited as “self-medication” of negative symptoms by patients with schizophrenia, and indeed in one study those who abused these medications tended to have higher Brief Psychiatric Rating Scale (BPRS) scores and more negative symptoms than those who did not abuse anticholinergic medications (Zemishlany *et al.*, 1996).

One might speculate that actions of anticholinergic medications against negative symptoms reflect activity in psychomotor circuits that are parallel in some way to the motor circuits in which anticholinergic medications oppose the parkinsonian actions of neuroleptics.

Conversely, muscarinic *agonists* have been proposed to have antipsychotic activity and may have potential effects against positive symptoms with particular attention paid to the compound xanomeline (reviewed in Bymaster *et al.*, 2002, 2003).

Recent findings on the function of muscarinic receptors in the striatum may shed some light on these apparent paradoxes. The recent paper by Wang and coworkers (also discussed in Section III.B of this chapter) shows that D2 dopamine receptors serve to inactivate striatal cholinergic interneurons which signal to M1 muscarinic receptors on MSpNs. In essence, muscarinic stimulation and blockade of dopamine are functional equivalents in this circuit. The muscarinic receptors reduce intracellular calcium in the MSpNs, which in turn reduces the production of endocannabinoids, reducing depolarization induced suppression

and long-term depression of neurotransmission. In summary, cholinergic activation in this circuit results in more activity of MSpNs in the indirect pathway—and subsequently less release of inhibition of the corticostriatal pathways that may convert drives and feelings into actions and perceptions. One might predict that this action would reduce positive symptoms in schizophrenia in a manner analogous to that of D2 blockade by neuroleptics, but one might also predict that in some components of these pathways, the same set of actions could result in an increase in negative symptoms (Wang *et al.*, 2006).

While far from clear, the body of evidence on effects of muscarinic agonists and antagonists in schizophrenia, and on the cholinergic binding properties of antipsychotic medications seems to point to a complex role for cholinergic transmission in different domains of psychopathology with loci of action in the brain likely to depend on the specific symptom domain examined. A challenge to neuropsychopharmacologists will be to find ways to balance and dissociate beneficial and harmful effects of blocking and enhancing cholinergic transmission at muscarinic receptors.

C. NICOTINE AMELIORATES A WIDE RANGE OF DEFICITS SEEN IN SCHIZOPHRENIA

A large part of the pharmacological evidence pointing to potential cholinergic roles in schizophrenia pathophysiology concerns salutary effects of nicotine in patients with schizophrenia. Nicotine affects a wide range of symptom domains and neuropsychological findings. We will give a broad sampling of the data here, and refer the reader to the subsequent chapter by Martin and Freedman, this volume, for an in-depth review of data on nicotinic receptors and the processing of sensory information in schizophrenia.

Nicotine has been shown to improve abnormalities in smooth pursuit eye movement and saccades during visual tracking (Avila *et al.*, 2003; Depatie *et al.*, 2002; Larrison-Faucher *et al.*, 2004; Sherr *et al.*, 2002). The improvement in saccades was independent of the smoking status of the patients, thus addressing the possibility that nicotine's effect resulted directly from the elevated incidence of smoking by people with schizophrenia. Nicotine also improved sustained attention in these visual tasks (Avila *et al.*, 2003; Depatie *et al.*, 2002). The effects of nicotine on performance of visual tracking and tasks of visual attention may involve hippocampus and cingulate gyrus (Levin *et al.*, 2006; Newhouse *et al.*, 2004; Tanabe *et al.*, 2006).

In another domain, nicotine improved performance of tasks involving working memory in schizophrenic subjects, enhancing task-related activation of thalamus and anterior cingulate cortex as seen on fMRI (Jacobsen *et al.*, 2004).

Nicotine has also been shown to reverse haloperidol-induced impairments in reaction time and working memory (Levin *et al.*, 1996). Nicotine nasal

spray improved delayed recognition and spatial working memory in schizophrenic patients (Myers *et al.*, 2004; Smith *et al.*, 2006). Interestingly, nicotine may actually impair working memory in otherwise healthy smokers (Park *et al.*, 2000) suggesting an inherent difference in nicotine responses in the brains of persons with schizophrenia (discussed in Mansvelder *et al.*, 2006; Newhouse *et al.*, 2004).

While numerous studies show beneficial effects of administered nicotine in schizophrenia, the reverse is not true, that is to say, nicotine withdrawal did not increase positive symptoms in a group of patients with schizophrenia who quit smoking. Increases in negative symptoms were modest and transient (Dalack *et al.*, 1999). In considering these results, it may be important to take into account that the frequent high peaks of nicotine delivered by smoking may not be of sustained therapeutic benefit as compared to other systems of delivery.

Finally, the fairly extensive data on sensory processing and nicotine is perhaps the best pharmacological evidence for a role of cholinergic systems in schizophrenia. It is made stronger by the linkage of a polymorphism in the $\alpha 7$ nicotinic receptor subunit gene to sensory gating deficits in patients with schizophrenia and their relatives (Freedman *et al.*, 2003; see chapter by Martin and Freedman, this volume). This topic is reviewed in detail in the subsequent chapter by Martin and Freedman, this volume. Overall, and in contrast to the data on muscarinic AChRs, evidence to date strongly supports the notion that treatments that interact with nAChRs have almost uniformly ameliorative effects on symptoms of schizophrenia.

D. DESPITE CLEAR EFFECTS OF OTHER CHOLINERGIC COMPOUNDS, CHOLINESTERASE INHIBITORS ARE NOT PROVEN ADJUNCTS IN THE TREATMENT OF SCHIZOPHRENIA

Further pharmacological evidence concerning cholinergic participation in the pathophysiology of schizophrenia comes from studies in which patients were treated with medications from the family of AChE inhibitors which were initially developed to treat Alzheimer's disease (Coyle and Kershaw, 2001; Crismon, 1994; Dooley and Lamb, 2000; Jann, 2000). The effects of these medications in patients with schizophrenia are equivocal at best.

There are several case reports describing improvement in negative and cognitive symptoms of individual patients with cholinesterase inhibitors (Rosse and Deutsch, 2002, using galantamine). There are some positive open label trials of rivastigmine (Lenzi *et al.*, 2003; Mendelsohn *et al.*, 2004) in which patients show improvements on standard-rating scales such as Positive and Negative Symptom

Scale (PAANS) or BPRS. The study by Lenzi *et al.* (2003) was focused on quality of life measures. The study by Mendelsohn *et al.* (2004) was limited to patients with comorbid dementia, so it may be difficult to generalize the result to the schizophrenic population as a whole.

Some positive findings involving cholinesterase inhibitors were in studies that focused on a specific neurocognitive endophenotype rather than on clinical outcome as a whole. One group found that donepezil normalized fMRI findings on a verbal fluency task (Nahas *et al.*, 2003) and another group found that rivastigmine improved performance on a sustained attention task (Aasen *et al.*, 2005).

On the negative side, several reports of placebo-controlled, double-blind crossover trials of donepezil show no efficacy against symptoms of schizophrenia. These studies were all done using donepezil as a neuroleptic augmentation treatment. Some (Mazeh *et al.*, 2006; Stryjer *et al.*, 2003) were in elderly patients or patients with known comorbid dementia. Others were in a general population of stable schizophrenic patients (Friedman *et al.*, 2002; Stryjer *et al.*, 2004; Tugal *et al.*, 2004). No effects were seen on positive, negative, or cognitive symptoms. All of the studies were fairly small, and in some cases there is concern about potential confounding effects of concurrent nicotine use by patients.

Overall, at this time there is little evidence to suggest significant benefits of cholinesterase inhibitors in schizophrenia, especially to patients who do not suffer from comorbid dementia. In some respects, given what we have outlined about the complexity of cholinergic systems, it is not surprising that a class of medications which brings about global increases in ACh levels would have modest effects; in many brain locations, presynaptic inhibition of release may compensate for decreased degradation when ACh levels rise. Thus, given the evidence of effects of treatments targeting nicotinic and muscarinic receptors, it would seem unwarranted to view the modest effects of cholinesterase inhibitors as evidence against participation of cholinergic systems in schizophrenia pathophysiology.

VII. Conclusions

Proper function of cholinergic systems in the brain is essential for a variety of neurocognitive tasks that are impaired in schizophrenia including attention, volition, working memory, assignment of salience, and the processing of sensory information.

While it is unlikely that cholinergic deficits alone account for any particular symptom domain in schizophrenia, there is ample evidence that schizophrenia is

associated with genetic changes and brain abnormalities that can influence both the development and function of cholinergic systems, and that interaction of cholinergic deficits with deficits in other systems has the potential to produce disease symptoms.

Perhaps the clearest case of a cholinergic deficit is that of abnormality in the control of $\alpha 7$ nicotinic receptor expression conferring deficits in sensory gating and vulnerability to schizophrenia (cf chapter by Martin and Freedman, this volume). However, it is likely that nicotine has other important sites of action relating to schizophrenia, and that muscarinic effects on corticostriatal and other circuits are of independent import.

Inferences made from clinical and preclinical psychopharmacological data about the performance of cholinergic systems in schizophrenia are fraught with difficulty and do not point to a simple dysregulation of cholinergic transmission at a single brain location. Rather, there are numerous points where dysfunction of particular components of cholinergic signaling can contribute to symptoms or where medications can ameliorate (or worsen) symptoms regardless of an intrinsic cholinergic deficit. In some instances, these points are uncomfortably close to one another and the effects of cholinergic signaling may be arrayed in opposite directions. In other instances, manipulation of cholinergic systems at one site may be undone by effects of the same manipulation at a distant site. The challenge for neurobiologists and psychopharmacologists is to find ways to refine our interventions in this complex system and to develop compounds or combinations of compounds which can target sites of interest, without substituting one set of impairments for another.

References

- Aasen, I., Kumari, V., and Sharma, T. (2005). Effects of rivastigmine on sustained attention in schizophrenia: An fMRI study. *J. Clin. Psychopharmacol.* **25**, 311–317.
- Albuquerque, E. X., Pereira, E. F., Bonfante-Cabarcas, R., Marchioro, M., Matsubayashi, H., Alkondon, M., and Maelicke, A. (1996). Nicotinic acetylcholine receptors on hippocampal neurons: Cell compartment-specific expression and modulatory control of channel activity. *Prog. Brain Res.* **109**, 111–124.
- Apicella, P. (2002). Tonically active neurons in the primate striatum and their role in the processing of information about motivationally relevant events. *Eur. J. Neurosci.* **16**, 2017–2026.
- Avila, M. T., Sherr, J. D., Hong, E., Myers, C. S., and Thaker, G. K. (2003). Effects of nicotine on leading saccades during smooth pursuit eye movements in smokers and nonsmokers with schizophrenia. *Neuropsychopharmacology* **28**, 2184–2191.
- Aznavour, N., Watkins, K. C., and Descarries, L. (2005). Postnatal development of the cholinergic innervation in the dorsal hippocampus of rat: Quantitative light and electron microscopic immunocytochemical study. *J. Comp. Neurol.* **486**, 61–75.

- Bachy, I., and Retaux, S. (2006). GABAergic specification in the basal forebrain is controlled by the LIM-hd factor Lhx7. *Dev. Biol.* **291**, 218–226.
- Berger-Sweeney, J. (2003). The cholinergic basal forebrain system during development and its influence on cognitive processes: Important questions and potential answers. *Neurosci. Biobehav. Rev.* **27**, 401–411.
- Borda, T., Perez Rivera, R., Joensen, L., Gomez, R. M., and Sterin-Borda, L. (2002). Antibodies against cerebral M1 cholinergic muscarinic receptor from schizophrenic patients: Molecular interaction. *J. Immunol.* **168**, 3667–3674.
- Brady, D. R., Phelps, P. E., and Vaughn, J. E. (1989). Neurogenesis of basal forebrain cholinergic neurons in rat. *Brain Res. Dev. Brain Res.* **47**, 81–92.
- Brebion, G., Bressan, R. A., Amador, X., Malaspina, D., and Gorman, J. M. (2004). Medications and verbal memory impairment in schizophrenia: The role of anticholinergic drugs. *Psychol. Med.* **34**, 369–374.
- Breese, C. R., Lee, M. J., Adams, C. E., Sullivan, B., Logel, J., Gillen, K. M., Marks, M. J., Collins, A. C., and Leonard, S. (2000). Abnormal regulation of high affinity nicotinic receptors in subjects with schizophrenia. *Neuropsychopharmacology* **23**, 351–364.
- Bresnahan, M., Schaefer, C. A., Brown, A. S., and Susser, E. S. (2005). Prenatal determinants of schizophrenia: What we have learned thus far? *Epidemiol. Psychiatr. Soc.* **14**, 194–197.
- Buzsaki, G. (2002). Theta oscillations in the hippocampus. *Neuron* **33**, 325–340.
- Bymaster, F. P., Calligaro, D. O., Falcone, J. F., Marsh, R. D., Moore, N. A., Tyne, N. C., Seeman, P., and Wong, D. T. (1996). Radioreceptor binding profile of the atypical antipsychotic olanzapine. *Neuropsychopharmacology* **14**, 87–96.
- Bymaster, F. P., Shannon, H. E., Rasmussen, K., DeLapp, N. W., Ward, J. S., Calligaro, D. O., Mitch, C. H., Whitesitt, C., Ludvigsen, T. S., Sheardown, M., Swedberg, M., Rasmussen, T., et al. (1999). Potential role of muscarinic receptors in schizophrenia. *Life Sci.* **64**, 527–534.
- Bymaster, F. P., Felder, C. C., Ahmed, S., and McKinzie, D. (2002). Muscarinic receptors as a target for drugs treating schizophrenia. *Curr. Drug. Targets CNS Neurol. Disord.* **1**, 163–181.
- Bymaster, F. P., Felder, C. C., Tzavara, E., Nomikos, G. G., Calligaro, D. O., and McKinzie, D. L. (2003). Muscarinic mechanisms of antipsychotic atypicality. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **27**, 1125–1143.
- Calabresi, P., Centonze, D., Gubellini, P., Pisani, A., and Bernardi, G. (2000). Acetylcholine-mediated modulation of striatal function. *Trends Neurosci.* **23**, 120–126.
- Clancy, B., Darlington, R. B., and Finlay, B. L. (2001). Translating developmental time across mammalian species. *Neuroscience* **105**, 7–17.
- Cobb, S. R., and Davies, C. H. (2005). Cholinergic modulation of hippocampal cells and circuits. *J. Physiol.* **562**, 81–88.
- Colgin, L. L., Kramar, E. A., Gall, C. M., and Lynch, G. (2003). Septal modulation of excitatory transmission in hippocampus. *J. Neurophysiol.* **90**, 2358–2366.
- Corcoran, C., Gallitano, A., Leitman, D., and Malaspina, D. (2001). The neurobiology of the stress cascade and its potential relevance for schizophrenia. *J. Psychiatr. Pract.* **7**, 3–14.
- Corcoran, C., Walker, E., Huot, R., Mittal, V., Tessner, K., Kestler, L., and Malaspina, D. (2003). The stress cascade and schizophrenia: Etiology and onset. *Schizophr. Bull.* **29**, 671–692.
- Court, J., Spurden, D., Lloyd, S., McKeith, I., Ballard, C., Cairns, N., Kerwin, R., Perry, R., and Perry, E. (1999). Neuronal nicotinic receptors in dementia with Lewy bodies and schizophrenia: Alpha-bungarotoxin and nicotine binding in the thalamus. *J. Neurochem.* **73**, 1590–1597.
- Court, J. A., Piggott, M. A., Lloyd, S., Cookson, N., Ballard, C. G., McKeith, I. G., Perry, R. H., and Perry, E. K. (2000). Nicotine binding in human striatum: Elevation in schizophrenia and reductions in dementia with Lewy bodies, Parkinson's disease and Alzheimer's disease and in relation to neuroleptic medication. *Neuroscience* **98**, 79–87.

- Coyle, J., and Kershaw, P. (2001). Galantamine, a cholinesterase inhibitor that allosterically modulates nicotinic receptors: Effects on the course of Alzheimer's disease. *Biol. Psychiatry* **49**, 289–299.
- Cragg, S. J. (2006). Meaningful silences: How dopamine listens to the ACh pause. *Trends Neurosci.* **29**, 125–131.
- Crismon, M. L. (1994). Tacrine: First drug approved for Alzheimer's disease. *Ann. Pharmacother.* **28**, 744–751.
- Crook, J. M., Tomaskovic-Crook, E., Copolov, D. L., and Dean, B. (2000). Decreased muscarinic receptor binding in subjects with schizophrenia: A study of the human hippocampal formation. *Biol. Psychiatry* **48**, 381–388.
- Dalack, G. W., Becks, L., Hill, E., Pomerleau, O. F., and Meador-Woodruff, J. H. (1999). Nicotine withdrawal and psychiatric symptoms in cigarette smokers with schizophrenia. *Neuropsychopharmacology* **21**, 195–202.
- De Luca, V., Wong, A. H., Muller, D. J., Wong, G. W., Tyndale, R. F., and Kennedy, J. L. (2004). Evidence of association between smoking and alpha7 nicotinic receptor subunit gene in schizophrenia patients. *Neuropsychopharmacology* **29**, 1522–1526.
- Dean, B., Crook, J. M., Pavey, G., Opeskin, K., and Copolov, D. L. (2000). Muscarinic1 and 2 receptor mRNA in the human caudate-putamen: No change in m1 mRNA in schizophrenia. *Mol. Psychiatry* **5**, 203–207.
- Dean, B., McLeod, M., Keriakous, D., McKenzie, J., and Scarr, E. (2002). Decreased muscarinic1 receptors in the dorsolateral prefrontal cortex of subjects with schizophrenia. *Mol. Psychiatry* **7**, 1083–1091.
- DeBoer, P., Heeringa, M. J., and Abercrombie, E. D. (1996). Spontaneous release of acetylcholine in striatum is preferentially regulated by inhibitory dopamine D2 receptors. *Eur. J. Pharmacol.* **317**, 257–262.
- Depatie, L., O'Driscoll, G. A., Holahan, A. L., Atkinson, V., Thavundayil, J. X., Kin, N. N., and Lal, S. (2002). Nicotine and behavioral markers of risk for schizophrenia: A double-blind, placebo-controlled, cross-over study. *Neuropsychopharmacology* **27**, 1056–1070.
- Dooley, M., and Lamb, H. M. (2000). Donepezil: A review of its use in Alzheimer's disease. *Drugs Aging* **16**, 199–226.
- Eriksson, P., Ankarberg, E., Viberg, H., and Fredriksson, A. (2001). The developing cholinergic system as target for environmental toxicants, nicotine and polychlorinated biphenyls (PCBs): Implications for neurotoxicological processes in mice. *Neurotox. Res.* **3**, 37–51.
- Fagan, A. M., Garber, M., Barbacid, M., Silos-Santiago, I., and Holtzman, D. M. (1997). A role for TrkA during maturation of striatal and basal forebrain cholinergic neurons *in vivo*. *J. Neurosci.* **17**, 7644–7654.
- Falls, D. L., Rosen, K. M., Corfas, G., Lane, W. S., and Fischbach, G. D. (1993). ARIA, a protein that stimulates acetylcholine receptor synthesis, is a member of the neu ligand family. *Cell* **72**, 801–815.
- Flames, N., Long, J. E., Garratt, A. N., Fischer, T. M., Gassmann, M., Birchmeier, C., Lai, C., Rubenstein, J. L., and Marin, O. (2004). Short- and long-range attraction of cortical GABAergic interneurons by neuregulin-1. *Neuron* **44**, 251–261.
- Frazier, C. J., Rollins, Y. D., Breese, C. R., Leonard, S., Freedman, R., and Dunwiddie, T. V. (1998). Acetylcholine activates an alpha-bungarotoxin-sensitive nicotinic current in rat hippocampal interneurons, but not pyramidal cells. *J. Neurosci.* **18**, 1187–1195.
- Freedman, R., Olincy, A., Ross, R. G., Waldo, M. C., Stevens, K. E., Adler, L. E., and Leonard, S. (2003). The genetics of sensory gating deficits in schizophrenia. *Curr. Psychiatry Rep.* **5**, 155–161.

- Friedman, J. I., Adler, D. N., Howanitz, E., Harvey, P. D., Brenner, G., Temporini, H., White, L., Parrella, M., and Davis, K. L. (2002). A double blind placebo controlled trial of donepezil adjunctive treatment to risperidone for the cognitive impairment of schizophrenia. *Biol. Psychiatry* **51**, 349–357.
- Furusho, M., Ono, K., Takebayashi, H., Masahira, N., Kagawa, T., Ikeda, K., and Ikenaka, K. (2006). Involvement of the Olig2 transcription factor in cholinergic neuron development of the basal forebrain. *Dev. Biol.* **293**, 348–357.
- Gotti, C., and Clementi, F. (2004). Neuronal nicotinic receptors: From structure to pathology. *Prog. Neurobiol.* **74**, 363–396.
- Graybiel, A. M., Aosaki, T., Flaherty, A. W., and Kimura, M. (1994). The basal ganglia and adaptive motor control. *Science* **265**, 1826–1831.
- Grosse, G., Djalali, S., Deng, D. R., Holtje, M., Hinz, B., Schwartzkopff, K., Cygon, M., Rothe, T., Stroth, T., Hellweg, R., Ahnert-Hilger, G., and Hortnag, H. (2005). Area-specific effects of brain-derived neurotrophic factor (BDNF) genetic ablation on various neuronal subtypes of the mouse brain. *Brain Res. Dev. Brain Res.* **156**, 111–126.
- Guijarro, C., Rutz, S., Rothmaier, K., Turiault, M., Zhi, Q., Naumann, T., Frotscher, M., Tronche, F., Jackisch, R., and Kretz, O. (2006). Maturation and maintenance of cholinergic medial septum neurons require glucocorticoid receptor signaling. *J. Neurochem.* **97**, 747–758.
- Haroutunian, V., Davidson, M., Kanof, P. D., Perl, D. P., Powchik, P., Losonczy, M., McCrystal, J., Purohit, D. P., Bierer, L. M., and Davis, K. L. (1994). Cortical cholinergic markers in schizophrenia. *Schizophr. Res.* **12**, 137–144.
- Harrison, P. J., and Weinberger, D. R. (2005). Schizophrenia genes, gene expression, and neuropathology: On the matter of their convergence. *Mol. Psychiatry* **10**, 40–68; image 5.
- Hasselmo, M. E. (2005). What is the function of hippocampal theta rhythm?—Linking behavioral data to phasic properties of field potential and unit recording data. *Hippocampus* **15**, 936–949.
- Hohmann, C. F. (2003). A morphogenetic role for acetylcholine in mouse cerebral neocortex. *Neurosci. Biobehav. Rev.* **27**, 351–363.
- Hohmann, C. F., and Berger-Sweeney, J. (1998). Cholinergic regulation of cortical development and plasticity. New twists to an old story. *Perspect. Dev. Neurobiol.* **5**, 401–425.
- Holt, D. J., Herman, M. M., Hyde, T. M., Kleinman, J. E., Sinton, C. M., German, D. C., Hersh, L. B., Graybiel, A. M., and Saper, C. B. (1999). Evidence for a deficit in cholinergic interneurons in the striatum in schizophrenia. *Neuroscience* **94**, 21–31.
- Holt, D. J., Bachus, S. E., Hyde, T. M., Wittie, M., Herman, M. M., Vangel, M., Saper, C. B., and Kleinman, J. E. (2005). Reduced density of cholinergic interneurons in the ventral striatum in schizophrenia: An *in situ* hybridization study. *Biol. Psychiatry* **58**, 408–416.
- Hyde, T. M., and Crook, J. M. (2001). Cholinergic systems and schizophrenia: Primary pathology or epiphenomena? *J. Chem. Neuroanat.* **22**, 53–63.
- Ichikawa, J., Dai, J., O’Laughlin, I. A., Fowler, W. L., and Meltzer, H. Y. (2002). Atypical, but not typical, antipsychotic drugs increase cortical acetylcholine release without an effect in the nucleus accumbens or striatum. *Neuropsychopharmacology* **26**, 325–339.
- Jacobsen, L. K., D’Souza, D. C., Mencl, W. E., Pugh, K. R., Skudlarski, P., and Krystal, J. H. (2004). Nicotine effects on brain function and functional connectivity in schizophrenia. *Biol. Psychiatry* **55**, 850–888.
- Jann, M. W. (2000). Rivastigmine, a new-generation cholinesterase inhibitor for the treatment of Alzheimer’s disease. *Pharmacotherapy* **20**, 1–12.
- Ji, D., Lape, R., and Dani, J. A. (2001). Timing and location of nicotinic activity enhances or depresses hippocampal synaptic plasticity. *Neuron* **31**, 131–141.

- Johnson, D. E., Nedza, F. M., Spracklin, D. K., Ward, K. M., Schmidt, A. W., Iredale, P. A., Godek, D. M., and Rollema, H. (2005). The role of muscarinic receptor antagonism in antipsychotic-induced hippocampal acetylcholine release. *Eur. J. Pharmacol.* **506**, 209–219.
- Johnston, M. V., Rutkowski, J. L., Wainer, B. H., Long, J. B., and Mobley, W. C. (1987). NGF effects on developing forebrain cholinergic neurons are regionally specific. *Neurochem. Res.* **12**, 985–994.
- Johnstone, E. C., Crow, T. J., Ferrier, I. N., Frith, C. D., Owens, D. G., Bourne, R. C., and Gamble, S. J. (1983). Adverse effects of anticholinergic medication on positive schizophrenic symptoms. *Psychol. Med.* **13**, 513–527.
- Jones, S., and Yakel, J. L. (1997). Functional nicotinic ACh receptors on interneurons in the rat hippocampus. *J. Physiol.* **504**(Pt. 3), 603–610.
- Kane, J., Honigfeld, G., Singer, J., and Meltzer, H. (1988). Clozapine for the treatment-resistant schizophrenic. A double-blind comparison with chlorpromazine. *Arch. Gen. Psychiatry* **45**, 789–796.
- Kane, J. M., Marder, S. R., Schooler, N. R., Wirshing, W. C., Umbricht, D., Baker, R. W., Wirshing, D. A., Safferman, A., Ganguli, R., McMeniman, M., and Borenstein, M. (2001). Clozapine and haloperidol in moderately refractory schizophrenia: A 6-month randomized and double-blind comparison. *Arch. Gen. Psychiatry* **58**, 965–972.
- Kawai, H., Zago, W., and Berg, D. K. (2002). Nicotinic alpha 7 receptor clusters on hippocampal GABAergic neurons: Regulation by synaptic activity and neurotrophins. *J. Neurosci.* **22**, 7903–7912.
- Laplanche, F., Srivastava, L. K., and Quirion, R. (2004). Alterations in dopaminergic modulation of prefrontal cortical acetylcholine release in post-pubertal rats with neonatal ventral hippocampal lesions. *J. Neurochem.* **89**, 314–323.
- Larrison-Faucher, A. L., Matorin, A. A., and Sereno, A. B. (2004). Nicotine reduces antisaccade errors in task impaired schizophrenic subjects. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **28**, 505–516.
- Lavolette, S. R., and van der Kooy, D. (2004). The neurobiology of nicotine addiction: Bridging the gap from molecules to behaviour. *Nat. Rev. Neurosci.* **5**, 55–65.
- Lee, M. G., Hassani, O. K., Alonso, A., and Jones, B. E. (2005). Cholinergic basal forebrain neurons burst with theta during waking and paradoxical sleep. *J. Neurosci.* **25**, 4365–4369.
- Lenzi, A., Maltinti, E., Poggi, E., Fabrizio, L., and Coli, E. (2003). Effects of rivastigmine on cognitive function and quality of life in patients with schizophrenia. *Clin. Neuropharmacol.* **26**, 317–321.
- Leonard, S., Adams, C., Breese, C. R., Adler, L. E., Bickford, P., Byerley, W., Coon, H., Griffith, J. M., Miller, C., Myles-Worsley, M., Nagamoto, H. T., Rollins, Y., *et al.* (1996). Nicotinic receptor function in schizophrenia. *Schizophr. Bull.* **22**, 431–445.
- Leonard, S., Gault, J., Hopkins, J., Logel, J., Vianzon, R., Short, M., Drebing, C., Berger, R., Venn, D., Sirota, P., Zerbe, G., Olincy, A., *et al.* (2002). Association of promoter variants in the alpha7 nicotinic acetylcholine receptor subunit gene with an inhibitory deficit found in schizophrenia. *Arch. Gen. Psychiatry* **59**, 1085–1096.
- Levin, E. D., Wilson, W., Rose, J. E., and McEvoy, J. (1996). Nicotine-haloperidol interactions and cognitive performance in schizophrenics. *Neuropsychopharmacology* **15**, 429–436.
- Levin, E. D., McClernon, F. J., and Rezvani, A. H. (2006). Nicotinic effects on cognitive function: Behavioral characterization, pharmacological specification, and anatomic localization. *Psychopharmacology (Berl.)* **184**, 523–539.
- Lewis, D. A. (1990). The organization of chemically-identified neural systems in monkey prefrontal cortex: Afferent systems. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **14**, 371–377.

- Li, Z., Huang, M., Ichikawa, J., Dai, J., and Meltzer, H. Y. (2005). N-desmethylozapine, a major metabolite of clozapine, increases cortical acetylcholine and dopamine release *in vivo* via stimulation of M1 muscarinic receptors. *Neuropsychopharmacology* **30**, 1986–1995.
- Liao, D. L., Hong, C. J., Chen, H. M., Chen, Y. E., Lee, S. M., Chang, C. Y., Chen, H., and Tsai, S. J. (2003). Association of muscarinic m1 receptor genetic polymorphisms with psychiatric symptoms and cognitive function in schizophrenic patients. *Neuropsychobiology* **48**, 72–76.
- Lieberman, J. A., Phillips, M., Gu, H., Stroup, S., Zhang, P., Kong, L., Ji, Z., Koch, G., and Hamer, R. M. (2003). Atypical and conventional antipsychotic drugs in treatment-naïve first-episode schizophrenia: A 52-week randomized trial of clozapine vs chlorpromazine. *Neuropsychopharmacology* **28**, 995–1003.
- Lieberman, J. A., Stroup, T. S., McEvoy, J. P., Swartz, M. S., Rosenheck, R. A., Perkins, D. O., Keefe, R. S., Davis, S. M., Davis, C. E., Lebowitz, B. D., Severe, J., and Hsiao, J. K. (2005). Effectiveness of antipsychotic drugs in patients with chronic schizophrenia. *N. Engl. J. Med.* **353**, 1209–1223.
- Liu, Y., Ford, B., Mann, M. A., and Fischbach, G. D. (2001). Neuregulins increase $\alpha 7$ nicotinic acetylcholine receptors and enhance excitatory synaptic transmission in GABAergic interneurons of the hippocampus. *J. Neurosci.* **21**, 5660–5669.
- Lopez-Bendito, G., Cautinat, A., Sanchez, J. A., Bielle, F., Flames, N., Garratt, A. N., Talmage, D. A., Role, L. W., Charnay, P., Marin, O., and Garel, S. (2006). Tangential neuronal migration controls axon guidance: A role for neuregulin-1 in thalamocortical axon navigation. *Cell* **125**, 127–142.
- Lucas-Meunier, E., Fossier, P., Baux, G., and Amar, M. (2003). Cholinergic modulation of the cortical neuronal network. *Pflügers Arch.* **446**, 17–29.
- MacDermott, A. B., Role, L. W., and Siegelbaum, S. A. (1999). Presynaptic ionotropic receptors and the control of transmitter release. *Annu. Rev. Neurosci.* **22**, 443–485.
- Mansvelder, H. D., van Aerde, K. I., Couey, J. J., and Brussaard, A. B. (2006). Nicotinic modulation of neuronal networks: From receptors to cognition. *Psychopharmacology (Berl)* **184**, 292–305.
- Marin, O., Anderson, S. A., and Rubenstein, J. L. (2000). Origin and molecular specification of striatal interneurons. *J. Neurosci.* **20**, 6063–6076.
- Maurice, N., Mercer, J., Chan, C. S., Hernandez-Lopez, S., Held, J., Tkatch, T., and Surmeier, D. J. (2004). D2 dopamine receptor-mediated modulation of voltage-dependent Na^+ channels reduces autonomous activity in striatal cholinergic interneurons. *J. Neurosci.* **24**, 10289–10301.
- Mazeh, D., Zemishlani, H., Barak, Y., Mirecki, I., and Paleacu, D. (2006). Donepezil for negative signs in elderly patients with schizophrenia: An add-on, double-blind, crossover, placebo-controlled study. *Int. Psychogeriatr.* **18**(3), 429–436.
- McEvoy, J. P., and Freter, S. (1989). The dose-response relationship for memory impairment by anticholinergic drugs. *Compr. Psychiatry* **30**, 135–138.
- Mechawar, N., and Descarries, L. (2001). The cholinergic innervation develops early and rapidly in the rat cerebral cortex: A quantitative immunocytochemical study. *Neuroscience* **108**, 555–567.
- Mendelsohn, E., Rosenthal, M., Bohiri, Y., Werber, E., Kotler, M., and Strous, R. D. (2004). Rivastigmine augmentation in the management of chronic schizophrenia with comorbid dementia: An open-label study investigating effects on cognition, behaviour and activities of daily living. *Int. Clin. Psychopharmacol.* **19**, 319–324.
- Mesulam, M. M. (1999). Spatial attention and neglect: Parietal, frontal and cingulate contributions to the mental representation and attentional targeting of salient extrapersonal events. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **354**, 1325–1346.
- Mesulam, M. M. (2004). The cholinergic innervation of the human cerebral cortex. *Prog. Brain Res.* **145**, 67–78.

- Mesulam, M. M., Mufson, E. J., Levey, A. I., and Wainer, B. H. (1984). Atlas of cholinergic neurons in the forebrain and upper brainstem of the macaque based on monoclonal choline acetyltransferase immunohistochemistry and acetylcholinesterase histochemistry. *Neuroscience* **12**, 669–686.
- Michal, P., Lysikova, M., El-Fakahany, E. E., and Tucek, S. (1999). Clozapine interaction with the M2 and M4 subtypes of muscarinic receptors. *Eur. J. Pharmacol.* **376**, 119–125.
- Mirza, N. R., Peters, D., and Sparks, R. G. (2003). Xanomeline and the antipsychotic potential of muscarinic receptor subtype selective agonists. *CNS Drug Rev.* **9**, 159–186.
- Mobley, W. C., Rutkowski, J. L., Tennekoon, G. I., Gemski, J., Buchanan, K., and Johnston, M. V. (1986). Nerve growth factor increases choline acetyltransferase activity in developing basal forebrain neurons. *Brain Res.* **387**, 53–62.
- Moore, H., Fadel, J., Sarter, M., and Bruno, J. P. (1999). Role of accumbens and cortical dopamine receptors in the regulation of cortical acetylcholine release. *Neuroscience* **88**, 811–822.
- Mori, T., Yuxing, Z., Takaki, H., Takeuchi, M., Iseki, K., Hagino, S., Kitanaka, J., Takemura, M., Misawa, H., Ikawa, M., Okabe, M., and Wanaka, A. (2004). The LIM homeobox gene, L3/Lhx8, is necessary for proper development of basal forebrain cholinergic neurons. *Eur. J. Neurosci.* **19**, 3129–3141.
- Myers, C. S., Robles, O., Kakoyannis, A. N., Sherr, J. D., Avila, M. T., Blaxton, T. A., and Thaker, G. K. (2004). Nicotine improves delayed recognition in schizophrenic patients. *Psychopharmacology (Berl.)* **174**, 334–340.
- Nahas, Z., George, M. S., Horner, M. D., Markowitz, J. S., Li, X., Lorberbaum, J. P., Owens, S. D., McGurk, S., DeVane, L., and Risch, S. C. (2003). Augmenting atypical antipsychotics with a cognitive enhancer (donepezil) improves regional brain activity in schizophrenia patients: A pilot double-blind placebo controlled BOLD fMRI study. *Neurocase* **9**, 274–282.
- Newhouse, P. A., Potter, A., and Singh, A. (2004). Effects of nicotinic stimulation on cognitive performance. *Curr. Opin. Pharmacol.* **4**, 36–46.
- Ninan, I., and Kulkarni, S. K. (1996). Clozapine-induced cognitive dysfunction in mice. *Methods Find. Exp. Clin. Pharmacol.* **18**, 367–372.
- Olsson, M., Bjorklund, A., and Campbell, K. (1998). Early specification of striatal projection neurons and interneuronal subtypes in the lateral and medial ganglionic eminence. *Neuroscience* **84**, 867–876.
- Parikh, V., Evans, D. R., Khan, M. M., and Mahadik, S. P. (2003). Nerve growth factor in never-medicated first-episode psychotic and medicated chronic schizophrenic patients: Possible implications for treatment outcome. *Schizophr. Res.* **60**, 117–123.
- Park, S., Knopick, C., McGurk, S., and Meltzer, H. Y. (2000). Nicotine impairs spatial working memory while leaving spatial attention intact. *Neuropsychopharmacology* **22**, 200–209.
- Partridge, J. G., Apparsundaram, S., Gerhardt, G. A., Ronesi, J., and Lovinger, D. M. (2002). Nicotinic acetylcholine receptors interact with dopamine in induction of striatal long term depression. *J. Neurosci.* **22**, 2541–2549.
- Phillips, H. S., Nishimura, M., Armanini, M. P., Chen, K., Albers, K. M., and Davis, B. M. (2004). Rescue of NGF-deficient mice II: Basal forebrain cholinergic projections require NGF for target innervation but not guidance. *Brain Res. Mol. Brain Res.* **124**, 1–11.
- Picciotto, M. R. (2003). Nicotine as a modulator of behavior: Beyond the inverted U. *Trends Pharmacol. Sci.* **24**, 493–499.
- Raedler, T. J., Knable, M. B., Jones, D. W., Urbina, R. A., Gorey, J. G., Lee, K. S., Egan, M. F., Coppola, R., and Weinberger, D. R. (2003). *In vivo* determination of muscarinic acetylcholine receptor availability in schizophrenia. *Am. J. Psychiatry* **160**, 118–127.
- Reddy, G. R., Basha, M. R., Devi, C. B., Suresh, A., Baker, J. L., Shafeek, A., Heinz, J., and Chetty, C. S. (2003). Lead induced effects on acetylcholinesterase activity in cerebellum and hippocampus of developing rat. *Int. J. Dev. Neurosci.* **21**, 347–352.

- Rice, M. E., and Cragg, S. J. (2004). Nicotine amplifies reward-related dopamine signals in striatum. *Nat. Neurosci.* **7**, 583–584.
- Robinson, S. E. (2002). Effects of perinatal buprenorphine and methadone exposures on striatal cholinergic ontogeny. *Neurotoxicol. Teratol.* **24**, 137–142.
- Role, L. W., and Berg, D. K. (1996). Nicotinic receptors in the development and modulation of CNS synapses. *Neuron* **16**, 1077–1085.
- Rosse, R. B., and Deutsch, S. I. (2002). Adjuvant galantamine administration improves negative symptoms in a patient with treatment-refractory schizophrenia. *Clin. Neuropharmacol.* **25**, 272–275.
- Sacco, K. A., Bannon, K. L., and George, T. P. (2004). Nicotinic receptor mechanisms and cognition in normal states and neuropsychiatric disorders. *J. Psychopharmacol.* **18**, 457–474.
- Sarter, M., and Parikh, V. (2005). Choline transporters, cholinergic transmission and cognition. *Nat. Rev. Neurosci.* **6**, 48–56.
- Sarter, M., Nelson, C. L., and Bruno, J. P. (2005). Cortical cholinergic transmission and cortical information processing in schizophrenia. *Schizophr. Bull.* **31**, 117–138.
- Scarr, E., Keriakous, D., Crossland, N., and Dean, B. (2006). No change in cortical muscarinic M2, M3 receptors or [35S]GTPgammaS binding in schizophrenia. *Life Sci.* **78**, 1231–1237.
- Semba, K., Vincent, S. R., and Fibiger, H. C. (1988). Different times of origin of choline acetyltransferase- and somatostatin-immunoreactive neurons in the rat striatum. *J. Neurosci.* **8**, 3937–3944.
- Sherr, J. D., Myers, C., Avila, M. T., Elliott, A., Blaxton, T. A., and Thaker, G. K. (2002). The effects of nicotine on specific eye tracking measures in schizophrenia. *Biol. Psychiatry* **52**, 721–728.
- Shirazi-Southall, S., Rodriguez, D. E., and Nomikos, G. G. (2002). Effects of typical and atypical antipsychotics and receptor selective compounds on acetylcholine efflux in the hippocampus of the rat. *Neuropsychopharmacology* **26**, 583–594.
- Singh, M. M., Kay, S. R., and Opler, L. A. (1987). Anticholinergic-neuroleptic antagonism in terms of positive and negative symptoms of schizophrenia: Implications for psychobiological subtyping. *Psychol. Med.* **17**, 39–48.
- Smiley, J. F., Morrel, F., and Mesulam, M. M. (1997). Cholinergic synapses in human cerebral cortex: An ultra structural study in serial sections. *Exp. Neurol.* **144**(2), 361–368.
- Smiley, J. F., Subramanian, M., and Mesulam, M.-M. (1999). Monoaminergic-cholinergic interactions in the primate basal forebrain. *Neuroscience* **93**, 817–829.
- Smith, R. C., Warner-Cohen, J., Matute, M., Butler, E., Kelly, E., Vaidhyathanaswamy, S., and Khan, A. (2006). Effects of nicotine nasal spray on cognitive function in schizophrenia. *Neuropsychopharmacology* **31**, 637–643.
- Smythies, J. (2005). Section I. The cholinergic system. *Int. Rev. Neurobiol.* **64**, 1–122.
- Sofroniew, M. V., Howe, C. L., and Mobley, W. C. (2001). Nerve growth factor signaling, neuroprotection, and neural repair. *Annu. Rev. Neurosci.* **24**, 1217–1281.
- Strauss, M. E., Reynolds, K. S., Jayaram, G., and Tune, L. E. (1990). Effects of anticholinergic medication on memory in schizophrenia. *Schizophr. Res.* **3**, 127–129.
- Stryjer, R., Strous, R. D., Bar, F., Werber, E., Shaked, G., Buhiri, Y., Kotler, M., Weizman, A., and Rabey, J. M. (2003). Beneficial effect of donepezil augmentation for the management of comorbid schizophrenia and dementia. *Clin. Neuropharmacol.* **26**, 12–17.
- Stryjer, R., Strous, R., Bar, F., Shaked, G., Shiloh, R., Rozencwaig, S., Grupper, D., Buchman, N., Kotler, M., Rabey, J. M., and Weizman, A. (2004). Donepezil augmentation of clozapine monotherapy in schizophrenia patients: A double blind cross-over study. *Hum. Psychopharmacol.* **19**, 343–346.
- Takahashi, L. K. (1998). Prenatal stress: Consequences of glucocorticoids on hippocampal development and function. *Int. J. Dev. Neurosci.* **16**, 199–207.
- Takahashi, L. K., and Goh, C. S. (1998). Glucocorticoid facilitation of cholinergic development in the rat hippocampus. *Neuroscience* **83**, 1145–1153.

- Tanabe, J., Tregellas, J. R., Martin, L. F., and Freedman, R. (2006). Effects of nicotine on hippocampal and cingulate activity during smooth pursuit eye movement in schizophrenia. *Biol. Psychiatry* **59**, 754–761.
- Tandon, R., Shipley, J. E., Greden, J. F., Mann, N. A., Eisner, W. H., and Goodson, J. A. (1991). Muscarinic cholinergic hyperactivity in schizophrenia. Relationship to positive and negative symptoms. *Schizophr. Res.* **4**, 23–30.
- Tandon, R., DeQuardo, J. R., Goodson, J., Mann, N. A., and Greden, J. F. (1992). Effect of anticholinergics on positive and negative symptoms in schizophrenia. *Psychopharmacol. Bull.* **28**, 297–302.
- Tapper, A. R., McKinney, S. L., Nashmi, R., Schwarz, J., Deshpande, P., Labarca, C., Whiteaker, P., Marks, M. J., Collins, A. C., and Lester, H. A. (2004). Nicotine activation of $\alpha 4$ receptors: Sufficient for reward, tolerance, and sensitization. *Science* **306**, 1029–1032.
- Thomas, J. D., La Fiette, M. H., Quinn, V. R., and Riley, E. P. (2000). Neonatal choline supplementation ameliorates the effects of prenatal alcohol exposure on a discrimination learning task in rats. *Neurotoxicol. Teratol.* **22**, 703–711.
- Tugal, O., Yazici, K. M., Anil Yagcioglu, A. E., and Gogus, A. (2004). A double-blind, placebo controlled, cross-over trial of adjunctive donepezil for cognitive impairment in schizophrenia. *Int. J. Neuropsychopharmacol.* **7**, 117–123.
- Tune, L. E., Strauss, M. E., Lew, M. F., Breitlinger, E., and Coyle, J. T. (1982). Serum levels of anticholinergic drugs and impaired recent memory in chronic schizophrenic patients. *Am. J. Psychiatry* **139**, 1460–1462.
- van Vulpén, E. H., and van der Kooy, D. (1998). Striatal cholinergic interneurons: Birthdates predict compartmental localization. *Brain Res. Dev. Brain Res.* **109**, 51–58.
- Vizi, E. S., and Kiss, J. P. (1998). Neurochemistry and pharmacology of the major hippocampal transmitter systems: Synaptic and nonsynaptic interactions. *Hippocampus* **8**, 566–607.
- Wang, Z., Kai, L., Day, M., Ronesi, J., Yin, H. H., Ding, J., Tkatch, T., Lovinger, D. M., and Surmeier, D. J. (2006). Dopaminergic control of corticostriatal long-term synaptic depression in medium spiny neurons is mediated by cholinergic interneurons. *Neuron* **50**, 443–452.
- Ward, N. L., and Hagg, T. (2000). BDNF is needed for postnatal maturation of basal forebrain and neostriatum cholinergic neurons *in vivo*. *Exp. Neurol.* **162**, 297–310.
- Weickert, C. S., Hyde, T. M., Lipska, B. K., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2003). Reduced brain-derived neurotrophic factor in prefrontal cortex of patients with schizophrenia. *Mol. Psychiatry* **8**, 592–610.
- Weickert, C. S., Ligons, D. L., Romanczyk, T., Ungaro, G., Hyde, T. M., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2005). Reductions in neurotrophin receptor mRNAs in the prefrontal cortex of patients with schizophrenia. *Mol. Psychiatry* **10**, 637–650.
- Wilson, C. J. (2006). Striatal D2 receptors and LTD: Yes, but not where you thought they were. *Neuron* **50**, 347–348.
- Wolpowitz, D., Mason, T. B., Dietrich, P., Mendelsohn, M., Talmage, D. A., and Role, L. W. (2000). Cysteine-rich domain isoforms of the neuregulin-1 gene are required for maintenance of peripheral synapses. *Neuron* **25**, 79–91.
- Wonnacott, S., Sidhpura, N., and Balfour, D. J. (2005). Nicotine: From molecular mechanisms to behaviour. *Curr. Opin. Pharmacol.* **5**, 53–59.
- Woolf, N. (1991). Cholinergic systems in mammalian brain and spinal cord. *Prog. Neurobiol.* **37**, 475–524.
- Yang, X., Kuo, Y., Devay, P., Yu, C., and Role, L. (1998). A cysteine-rich isoform of neuregulin controls the level of expression of neuronal nicotinic receptor channels during synaptogenesis. *Neuron* **20**, 255–270.
- Zemishlany, Z., Aizenberg, D., Weiner, Z., and Weizman, A. (1996). Trihexyphenidyl (Artane) abuse in schizophrenic patients. *Int. Clin. Psychopharmacol.* **11**, 199–202.

- Zhang, H., and Sulzer, D. (2004). Frequency-dependent modulation of dopamine release by nicotine. *Nat. Neurosci.* **7**, 581–582.
- Zhao, Y., Marin, O., Hermesz, E., Powell, A., Flames, N., Palkovits, M., Rubenstein, J. L., and Westphal, H. (2003). The LIM-homeobox gene *Lhx8* is required for the development of many cholinergic neurons in the mouse forebrain. *Proc. Natl. Acad. Sci. USA* **100**, 9005–9010.
- Zhou, F. M., Liang, Y., and Dani, J. A. (2001). Endogenous nicotinic cholinergic activity regulates dopamine release in the striatum. *Nat. Neurosci.* **4**, 1224–1229.
- Zwart, R., Van Kleef, R. G., and Vijverberg, H. P. (1999). Physostigmine and atropine potentiate and inhibit neuronal alpha 4 beta 4 nicotinic receptors. *Ann. NY Acad. Sci.* **868**, 636–639.

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SCHIZOPHRENIA AND THE $\alpha 7$ NICOTINIC ACETYLCHOLINE RECEPTOR

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In addition to the devastating symptoms of psychosis, many people with schizophrenia also suffer from cognitive impairment. These cognitive symptoms lead to marked dysfunction and can impact employability, treatment adherence, and social skills. Deficits in P50 auditory gating are associated with attentional impairment and may contribute to cognitive symptoms and perceptual disturbances. This nicotinic cholinergic-mediated inhibitory process represents a potential new target for therapeutic intervention in schizophrenia. This chapter will review evidence implicating the nicotinic cholinergic, and specifically, the $\alpha 7$ nicotinic receptor system in the pathology of schizophrenia. Impaired auditory sensory gating has been linked to the $\alpha 7$ nicotinic receptor gene on the chromosome 15q14 locus. A majority of persons with schizophrenia are heavy smokers. Although nicotine can acutely reverse diminished auditory sensory gating in people with schizophrenia, this effect is lost on a chronic basis due to receptor desensitization. The $\alpha 7$ nicotinic agonist 3-(2,4 dimethoxy)benzylidene-anabaseine (DMXBA) can also enhance auditory sensory gating in animal models. DMXBA is well tolerated in humans and a new study in persons with schizophrenia has found that DMXBA enhances both P50 auditory gating and cognition. $\alpha 7$ Nicotinic acetylcholine receptor agonists appear to be viable candidates for the treatment of cognitive disturbances in schizophrenia.

I. Introduction

In addition to the more obvious symptoms of hallucinations and delusions, people with schizophrenia frequently suffer from cognitive symptoms such as the inability to focus attention. This results in a “flooding” with extraneous sensory stimuli which overwhelms the person’s ability to think coherently (Venables, 1992). Poor cognitive functioning contributes to both poor role-functioning and high costs of care through its association with activities of daily living, productivity, rate of inpatient hospitalization and outpatient utilization, independence, trainability/education levels, employability, and the lost productivity of family members spent caring for their ill relatives (reviewed in Sevy and Davidson, 1995). Cognitive impairment also contributes to poor medication adherence (Jeste *et al.*, 2003) and limits the efficacy of rehabilitative therapies (reviewed in Sharma and Antonova, 2003). Cognitive deficits improve slightly with current antipsychotic medications, but they are not normalized and therefore remain a target for new treatment efforts (Weickert *et al.*, 2003). Given increasing evidence for a role of the nicotinic cholinergic system’s role in the cognitive symptoms of schizophrenia, the $\alpha 7$ nicotinic acetylcholine receptor has been proposed as a candidate for the development of medications specifically targeting cognitive deficits in schizophrenia (Martin *et al.*, 2004). This chapter will review the neurobiological findings that led to the development of this promising new drug treatment for schizophrenia as well as new evidence for the beneficial effect of an $\alpha 7$ nicotinic receptor agonist on cognitive impairment in schizophrenia.

II. Neurobiological and Neurogenetic Evidence for a Link Between the $\alpha 7$ Nicotinic Acetylcholine Receptor and Schizophrenia

Sensory gating, measured using the P50 auditory-evoked response, is impaired in persons with schizophrenia (Adler *et al.*, 1985). The P50 auditory-evoked response occurs 40–75 ms following an auditory stimulus. When a second auditory stimulus is presented in close proximity (500 ms), the P50 auditory-evoked response to the second stimulus is diminished, which is evidence for the activity of an inhibitory process. This impairment has been replicated in multiple independent laboratories (Boutros *et al.*, 1991; Clementz *et al.*, 1997; Judd *et al.*, 1992; Louchart-de la Chapelle *et al.*, 2005; Ward *et al.*, 1996) and is present by the first episode of psychosis (Yee *et al.*, 1998). This inhibitory failure is associated with poor sustained attention, as measured by diminished performance on the Digit Vigilance Test and other tests of attentional dysfunction (Cullum *et al.*, 1993; Yee *et al.*, 1998).

Evidence for the role of the $\alpha 7$ nicotinic acetylcholine receptor in auditory gating was initially established using multiple animal models. The auditory-evoked response of hippocampal CA3 pyramidal neurons in the rat, the P20-N40 field potential, parallels the properties of the human P50 auditory-evoked response. The $\alpha 7$ nicotinic receptor antagonist α -bungarotoxin disrupts P20-N40 gating, while the nicotinic receptor channel blocker mecamylamine and the muscarinic antagonist scopolamine have no effect on P20-N40 gating (Luntz-Leybman *et al.*, 1992). The DBA/2 strain of mice has genetically decreased levels of $\alpha 7$ nicotinic receptors in the CA3 region and impaired auditory gating (Stevens *et al.*, 1996). Finally, nicotine restores auditory gating in fimbria-fornix lesioned rats with impaired auditory gating due to the loss of cholinergic innervation to the hippocampus (Bickford and Wear, 1995).

$\alpha 7$ Nicotinic receptors mediate this inhibitory processing by enhancing the release of gamma-aminobutyric acid (GABA) from GABAergic interneurons via a postsynaptic, calcium-dependent mechanism (Albuquerque *et al.*, 1998; Frazier *et al.*, 1998). Nitric oxide prolongs this effect through a second messenger system (Adams *et al.*, 2000). This enhanced release of GABA stimulates GABA_B receptors which in turn decreases the release of glutamate (Hershman *et al.*, 1995). This effect is thought to prevent hippocampal neurons from responding to the second stimulus in the auditory gating paradigm. These nicotinic receptor-mediated interactions between inhibitory (GABA) and excitatory (glutamate) neurons are also proposed to play a role in the efficiency and patterning of neuronal functioning within the hippocampus and cortex (Albuquerque *et al.*, 2000; Alkonon *et al.*, 2000; Ji and Dani, 2000; Jones *et al.*, 1999).

A parallel series of studies in humans also implicated the $\alpha 7$ nicotinic acetylcholine receptor in the physiology of P50 auditory gating. Nicotine gum and physostigmine were found to improve gating in the relatives of persons with schizophrenia who also had impaired auditory gating (Adler *et al.*, 1992). The study of this group of relatives was especially useful as it was able to avoid the confounds of the additional pathological effects of schizophrenia, the effects of chronic neuroleptic treatment as well as the effects of chronic smoking on nicotinic receptor levels. These findings were extended to persons with schizophrenia (Adler *et al.*, 1993). Next, mecamylamine was administered with nicotine at a dose which blocks $\alpha 4/\beta 2$ receptors. Mecamylamine did not attenuate the nicotine induced enhancement of auditory gating. Therefore, the $\alpha 7$ nicotinic receptor appears to be the primary cholinergic receptor responsible for P50 auditory gating in humans as well (Freedman *et al.*, 1994).

In addition to the $\alpha 7$ -mediated deficits in P50 auditory gating, people with schizophrenia also have abnormalities in the expression of central nervous system nicotinic receptors. Decreased $\alpha 7$ nicotinic receptor binding has been noted in the reticular nucleus of the thalamus (Court *et al.*, 1999), the hippocampus (Freedman *et al.*, 1995), and the cingulate cortex (Marutle *et al.*, 2001).

Reduced $\alpha 7$ subunit levels have been noted in frontal lobe regions (Guan *et al.*, 1999), including the dorsolateral prefrontal cortex (Martin-Ruiz *et al.*, 2003). Reduced levels of mRNA are also seen in peripheral blood lymphocytes (Perl *et al.*, 2003).

The relatives of persons with schizophrenia also have poor P50 auditory gating (Clementz *et al.*, 1998; Ross *et al.*, 1999; Siegal *et al.*, 1984), consistent with a genetically determined trait (Waldo *et al.*, 1991). An initial genome scan using poor P50 auditory gating as a phenotype gave only suggestive results at several chromosomes (Coon *et al.*, 1993). Following the identification of genetic markers specific to the $\alpha 7$ nicotinic acetylcholine receptor gene (CHRNA-7) at 15q13–14 (Chini *et al.*, 1994), P50 auditory gating was linked to the chromosome 15q14 locus of CHRNA-7 (Freedman *et al.*, 1997). Families from the NIMH Schizophrenia Genetics Initiative database have since been utilized to find linkage to the diagnosis of schizophrenia itself (Leonard *et al.*, 1998). Since that time, replications of these findings have occurred in North American families of African descent (Kaufmann *et al.*, 1998) and European descent (Tsuang *et al.*, 2001), German families (Stöber *et al.*, 2000), South African families (Riley *et al.*, 2000), Azorean families (Xu *et al.*, 2001), Taiwanese families (Liu *et al.*, 2001) and Canadian families (De Luca *et al.*, 2004). Other studies, including those that have looked specifically at this region, have not found linkage (Curtis *et al.*, 1999; Neves-Pereira *et al.*, 1998).

Although no amino acid-coding region polymorphisms have been identified, multiple single nucleotide polymorphisms in the promoter region of CHRNA-7 as well as a partial duplication of the CHRNA-7 gene have been characterized (Gault *et al.*, 1998). Certain alleles are more frequently present in people with schizophrenia and their family members (Houy *et al.*, 2004; Leonard *et al.*, 2002). Furthermore, as some of these alleles are associated with both decreased promoter region activity *in vitro* and impaired P50 auditory gating, they represent functional polymorphisms that may be related to brain inhibitory pathway failure (Leonard *et al.*, 2002).

III. The Prototypic $\alpha 7$ Nicotinic Agonist, Nicotine, and Schizophrenia

The frequency of tobacco smoking is elevated in people with schizophrenia in both inpatient (De Leon *et al.*, 1995; Llerena *et al.*, 2003) and outpatient settings (Diwan *et al.*, 1998; Hughes *et al.*, 1986). They are heavier smokers (De Leon *et al.*, 1995; Kelly and McCreadie, 1999; Lasser *et al.*, 2000; Masterson and O'Shea, 1984) and they extract more nicotine per cigarette smoked than the general population (Olinicy *et al.*, 1997; Strand and Nybäck, 2005 but see Bozikas *et al.*, 2005). In addition to the health implications of smoking (Goff *et al.*, 2005), the burden of this heavy use includes spending 27% of an already limited income on

the purchase of cigarettes (Steinberg *et al.*, 2005). Their motivation to quit smoking is low (Addington *et al.*, 1997; Ziedonis and George, 1997), and the smoking cessation rate is lower than the rates of other mentally ill populations (Diwan *et al.*, 1998) and the general population (Kelly and McCreadie, 1999). Fortunately, interventions targeted specifically for persons with schizophrenia are being developed (Steinberg *et al.*, 2004; Ziedonis *et al.*, 2003). Successful interventions have utilized cognitive behavioral therapy and sustained release bupropion (Evins *et al.*, 2001; Weiner *et al.*, 2001), nicotine replacement therapy (Breckenridge, 1990; Chou *et al.*, 2004; George *et al.*, 2000; Williams *et al.*, 2004; Ziedonis and George, 1997), cognitive behavioral therapy alone (Addington *et al.*, 1998), and contingent monetary reinforcement (Tidey *et al.*, 2002) to reduce smoking or promote abstinence. The reduction in smoking achieved may last for up to 2 years following the cessation treatment and is associated with a greater likelihood of abstaining in the future (Evins *et al.*, 2004).

The high rate and heavy level of smoking seen in this population may be related to the illness or its treatment (reviewed in Dalack *et al.*, 1998). Patients report that they smoke as a sedative, to reduce negative symptoms, and to counteract medication side effects (Forchuk *et al.*, 2002). Some investigators have hypothesized that smoking in people with schizophrenia may be their striving to reduce neuroleptic-induced side effects such as iatrogenic parkinsonism (Decina *et al.*, 1990; Goff *et al.*, 1992). Others have hypothesized that smoking may be an attempt to prevent worsening of their symptoms during nicotine withdrawal (Dalack *et al.*, 1999; Dalack and Meador-Woodruff, 1996) or an endeavor to alleviate symptoms of depression, anxiety, anhedonia, or amotivation (Glassman, 1993; Nisell *et al.*, 1995; Svensson *et al.*, 1990; Tung *et al.*, 1990). Finally, smoking may be a strategy to improve cognition (Nomikos *et al.*, 2000; Taiminen *et al.*, 1998) and sensory gating (Adler *et al.*, 1993).

Systematic studies of these hypotheses involving the administration (or withdrawal) of nicotine have demonstrated positive effects on movement disorders, negative symptoms, some cognitive tasks, sensory gating, and eye movement performance. Nicotine patch administration can improve tremor, bradykinesia-rigidity, and akathisia (Anfang and Pope, 1997; Yang *et al.*, 2002). However, one study found a worsening of Abnormal Involuntary Movement Scale scores following nicotine patch administration (Dalack *et al.*, 1999) and another study found no effect of smoking on tardive dyskinesia, extrapyramidal or parkinsonian symptoms (Smith *et al.*, 2002). Nicotine withdrawal as part of an abstinence or harm reduction treatment may exacerbate psychotic and depressive symptoms (Evins *et al.*, 2001), although this exacerbation may be prevented by the use of nicotine replacement (George *et al.*, 2000) or bupropion (Evins *et al.*, 2001; Weiner *et al.*, 2001). The use of a nicotine patch in a research setting does not affect Brief Psychiatric Rating Scale scores or Scale for the Assessment of Negative Symptoms SANS; (Dalack *et al.*, 1999; Yang *et al.*, 2002). A case report

found that adjuvant galantamine, the anticholinesterase inhibitor and allosteric nicotinic receptor modulator, improved the SANS score (Rosse and Deutsch, 2002).

The effects of nicotine on neuropsychological measures in persons with schizophrenia have been mixed. Abstinence and then reinitiation of smoking had no effect on attentional measures (Sacco *et al.*, 2005), the nicotine patch improved attention (Dépatie *et al.*, 2002; Levin *et al.*, 1996), nicotine gum worsened attention in smokers and improved it in nonsmokers (Harris *et al.*, 2004), and nicotine nasal spray had no effect on attention (Sherr *et al.*, 2002). Smoking abstinence impaired working memory (George *et al.*, 2001; Sacco *et al.*, 2005) and the reinstatement of smoking improved performance (Sacco *et al.*, 2005). The nicotine patch also improved haloperidol-induced deficits on another test of working memory (Levin *et al.*, 1996). A functional magnetic resonance imaging study of an auditory working memory task found a behavioral improvement following nicotine patch that was associated with increased activation within the insula, putamen, and thalamus (Jacobsen *et al.*, 2004). Nicotine nasal spray, however, had no effect on working memory (Myers *et al.*, 2004). While one study found a positive effect for nicotine nasal spray on verbal memory (Smith *et al.*, 2002), this effect was not seen for smoking (Sacco *et al.*, 2005; Smith *et al.*, 2002), nicotine gum (Harris *et al.*, 2004), or the nicotine patch (Levin *et al.*, 1996). Both nicotine nasal spray and the patch appear to improve complex reaction times (Levin *et al.*, 1996; Smith *et al.*, 2002), but there is no effect on simple reaction time (Levin *et al.*, 1996). Neither abstinence (George *et al.*, 2001) nor the reinitiation of smoking affects executive functioning (Sacco *et al.*, 2005). Finally, one study found an improvement in a visuospatial delayed recognition task following nicotine nasal spray in smokers (Myers *et al.*, 2004) while another study found no effect of nicotine gum on visuospatial abilities (Harris *et al.*, 2004).

Studies of the effect of nicotine on physiological abnormalities such as sensory gating and eye tracking have been more consistent. One of the first investigations of the effect of nicotine in persons with schizophrenia found that abnormal P50 auditory gating was normalized in persons with schizophrenia after smoking (Adler *et al.*, 1993). This finding was replicated using the nicotine patch (Griffith *et al.*, 1998). In a different paradigm of sensory gating, prepulse inhibition, smoking prior to testing results in better test performance than not smoking (Kumari *et al.*, 2001). Studies of smooth pursuit eye movements have been equally robust, with every study to date finding significant enhancement of performance using cigarette smoking (Olincy *et al.*, 1998, 2003), nicotine nasal spray (Avila *et al.*, 2003; Sherr *et al.*, 2002), and nicotine patch (Dépatie *et al.*, 2002). A functional magnetic resonance imaging study of the effects of nicotine on smooth pursuit eye movements found that nicotine enhanced cingulate and precuneus activation and decreased abnormally elevated hippocampal activation

(Tregellas *et al.*, 2004, 2005). Antisaccade task performance also improved following the administration of nicotine gum (Larrison-Faucher *et al.*, 2004).

While the physiological studies are all positive, the neurocognitive findings are less consistent. These studies are limited by the difficulties inherent in studies of any pharmacological agent, such as dosing and administrative route, as well as the specific difficulty present in administering nicotine in persons who are already dependent on the substance. One way to control for these difficulties is to use a population that is not dependent on nicotine such as nonsmokers with schizophrenia. While diminishing the generalizability of the results as well as making recruitment more difficult, this avoids the confounds of withdrawal and different long-term biological effects of smoking such as receptor upregulation and desensitization. Adler *et al.* (1992) took this approach one step further by first examining the effects of nicotine gum in the first-degree relatives of persons with schizophrenia who were impaired on the P50 auditory gating paradigm, thereby avoiding the additional confounds of the illness itself and the medications used to treat the illness. If one chooses to use schizophrenics who smoke, two types of difficulties arise. The first is how to deal with the issue of withdrawal. One must find a balance between clearing the system of the acute effects of nicotine while not precipitating symptoms of withdrawal that might affect performance. Our laboratory has advocated the use of a 2-h period of abstinence to balance these demands. The second issue is how to control for smoking status if using a comparison group of control smokers. While the test-retest reliability of the reported smoking history in persons with schizophrenia is quite high (0.92–0.99) and the intercorrelation of objective measures of smoking heaviness such as carbon monoxide, urine cotinine, and nicotine are fairly similar between persons with schizophrenia and controls (0.52–0.80), the relationship between the reported number of cigarettes smoked per day and these objective measures is much lower for persons with schizophrenia (0.02–0.37 vs 0.61–0.65; Yang *et al.*, 2003). Although not studied directly, this may be due to the greater extraction of nicotine in persons with schizophrenia (Olincy *et al.*, 1997). Despite these difficulties, however, there appears to be clear normalization of deficits in persons with schizophrenia following cigarette smoking or the administration of nicotine.

Nicotine, however, has several limitations as a therapeutic agent. Nicotine induces tachyphylaxis, as demonstrated by the inability of repeated dosing of nicotine to enhance impaired P50 auditory gating. Therefore, sustained benefit does not occur. While nicotine replacement eliminates many of the risks of the other ingredients and additives in tobacco, the long-term risks of chronic nicotine use are unknown and may include carcinogenic risk (Crowley-Weber *et al.*, 2003; Heusch and Maneckjee, 1998) and cerebro- or cardiovascular risk (Benowitz, 2003; Benowitz and Gourlay, 1997; Chalon *et al.*, 2000; Elliott *et al.*, 2003;

Fang *et al.*, 2003; Hakki *et al.*, 2002; West *et al.*, 2003). Furthermore, nicotine is an addictive agent, and the development of tolerance can lead to the stressful symptoms of withdrawal in the absence of continued nicotine dosing (Benowitz, 1998). Less potentially toxic and more chronically effective cholinergic treatments are needed.

An alternative to the use of nicotine as a nicotinic agonist would be to increase endogenous release of acetylcholine. For instance, clozapine is able to increase acetylcholine levels in hippocampus (Shirazi-Southall *et al.*, 2002). Consistent with this acetylcholine-enhancing effect, patients on clozapine have near normal levels of P50 suppression. The normalization of auditory gating over time parallels clinical improvement (Nagamoto *et al.*, 1999). Typical neuroleptics and the majority of atypical neuroleptics, however, have no effect on P50 auditory gating (Adler *et al.*, 2004; Freedman *et al.*, 1983; Yee *et al.*, 1998). Clozapine-induced normalization of auditory gating in DBA/2 mice is blocked by α -bungarotoxin, implicating an $\alpha 7$ nicotinic receptor mechanism (Simosky *et al.*, 2003). Ondansetron, an antiemetic, also increases acetylcholine levels via 5HT-3 receptor antagonism. Similar to clozapine, it enhances P50 auditory gating in persons with schizophrenia (Adler *et al.*, 2005). The anticholinesterase inhibitor donepezil nonsignificantly enhanced the P50 auditory gating in persons with schizophrenia (Buchanan *et al.*, 2002). Although olanzapine is also able to increase acetylcholine levels in the hippocampus (Shirazi-Southall *et al.*, 2002), and a cross-sectional study has shown less impaired levels of auditory gating in people with schizophrenia treated with olanzapine (Light *et al.*, 2000), a more definitive cause and effect relationship has not been demonstrated with a longitudinal study (Arango *et al.*, 2003) and a second cross-sectional study found no differences between unmedicated schizophrenics and patients taking olanzapine (Adler *et al.*, 2004).

Interestingly, clozapine has also shown efficacy in its ability to reduce smoking levels in some (Combs and Advokat, 2000; George *et al.*, 1995; McEvoy *et al.*, 1999) but not every study of persons with schizophrenia (De Leon *et al.*, 2005). An additional study examining the effects of bupropion on smoking rates in persons with schizophrenia was confounded by clozapine use. The one abstinent person at 3 months and three of the four abstinent persons at a 2 year follow-up were also taking clozapine (Evins *et al.*, 2001, 2004). These findings may be consistent with a decreased need to self-medicate with cigarette smoking. However, this effect may not be unique to clozapine, as olanzapine and risperidone have also been shown to be associated with greater abstinence when compared to typical antipsychotics (George *et al.*, 2000). Despite its superior efficacy (Kane *et al.*, 1988) and these additional proposed benefits, treatment with clozapine is limited given the significant side effects of sedation, drooling, tachycardia, and weight gain as well as the serious potential side effects of seizures and agranulocytosis.

IV. The Search for an $\alpha 7$ Nicotinic Acetylcholine Receptor Agonist

Two compounds in current clinical use may have direct effects on $\alpha 7$ nicotinic receptors. The anticholinesterase inhibitor galantamine, which has additional modulatory effects on the $\alpha 7$ nicotinic receptor, has been reported to be beneficial for schizophrenia in a case study (Rosse and Deutsch, 2002). Tropisetron, a 5-HT₃ antagonist marketed outside the United States as an anti-nausea drug, also has efficacy as an $\alpha 7$ nicotinic receptor agonist (Macor *et al.*, 2001; Papke *et al.*, 2005). Tropisetron increases the inhibition of P50 auditory gating in schizophrenia (Koike *et al.*, 2005), an effect due to actions at the $\alpha 7$ nicotinic receptor (Hashimoto *et al.*, 2005).

In addition to these medications already being clinically utilized, several cholinergic receptor agonists have been developed to further characterize central nervous system cholinergic function and as potential candidates for the treatment of dementia of the Alzheimer's type (Kem, 2000). Drugs currently in development include a 1,4-diaza-bicyclo[3.2.2]nonane-4-carboxylic acid 4-pyridin-2-yl-phenyl ester at Pfizer Inc., an (E)-N-methyl-5 (3-pyridinyl)-4-penten-2-amine at Targacept Inc., and a substituted-heteroaryl-7-aza[2.2.1]bicycloheptanes at Pharmacia & Upjohn Company. AR-R 17779, an Astra Arcus product, is an acetylcholine analogue with full agonist properties at the $\alpha 7$ nicotinic receptor (Mullen *et al.*, 2000). ABT-418, while primarily functioning as an $\alpha 4/\beta 2$ agonist, also has some agonist properties at the $\alpha 7$ nicotinic receptor (Briggs *et al.*, 1995). Derivatives of the 5-HT₃ receptor antagonist tropisetron are currently in development (Macor *et al.*, 2001). 3-(2,4 Dimethoxy)benzylidene-anabaseine (DMXBA) is one of a series of compounds derived from anabaseine, an alkaloid found in marine worms (Kem *et al.*, 1971, 1997; Meyer *et al.*, 1998c). DMXBA is a partial agonist at the $\alpha 7$ receptor (Briggs *et al.*, 1995; De Fiebre *et al.*, 1995) and is a weak competitive antagonist at $\alpha 4/\beta 2$ nicotinic (Kem *et al.*, 1996; Meyer *et al.*, 1998a; Papke *et al.*, 2000) and 5HT-3 receptors. Although the metabolites of DMXBA are also active at these receptors, their biological effect may be limited by their greater polarity and consequently, greater difficulty in crossing the blood-brain barrier (Kem *et al.*, 2004).

The efficacy of $\alpha 7$ nicotinic receptor agonists has also been assessed in multiple animal paradigms of learning and memory (Levin and Rezvani, 2000; Levin and Simon, 1998). DMXBA improves memory-related behaviors in multiple paradigms, including a delayed matching to sample task (Briggs *et al.*, 1997), nonspatial avoidance task (Arendash *et al.*, 1995; Meyer *et al.*, 1994, 1997, 1998b), a 17-arm maze (Arendash *et al.*, 1995), and the Morris water maze (Meyer *et al.*, 1997). DMXBA also improves learning behavior as evidenced by enhanced performance during eye blink classical conditioning acquisition (Woodruff-Pak, 2003; Woodruff-Pak *et al.*, 1994) and performance in the Lashley III maze

(Arendash *et al.*, 1995). Some of these beneficial effects may be mediated by enhancement of long-term potentiation in hippocampal cells, a process important in learning and memory formation (Hunter *et al.*, 1994). Finally, a mouse model of schizophrenia-like cognitive and deficit symptoms, "popping," induced by the administration of a *N*-methyl-D-aspartic acid receptor antagonist is reduced following the administration of anabaseine (Mastropaolo *et al.*, 2004).

Given the known role of $\alpha 7$ nicotinic receptors in auditory gating, these drug candidates have also been tested for their ability to reverse auditory gating in animal models. As hypothesized, subcutaneous administration of DMXBA normalizes auditory gating in DBA/2 mice (Stevens *et al.*, 1998). Furthermore, a second injection of DMXBA produces a similar enhancement of inhibition. This lack of tachyphylaxis may represent improved efficacy of DMXBA in normalizing auditory gating on a chronic basis (Stevens *et al.*, 1998, 1999). Intragastrically administered DMXBA also enhanced impaired auditory gating, demonstrating that the medication can be effectively administered on an oral basis and is still efficacious at normalizing impaired auditory gating (Simosky *et al.*, 2001). DMXBA failed in another more complex and strain-dependent model of sensory gating, prepulse inhibition (Olivier *et al.*, 2001; Schreiber *et al.*, 2002). Despite this lack of effect of $\alpha 7$ nicotinic receptor agonists on prepulse inhibition measures, the robust reversal of P50 auditory gating deficits in these animal models is very promising for a similar effect in studies of auditory gating and cognition in schizophrenia.

V. The Phase 1 Study of DMXBA in Schizophrenia

On the basis of the success of preclinical trials of $\alpha 7$ agonists in animal models of learning and memory and the safety of these drugs, DMXBA was initially evaluated in normal subjects with a planned development for the treatment of dementia of the Alzheimer's type. DMXBA was found to significantly improve simple reaction time, correct detection during digit vigilance, both word and picture recognition memory, and both immediate and delayed word recall. Additionally, DMXBA improved subject performance speed on a numeric and spatial working memory task. Improvement was seen at doses from 25 to 150 mg with minimal adverse events (Kitagawa *et al.*, 2003). Despite these promising results, further development of DMXBA was not pursued by Taiho Pharmaceuticals. However, following the correction of the P50 auditory gating deficit by nicotine in persons with schizophrenia, the evidence of the $\alpha 7$ nicotinic receptor's role in this gating deficit in animal studies, as well as the reversal of sensory gating abnormalities in an animal model by the $\alpha 7$ nicotinic receptor agonist DMXBA,

this drug was identified as a potential candidate in the treatment of cognitive dysfunction in schizophrenia (Martin *et al.*, 2004).

A second phase I trial of DMXBBA has been conducted in persons with schizophrenia (Olincy *et al.*, 2006). During this 3-visit study, DMXBBA was administered in a double-blind fashion to 12 persons with schizophrenia. Doses were either placebo, a 75-mg dose with a 37.5-mg follow-up dose, or a 150-mg dose with a 75-mg follow-up dose. Subjects then underwent P50 auditory gating as well as neurocognitive testing. DMXBBA normalized the P50 ratio (effect size of 2.36) as well as the test wave amplitude (effect size 1.45), a more specific measure of inhibition (Fig. 1). These findings are an improvement over the study of nicotine on P50 auditory gating in relatives (effect size 0.86). DMXBBA was also

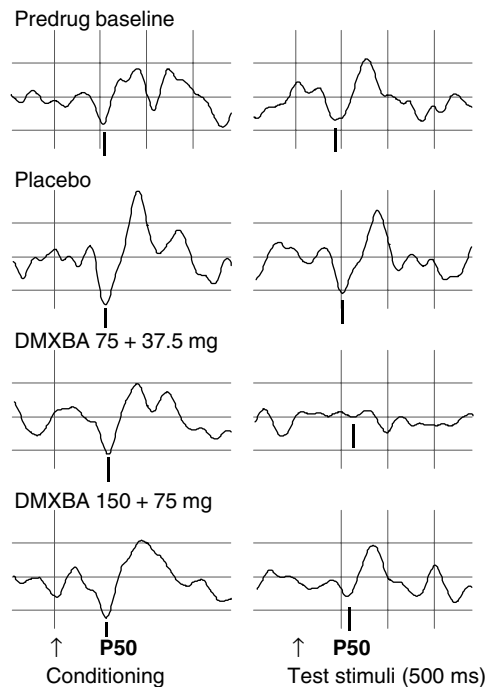


FIG. 1. Auditory-evoked responses of a subject with schizophrenia. Stimuli were a conditioning auditory stimulus and an identical test stimulus, delivered 500 ms apart. Inhibition of the test P50 response is increased by DMXBBA administration, particularly during the lower dose (third row), compared to baseline and placebo responses above it. Arrows show the timing of the stimuli and vertical bars mark the location of the P50 wave in the tracings above. Positive polarity is downwards; vertical grid interval is 2 μ V, and horizontal is 50 ms. This figure is reproduced with permission from Olincy *et al.* (2006). Proof-of-concept trial of an $\alpha 7$ nicotinic agonist in schizophrenia. *Arch. Gen. Psychiatry*, June 2006, **63**, 630–638; Copyright © 2006, American Medical Association. All rights reserved.

able to improve performance on both the Repeatable Battery for the Assessment of Neuropsychological Status' (RBANS) Total Scale score (effect size 1.8) as well as the Attention subscale (effect size 2.17; Fig. 2). These effect sizes are much larger than those seen for nicotine on the RBANS (0.6 and 0.25 for the Total scale score and Attention subscale score, respectively; Harris *et al.*, 2004) as well as for multiple other tests of the actions of nicotine on cognition in schizophrenia (effect sizes of 0.27–1.3; Dépatie *et al.*, 2002; Levin *et al.*, 1996; Myers *et al.*, 2004; Sacco *et al.*, 2005; Smith *et al.*, 2002). Furthermore, the effect sizes seen with DMXB A were also favorable when compared to the typical effect sizes of 0.2–0.5 for the effect of second generation antipsychotics on attentional and composite cognitive scores in persons with schizophrenia (Keefe *et al.*, 2004). The positive effects of DMXB A on sensory gating and cognition were not related to any changes in Brief Psychiatric Rating Scale scores and were therefore not due to changes in positive, negative, or anxiety-related symptoms.

These findings provide further evidence for a role of the nicotinic cholinergic system in the pathology of schizophrenia. Furthermore, specific $\alpha 7$ nicotinic cholinergic agonism is a therapeutic mechanism that provides hope for the

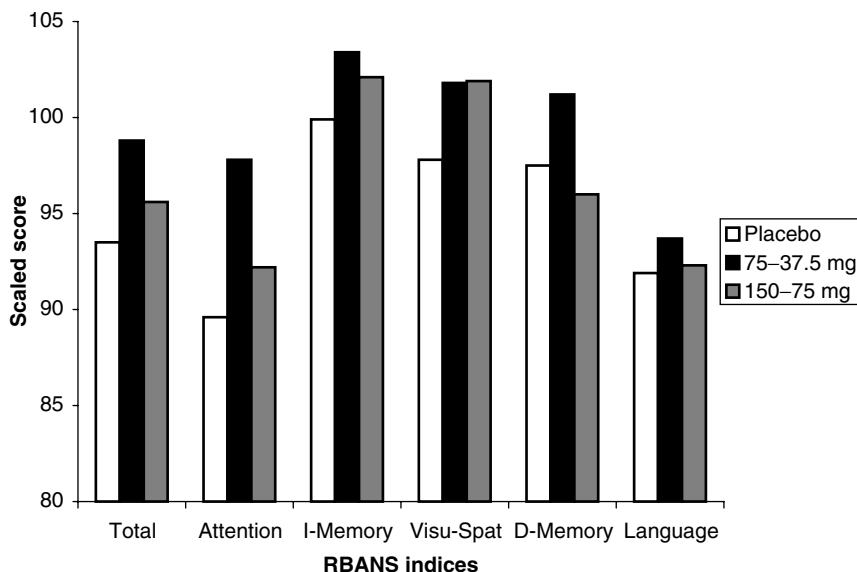


FIG. 2. Effects of DMXB A and placebo on the RBANS Total Scale score and its specific indices. I is immediate and D is delayed memory. This figure is reproduced with permission from Olincy *et al.* (2006). Proof-of-concept trial of an $\alpha 7$ nicotinic agonist in schizophrenia. *Arch. Gen. Psychiatry*, June 2006, **63**, 630–638; Copyright © 2006, American Medical Association. All rights reserved.

treatment of cognitive deficits in schizophrenia. As cognitive symptoms are more closely related to psychosocial dysfunction than traditional positive symptoms, such as hallucinations and delusions (Green, 1996), such a treatment could substantially increase the quality of life for persons with this devastating illness and reduce the financial burden of this disease.

References

- Adams, C. E., Stevens, K. E., Kem, W. R., and Freedman, R. (2000). Inhibition of nitric oxide synthase prevents alpha-7 nicotinic receptor-mediated restoration of inhibitory auditory gating in rat hippocampus. *Brain Res.* **877**, 235–244.
- Addington, J., el-Guebaly, N., Addington, D., and Hodgins, D. (1997). Readiness to stop smoking in schizophrenia. *Can. J. Psychiatry* **42**, 49–52.
- Addington, J., el-Guebaly, N., Campbell, W., Hodgins, D., and Addington, D. (1998). Smoking cessation treatment for patients with schizophrenia. *Am. J. Psychiatry* **155**, 974–976.
- Adler, L. E., Waldo, M. C., and Freedman, R. (1985). Neurophysiologic studies of sensory gating in schizophrenia: Comparison of auditory and visual responses. *Biol. Psychiatry* **20**, 1284–1296.
- Adler, L. E., Hoffer, L. D., Griffith, J., Waldo, M. C., and Freedman, R. (1992). Normalization by nicotine of deficient auditory sensory gating in the relatives of schizophrenics. *Biol. Psychiatry* **32**, 607–616.
- Adler, L. E., Hoffer, L. D., Wiser, A., and Freedman, R. (1993). Normalization of auditory physiology by cigarette smoking in schizophrenic patients. *Am. J. Psychiatry* **150**, 1856–1861.
- Adler, L. E., Olincy, A., Cawthra, E. M., McRae, K. A., Harris, J. G., Nagamoto, H. T., Waldo, M. C., Hall, M.-H., Bowles, A., Woodward, L., Ross, R. G., and Freedman, R. (2004). Varied effects of atypical neuroleptics on P50 auditory gating in schizophrenia patients. *Am. J. Psychiatry* **161**, 1822–1828.
- Adler, L. E., Cawthra, E. M., Donovan, K. A., Harris, J. G., Nagamoto, H. T., Olincy, A., and Waldo, M. C. (2005). Improved P50 auditory gating with ondansetron in medicated schizophrenia patients. *Am. J. Psychiatry* **162**, 386–388.
- Albuquerque, E. X., Pereira, E. F. R., Braga, M. F. M., and Alkondon, M. (1998). Contribution of nicotinic receptors to the function of synapses in the central nervous system: The action of choline as a selective agonist of alpha-7 receptors. *J. Physiol. (Paris)* **92**, 309–316.
- Albuquerque, E. X., Pereira, E. F. R., Mike, A., Eisenbreg, H. M., Maelicke, A., and Alkondon, M. (2000). Neuronal nicotinic receptors in synaptic functions in humans and rats: Physiological and clinical relevance. *Behav. Brain Res.* **113**, 131–141.
- Alkondon, M., Braga, M. F. M., Pereira, E. F. R., Maelicke, A., and Albuquerque, E. X. (2000). Alpha-7 nicotinic acetylcholine receptors and modulation of gabaergic synaptic transmission in the hippocampus. *Eur. J. Pharmacol.* **393**, 59–67.
- Anfang, M. K., and Pope, H. G., Jr. (1997). Treatment of neuroleptic-induced akathisia with nicotine patches. *Psychopharmacology* **134**, 153–156.
- Arango, C., Summerfelt, A., and Buchanan, R. W. (2003). Olanzapine effects on auditory sensory gating in schizophrenia. *Am. J. Psychiatry* **160**, 2066–2068.
- Arendash, G. W., Sengstock, G. J., Sanberg, P. R., and Kem, W. R. (1995). Improved learning and memory in aged rats with chronic administration of the nicotinic receptor agonist GTS-21. *Brain Res.* **674**, 252–259.

- Avila, M. T., Sherr, J. D., Hong, E., Myers, C., and Thaker, G. K. (2003). Effects of nicotine on leading saccades during smooth pursuit eye movements in smokers and nonsmokers with schizophrenia. *Neuropsychopharmacology* **28**, 2184–2191.
- Benowitz, N. L. (1998). Summary: Risks and benefits of nicotine. In "Nicotine Safety and Toxicity" (N. L. Benowitz, Ed.), pp. 185–188. Oxford University Press, New York.
- Benowitz, N. L. (2003). Basic cardiovascular research and its implications for the medicinal use of nicotine. *J. Am. Coll. Cardiol.* **41**, 497–498.
- Benowitz, N. L., and Gourlay, S. G. (1997). Cardiovascular toxicity of nicotine: Implications for nicotine replacement therapy. *J. Am. Acad. Cardiol.* **29**, 1422–1431.
- Bickford, P. C., and Wear, K. D. (1995). Restoration of sensory gating of auditory evoked response by nicotine in fimbria-fornix lesioned rats. *Brain Res.* **705**, 235–240.
- Boutros, N. N., Zouridakis, G., and Overall, J. (1991). Replication and extension of P50 findings in schizophrenia. *Clin. Electroencephalogr.* **22**, 40–45.
- Bozikas, V. P., Niopas, I., Kafantari, A., Kanaze, F. I., Gabrieli, C., Melissidis, P., Gamvrula, K., Fokas, K., and Karavatos, A. (2005). No increased levels of the nicotine metabolite cotinine in smokers with schizophrenia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **29**, 1–6.
- Breckenridge, J. S. (1990). Smoking by outpatients. *Hosp. Commun. Psychiatry* **41**, 454–455.
- Briggs, C. A., McKenna, D. G., and Piattoni-Kaplan, M. (1995). Human alpha-7 nicotinic acetylcholine receptor responses to novel ligands. *Neuropharmacology* **34**, 583–590.
- Briggs, C. A., Anderson, D. J., Brioni, J. D., Buccafusco, J. J., Buckley, M. J., Campbell, J. E., Decker, M. W., Donnelly-Roberts, D., Elliot, R. L., Gopalakrishnan, M., Holladay, M. W., Hui, Y.-H., et al. (1997). Functional characterization of the novel neuronal nicotinic acetylcholine receptor ligand GTS-21 *in vitro* and *in vivo*. *Pharmacol. Biochem. Behav.* **57**, 231–241.
- Buchanan, R. W., Summerfelt, A., Tek, C., and Gold, J. (2002). An open-labeled trial of adjunctive donepezil for cognitive impairments in patients with schizophrenia. *Schizophr. Res.* **59**, 29–33.
- Chalon, S., Moreno, H., Jr., Benowitz, N. L., Hoffman, B. B., and Blaschke, T. F. (2000). Nicotine impairs endothelium-dependent dilatation in human veins *in vivo*. *Clin. Pharmacol. Ther.* **67**, 391–397.
- Chini, B., Raimond, E., Elgoyhen, A. B., Moralli, D., Balzaretto, M., and Heinemann, S. (1994). Molecular cloning and chromosomal localization of the human alpha-7-nicotinic receptor subunit gene (CHRNA7). *Genomics* **19**, 379–381.
- Chou, K.-R., Chen, R., Lee, J.-F., Ku, C.-H., and Lu, R.-B. (2004). The effectiveness of nicotine-patch therapy for smoking cessation in patients with schizophrenia. *Int. J. Nurs. Stud.* **41**, 321–330.
- Clementz, B. A., Geyer, M. A., and Braff, D. L. (1997). P50 suppression among schizophrenia and normal comparison subjects: A methodological analysis. *Biol. Psychiatry* **41**, 1035–1044.
- Clementz, B. A., Geyer, M. A., and Braff, D. L. (1998). Poor P50 suppression among schizophrenia patients and their first-degree biological relatives. *Am. J. Psychiatry* **155**, 1691–1694.
- Combs, D. R., and Advokat, C. (2000). Antipsychotic medication and smoking prevalence in acutely hospitalized patients with chronic schizophrenia. *Schizophr. Res.* **46**, 129–137.
- Coon, H., Plaetke, R., Holik, J., Hoff, M., Myles-Worsley, M., Waldo, M. C., Freedman, R., and Byerley, W. (1993). Use of a neurophysiological trait in linkage analysis of schizophrenia. *Biol. Psychiatry* **34**, 277–289.
- Court, J., Spurdin, D., Lloyd, S., McKeith, I., Ballard, C., Cairns, N., Kerwin, R., Perry, R., and Perry, E. (1999). Neuronal nicotinic receptors in dementia with lewy bodies and schizophrenia: Alpha-bungarotoxin and nicotine binding in thalamus. *J. Neurochem.* **73**, 1590–1597.
- Crowley-Weber, C. L., Dvorakova, K., Crowley, C., Bernstein, H., Bernstein, C., Garewal, H., and Payne, C. M. (2003). Nicotine increases oxidative stress, activates NF-kB and GRP78, induces apoptosis and sensitizes cells to genotoxic/xenobiotic stresses by a multiple stress inducer, deoxycholate: Relevance to colon carcinogenesis. *Chem. Biol. Interact.* **145**, 53–66.

- Cullum, C. M., Harris, J. G., Waldo, M. C., Smernoff, E., Madison, A., Nagamoto, H. T., Griffith, J., Adler, L. E., and Freedman, R. (1993). Neurophysiological and neuropsychological evidence for attentional dysfunction in schizophrenia. *Schizophr. Res.* **10**, 131–141.
- Curtis, L., Blouin, J.-L., Radhakrishna, U., Gehrig, C., Lasseter, V. K., Wolyniec, P., Nestadt, G., Dombroski, B., Kazazian, H. H., Pulver, A. E., Housman, D., Bertrand, D., *et al.* (1999). No evidence for linkage between schizophrenia and markers at chromosome 15q13–14. *Am. J. Med. Genet.* **88**, 109–112.
- Dalack, G. W., and Meador-Woodruff, J. H. (1996). Smoking, smoking withdrawal and schizophrenia: Case reports and a review of the literature. *Schizophr. Res.* **22**, 133–141.
- Dalack, G. W., Healy, D. J., and Meador-Woodruff, J. H. (1998). Nicotine dependence in schizophrenia: Clinical phenomena and laboratory findings. *Am. J. Psychiatry* **155**, 1490–1501.
- Dalack, G. W., Becks, L., Hill, E., Pomerleau, O. F., and Meador-Woodruff, J. H. (1999). Nicotine withdrawal and psychiatric symptoms in cigarette smokers with schizophrenia. *Neuropsychopharmacology* **21**, 195–202.
- De Fiebre, C. M., Meyer, E. M., Henry, J. C., Muraskin, S. I., Kem, W. R., and Papke, R. L. (1995). Characterization of a series of anabaseine-derived compounds reveals that the 3-(4)-dimethylaminocinnamylidene derivative is a selective agonist at neuronal nicotinic $\alpha 7$ /125 I- α -bungarotoxin receptor subtypes. *Mol. Pharmacol.* **47**, 164–171.
- De Leon, J., Dadvand, M., Canuso, A., Odom White, A., Stanilla, J. K., and Simpson, G. M. (1995). Schizophrenia and smoking: An epidemiological survey in a state hospital. *Am. J. Psychiatry* **152**, 453–455.
- De Leon, J., Diaz, F. J., Josiassen, R. C., Cooper, T. B., and Simpson, G. M. (2005). Does clozapine decrease smoking? *Prog. Neuro-Psychopharmacology Biol. Psychiatry* **29**, 757–762.
- De Luca, V., Wang, H., Squassina, A., Wong, G. W., Yeomans, J., and Kennedy, J. L. (2004). Linkage of M5 muscarinic and $\alpha 7$ -nicotinic receptor genes on 15q13 to Schizophrenia. *Neuroschobiology* **50**, 124–127.
- Decina, P., Caracci, G., Sandik, R., Berman, W., Mukherjee, S., and Scapicchio, P. (1990). Cigarette smoking and neuroleptic-induced parkinsonism. *Biol. Psychiatry* **28**, 502–508.
- Dépatie, L., O'Driscoll, G. A., Holahan, A.-L., Atkinson, V., Thavundayil, J. X., Kin, N. N. Y., and Lal, S. (2002). Nicotine and behavioral markers of risk for schizophrenia: A double-blind, placebo-controlled, cross-over study. *Neuropsychopharmacology* **27**, 1056–1070.
- Diwan, A., Castine, M., Pomerleau, C. S., Meador-Woodruff, J. H., and Dalack, G. W. (1998). Differential prevalence of cigarette smoking in patients with schizophrenia vs mood disorders. *Schizophr. Res.* **33**, 113–118.
- Elliott, B. M., Faraday, M. M., and Grunberg, N. E. (2003). Effects of nicotine on heart dimensions and blood volume in male and female rats. *Nicotine Tobacco Res.* **5**, 341–348.
- Evins, A. E., Mays, V. K., Rigotti, N. A., Tisdale, T., Cather, C., and Goff, D. C. (2001). A pilot trial of bupropion added to cognitive behavioral therapy for smoking cessation in schizophrenia. *Nicotine Tobacco Res.* **3**, 397–403.
- Evins, A. E., Cather, C., Rigotti, N. A., Freudenreich, O., Henderson, D. C., Olm-Shipman, C. M., and Goff, D. C. (2004). Two-year follow-up of a smoking cessation trial in patients with schizophrenia: Increased rates of smoking cessation and reduction. *J. Clin. Psychiatry* **65**, 307–311.
- Fang, Q., Sun, H., and Mayhan, W. G. (2003). Impairment of nitric oxide synthase-dependent dilatation of cerebral arterioles during infusion of nicotine. *Am. J. Physiol. Heart Circ. Physiol.* **284**, H528–534.
- Forchuk, C., Norman, R., Malla, A., Martin, M.-L., McLean, T., Cheng, S., Diaz, K., McIntosh, E., Rickwood, A., Vos, S., and Gibney, C. (2002). Schizophrenia and the motivation for smoking. *Perspect. Psychiatr. Care* **38**, 41–49.

- Frazier, C. J., Rollins, Y. D., Breese, C. R., Leonard, S., Freedman, R., and Dunwiddle, T. V. (1998). Acetylcholine activates an alpha-bungarotoxin-sensitive nicotinic current in rat hippocampal interneurons, but not pyramidal cells. *J. Neurosci.* **18**, 1187–1195.
- Freedman, R., Adler, L. E., Waldo, M. C., Pachtman, E., and Franks, R. D. (1983). Neurophysiological evidence for a defect in inhibitory pathways in schizophrenia: Comparisons of medicated and drug-free patients. *Biol. Psychiatry* **18**, 537–551.
- Freedman, R., Adler, L. E., Bickford, P. C., Byerley, W., Coon, H., Cullum, C. M., Griffith, J., Harris, J. G., Leonard, S., Miller, C., Myles-Worsley, M., Nagamoto, H. T., *et al.* (1994). Schizophrenia and nicotinic receptors. *Harv. Rev. Psychiatry* **2**, 179–192.
- Freedman, R., Hall, M., Adler, L. E., and Leonard, S. (1995). Evidence in postmortem brain tissue for decreased numbers of hippocampal nicotinic receptors in schizophrenia. *Biol. Psychiatry* **38**, 22–33.
- Freedman, R., Coon, H., Myles-Worsley, M., Orr-Urtreger, A., Olincy, A., Davis, A., Polymeropoulos, M., Holik, J., Hopkins, J., Hoff, M., Rosenthal, J., Waldo, M. C., *et al.* (1997). Linkage of a neurophysiological deficit in schizophrenia to a chromosome 15 locus. *Proc. Natl. Acad. Sci. USA* **94**, 587–592.
- Gault, J., Robinson, M., Berger, R., Drebing, C., Logel, J., Hopkins, J., Moore, T., Jacobs, S., Meriwether, J., Choi, M. J., Kim, E. J., Walton, K., *et al.* (1998). Genomic organization and partial duplication of the human alpha-7 neuronal nicotinic acetylcholine receptor gene (CHRNA7). *Genomics* **52**, 173–185.
- George, T. P., Serynak, M. J., Ziedonis, D. M., and Woods, S. W. (1995). Effects of clozapine on smoking in chronic schizophrenic outpatients. *J. Clin. Psychiatry* **56**, 344–346.
- George, T. P., Ziedonis, D. M., Feingold, A., Pepper, W. T., Satterburg, C. A., Winkel, J., Rounsaville, B. J., and Kosten, T. R. (2000). Nicotine transdermal patch and atypical antipsychotic medications for smoking cessation in schizophrenia. *Am. J. Psychiatry* **157**, 1835–1842.
- George, T. P., Vessicchio, J. C., Termine, A., Sahady, D. M., Head, C. A., Pepper, W. T., Kosten, T. R., and Wexler, B. E. (2001). Effects of smoking abstinence on visuospatial working memory function in schizophrenia. *Neuropsychopharmacology* **26**, 75–85.
- Glassman, A. H. (1993). Cigarette smoking: Implications for psychiatric illness. *Am. J. Psychiatry* **150**, 546–553.
- Goff, D. C., Henderson, D. C., and Amico, E. (1992). Cigarette smoking in schizophrenia: Relationship to psychopathology and medication side effects. *Am. J. Psychiatry* **149**, 1189–1194.
- Goff, D. C., Cather, C., Evins, A. E., Henderson, D. C., Freudenreich, O., Copeland, P. M., Bierer, M., Duckworth, K., and Sacks, F. M. (2005). Medical morbidity and mortality in schizophrenia: Guidelines for psychiatrists. *J. Clin. Psychiatry* **66**(2), 183–194.
- Green, M. F. (1996). What are the functional consequences of neurocognitive deficits in schizophrenia? *Am. J. Psychiatry* **153**, 321–330.
- Griffith, J. M., O'Neill, J., Petty, F., Garver, D., Young, D., and Freedman, R. (1998). Nicotinic receptor desensitization and sensory gating deficits in schizophrenia. *Biol. Psychiatry* **44**, 98–106.
- Guan, Z.-Z., Zhang, X., Blennow, K., and Nordberg, A. (1999). Decreased protein level of nicotinic receptor alpha-7 subunit in the frontal cortex from schizophrenic brain. *Neuroreport* **10**, 1779–1782.
- Hakki, A., Friedman, H., and Pross, S. (2002). Nicotine modulation of apoptosis in human coronary artery endothelial cells. *Int. Immunopharmacol.* **2**, 1403–1409.
- Harris, J. G., Kongs, S., Allensworth, D., Martin, L., Tregellas, J., Sullivan, B., Zerbe, G., and Freedman, R. (2004). Effects of nicotine on cognitive deficits in schizophrenia. *Neuropsychopharmacology* **29**, 1378–1385.
- Hashimoto, K., Iyo, M., Freedman, R., and Stevens, K. E. (2005). Tropisetron improves deficient inhibitory auditory processing in DBA/2 mice: Role of alpha-7 nicotinic acetylcholine receptors. *Psychopharmacology* **183**, 13–19.

- Hershman, K. M., Freedman, R., and Bickford, P. C. (1995). GABA-B antagonists diminish the inhibitory gating of auditory response in the rat hippocampus. *Neurosci. Lett.* **190**, 133–136.
- Heusch, W. L., and Maneckjee, R. (1998). Signaling pathways involved in nicotine regulation of apoptosis of human lung cancer cells. *Carcinogenesis* **19**, 551–556.
- Houy, E., Raux, G., Thibaut, F., Belmont, A., Demily, C., Allio, G., Haouzir, S., Fouldrin, G., Petit, M., Frebourg, T., and Campion, D. (2004). The promoter-194 C polymorphism of the nicotinic alpha 7 receptor gene has a protective effect against the P50 sensory gating deficit. *Mol. Psychiatry* **9**, 320–322.
- Hughes, J. R., Hatsukami, D. K., Mitchell, J. E., and Dahlgren, L. A. (1986). Prevalence of smoking among psychiatric outpatients. *Am. J. Psychiatry* **143**, 993–997.
- Hunter, B. E., De Fiebre, C. M., Papke, R. L., Kem, W. R., and Meyer, E. (1994). A novel nicotinic agonist facilitates induction of long-term potentiation in the rat hippocampus. *Neurosci. Lett.* **168**, 130–134.
- Jacobsen, L. K., D'Souza, D. C., Mencl, W. E., Pugh, K. R., Skudlarski, P., and Krystal, J. (2004). Nicotine effects on brain function and functional connectivity in schizophrenia. *Biol. Psychiatry* **55**, 850–858.
- Jeste, S. D., Patterson, T. L., Palmer, B. W., Dolder, C. R., Goldman, S., and Jeste, D. V. (2003). Cognitive predictors of medication adherence among middle-aged and older outpatients with schizophrenia. *Schizophr. Res.* **63**, 49–58.
- Ji, D., and Dani, J. A. (2000). Inhibition and disinhibition of pyramidal neurons by activation of nicotinic receptors on hippocampal interneurons. *J. Neurophysiol.* **83**, 2682–2690.
- Jones, S., Sudweeks, S., and Yakel, J. L. (1999). Nicotinic receptors in the brain: Correlating physiology with function. *TINS* **22**, 555–561.
- Judd, L. L., McAdams, L., Budnick, B., and Braff, D. L. (1992). Sensory gating deficits in schizophrenia: New results. *Am. J. Psychiatry* **149**, 488–493.
- Kane, J., Honigfeld, G., Singer, J., and Meltzer, H. (1988). Clozapine for the treatment-resistant schizophrenic. *Arch. Gen. Psychiatry* **45**, 789–796.
- Kaufmann, C. A., Suarez, B., Malaspina, D., Pepple, J., Svrakic, D. M., Markel, P. D., Meyer, J., Zambuto, C. T., Schmitt, K., Matise, T. C., Harkavy Friedman, J. M., Hampe, C., *et al.* (1998). NIMH Genetics initiative millenium schizophrenia consortium: Linkage analysis of African-American pedigrees. *Am. J. Med. Genet.* **81**, 282–289.
- Keefe, R. S. E., Seidman, L. J., Christensen, B. K., Hamer, R. M., Sharma, T., Sitskoorn, M. M., Lewine, R. R. J., Yurgelun-Todd, D., Gur, R. C., Tohen, M., Tollefson, G. D., Sanger, T. M., *et al.* (2004). Comparative Effect of atypical and conventional antipsychotic drugs on neuro-cognition in first-episode psychosis: A randomized, double-blind trial of olanzapine versus low doses of haloperidol. *Am. J. Psychiatry* **161**, 985–995.
- Kelly, C., and McCreadie, R. G. (1999). Smoking habits, current symptoms, and premorbid characteristics of schizophrenic patients in Nithsdale, Scotland. *Am. J. Psychiatry* **156**, 1751–1757.
- Kem, W. R. (2000). The brain alpha-7 nicotinic receptor may be an important therapeutic target for the treatment of Alzheimer's disease: Studies with DMXBA (GTS-21). *Behav. Brain Res.* **113**, 169–181.
- Kem, W. R., Abbott, B. C., and Coates, R. M. (1971). Isolation and structure of a hoplonemertine toxin. *Toxicon* **9**, 15–22.
- Kem, W. R., Mahnir, V. M., Lin, B., and Prokai-Tartrai, K. (1996). Two primary GTS-21 metabolites are potent partial agonists at alpha-7 nicotinic receptors expressed in the *Xenopus* oocyte. *Soc. Neurosci. Abstr.* **22**, 268.
- Kem, W. R., Mahnir, V. M., Papke, R. L., and Lingle, C. J. (1997). Anabaseine is a potent agonist on muscle and neuronal alpha-bungarotoxin-sensitive nicotinic receptors. *J. Pharmacol. Exp. Ther.* **283**, 979–992.

- Kem, W. R., Mahnir, V. M., Prokai, L., Papke, R. L., Cao, X., LeFrancois, S., Wildeboer, K., Prokai-Tatrai, K., Porter-Papke, J., and Soti, F. S. (2004). Hydroxy metabolites of the Alzheimer's drug candidate 3-[(2,4-dimethoxy)benzylidene]-anabaseine dihydrochloride (GTS-21): Their molecular properties, interactions with brain nicotinic receptors, and brain penetration. *Mol. Pharmacol.* **65**, 56–67.
- Kitagawa, H., Takenouchi, T., Azuma, R., Wesnes, K. A., Kramer, W. G., Clody, D. E., and Burnett, A. L. (2003). Safety, pharmacokinetics, and effects on cognitive function of multiple doses of gts-21 in healthy, male volunteers. *Neuropsychopharmacology* **28**, 542–551.
- Koike, K., Hashimoto, K., Takai, N., Shimizu, E., Komatsu, N., Watanabe, H., Nakazato, M., Okamura, N., Stevens, K. E., Freedman, R., and Iyo, M. (2005). Tropisetron improves deficits in auditory P50 suppression in schizophrenia. *Schizophr. Res.* **76**, 67–72.
- Kumari, V., Soni, W., and Sharma, T. (2001). Influence of cigarette smoking on prepulse inhibition of the acoustic startle response in schizophrenia. *Hum. Psychopharmacol.* **16**, 321–326.
- Larrison-Faucher, A. L., Matorin, A. A., and Sereno, A. B. (2004). Nicotine reduces antisaccade errors in task impaired schizophrenic subjects. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **28**, 505–516.
- Lasser, K., Boyd, J. W., Woolhandler, S., Himmelstein, D. U., McCormick, D., and Bor, D. H. (2000). Smoking and mental illness: A population-based prevalence study. *JAMA* **284**, 2606–2610.
- Leonard, S., Gault, J., Moore, T., Hopkins, J., Robinson, M., Olincy, A., Adler, L. E., Cloninger, C. R., Kaufmann, C. A., Tsuang, M. T., Faraone, S. V., Malaspina, D., *et al.* (1998). Further investigation of a chromosome 15 locus in schizophrenia: Analysis of affected sibpairs from the NIMH genetics initiative. *Am. J. Med. Genet.* **81**, 308–312.
- Levin, E. D., and Simon, B. B. (1998). Nicotinic acetylcholine involvement in cognitive function in animals. *Psychopharmacology* **138**, 217–230.
- Levin, E. D., and Rezvani, A. H. (2000). Development of nicotinic drug therapy for cognitive disorders. *Eur. J. Pharmacol.* **393**, 141–146.
- Levin, E., Wilson, W. H., Rose, J. E., and McEvoy, J. P. (1996). Nicotine-haloperidol interaction and cognitive performance in schizophrenics. *Neuropsychopharmacology* **15**, 429–436.
- Leonard, S., Gault, J., Hopkins, J., Logel, J., Vianzon, R., Short, M., Drebing, C., Berger, R., Venn, D., Sirota, P., Zerbe, G. O., Olincy, A., *et al.* (2002). Association of promoter variants in the alpha-7 nicotinic acetylcholine receptor subunit gene with an inhibitory deficit found in schizophrenia. *Arch. Gen. Psychiatry* **59**, 1085–1096.
- Light, G. A., Geyer, M. A., Clementz, B. A., Cadenhead, K. S., and Braff, D. L. (2000). Normal P50 suppression in schizophrenia patients treated with atypical antipsychotic medications. *Am. J. Psychiatry* **157**, 767–771.
- Liu, C.-M., Hwu, H.-G., Lin, M.-W., Ou-Yang, W.-C., Lee, S. F.-C., Fann, C. S. J., Wong, S.-H., and Hsieh, S.-H. (2001). Suggestive evidence for linkage of schizophrenia to markers at chromosome 15q13–14 in Taiwanese families. *Am. J. Med. Genet.* **105**, 658–661.
- Llerena, A., De la Rubia, A., Peñas-Lledo, E. M., Diaz, F. J., and De Leon, J. (2003). Schizophrenia and tobacco smoking in a Spanish psychiatric hospital. *Schizophr. Res.* **60**, 313–317.
- Louchart-de la Chapelle, S., Nkam, I., Houy, E., Belmont, A., Ménard, J.-F., Roussignol, A.-C., Siwek, O., Mezeraï, M., Guillemmou, M., Fouldrin, G., Levillain, D., Dollfus, S., *et al.* (2005). A Concordance study of three electrophysiological measures in schizophrenia. *Am. J. Psychiatry* **162**, 466–474.
- Luntz-Leybman, V., Bickford, P. C., and Freedman, R. (1992). Cholinergic gating of response to auditory stimuli in rat hippocampus. *Brain Res.* **587**, 130–136.
- Macor, J., Gurley, D., Lanthorn, T., Loch, J., III, Mack, R. A., Mullen, G., Tran, O., Wright, N., and Gordon, J. C. (2001). The 5-HT₃ Antagonist tropisetron (ICS 205–930) is a potent selective alpha-7 nicotinic receptor partial agonist. *Bioorg. Med. Chem. Lett.* **11**, 319–321.

- Martin, L. F., Kem, W. R., and Freedman, R. (2004). Alpha-7 nicotinic receptor agonists: Potential new candidates for the treatment of schizophrenia. *Psychopharmacology* **174**, 54–64.
- Martin-Ruiz, C. M., Haroutunian, V. H., Long, P., Young, A. H., Davis, K. L., Perry, E. K., and Court, J. A. (2003). Dementia rating and nicotinic receptor expression in the prefrontal cortex in schizophrenia. *Biol. Psychiatry* **54**, 1222–1233.
- Marutle, A., Zhang, X., Court, J., Piggot, M., Johnson, M., Perry, R., Perry, E., and Nordberg, A. (2001). Laminar distribution of nicotinic receptor subtypes in cortical regions in schizophrenia. *J. Chem. Neuroanat.* **22**, 115–126.
- Masterson, E., and O'Shea, B. (1984). Smoking and malignancy in schizophrenia. *Br. J. Psychiatry* **145**, 429–432.
- Mastropaolo, J., Rosse, R. B., and Deutsch, S. I. (2004). Anabaseine, a selective nicotinic acetylcholine receptor agonist, antagonizes MK-801-elicited mouse popping behavior, an animal model of schizophrenia. *Behav. Brain Res.* **153**, 419–422.
- McEvoy, J. P., Freudenreich, O., and Wilson, W. (1999). Smoking and therapeutic response to clozapine in patients with schizophrenia. *Biol. Psychiatry* **46**, 125–129.
- Meyer, E. M., De Fiebre, C. M., Hunter, B. E., Simpkins, C. E., Frauworth, N., and De Fiebre, N. E. C. (1994). Effects of anabaseine-related analogs on rat brain nicotinic receptor binding and on avoidance behaviors. *Drug Dev. Res.* **31**, 127–134.
- Meyer, E. M., Tay, E. T., Papke, R. L., Meyers, C., Huang, G.-L., and De Fiebre, C. M. (1997). 3-2,4-Dimethoxybenzylidene-anabaseine (DMXBA) selectively activates rat alpha-7 receptors and improves memory-related behaviors in a mecamylamine-sensitive manner. *Brain Res.* **768**, 49–56.
- Meyer, E. M., Kuryatov, A., Gerzanich, V., Lindstrom, J., and Papke, R. L. (1998a). Analysis of 3-(4-Hydroxy, 2-Methoxybenzylidene)anabaseine selectivity and activity at human and rat alpha-7 nicotinic receptors. *J. Pharmacol. Exp. Ther.* **287**, 918–925.
- Meyer, E. M., King, M. A., and Meyers, C. (1998b). Neuroprotective effects of 2,4-dimethoxybenzylidene anabaseine (DMXB) and tetrahydroaminoacridine (THA) in neocortices of nucleus basalis lesioned rats. *Brain Res.* **786**, 252–254.
- Meyer, E. M., Tay, E. T., Zoltevicz, J. A., Meyers, C., King, M. A., Papke, R. L., and De Fiebre, C. M. (1998c). Neuroprotective and memory-related actions of novel Alpha-7 nicotinic agents with different mixed agonist/antagonist properties. *J. Pharmacol. Exp. Ther.* **284**, 1026–1032.
- Mullen, G., Napier, J., Balestra, M., Decory, T., Hale, G., Macor, J., Mack, R., Loch III, J., Wu, E., Kover, A., et al. (2000). (–)-Spiro [1-azabicyclo[2.2.2.] octane-3, 5'-oxazolidin-2'-one], a conformationally restricted analogue of acetylcholine, is a highly selective full agonist at the alpha-7 nicotinic acetylcholine Receptor. *J. of Med. Chem.* **43**, 4045–4050.
- Myers, C. S., Robles, O., Kakoyannis, A. N., Sherr, J. D., Avila, M. T., Blaxton, T. A., and Thaker, G. K. (2004). Nicotine improves delayed recognition in schizophrenic patients. *Psychopharmacology* **174**, 334–340.
- Nagamoto, H. T., Adler, L. E., McRae, K. A., Huettl, P., Cawthra, E., Gerhardt, G., Hea, R., and Griffith, J. (1999). Auditory P50 in schizophrenics on clozapine: Improved gating parallels clinical improvement and changes in plasma 3-methoxy-4-hydroxyphenylglycol. *Neuropsychobiology* **39**, 10–17.
- Neves-Pereira, M., Bassett, A. S., Honer, W. G., Lang, D., King, N. A., and Kennedy, J. L. (1998). No evidence for linkage of the CHR7 gene region in Canadian schizophrenia families. *Am. J. Med. Genet.* **81**, 361–363.
- Nisell, M., Nomikos, G. G., and Svensson, T. H. (1995). Nicotine dependence, midbrain dopamine systems and psychiatric disorders. *Pharmacol. Toxicol.* **76**, 157–162.
- Nomikos, G. G., Schilström, B., Hildebrand, B. E., Panagis, G., Grenhoff, J., and Svensson, T. H. (2000). Role of alpha-7 nicotinic receptors in nicotine dependence and implications for psychiatric illness. *Behav. Brain Res.* **113**, 97–103.

- Olincy, A., Young, D. A., and Freedman, R. (1997). Increased levels of the nicotine metabolite cotinine in schizophrenic smokers compared to other smokers. *Biol. Psychiatry* **42**, 1–5.
- Olincy, A., Ross, R. G., Young, D. A., Roath, M., and Freedman, R. (1998). Improvement in smooth pursuit eye movements after cigarette smoking in schizophrenic patients. *Neuropsychopharmacology* **18**, 175–185.
- Olincy, A., Johnson, L. L., and Ross, R. G. (2003). Differential effects of cigarette smoking on performance of a smooth pursuit and a saccadic eye movement task in schizophrenia. *Psychiatry Res.* **117**, 223–236.
- Olincy, A., Harris, J. G., Johnson, L. L., Pender, V., Kongs, S., Allensworth, D., Ellis, J., Zerbe, G. O., Leonard, S., Stevens, K. E., Stevens, J. O., Martin, L., *et al.* (2006). Proof-of-concept trial of an $\alpha 7$ nicotinic agonist in schizophrenia. *Arch. Gen. Psychiatry* **63**, 630–638.
- Olivier, B., Leachy, C., Mullen, T., Paylor, R., Groppi, V. E., Sarnyai, Z., and Brunner, D. (2001). The DBA/2J strain and prepulse inhibition of startle: A model system to test antipsychotics? *Psychopharmacology* **156**, 284–290.
- Papke, R. L., Meyer, E., Nutter, T., and Uteshev, V. V. (2000). Alpha-7 receptor-selective agonists and modes of alpha-7 receptor activation. *Eur. J. Pharmacol.* **393**, 179–195.
- Papke, R. L., Schiff, H. C., Jack, B. A., and Horenstein, N. A. (2005). Molecular dissection of tropisetron, an alpha-7 nicotinic acetylcholine receptor-selective partial agonist. *Neurosci. Lett.* **378**, 140–144.
- Perl, O., Ilani, T., Strous, R. D., Lapidus, R., and Fuchs, S. (2003). The alpha-7 nicotinic acetylcholine receptor in schizophrenia: Decreased mRNA levels in peripheral blood lymphocytes. *FASEB* **17**, 1948–1950.
- Riley, B. P., Makoff, A., Mogudi-Carter, M., Jenkins, T., Williamson, R., Collier, D., and Murray, R. (2000). Haplotype transmission disequilibrium and evidence for linkage of the CHRNA-7 gene region to schizophrenia in southern African bantu families. *Am. J. Med. Genet.* **96**, 196–201.
- Ross, R. G., Olincy, A., Harris, J. G., Radant, A., Hawkins, M., Adler, L. E., and Freedman, R. (1999). Evidence for bilineal inheritance of physiological indicators of risk in childhood-onset schizophrenia. *Am. J. Med. Genet.* **88**, 188–199.
- Rosse, R. B., and Deutsch, S. I. (2002). Adjuvant galantamine administration improves negative symptoms in a patient with treatment-refractory schizophrenia. *Clin. Neuropharmacol.* **25**, 272–275.
- Sacco, K. A., Termine, A., Seyal, A., Dudas, M. M., Vessicchio, J. C., Krishnan-Sarin, S., Jatlow, P. I., Wexler, B. E., and George, T. P. (2005). Effects of cigarette smoking on spatial working memory and attentional deficits in schizophrenia. *Arch. Gen. Psychiatry* **62**, 649–659.
- Schreiber, R., Dalmus, M., and De Vry, J. (2002). Effects of the alpha-4/beta-2 and alpha-7 Nicotinic acetylcholine receptor agonists on prepulse inhibition of the acoustic startle response in rats and mice. *Psychopharmacology* **159**, 248–257.
- Sevy, S., and Davidson, M. (1995). The cost of cognitive impairment in schizophrenia. *Schizophr. Res.* **17**, 1–3.
- Sharma, T., and Antonova, L. (2003). Cognitive function in schizophrenia: Deficits, function consequences, and future treatment. *Psychiatr. Clin. North Am.* **26**, 25–40.
- Sherr, J. D., Myers, C., Avila, M. T., Elliot, A., Blaxton, T. A., and Thaker, G. K. (2002). The effects of nicotine on specific eye tracking measures in schizophrenia. *Biol. Psychiatry* **52**, 721–728.
- Shirazi-Southall, S., Rodriguez, D. E., and Nomikos, G. G. (2002). Effects of typical and atypical antipsychotics and receptor selective compounds on acetylcholine efflux in the hippocampus. *Neuropsychopharmacology* **26**, 583–594.
- Siegal, C., Waldo, M. C., Mizner, G., Adler, L. E., and Freedman, R. (1984). Deficits in sensory gating in schizophrenic patients and their relatives. *Arch. Gen. Psychiatry* **41**, 607–612.
- Simosky, J. K., Stevens, K. E., Kem, W. R., and Freedman, R. (2001). Intragastric DMXB, An Alpha-7 nicotinic agonist, improves deficient sensory inhibition in DBA/2 mice. *Biol. Psychiatry* **50**, 493–500.

- Simosky, J. K., Stevens, K. E., Adler, L. E., and Freedman, R. (2003). Clozapine improves deficient inhibitory auditory processing in DBA/2 mice, via a nicotinic cholinergic mechanism. *Psychopharmacology* **165**, 386–396.
- Smith, R. C., Singh, A., Infante, M., Khandat, A., and Kloos, A. (2002). Effects of cigarette smoking and nicotine nasal spray on psychiatric symptoms and cognition in schizophrenia. *Neuropsychopharmacology* **27**, 479–497.
- Steinberg, M. L., Ziedonis, D. M., Krejci, J. A., and Brandon, T. H. (2004). Motivational Interviewing with personalized feedback: A brief intervention for motivating smokers with schizophrenia to seek treatment for tobacco dependence. *J. Consult. Clin. Psychol.* **72**, 723–728.
- Steinberg, M. L., Williams, J. M., and Ziedonis, D. M. (2005). Financial implications of cigarette smoking among individuals with schizophrenia. *Tob. Control* **13**, 206.
- Stevens, K. E., Freedman, R., Collins, A. C., Hall, M., Leonard, S., Marks, M. J., and Rose, G. M. (1996). Genetic correlation of inhibitory gating of hippocampal auditory evoked response and alpha-bungarotoxin-binding nicotinic cholinergic receptors in inbred mouse strains. *Neuropsychopharmacology* **15**, 152–162.
- Stevens, K. E., Kem, W. R., Mahnir, V. M., and Freedman, R. (1998). Selective alpha-7 nicotinic agonists normalize inhibition of auditory response in DBA mice. *Psychopharmacology* **136**, 320–327.
- Stevens, K. E., Kem, W. R., and Freedman, R. (1999). Selective alpha-7 nicotinic receptor stimulation normalizes Chronic Cocaine-induced loss of hippocampal sensory inhibition in C3H mice. *Biological Psychiatry* **46**, 1443–1450.
- Stöber, G., Saar, K., Rüschemdorf, F., Meyer, J., Nürnberg, G., Jatzke, S., Franzek, E., Reis, A., Lesch, K.-P., Wienker, T. F., and Beckmann, H. (2000). Splitting schizophrenia: Periodic catatonia-susceptibility locus on chromosome 15q15. *Am. J. Hum. Genet.* **67**, 1201–1207.
- Strand, J.-E., and Nybäck, H. (2005). Tobacco use in schizophrenia: A study of cotinine concentrations in the saliva of patients and controls. *Eur. Psychiatry* **20**, 50–54.
- Svensson, T. H., Grenhoff, J., and Engberg, G. (1990). Effect of nicotine on dynamic function of brain catecholamine neurons. *Ciba Found. Symp.* **152**, 169–180.
- Taiminen, T. J., Salokangas, R. K. R., Saarijärvi, S., Niemi, H., Lehto, H., Ahola, V., and Syvälahti, E. (1998). Smoking and cognitive deficits in schizophrenia: A pilot study. *Addict. Behav.* **23**, 263–266.
- Tidey, J. W., O'Neill, S. C., and Higgins, S. T. (2002). Contingent monetary reinforcement of smoking reductions, with and without transdermal nicotine, in outpatients with schizophrenia. *Exp. Clin. Psychopharmacol.* **10**, 241–247.
- Tregellas, J. R., Tanabe, J. L., Miller, D. E., Ross, R. G., Olincy, A., and Freedman, R. (2004). Neurobiology of smooth pursuit eye movement deficits in schizophrenia: An fMRI study. *Am. J. Psychiatry* **161**, 315–321.
- Tregellas, J. R., Tanabe, J. L., Martin, L. F., and Freedman, R. (2005). fMRI of response to nicotine during a smooth pursuit eye movement task in schizophrenia. *Am. J. Psychiatry* **162**, 391–393.
- Tsuang, D. W., Skol, A. D., Faraone, S. V., Bingham, S., Young, K. A., Prabhudesai, S., Haverstock, S. L., Mena, F., Menon, A. S., Bisset, D., Pepple, J., Sauter, F., et al. (2001). Examination of genetic linkage of chromosome 15 to schizophrenia in a large veterans affairs cooperative study sample. *Am. J. Med. Genet.* **105**, 662–668.
- Tung, C.-S., Grenhoff, J., and Svensson, T. H. (1990). Nicotine counteracts midbrain dopamine cell dysfunction induced by prefrontal cortex inactivation. *Acta Physiol. Scand.* **138**, 427–428.
- Venables, P. H. (1992). Hippocampal function and schizophrenia: Experimental psychological evidence. *Ann. NY Acad. Sci.* **658**, 111–127.
- Waldo, M. C., Carey, G., Myles-Worsley, M., Cawthra, E., Adler, L. E., Nagamoto, H. T., Wender, P., Byerley, W., Plautke, R., and Freedman, R. (1991). Codistribution of a sensory gating deficit and schizophrenia in multi-affected families. *Psychiatry Res.* **39**, 257–268.

- Ward, P. B., Hoffer, L. D., Liebert, B., Catts, S. V., O'Donnell, M., and Adler, L. E. (1996). Replication of a P50 auditory sensory gating deficit in Australian patients with schizophrenia. *Psychiatry Res.* **64**, 121–135.
- Weickert, T. W., Goldberg, T. E., Marenco, S., Bigelow, L. B., Egan, M. F., and Weinberger, D. R. (2003). Comparison of cognitive performances during a placebo period and an atypical antipsychotic treatment period in schizophrenia: Critical examination of confounds. *Neuropsychopharmacology* **28**, 1491–1500.
- Weiner, E., Ball, M. P., Summerfelt, A., Gold, J., and Buchanan, R. W. (2001). Effects of sustained-release bupropion and supportive group therapy on cigarette consumption in patients with schizophrenia. *Am. J. Psychiatry* **158**, 635–637.
- West, K. A., Brognard, J., Clark, A. S., Linnoila, I. R., Yang, X., Swain, S. M., Harris, C., Belinsky, S., and Dennis, P. A. (2003). Rapid Akt activation by nicotine and a tobacco carcinogen modulates the phenotype of normal human airway epithelial cells. *J. Clin. Invest.* **111**, 81–90.
- Williams, J. M., Ziedonis, D. M., and Foulds, J. (2004). A case series of nicotine nasal spray in the treatment of tobacco dependence among patients with schizophrenia. *Psychiatr. Serv.* **55**, 1064–1066.
- Woodruff-Pak, D. S. (2003). Mecamylamine reversal by nicotine and by a partial alpha-7 nicotinic acetylcholine receptor agonist (GTS-21) in rabbits tested with delay eye blink classical conditioning. *Behav. Brain Res.* **143**, 159–167.
- Woodruff-Pak, D. S., Li, Y.-T., and Kem, W. R. (1994). A nicotinic agonist (GTS-21), eye blink classical conditioning, and nicotinic receptor binding in rabbit brain. *Brain Res.* **645**, 309–317.
- Xu, J., Pato, M. T., Dalla Torre, C., Medeiros, H., Carvalho, C., Basile, V. S., Bauer, A., Dourado, A., Valente, J., Soares, M. J., Macedo, A. A., Coelho, I., *et al.* (2001). Evidence of linkage disequilibrium between the alpha-7-nicotinic receptor gene (CHRNA-7) locus and schizophrenia in Azorean families. *Am. J. Med. Genet.* **105**, 669–674.
- Yang, Y. K., Nelson, L., Kamaraju, L., Wilson, W., and McEvoy, J. P. (2002). Nicotine decreases bradykinesia-rigidity in haloperidol-treated patients with schizophrenia. *Neuropsychopharmacology* **27**, 684–686.
- Yang, Y. K., McEvoy, J., Wilson, W. H., Levin, E. D., and Rose, J. E. (2003). Reliabilities and intercorrelations of reported and objective measures of smoking in patients with schizophrenia. *Schizophr. Res.* **60**, 9–12.
- Yee, C. M., Nuechterlein, K. H., Morris, S. E., and White, P. M. (1998). P50 Suppression in recent-onset schizophrenia: Clinical correlates and risperidone effects. *J. Abnorm. Psychol.* **107**, 691–698.
- Ziedonis, D. M., and George, T. P. (1997). Schizophrenia and nicotine use: Report of a pilot smoking cessation program and review of neurobiological and clinical issues. *Schizophr. Bull.* **23**, 247–254.
- Ziedonis, D. M., Williams, J. M., and Smelson, D. (2003). Serious mental illness and tobacco addiction: A model program to address this common but neglected issue. *Am. J. Med. Sci.* **326**, 223–230.

HISTAMINE AND SCHIZOPHRENIA

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With the availability of an increased number of experimental tools, for example potent and brain-penetrating H_1 -, H_2 -, and H_3 -receptor ligands and mutant mice lacking the histamine synthesis enzyme or the histamine receptors, the functional roles of histaminergic neurons in the brain have been considerably clarified during the recent years, particularly their major role in the control of arousal, cognition, and energy balance. Various approaches tend to establish the implication of histaminergic neurons in schizophrenia. A strong hyperactivity of histamine neurons is induced in rodent brain by administration of methamphetamine or NMDA-receptor antagonists. Histamine neuron activity is modulated by typical and atypical neuroleptics. H_3 -receptor antagonists/inverse agonists display antipsychotic-like properties in animal models of the disease. Because of the limited predictability value of most animal models and the paucity of drugs affecting histaminergic transmission that were tried so far in human, the evidence remains therefore largely indirect, but supports a role of histamine neurons in schizophrenia.

I. Introduction

The idea that histamine may have a function as a neurotransmitter in brain emerged only slowly during the preceding century, essentially at the beginning of the 1970s (Schwartz, 1975), although it had been detected therein much earlier. The main landmarks in this history can be summarized as follows. The development of reliable and sensitive methods to assay the amine and its synthesizing enzyme (Schwartz *et al.*, 1970; Taylor and Snyder, 1971b) was instrumental in allowing to establish its localization in neurons, as well as its presence in a neural pathway traveling in the medial forebrain bundle as evidenced indirectly by lesion studies (Garbarg *et al.*, 1974). The turnover of the amine in cerebral neurons was found to be rapid and almost instantaneously modified by drugs like barbiturates or reserpine (Pollard *et al.*, 1973a,b; Taylor and Snyder, 1971a). The demonstration of depolarization-induced release and enhanced synthesis via calcium-dependent mechanisms (Atack and Carlsson, 1972; Taylor and Snyder, 1973; Verdiere *et al.*, 1975), the elucidation of inactivating metabolic pathways (Reilly and Schayer, 1970; Schwartz *et al.*, 1971), and the characterization in brain of the H₁ and H₂ receptors by biochemical and electrophysiological approaches (Baudry *et al.*, 1975; Haas and Bucher, 1975) completed by the mid-1970s the "picture" of histamine as a typical monoaminergic neurotransmitter. Even more, taking into account a variety of features of the system made available at this time, it was proposed that histaminergic neurons were critically involved in the control of arousal (Schwartz, 1977).

Nevertheless, it took nearly 10 years to develop reliable immunohistochemical tools that permitted to identify a tiny posterior hypothalamic area, the tuberomammillary nucleus, as the origin of the histaminergic pathways (Panula *et al.*, 1984; Watanabe *et al.*, 1983) and, thereby, fully convince the neurobiological community of their existence. At approximately the same time, the third histamine receptor was identified in our laboratory, which is almost exclusively present in brain where it controls the neurotransmitter release and synthesis (Arrang *et al.*, 1983) and developed the first selective and brain-penetrating ligands (Arrang *et al.*, 1987); these agents were used, thereafter, in hundreds of studies, to modify the activity of histaminergic neurons and, thereby, disclose their functions. These basic aspects, which were covered in detail in several comprehensive reviews (Brown *et al.*, 2001; Haas and Panula, 2003; Schwartz *et al.*, 1991; Watanabe and Yanai, 2001), will be briefly presented in the first part of the present chapter.

The evidence for the implication of histaminergic neurons in neuropsychiatric diseases remains largely indirect due to the poor predictability value of most animal models and the paucity of drugs affecting histaminergic transmission that were tried in these human diseases, so far. However, the changes in histamine

neuron activity, the modulation of the histaminergic system by neuroleptics, and the antipsychotic-like properties of H_3 receptor antagonists/inverse agonists support a role of histamine neurons in schizophrenia.

II. The Histaminergic Neuronal System

A. ORGANIZATION

One decade after the first evidence by Garbarg *et al.* (1974) of an ascending histaminergic pathway obtained by lesions of the medial forebrain bundle, the exact localization of corresponding perikarya in the posterior hypothalamus was revealed immunohistochemically in the rat using antibodies against histamine (Panula *et al.*, 1984) or L-histidine decarboxylase (EC 4.1.1.22, HDC), the enzyme responsible for the one-step histamine formation in the brain (Watanabe *et al.*, 1984). Data on the distribution, morphology, and connections of histamine and HDC-immunoreactive neurons were comprehensively reviewed (Panula and Airaksinen, 1991; Panula *et al.*, 2000; Schwartz *et al.*, 1991; Tohyama *et al.*, 1991; Wouterlood and Steinbusch, 1991) and will be only summarized briefly here.

All known histaminergic perikarya constitute a continuous group of mainly magnocellular neurons located in the posterior hypothalamus and collectively named the tuberomammillary nucleus (Fig. 1). In the rat brain, the tuberomammillary nucleus consists of about 2000 histaminergic neurons (Ericson *et al.*, 1987) and can be subdivided into medial, ventral, and diffuse subgroups extending longitudinally from the caudal end of the hypothalamus to the midportion of the third ventricle. A similar organization was described in humans, except that histaminergic neurons are more numerous (~64,000) and occupy a larger proportion of the hypothalamus (Airaksinen *et al.*, 1991). Neurons expressing mRNAs for histidine decarboxylase were found by *in situ* hybridization in the tuberomammillary nucleus, but not in any other brain area (Bayliss *et al.*, 1990). The histaminergic neurons are characterized by the presence of an unusually large variety of markers for other neurotransmitter systems. Most, if not all, contain γ -aminobutyric acid (GABA; Airaksinen *et al.*, 1992; Ericson *et al.*, 1991b), adenosine deaminase, a cytoplasmic enzyme possibly involved in adenosine inactivation (Patel *et al.*, 1986; Senba *et al.*, 1985), and a splice variant of choline acetyltransferase (Kanayama *et al.*, 2003). Some histaminergic neurons also express several neuropeptides but these colocalizations are observed in various proportions and display strong species differences (Airaksinen *et al.*, 1992; Trottier *et al.*, 2002).

In analogy with other monoaminergic neurons, histaminergic neurons constitute long and highly divergent systems projecting in a diffused manner to many cerebral areas (Panula and Airaksinen, 1991; Tohyama *et al.*, 1991; Wouterlood

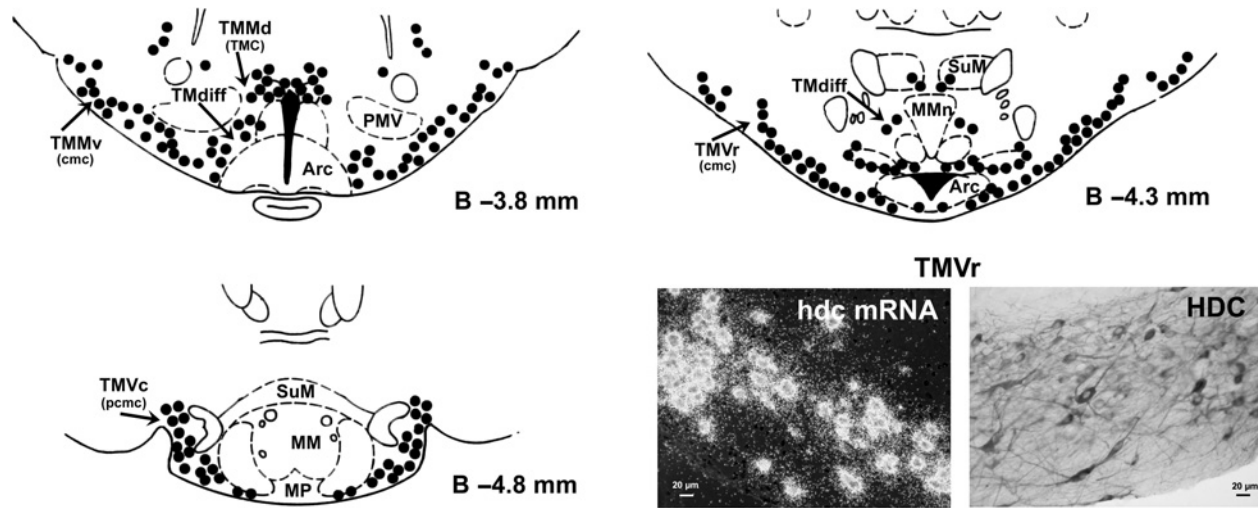


FIG. 1. Localization of histaminergic perikarya in the tuberomammillary nucleus. Histaminergic perikarya are represented by closed circles on frontal sections at indicated levels of caudal hypothalamus. Histidine decarboxylase (HDC) mRNA expression (left) and immunoreactivity (right) is shown in the ventral tuberomammillary subgroup (rostral part). Abbreviations: Arc, arcuate nucleus; cmc, caudal magnocellular nucleus; MM, medial mammillary nucleus medial part; MMn, medial mammillary nucleus median part; MP, medial mammillary nucleus posterior part; pcmc, posterior caudal magnocellular nucleus; PMV, premammillary nucleus ventral part; SuM, supramammillary nucleus; TMC, tuberal magnocellular nucleus; TMDiff, tuberomammillary nucleus diffuse part; TMMd, medial tuberomammillary subgroup dorsal part; TMMv, medial tuberomammillary subgroup ventral part; TMVc, ventral tuberomammillary subgroup caudal part; TMVr, ventral tuberomammillary subgroup rostral part.

and Steinbusch, 1991; Fig. 2). Immunoreactive, mostly unmyelinated, varicose or nonvaricose fibers are detected in almost all cerebral regions, particularly limbic structures. It was confirmed that individual neurons project to widely divergent areas, mainly in an ipsilateral fashion. Major areas of termination of these long ascending fibers arising from the tuberomammillary nucleus are all layers of the cerebral cortex, the olfactory bulb, the hippocampus, the nucleus accumbens, the globus pallidus, the thalamus, the amygdaloid complex, and many hypothalamic nuclei. A histaminergic neuronal system reminiscent of that described in rodents is present in the monkey and human brain with, for example, a dense network of fibers present in various cortical areas or thalamic nuclei (Jin *et al.*, 2002; Panula *et al.*, 1990; Wilson *et al.*, 1999).

Several anterograde and retrograde tracing studies established the existence of afferent connections to the histaminergic perikarya, namely from the infralimbic cortex, the septum-diagonal band complex, the preoptic region, the hypothalamus, and the hippocampal area (subiculum; Ericson *et al.*, 1991a; Wouterlood and Steinbusch, 1991). Sleep-active GABAergic neurons in the ventrolateral preoptic nucleus (VLPO) provide a major input to the tuberomammillary nucleus (Sherin *et al.*, 1996, 1998). The contacts between these two systems are reciprocal because the VLPO is densely innervated by histaminergic fibers (Chou *et al.*, 2002). Histaminergic neurons also receive very dense orexin innervation originating from the lateral hypothalamus (Chemelli *et al.*, 1999). Again, the relationships between the orexin and histamine systems seem to be reciprocal because the orexin neurons are heavily innervated by histaminergic axons (Eriksson *et al.*, 2001a). Supporting their role in the regulation of food intake, histaminergic neurons are densely innervated by nerve fiber varicosities immunoreactive for amylin and α -melanocyte stimulating hormone, two anorexigenic peptides (D'Este *et al.*, 2001; Fekete and Liposits, 2003). Projections from the brainstem to the tuberomammillary nucleus have also been demonstrated. Monoaminergic inputs to the tuberomammillary nucleus originate mainly from the medulla oblongata and from the raphe nuclei, with a lower innervation originating from the locus coeruleus, the ventral tegmental area, and the substantia nigra (Ericson *et al.*, 1989; Sakai *et al.*, 1990).

B. METABOLISM OF HISTAMINE

Histamine biosynthesis in the brain involves two steps: transport of the precursor L-histidine (His) into the cell and its subsequent decarboxylation by HDC (Schwartz *et al.*, 1991). The human HDC gene is composed of 12 exons and its transcripts are alternatively spliced but only the 2.4-kb mRNA, which is predominant in human brain, encodes functional HDC (Yatsunami *et al.*, 1994). The native HDC is a pyridoxal phosphate-dependent enzyme under a homodimeric form

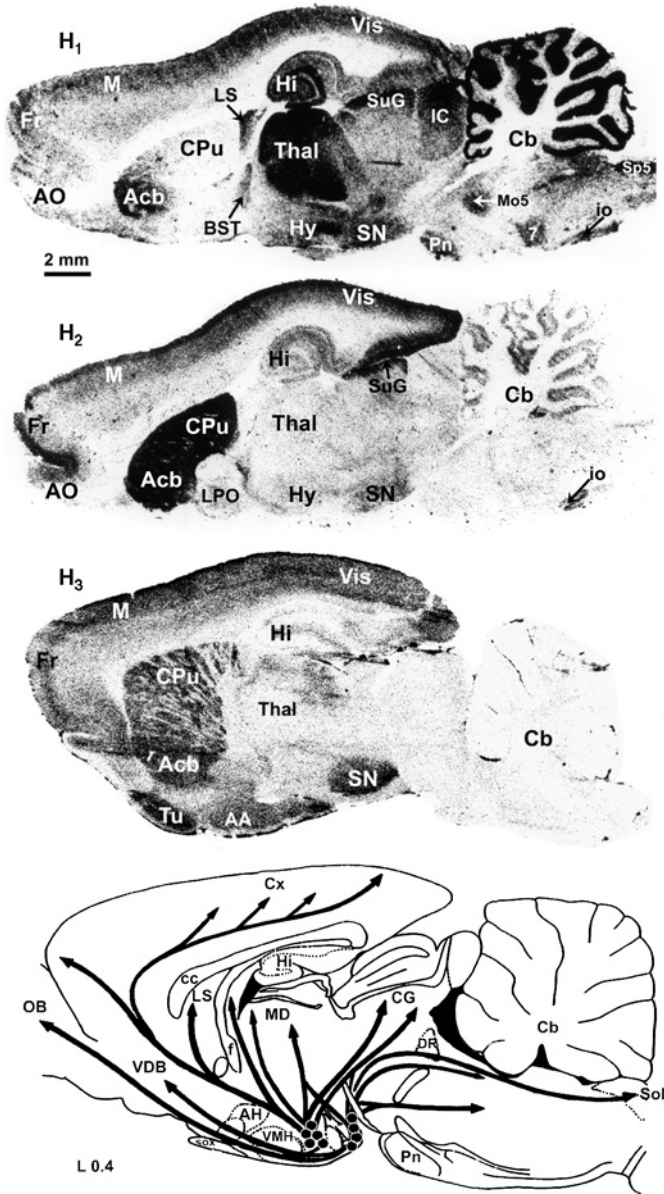


FIG. 2. Autoradiographic localization of histamine receptors and disposition of main histaminergic pathways on sagittal brain sections. H₁ and H₂ receptors were visualized on guinea pig brain sections using [¹²⁵I]iodobolpyramine and [¹²⁵I]iodoaminopotentidine, respectively. H₃ receptors were visualized on rat brain sections using [¹²⁵I]iodoproxyfan. Histaminergic pathways (arrows) were

constituted of two subunits of a 54-kDa isoform and mainly found in the cytoplasm of histaminergic neurons. The regional distribution of brain HDC activity is consistent with data derived from immunohistochemistry, with the highest activity being found in the hypothalamus, the lowest levels in the cerebellum and intermediate activity in telencephalic areas (Schwartz *et al.*, 1970, 1991). HDC-knockout mice provide a suitable model to investigate the involvement of histaminergic neurons in the regulation of fundamental functions such as sleep-wake control (Parmentier *et al.*, 2002), energy homeostasis (Fülöp *et al.*, 2003), learning (Dere *et al.*, 2003), seizure development (Chen *et al.*, 2003b), motor and emotional behaviors (Dere *et al.*, 2004; Iwabuchi *et al.*, 2004), or circadian rhythms (Abe *et al.*, 2004).

In neurons, the newly synthesized histamine is transported to vesicles by the vesicular monoamine transporter 2 (VMAT2) for which it displays high affinity (Peter *et al.*, 1994). Although histaminergic neurons constitute the major localization of HDC, at least two other types of histamine-producing cells, mast cells and microglial cells, have been reported by lesion, biochemical, and histochemical studies. Although they are rather scarce in the brain, mast cells are generally abundant in leptomeninges and also occur in the parenchyma of various brain areas, such as thalamus, where they are mainly distributed along cerebral vessels (Schwartz *et al.*, 1991). Microglial cells belong to the monocyte/macrophage lineage and also contain both HDC activity and mRNAs (Katoh *et al.*, 2001).

Brain histamine is metabolized via transmethylation into *tele*-methylhistamine (*tele*-MeHA) catalyzed by histamine *N*-methyltransferase (HMT, EC 2.1.1.8). *In vivo* inhibition of HMT increases neuronal histamine release, confirming that this enzyme plays a critical role in histamine inactivation (Itoh *et al.*, 1991). Its levels are decreased in Down syndrome and increased in Pick's disease (Kim *et al.*, 2002), and the drug tacrine, which is used in long-term palliative treatment of Alzheimer's disease, inhibits HMT, even more potently than acetylcholinesterase (Morisset *et al.*, 1996). HMT-like immunoreactivity within the CNS was found in the cytosol of a variety of neurons and in vascular walls, whereas astrocytes were not stained (Nishibori *et al.*, 2000). How extracellular histamine is transported

represented on a sagittal section of rat brain. Abbreviations: 7, facial nucleus; AA, amygdaloid area; Acb, accumbens nucleus; AH, anterior hypothalamic area; AO, anterior olfactory nuclei; BST, bed nucleus of the stria terminalis; Cb, cerebellum; cc, corpus callosum; CG, central gray; CPu, caudate putamen; Cx, cortex; DR, dorsal raphe nucleus; f, fornix; Fr, frontal cortex; Hi, hippocampus; Hy, hypothalamus; IC, inferior colliculus; io, inferior olive; LPO, lateral preoptic area; LS, lateral septum; M, motor cortex; MD, mediodorsal thalamic nucleus; Mo5, motor trigeminal nucleus; OB, olfactory bulb; Pn, pontine nucleus; SN, substantia nigra; Sol, nucleus of the solitary tract; sox, supraoptic decussation; Sp5, spinal trigeminal nucleus; SuG, superficial gray layer of superior colliculus; Thal, thalamus; Tu, olfactory tubercle; VDB, nucleus of the vertical limb diagonal band; Vis, visual cortex; VMH, ventromedial hypothalamic nucleus.

into these HMT-containing cells is still unclear. In contrast with other monoaminergic systems, no clear evidence for a high-affinity uptake system for histamine could be found (Schwartz *et al.*, 1991). There is, however, some evidence that histamine could be transported by a low-affinity, low-specificity, high-capacity system (Jonker and Schinkel, 2004).

C. HISTAMINE RECEPTORS

In the brain, the effects of histamine are mediated by three histamine receptor subtypes (H_1 , H_2 , and H_3), which have been defined by means of functional assays followed by design of selective agonists and antagonists and cloning of their genes (Hill *et al.*, 1997; Schwartz and Arrang, 2002). All three belong to the superfamily of receptors with seven transmembrane domains and coupled to guanylnucleotide-sensitive G-proteins (Table I).

1. *The Histamine H_1 Receptor*

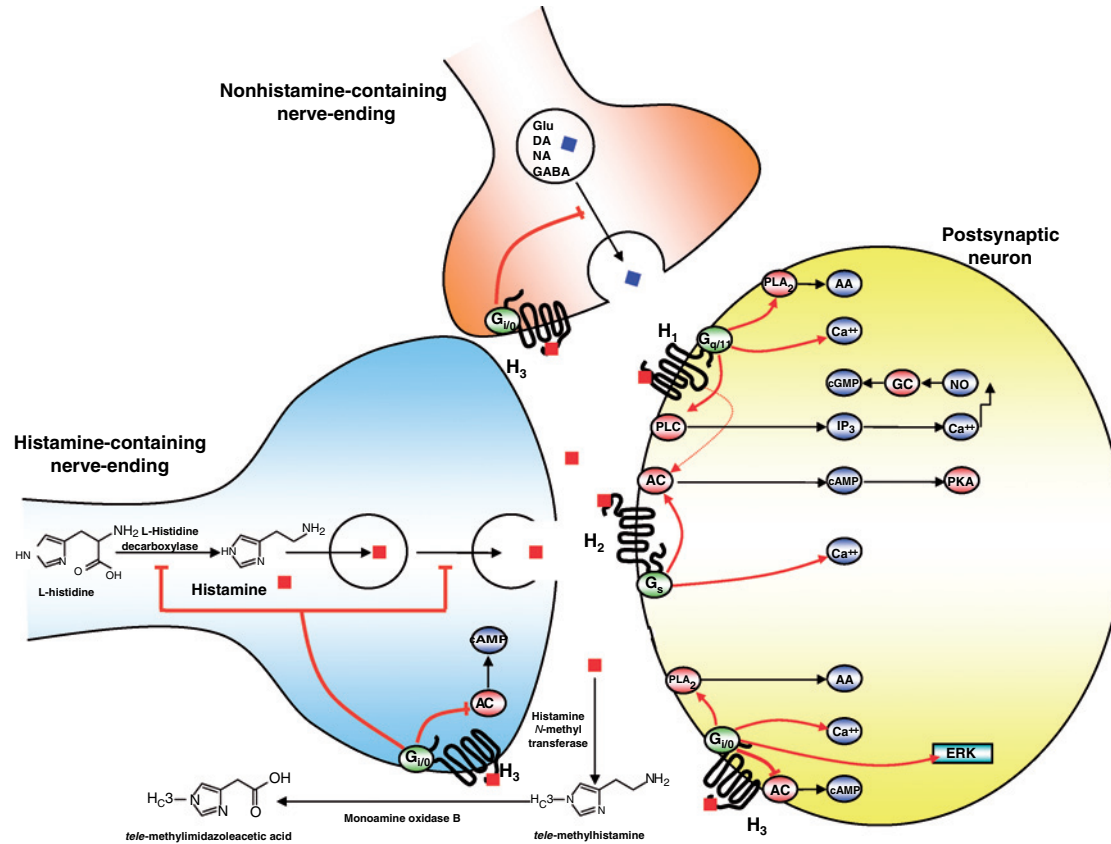
The H_1 receptor was initially defined in functional assays (e.g., smooth muscle contraction) and the design of potent antagonists, the so-called “antihistamines” (e.g., mepyramine), most of which display prominent sedative properties. It was first cloned from cow by expression cloning (Yamashita *et al.*, 1991), and subsequently, from a variety of species, including man (Hill *et al.*, 1997). The human gene contains an intron in the 5'-flanking untranslated region, close to the translation initiation codon, but the translated region is intronless (De Backer *et al.*, 1998). The histamine H_1 receptor produces its intracellular effects via the activation of $G_{q/11}$ proteins (Hill *et al.*, 1997; Leopoldt *et al.*, 1997; Fig. 3). In brain tissues and various cell systems (Leurs *et al.*, 1994; Schwartz *et al.*, 1991), H_1 receptor activation leads to stimulation of phospholipase $C\beta$ and inositol phosphate release. The subsequent mobilization of Ca^{2+} from intracellular stores followed by an influx of extracellular Ca^{2+} induces an increase in intracellular Ca^{2+} levels. This process is presumably responsible for the activation of various Ca^{2+} -dependent pathways by the recombinant or native H_1 receptor, such as potentiation of cAMP accumulation, cGMP accumulation, arachidonic acid release, and glycogenolysis (Leurs *et al.*, 1994; Schwartz *et al.*, 1991). The H_1 receptor mediates mainly excitatory responses in brain, leading to a depolarization and/or an increase in firing frequency in many neurons (Brown *et al.*, 2001; Haas and Panula, 2003).

Biochemical and localization studies of the H_1 receptor were made feasible with the design of reversible and irreversible radiolabeled probes such as [3H] mepyramine and [^{125}I]iodobolpyramine (Garbarg *et al.*, 1992; Pollard and Bouthenet, 1992). The distribution in the brain of H_1 receptor-binding sites is consistent with a predominant neuronal localization of H_1 receptors. They are abundant in guinea pig thalamus, hypothalamic nuclei (e.g., ventromedial

TABLE I
PROPERTIES OF FOUR HISTAMINE RECEPTOR SUBTYPES

	H ₁	H ₂	H ₃	H ₄
Coding sequence	491 a.a. (b) 488 a.a. (gp)	358 a.a. (r) 359 a.a. (d, h, gp)	445 a.a. (h) Shorter variants (h, r, m, gp)	390 a.a. (h) 389 a.a. (gp)
Chromosome localization	486 a.a. (r) 3p25	5	20qTEL	391 a.a. (m, r) 18q11.2
Highest brain densities	Thalamus Cerebellum Hippocampus	Striatum Cerebral cortex Amygdala	Striatum Frontal cortex Substantia nigra	Very low density
Autoreceptor	No	No	Yes	No
Affinity for histamine	Micromolar	Micromolar	Nanomolar	Nanomolar
Characteristic agonists	Histaprodifen	Impromidine	R- α -methylhistamine	4-methylhistamine
Characteristic antagonists	Mepyramine	Cimetidine	Thioperamide	JNJ 7777120
Radioligands	[³ H]Mepyramine	[³ H]Tiotidine	[³ H] R- α -methylhistamine	[³ H]Histamine
Second messengers	[¹²⁵ I]Iodobolpyramine Inositol phosphates (+) Ca ²⁺ (+) Arachidonic acid (+) cAMP (potentiation)	[¹²⁵ I]Iodoamino-potentidine cAMP (+) Ca ²⁺ (+)	[¹²⁵ I]Iodoproxyfan cAMP (-) Inositol phosphates (-) Arachidonic acid (+) Ca ²⁺ (-)	cAMP (-)

a.a., amino acid.



nuclei), nucleus accumbens, amygdaloid nuclei, and frontal cortex but not in caudate-putamen (Pollard and Bouthenet, 1992; Fig. 2). In the human brain, binding sites are more abundant in the neostriatum than in the guinea pig (Martinez-Mir *et al.*, 1990), and, in agreement with the H_1 receptor-mediated modulation of thalamocortical functions by histamine, H_1 receptor-binding sites and mRNAs are abundant in the human thalamus and prefrontal cortex (Jin and Panula, 2005; Jin *et al.*, 2002).

Blockade of H_1 receptors in brain is presumably involved in the sedative, pro-obesity and proconvulsant properties of many drugs displaying high H_1 receptor affinity. The behavioral data obtained with H_1 receptor knockout mice largely support a role of H_1 receptors in arousal and cognition (Huang *et al.*, 2001; Inoue *et al.*, 1996; Lin *et al.*, 2002), anxiety and aggressive behavior (Yanai *et al.*, 1998), nociception (Mobarakeh *et al.*, 2000; Sakurada *et al.*, 2004), anticonvulsant action (Chen *et al.*, 2003b; Hirai *et al.*, 2004), and regulation of food intake and body weight (Masaki *et al.*, 2004; Morimoto *et al.*, 1999).

2. The Histamine H_2 Receptor

Studies of the molecular properties of the H_2 receptor have been greatly facilitated in the 1990s by the design of its first potent and selective radioligand, [125 I]iodoaminopotentidine (Ruat *et al.*, 1990; Traiffort *et al.*, 1992), as well as by the cloning of its gene in various species including man (Gantz *et al.*, 1991a,b; Hill *et al.*, 1997). The recombinant or native H_2 receptor is coupled to G_s proteins and mediates activation of adenylyl cyclase with subsequent increases in cAMP formation and protein kinase A activation (Green *et al.*, 1977; Hegstrand *et al.*, 1976; Schwartz *et al.*, 1991; Fig. 3). As the H_1 receptor, the H_2 receptor usually mediates excitatory responses in neurons (Brown *et al.*, 2001; Haas and Panula, 2003). Its distribution in the brain is consistent with a predominant neuronal localization. Autoradiographic localization of the H_2 receptor using [125 I]iodoaminopotentidine in the guinea pig (Vizuete *et al.*, 1997), monkey, and human brain (Honrubia *et al.*, 2000; Martinez-Mir *et al.*, 1990; Traiffort *et al.*, 1992) shows it distributed heterogeneously (Fig. 2). The H_2 receptor is found in most areas of the cerebral cortex. The caudate putamen, ventral striatal complex and amygdaloid nuclei (bed nucleus of the stria terminalis) are among the richest brain areas. The distribution of the mRNAs is generally in agreement with that of the corresponding binding sites. In the striatum, the absence of mRNAs in the

FIG. 3. Histamine metabolism and signaling in brain. Abbreviations: AA, arachidonic acid; AC, adenylyl cyclase; DA, dopamine; ERK, extracellular signal-related kinase; GABA, γ -aminobutyric acid; GC, guanylate cyclase; Glu, glutamate; IP_3 , inositol-1,4,5-triphosphate; NA, noradrenaline; PKA, protein kinase A; PLA₂, phospholipase A₂; PLC, phospholipase C; NO, nitric oxide.

substantia nigra together with the loss of binding sites in Huntington's disease (Martinez-Mir *et al.*, 1993) indicate that H₂ receptors are expressed by intrinsic neurons. The brain-penetrating H₂ receptor antagonist, zolantidine, has been used to investigate the involvement of H₂ receptors in various neurochemical and behavioral responses (Mori *et al.*, 2004; Nalwalk *et al.*, 1995). A number of tricyclic antidepressants are very potent inhibitors of the H₂ receptor-linked adenylyl cyclase on brain membranes (Green and Maayani, 1977; Kanof and Greengard, 1978), but not on intact cell preparations (Dam Trung Tuong *et al.*, 1980). In addition, the idea that antidepressants derive their clinical efficacy from blockade of cerebral H₂ receptors seems unlikely because such a blockade was not observed after chronic treatments (Nowak *et al.*, 1983).

3. The Histamine H₃ Receptor

The H₃ receptor was initially detected and identified by traditional pharmacological approaches as an autoreceptor controlling histamine synthesis and release in the rat and human brain (Arrang *et al.*, 1983, 1987, 1988). The cloning of its cDNAs in various species including human and rat confirmed that the H₃ receptor is coupled to G_i/G_o proteins (Drutel *et al.*, 2001; Lovenberg *et al.*, 1999; Morisset *et al.*, 2000). In various cell lines, activation of the recombinant H₃ receptor inhibits adenylyl cyclase, activates phospholipase A₂, and activates the ERK signaling pathway (Fig. 3). Although a direct inhibition of adenylyl cyclase could not be observed in various brain regions (Garbarg *et al.*, 1989; Schlicker *et al.*, 1994a), H₃ autoreceptors modulate histamine synthesis through the cAMP pathway (Gomez-Ramirez *et al.*, 2002; Torrent *et al.*, 2005) and H₃ receptor activation inhibits dopamine D₁ receptor-mediated cAMP formation in the rat striatum (Sanchez-Lemus and Arias-Montano, 2004). Recombinant H₃ receptors display a high level of constitutive (or spontaneous) activity and most antagonists act in fact as inverse agonists on various responses (Esbenshade *et al.*, 2005; Morisset *et al.*, 2000). Consistent with the physiological relevance of the process, constitutive activity of native H₃ receptors is detected in rodent brain (Morisset *et al.*, 2000; Rouleau *et al.*, 2002). Inverse agonists at H₃ receptors enhance histamine neuron activity by abrogating the brake triggered by constitutive activity of H₃ autoreceptors (Gbahou *et al.*, 2003; Morisset *et al.*, 2000) and are, therefore, important tools to delineate the functions of histaminergic neurons. The recombinant human H₃ receptors expressed at physiological densities also display constitutive activity, suggesting it is present in human brain (Rouleau *et al.*, 2002).

After its characterization as an autoreceptor present on histamine neurons, the H₃ receptor was shown to inhibit presynaptically the release of other monoamines in brain. H₃ receptors inhibit the *in vitro* release of various neurotransmitters, including histamine itself, noradrenaline, serotonin, dopamine, glutamate, GABA, and

tachykinins (Schlicker *et al.*, 1994b; Schwartz and Arrang, 2002). This presynaptic inhibition presumably results from a direct G-protein-mediated blockade of voltage-gated calcium channels (Brown and Haas, 1999; Jang *et al.*, 2001; Silver *et al.*, 2002). In brain slices or isolated neurons, somatodendritic H₃ autoreceptors also inhibit the firing rate of histaminergic neurons by inhibiting multiple high-threshold calcium channels (Stevens *et al.*, 2001; Takeshita *et al.*, 1998).

The inhibition mediated by H₃ autoreceptors is now well established as a major control mechanism for the activity and functions of histaminergic neurons. However, the physiological role of the H₃ receptors present on other neuronal populations remains largely unknown. In the striatum, H₃ heteroreceptors inhibit dopamine synthesis (Molina-Hernandez *et al.*, 2000) and release (Schlicker *et al.*, 1993) but do not regulate dopamine neuron activity *in vivo* under basal conditions (Imaizumi and Onodera, 1993; Miyazaki *et al.*, 1997; Oishi *et al.*, 1990b). In the striatum, dentate gyrus, and amygdala, H₃ receptor activation inhibits glutamatergic transmission *in vitro* (Brown and Reymann, 1996; Doreulee *et al.*, 2001; Jiang *et al.*, 2005; Molina-Hernandez *et al.*, 2001), but the standard antagonist/inverse agonist thioperamide does not increase synaptic potentials in the freely moving rat (Manahan-Vaughan *et al.*, 1998). H₃ receptors inhibit GABA release in rat striatum, substantia nigra, and hypothalamus (Arias-Montano *et al.*, 2001; Garcia *et al.*, 1997; Jang *et al.*, 2001).

A detailed autoradiographic mapping of the rat H₃ receptor was achieved with the selective antagonist [¹²⁵I]iodoproxyfan (Pillot *et al.*, 2002a; Fig. 2). The comparison with the distribution of H₃ receptor mRNAs provides evidence for the presence of H₃ receptors on many neuronal perikarya, dendrites, and projections. The highest receptor densities are found in the cerebral cortex, basal ganglia, olfactory tubercles, amygdala, and tuberomammillary nucleus. Receptor densities are particularly high in the striatum where lesions indicated that most H₃ receptors are present on projection neurons (Anichtchik *et al.*, 2000a; Cumming *et al.*, 1991; Pollard *et al.*, 1993; Ryu *et al.*, 1994a,b, 1995). In agreement, high densities of H₃ receptor mRNAs are also found in the striatum from rat, guinea pig, and human (Anichtchik *et al.*, 2001; Pillot *et al.*, 2002a; Tardivel-Lacombe *et al.*, 2000). H₃ receptors present on striatonigral neurons account for the dense binding in the substantia nigra pars reticulata (Ryu *et al.*, 1996). H₃ receptors are also expressed in striatopallidal projection neurons (Pillot *et al.*, 2002b) and account for the dense binding in the external globus pallidus in rat (Pillot *et al.*, 2002a) and human (Anichtchik *et al.*, 2001; Martinez-Mir *et al.*, 1990). This expression in the external pallidum is increased in Parkinson's disease (Anichtchik *et al.*, 2001) and dramatically reduced in Huntington's disease (Goodchild *et al.*, 1999). H₃ receptor functions have primarily been studied with standard agonists and antagonists/inverse agonists and with mice lacking H₃ receptors (Koyama *et al.*, 2003; Rizk *et al.*, 2004; Takahashi *et al.*, 2002; Toyota *et al.*, 2002).

4. *The Histamine H₄ Receptor*

Several groups cloned an additional histamine receptor subtype, termed H₄, via *in silico* analysis of human genomic databases (Hough, 2001). Its gene structure, coding sequence, and pharmacology are clearly related to the H₃ receptor. Only few selective ligands have so far been designed for this novel receptor (Gbahou *et al.*, 2006; Lim *et al.*, 2005; Thurmond *et al.*, 2004) and antagonists mainly display anti-inflammatory properties (de Esch *et al.*, 2005). Its expression in the brain is very low, if any, and was detected only in some studies (Coge *et al.*, 2001; Liu *et al.*, 2001a). Its highest level of expression is observed in hematopoietic tissues and cells (bone marrow, spleen, leucocytes; Hough, 2001).

D. HISTAMINERGIC NEURON ACTIVITY AND ITS CONTROL

The autoreceptor-regulated modulation of histamine synthesis in, and release from, brain neurons is well documented (Schwartz *et al.*, 1991). It was initially evidenced in brain slices or synaptosomes (Arrang *et al.*, 1985) after labeling the endogenous pool of histamine using the [³H]-precursor (Verdiere *et al.*, 1975). Exogenous histamine decreases the release and formation of [³H]histamine induced by depolarization, and analysis of these responses led to the pharmacological definition of H₃ receptors. The inhibition mediated by the H₃ autoreceptor constitutes a major regulatory mechanism for histaminergic neuron activity under physiological conditions (Arrang *et al.*, 1987; Schwartz *et al.*, 1991). Administration of selective H₃ receptor agonists reduces histamine turnover and release *in vivo* (Garbarg *et al.*, 1989; Itoh *et al.*, 1992). In contrast, H₃ receptor antagonists/inverse agonists enhance histamine turnover and release *in vivo*, indicating that autoreceptors are tonically activated (Itoh *et al.*, 1991; Ligneau *et al.*, 1998; Mochizuki *et al.*, 1991; Morisset *et al.*, 2000).

In vivo, both neurochemical and electrophysiological studies indicate that the activity of histaminergic neurons is maximal during arousal (Schwartz *et al.*, 1991). Neurons identified as histaminergic neurons exhibit a circadian rhythm of their firing rate, highest during waking and falling silent during deep slow-wave or paradoxical sleep (hence referred to as “waking-on” or “REM-off” neurons; Steininger *et al.*, 1999; Vanni-Mercier *et al.*, 2003). An important determinant of this circadian rhythm of tuberomammillary histaminergic neuron activity is the GABAergic inhibitory input from the VLPO which is activated during sleep (Sherin *et al.*, 1996, 1998; Yang and Hatton, 1997). GABAergic inhibitory postsynaptic potentials are mediated by GABA_A receptors located on histaminergic neurons (Stevens *et al.*, 1999). Histaminergic neurons contain GABA (Airaksinen *et al.*, 1992; Ericson *et al.*, 1991b), but to what extent these receptors play an autoinhibitory role is unclear. Histamine turnover in the brain is rapidly reduced after administration of GABAergic sedative drugs such as barbiturates and

benzodiazepines (Oishi *et al.*, 1986; Pollard *et al.*, 1974), presumably as a result of their interaction with these GABA_A receptors. *In vivo* microdialysis shows that endogenous GABA as well as systemic administration of muscimol, pentobarbital, diazepam, and halothane inhibits histamine release in the rat brain (Chikai *et al.*, 1993; Mammoto *et al.*, 1997; Okakura-Mochizuki *et al.*, 1996). GABA_A receptors present in the tuberomammillary nucleus also play a key role in the sedative component of anesthesia. Microinjection of muscimol in the tuberomammillary nucleus produces sedation in rats and cats (Lin *et al.*, 1989; Nelson *et al.*, 2002), and systemic administration of muscimol, propofol, or pentobarbital decreases fos expression in the tuberomammillary nucleus (Nelson *et al.*, 2002).

Orexins directly excite the histaminergic neurons *in vitro* (Bayer *et al.*, 2001; Eriksson *et al.*, 2001a). Most histamine neurons express mRNAs and immunoreactivity for both orexin receptors (Eriksson *et al.*, 2001a; Marcus *et al.*, 2001), but the orexin 2 receptor seems mainly involved (Willie *et al.*, 2003). Orexins released from neurons emanating from the lateral hypothalamus enhance histamine neuron activity. Orexin levels are not altered by circadian time but their arousal effect depends on activation of histaminergic neurons (Huang *et al.*, 2001; Shigemoto *et al.*, 2004). In agreement, an altered histamine level was reported in orexin receptor-2-mutated dogs, an animal model of narcolepsy (Nishino *et al.*, 2001).

Several other systems activate or inhibit histamine neurons. Serotonin depolarizes histamine neurons and increases their firing rate *in vitro* by activation of 5-HT_{2C} receptors and an NCX1 Na⁺/Ca²⁺ exchanger (Eriksson *et al.*, 2001b). Several types of serotonergic receptors are likely to modulate histamine neuron activity *in vivo* (Laitinen *et al.*, 1995a; Morisset *et al.*, 1999; Oishi *et al.*, 1992). Ghrelin, a potent orexigenic peptide, activates histamine neurons by inhibiting G-protein-coupled inward rectifier K⁺ (GIRK) channels (Bajic *et al.*, 2004; Nakazato *et al.*, 2001). Intracellular recordings from rat hypothalamic slices indicate that morphine increases the firing of histaminergic neurons (Eriksson *et al.*, 2000), and histamine release in mouse cerebral cortex is enhanced by stimulation of κ -opioid receptors (Itoh *et al.*, 1988). Morphine and μ -opioid receptor agonists (Chikai *et al.*, 1994; Itoh *et al.*, 1988) enhance histamine release and turnover *in vivo*.

Inhibitory actions on histaminergic neurons have also been found. Besides GABA, nociceptin inhibits the firing of histaminergic neurons by activating GIRK channels (Eriksson *et al.*, 2000). [³H]Histamine synthesis and release are inhibited in various brain regions by stimulation of not only autoreceptors but also α_2 -adrenergic receptors, M₁-muscarinic receptors and κ -opioid receptors (Schwartz *et al.*, 1991). Since these regulations are also observed with synaptosomes, all these receptors presumably represent true presynaptic heteroreceptors. Agents inhibiting histamine release *in vitro* via stimulation of presynaptic α_2 -adrenergic receptors reduce histamine release and turnover *in vivo* (Gulat-Marnay *et al.*, 1989a; Prast *et al.*, 1991). A similar inhibition is induced by activation of muscarinic heteroreceptors

(Gulat-Marnay *et al.*, 1989b; Oishi *et al.*, 1990a; Prast *et al.*, 1994). Whether these heteroreceptors are tonically activated under basal conditions remains unclear: systemic administration of antagonists of these receptors does not enhance histamine turnover, but *in vivo* microdialysis studies show that their local perfusion increases histamine release (Laitinen *et al.*, 1995b; Prast *et al.*, 1994).

An increase of *tele*-methylhistamine levels with age has been reported in human cerebrospinal fluid (Prell *et al.*, 1990). Changes in histamine neuron activity also occur in various neuropsychiatric diseases. *tele*-Methylhistamine levels are increased in the cerebrospinal fluid of schizophrenic patients (Prell *et al.*, 1995). Histamine levels are unaffected in the brain of MPTP-treated mice (Cumming *et al.*, 1989) but increased in the brain of patients with Parkinson's disease, where they are associated with a strong increase in histaminergic innervation in the substantia nigra (Anichtchik *et al.*, 2000b; Rinne *et al.*, 2002). A reduced number of histamine neurons (Nakamura *et al.*, 1993) and a significant decrease of HDC activity and histamine levels in the hypothalamus, hippocampus, and cortex (Panula *et al.*, 1998; Schneider *et al.*, 1997) have been found in Alzheimer's brains. Similar deficits have been reported in frontal cortex of patients with Down syndrome (Schneider *et al.*, 1997).

E. PHYSIOLOGICAL ROLES OF HISTAMINERGIC NEURONS

Owing to the use of an increased number of experimental tools, for example, histamine H₃ receptor antagonists to activate histaminergic neurons (Arrang *et al.*, 1987), α -fluoromethylhistidine to block brain histamine synthesis (Garbarg *et al.*, 1980), mutant mice lacking L-histidine decarboxylase (Parmentier *et al.*, 2002) or the histamine H₁ receptor (Inoue *et al.*, 1996), the functional role of histaminergic neurons has been considerably clarified during the recent years.

Since our initial proposal in 1977 (Schwartz, 1977), a large variety of studies are now available to strengthen the view that the histaminergic system is one of the major neuronal systems controlling cortical activation and wakefulness (Lin, 2000). In agreement, ablation of these neurons, inhibition of histamine synthesis, release or action via the H₁ receptor all decrease wakefulness and increase deep slow-wave sleep; conversely, inhibition of histamine methylation or facilitation of histamine release via H₃ autoreceptor blockade all increase arousal. The major part played by the H₁ receptor in histamine-induced arousal, confirmed in mutant mice lacking this receptor (Inoue *et al.*, 1996), accounts for the sedating effects of the first generation of "antihistamines," that is, antagonists which easily enter the brain and are still ingredients of over-the-counter sleeping pills (Sangalli, 1997; Tashiro *et al.*, 2002).

The idea that activation of histaminergic neurons might improve cognitive performances is consistent with their projections to brain areas such as the

prefrontal and cingulate cortices or hippocampus, their projections to cholinergic perikarya, their excitatory influences therein, and their positive role in wakefulness. The procognitive and proattentional roles of histaminergic neuron activation were largely established in behavioral studies in rodents using thioperamide or other H_3 receptor inverse agonists (Hancock and Fox, 2004). For example, H_3 receptor antagonists/inverse agonists exert proattentional activity in a 5-trial acquisition test performed in spontaneously hypertensive rat pups, often considered as a model for attentional deficits and impulsivity in ADHD patients (Fox *et al.*, 2002). These effects of H_3 antagonists are reversed by H_1 antagonists, which suggests that the latter are attributable to enhanced histamine release. In agreement, H_3 receptor knockout mice display enhanced spatial learning and memory (Rizk *et al.*, 2004).

Histaminergic neurons affect secretion of several pituitary hormones and may also participate in the hormonal responses to stress (Knigge and Warberg, 1991; Schwartz *et al.*, 1991). They are activated during various forms of stress and heavily project to hypothalamic or limbic brain areas (e.g., amygdala or bed nucleus of the stria terminalis) involved in these responses. Furthermore, it seems that the activation of various subpopulations of histaminergic neurons within the tuberomammillary nucleus varies according to the nature of stressful stimuli (Ito, 2000; Miklos and Kovacs, 2003).

A satiating role of endogenous histamine is strongly suggested by several observations, although this view is not in agreement with all experimental data. Weight gain is often experienced by patients receiving first-generation H_1 antihistamines crossing the blood-brain barrier as well as antipsychotics or antidepressants displaying potent H_1 receptor antagonist properties. Central infusion of histamine reduces fat accumulation in leptin-resistant obese mice (Masaki *et al.*, 2001). Histamine neurons projecting to the hypothalamus may be responsible for the food intake suppression induced by the fat cell-produced hormone, leptin. In agreement, intracerebroventricular administration of leptin increases hypothalamic histamine release (Morimoto *et al.*, 2000), whereas histamine depletion by α -fluoromethylhistidine treatment attenuates leptin-induced feeding inhibition (Yoshimatsu *et al.*, 1999). Aged mice with gene disruption of either the H_1 receptor (Masaki *et al.*, 2004) or the histidine decarboxylase gene (Fülöp *et al.*, 2003) display adiposity; also both mice display hyperleptinemia, which suggests the existence of a regulatory loop between hypothalamic histamine neurons and leptin-producing cells, the nature of which remains elusive. Following the initial observation that the prototypical H_3 receptor antagonist/inverse agonist thioperamide decreases food intake in rats (Sakata *et al.*, 1990), studies using various compounds belonging to this drug class have confirmed that increased brain histamine release in rodents is associated with anorectic and antiobesity effects (Hancock, 2003). For instance, treatment of mice stabilized on a high-fat diet with A-331440, a potent nonimidazole H_3 receptor antagonist, decreased weight

similarly to dexfenfluramine, reduced body fat, and normalized insulin-resistance test (Hancock *et al.*, 2004). Nevertheless, not all H_3 receptor inverse agonists elicit such effects (Barbier *et al.*, 2004), thioperamide-induced appetite suppression was attributed to taste aversion to this compound (Sindelar *et al.*, 2004); in a mouse model of H_3 receptor disruption, a paradoxical increase in body weight, food intake, and adiposity, together with reductions in energy expenditure, has been reported (Takahashi *et al.*, 2002).

The anticonvulsant properties of endogenous histamine were initially suggested from the occurrence of seizures in epileptic patients, particularly children, following administration of high doses of H_1 receptor antagonists crossing the blood-brain barrier, even those devoid of anticholinergic activity (Sangalli, 1997). The role of histaminergic neurons in preventing seizures, or even the development of epileptogenesis, presumably, in most cases, via H_1 receptor activation, has been shown in several rodent models of epilepsy (Yokoyama, 2001). In agreement, drug-induced changes in histamine synthesis, release, or metabolism confirmed the role of the endogenous amine acting via the H_1 receptor in preventing seizure activity elicited in rodents by pentetrazole, transcranial electrical stimulation, or amygdaloid kindling. Consistently, H_3 antagonists inhibit amygdaloid-kindled seizures, an effect prevented by H_1 antagonists, which suggests the involvement of endogenous histamine (Kamei, 2001). These studies suggest that enhancing brain histamine release via H_3 receptor blockade should represent a novel therapeutic approach for epilepsies.

III. Changes in the Histaminergic System in Schizophrenia

A. GENETIC STUDIES

HMT may be an important target to modulate histaminergic neurotransmission in neuropsychiatric disorders. Among the various HMT polymorphisms that have been identified in the human gene (Aksoy *et al.*, 1996; Chen *et al.*, 2003a; Wang *et al.*, 2002), a common C314T transition located in exon 4 results in decreased levels of enzyme activity (Chen *et al.*, 2003a; Preuss *et al.*, 1998) but was not associated with schizophrenia (Yan *et al.*, 2000).

The human H_1 receptor gene contains an intron in the 5'-flanking untranslated region, close to the translation initiation codon, but the translated region is intronless (De Backer *et al.*, 1998). Several polymorphisms have been identified in the promoter and coding region, but none of them was found to be associated with schizophrenia or response to clozapine (Hong *et al.*, 2002; Mancama *et al.*, 2000, 2002).

The 5'-untranslated region of the human H_2 receptor gene contains an intron but the translated region is intronless. An initial study by Orange *et al.* (1996) has

reported six polymorphisms of the coding region in a UK population. Among them, a A649G transition was found to be associated with schizophrenia. However, none of these variants could be detected in other studies. Several other polymorphisms have been identified in various ethnic groups in the promoter region and one (543G/A) in the coding region of the gene, but none of them was found to be associated with schizophrenia or response to clozapine (Fukushima *et al.*, 2001; Ito *et al.*, 2000; Mancama *et al.*, 2000, 2002).

B. HISTAMINE NEURON ACTIVITY

Perfusion of the rat posterior hypothalamus, in which histaminergic cell bodies are located, with dopamine D2 receptor agonists enhances histamine release *in vivo* (Prast *et al.*, 1993). In addition, methamphetamine, a psychotogenic drug which enhances dopamine release in schizophrenic patients (Laruelle *et al.*, 1996), was shown to enhance histamine release in dialysates of rat striatum. This response was completely blocked by haloperidol, an antagonist at dopamine D2-like receptors (Ito *et al.*, 1996). Furthermore, the behavioral sensitization to dopamine agonists, a cardinal feature of schizophrenia, observed following repeated administration of methamphetamine was accompanied by an enhanced basal histamine release in rat striatum. This effect was again blocked by haloperidol, reflecting an increased tonic dopaminergic influence on histaminergic neurons (Ito *et al.*, 1996).

Consistent with these findings on histamine release, methamphetamine also enhances histaminergic neuron activity in mouse brain both acutely and in a long-term fashion. A single administration of methamphetamine markedly increases *tele*-MeHA levels, an index of histamine neuron activity, in the cerebral cortex, striatum, and hypothalamus (Morisset *et al.*, 2002). This enhancing effect of methamphetamine on *tele*-MeHA levels results from the stimulation of histaminergic neurons by endogenous dopamine activating selectively D2 receptors. In agreement, this effect was completely blocked by haloperidol, a D2/D3 receptor antagonist, but remained unchanged either after administration of nafadotride used at a dose inducing a selective blockade of the D3 receptor (Sautel *et al.*, 1995), or in the brain of mice lacking functional D3 receptors. Using [¹²⁵I]iodosulpride as a ligand, D2-like receptor-binding sites have been evidenced by autoradiography at the level of the tuberomammillary nucleus (Bouthenet *et al.*, 1987), an area in which D3 receptors could not be detected (J. Diaz, personal communication). Therefore, endogenous dopamine may directly activate histamine neurons by interacting with D2 receptors located on their perikarya or dendrites (Morisset *et al.*, 2002), as also supported by retrograde tracing studies combined with immunohistochemistry showing that dopamine-containing axons emanating from the ventral tegmental area or substantia nigra

project to the tuberomammillary nucleus (Ericson *et al.*, 1989). D2 receptors located on histaminergic nerve endings do not seem to be involved since apomorphine fails to significantly affect histamine release from slices of rat cerebral cortex (Schwartz *et al.*, 1990).

Basal *tele*-MeHA levels were enhanced in various brain regions of sensitized mice showing that repeated administration of methamphetamine induces a long-lasting enhancement of histaminergic neuron activity in the whole brain (Morisset *et al.*, 2002), which is consistent with the increase in histamine release observed in the striatum of sensitized rats (Ito *et al.*, 1996). Like the response to acute administration, this effect of chronic treatment with methamphetamine on *tele*-MeHA levels was blocked by haloperidol, strongly suggesting that it resulted from a sensitized release of dopamine from dopaminergic afferents, leading to a higher degree of activation of D2 receptors present on histaminergic neurons.

In addition to the dopaminergic hypothesis of psychotic disorders, a hypoactivity of glutamatergic transmission has been implicated in schizophrenia. Initially, this glutamatergic hypothesis of schizophrenia was mainly based on the schizophrenia-like psychotomimetic effects of phencyclidine (PCP), which are now mainly attributed to noncompetitive antagonism of the *N*-methyl-D-aspartate (NMDA) receptor (Javitt and Zukin, 1991; Jentsch and Roth, 1999). Consistent with a hyperactivity of histamine neurons in psychotic disorders, it was shown that PCP, in a dose range of 2–10 mg/kg, significantly enhances *tele*-MeHA levels in various mouse brain regions (Itoh *et al.*, 1985, 1986). It was initially suggested that this enhancing effect of PCP on histamine neuron activity involved stimulation of opiate receptors via release of endogenous opioid peptides because it was antagonized by a large dose of naloxone (Itoh *et al.*, 1987) and because direct stimulation of μ -opioid receptors had been shown to increase histamine turnover in the mouse brain (Itoh *et al.*, 1988; Nishibori *et al.*, 1985). However, although it was initially reported to interact with a large number of molecular target sites, PCP was subsequently found to act as a more potent noncompetitive antagonist of the NMDA receptor (Anis *et al.*, 1983) and many data demonstrate that blockade of the NMDA channel is the primary mechanism involved in the effects elicited by PCP (Javitt and Zukin, 1991; Jentsch and Roth, 1999). It could therefore be predicted from the previous studies with PCP that NMDA receptor blockade could activate histamine neurons. Consistent with this proposal, we showed that the effect of PCP is mimicked by MK-801, another NMDA open-channel blocker displaying high potency and selectivity (Wong *et al.*, 1986). MK-801 administration results in an enhancement of *tele*-MeHA levels in the same range as that elicited by PCP and which occurred with a low ED₅₀ value (~ 0.1 mg/kg; Faucard *et al.*, 2006). In addition, a significant increase in *hdc* mRNA expression is induced by PCP administration both in the rostral and caudal parts of the tuberomammillary nucleus (Faucard *et al.*, 2006). Therefore, the strong enhancement of *tele*-MeHA levels and *hdc*-mRNA expression

induced in rodent brain not only by methamphetamine but also by NMDA receptor antagonists further supports the existence of a hyperactivity of histamine neurons in psychotic disorders.

Consistent with this proposal, Prell *et al.* (1995) showed that *tele*-MeHA levels were significantly elevated in the cerebrospinal fluid of patients with chronic schizophrenia, either under neuroleptic treatment or not, indicating that hyperactivity of dopaminergic transmission is associated with an enhanced activity of histaminergic neurons in the disease (Prell *et al.*, 1995). In addition, the decrease in H_1 receptor-mediated responses consistently observed in schizophrenic patients (Nakai *et al.*, 1991; Rauscher *et al.*, 1980) is likely to result from the down-regulation of postsynaptic H_1 receptors induced by the chronic increase in histamine release. In agreement, positron emission tomography (PET) studies using [^{11}C]doxepin revealed a significant decrease in H_1 receptor binding in the frontal and prefrontal cortices and the cingulate gyrus brain of schizophrenic patients (Iwabuchi *et al.*, 2005).

IV. Interactions of Antipsychotic Drugs with the Histaminergic System

A. INTERACTIONS OF APDs WITH HISTAMINE RECEPTORS

A large number of antipsychotics are potent H_1 receptor antagonists and block [3H]mepyramine binding to the receptor in rodent and human brain at sub-therapeutic dosages (Quach *et al.*, 1979; Richelson and Souder, 2000). The major part played by the H_1 receptor in the arousal induced by histamine neurons suggests that this blockade of H_1 receptors in brain is involved in the sedative side-effects of many antipsychotic drugs (APDs). In addition, the inhibitory role of endogenous histamine on food intake mediated by the H_1 receptor, namely on the ventromedial nucleus (Sakata *et al.*, 1997), probably accounts for the weight gain that is often experienced by patients receiving antipsychotics displaying potent H_1 -receptor antagonist properties. This eventually results in an increased risk of developing a "metabolic syndrome" in patients chronically treated with such agents (Kroeze *et al.*, 2003; Richelson and Souder, 2000). Assuming that blockade of the dopamine D2 receptor is the mechanism of action of APDs, the ratio of K_i 's to this receptor and the H_1 receptor of various compounds, which varies largely (Table II), should reflect their ability to block the H_1 receptor at therapeutic dosages and, thereby, exert sedative and probesity side effects. Hence, for instance, olanzapine is one of the most potent H_1 receptor antagonist known so far (Richelson and Souder, 2000) and its marked sedative and weight-gain side-effects are well established. In agreement, sedative APDs at therapeutic dosages were shown to occupy a significant fraction

TABLE II
COMPARED POTENCIES (K_i , nM) OF SEVERAL APDs AT HUMAN HISTAMINE H_1 AND
DOPAMINE D2 RECEPTORS

Compounds	Dopamine D2	Histamine H_1	Ratio D2/ H_1
Clozapine	210	3.1	68
Haloperidol	2.6	260	0.01
Olanzapine	20	0.087	230
Quetiapine	770	19	40
Risperidone	3.8	5.2	0.7
Sertindole	2.7	320	0.01
Ziprasidone	2.6	4.6	0.6

Values are derived from Richelson and Souder (2000).

of cerebral H_1 receptors in living rodents (Quach *et al.*, 1979) and weight-gain propensity of several APDs in patients were significantly correlated to their relative potencies at these receptors (Kroeze *et al.*, 2003).

An intriguing observation is that several APDs and a number of tricyclic antidepressants are very potent and competitive inhibitors of the H_2 receptor-linked adenylyl cyclase on brain membranes (Green and Maayani, 1977; Green *et al.*, 1977; Kanof and Greengard, 1978). This has led to the suggestion that blockade of the H_2 receptor might account at least partially for the clinical activity. However, for unclear reasons, such a potent blockade could not be observed on intact cell preparations (Dam Trung Tuong *et al.*, 1980).

Whereas typical APDs were ineffective at the H_3 receptor, the atypical APD clozapine has been shown to block the rodent H_3 receptor as evidenced on the release of noradrenaline or serotonin from brain slices and in radioligand binding assays to the recombinant or native receptor (Alves-Rodrigues *et al.*, 1996; Kathmann *et al.*, 1994; Lovenberg *et al.*, 2000; Morisset *et al.*, 1999). This led to the speculation that some of its "atypical" properties might be due to H_3 receptor antagonism. However, consistent with a species-related heterogeneity of H_3 receptors (Schwartz *et al.*, 2001), clozapine does not significantly interact with the recombinant human H_3 receptor (Lovenberg *et al.*, 2000).

Clozapine displays a submicromolar affinity at the human H_4 R (de Esch *et al.*, 2005), but plasma and brain concentrations associated with clinical responses meet or exceed these values. Even more interesting is that clozapine, which acts as an antagonist at various receptors, behaves as a full agonist at the recombinant human H_4 R and at the H_4 R present on human eosinophils (Buckland *et al.*, 2003; Liu *et al.*, 2001a,b; Oda *et al.*, 2000). Because the H_4 R is mainly expressed on hematopoietic cells, one might therefore speculate that agranulocytosis, which often limits clozapine effectiveness, is related to H_4 R activation.

B. MODULATION OF HISTAMINE NEURON ACTIVITY BY APDs

Consistent with a tonic stimulation of histamine neurons by endogenous dopamine interacting with D2 receptors present on histaminergic cell bodies, typical APDs, for example, haloperidol, decrease histamine neuron activity (Morisset *et al.*, 1999).

In contrast, atypical neuroleptics, for example, clozapine, enhance histamine turnover, an effect related to 5-HT₂ receptor blockade. Ketanserin, a preferential 5-HT_{2A} receptor antagonist, mimicked the enhancing effect of atypical antipsychotics on *tele*-MeHA levels in mouse cerebral cortex, striatum, and hypothalamus. DOI, a 5-HT_{2A/2C} receptor agonist, did not modify *tele*-MeHA level but strongly reversed the effect of clozapine (Morisset *et al.*, 1999). These findings therefore show that endogenous serotonin tonically inhibits HA neurons via 5-HT_{2A} receptors, an effect blocked by clozapine. These 5-HT_{2A} receptors could be located on HA neurons themselves, on interneurons, or nearby axon terminals impinging on the formers. In agreement with the blockade of 5-HT_{2A} receptors by atypical neuroleptics, the effect of clozapine was not additive with that of ketanserin. This strongly suggests that the activation of histaminergic neurons by clozapine (and other novel antipsychotics) is entirely attributable to 5-HT_{2A} receptor blockade (Morisset *et al.*, 1999).

A recognized advantage of atypical APDs compared to typical APDs is their arousing and procognitive effects resulting in a significant efficacy against negative symptomatology. The positive functional role attributed to histaminergic neurons in wakefulness, attention, and cognition suggests that this property of atypical antipsychotics could be related at least partially to their unique ability to activate histamine neurons.

V. Role of Histaminergic Neurons in Schizophrenia

Overdose of a variety of classical H₁-antagonists was repeatedly reported to result in toxic psychoses with hallucinations resembling schizophrenia and the hallucinogenic potential of these drugs has even led to abuse (Sangalli, 1997). The increase in dopamine release and the blockade of dopamine uptake induced by such compounds in the striatum, rather than blockade of H₁ receptors, presumably explain their abuse potential (Dringenberg *et al.*, 1998). In agreement, dopamine turnover remains unchanged in the forebrain from H₁ receptor knockout mice (Yanai *et al.*, 1998).

In several open-label clinical trials, famotidine, an H₂R antagonist with limited brain penetration, was found to display an antipsychotic efficacy (Kaminsky

et al., 1990; Oyewumi *et al.*, 1994; Rosse *et al.*, 1996), a finding which remains to be confirmed in controlled studies.

In the search of new antipsychotic agents, increasing evidence supports the therapeutic potential of H₃ receptor antagonists/inverse agonists for the symptomatic treatment of schizophrenia. The latter do not change spontaneous locomotor activity when they are used alone (Clapham and Kilpatrick, 1994; Imaizumi and Onodera, 1993; Pillot *et al.*, 2002b) and do not induce locomotor sensitization (Komater *et al.*, 2003). However, the locomotor activation elicited in rat and mouse by various dopaminergic agonists such as amphetamine, methamphetamine, and apomorphine is attenuated by thioperamide and ciproxifan, two standard H₃ receptor antagonists/inverse agonists (Clapham and Kilpatrick, 1994; Morisset *et al.*, 2002). Also ciproxifan, a potent H₃ receptor antagonist, significantly decreases the stereotypies induced in mice by methamphetamine and apomorphine (Sadakhom, C.; Frances, H., and Arrang, J. M. in preparation). Consistent with these findings, the effect of methamphetamine on locomotor activity and stereotypic behavior was less pronounced in H₃ receptor knockout mice (Toyota *et al.*, 2002).

In another animal model of psychosis (Andine *et al.*, 1999; Carlsson and Carlsson, 1990), the locomotor hyperactivity induced in rodents by the NMDA receptor antagonist MK-801 is also markedly attenuated by H₃ receptor antagonists/inverse agonists (Faucard *et al.*, 2006).

In DBA/2 mice in which sensorimotor-gating deficits, which are considered as cardinal signs of the disease, occur naturally, H₃ receptor antagonists/inverse agonists also improve gating as shown by the increase that they induce in prepulse inhibition of startle and N40 auditory-evoked response (Browman *et al.*, 2004; Fox *et al.*, 2005).

The neurochemical mechanisms underlying these antipsychotic-like effects induced by H₃ receptor blockade remain unknown. Facilitation of histamine release via H₃ autoreceptor blockade may be involved. However, a histamine neuron hyperactivity being already observed in schizophrenia, this would imply that histaminergic neurons are not directly involved in psychotic symptoms but are involved in a compensatory manner. The further enhancement of histamine neuron activity induced by H₃ receptor antagonists/inverse agonists would therefore attenuate psychotic symptoms. Consistent with such a hypothesis, the time course of hyperlocomotion and activation of histamine neurons induced by methamphetamine do not parallel (Morisset *et al.*, 2002), and enhancement of histamine release induced by histidine loads or inhibitors of histamine catabolism have also been reported to reduce methamphetamine-induced locomotor activity (Itoh *et al.*, 1984; Ito *et al.*, 1997).

Alternatively, a direct involvement of histamine in schizophrenic symptoms cannot be ruled out. H₃ receptors are present at high densities on many perikarya

and/or dendrites of intrinsic neurons in the cerebral cortex, basal ganglia, and limbic areas (Pillot *et al.*, 2002a,b). Therefore, the hyperactivity of histamine neurons reported in schizophrenia and in animal models of the disease might enhance the activation of these postsynaptic H₃ receptors. Their blockade would lead to antipsychotic properties of H₃ receptor antagonists.

Whatever the mechanisms involved, recent data show the existence of strong but complex functional interactions between endogenous histamine and dopamine in the brain. Ciproxifan used alone has no effect but strongly modulates the effects of methamphetamine on neuropeptide mRNA expression not only in the caudate-putamen but also in the nucleus accumbens (Pillot *et al.*, 2003). The synergism between the two drugs on enkephalin neurons and their antagonism on substance P/dynorphin neurons may suggest direct interactions between H₃ receptors and dopamine receptors. H₃ receptor activation inhibits dopamine D₁ receptor-mediated cAMP formation in the rat striatum (Sanchez-Lemus and Arias-Montano, 2004) and the expression of H₃ receptors is influenced by endogenous activation of D₁ receptors (Ryu *et al.*, 1996). Presynaptic H₃ heteroreceptors inhibit dopamine synthesis (Molina-Hernandez *et al.*, 2000) and release (Schlicker *et al.*, 1993). Although they do not appear to regulate dopamine neuron activity *in vivo* under basal conditions (Imaizumi and Onodera, 1993; Miyazaki *et al.*, 1997; Oishi *et al.*, 1990b), the inhibition of dopamine neuron activity by H₃ heteroreceptors may become operating in schizophrenia because of an enhanced histamine release, as shown by the potentiation of methamphetamine-induced accumbal dopamine release induced by H₃ receptor antagonists/inverse agonists (Munzar *et al.*, 2004). In addition, in freely moving rat microdialysis studies, H₃ receptor antagonists do not enhance dopamine release in striatum but enhance it in frontal cortex (Fox *et al.*, 2005).

Both pharmacological interactions between H₃ and D2 receptors and pharmacokinetic drug-drug interactions may account for the complex interactions reported between H₃ receptor antagonists/inverse agonists and neuroleptics. In one study, the imidazole derivative ciproxifan potentiated the enkephalin, neurotensin, and c-fos expression induced in rat caudate-putamen and nucleus accumbens (Pillot *et al.*, 2002b). By contrast, thioperamide, another imidazole compound, decreased haloperidol-induced c-fos expression in the rat dorsolateral striatum but not in the nucleus accumbens (Hussain *et al.*, 2002). Similar discrepancies were also found in behavioral studies. Ciproxifan and thioperamide potentiated haloperidol-induced catalepsy in the rat (Pillot *et al.*, 2002b; Zhang *et al.*, 2005) but not in the mouse (Morisset *et al.*, 1999). In the rat, the potentiation of catalepsy was likely to result at least partially from an inhibition of cytochrome P450 enzymes by imidazole derivatives (Yang *et al.*, 2002), and two nonimidazole H₃ receptor antagonists/inverse agonists tended to attenuate risperidone-induced catalepsy (Zhang *et al.*, 2005).

VI. Conclusions

The present chapter testifies how our knowledge of the molecular neurobiology of cerebral histaminergic systems and their implications in physiological functions, for example, arousal, cognition, or control of food intake, has progressed during the last years. This appears as the result of the development of reliable research tools such as selective ligands for the various receptor subtypes or genetically modified mice. Recent findings support the possible implication of the histaminergic system in schizophrenia and therapeutic utility and/or side effects of APDs. H_3 receptor antagonists/inverse agonists raise a great interest as innovative therapeutics in various CNS disorders including schizophrenia and are currently undergoing clinical trials. The results of these clinical studies are now awaited to confirm this potential interest and they should teach us a lot about the role of the histaminergic system in the human brain.

References

- Abe, H., Honma, S., Ohtsu, H., and Honma, K. (2004). Circadian rhythms in behavior and clock gene expressions in the brain of mice lacking histidine decarboxylase. *Brain Res. Mol. Brain Res.* **124**, 178–187.
- Airaksinen, M. S., Paetau, A., Paljarvi, L., Reinikainen, K., Riekkinen, P., Suomalainen, R., and Panula, P. (1991). Histamine neurons in human hypothalamus: Anatomy in normal and Alzheimer diseased brains. *Neuroscience* **44**, 465–481.
- Airaksinen, M. S., Alanen, S., Szabat, E., Visser, T. J., and Panula, P. (1992). Multiple neurotransmitters in the tuberomammillary nucleus: Comparison of rat, mouse, and guinea pig. *J. Comp. Neurol.* **323**, 103–116.
- Aksoy, S., Raftogianis, R., and Weinshilboum, R. (1996). Human histamine *N*-methyltransferase gene: Structural characterization and chromosomal location. *Biochem. Biophys. Res. Commun.* **219**, 548–554.
- Alves-Rodrigues, A., Leurs, R., Willems, E., and Timmerman, H. (1996). Binding of clozapine metabolites and analogues to the histamine H_3 receptor in rat brain cortex. *Arch. Pharm. (Weinheim)* **329**, 413–416.
- Andine, P., Widermark, N., Axelsson, R., Nyberg, G., Olofsson, U., Martensson, E., and Sandberg, M. (1999). Characterization of MK-801-induced behavior as a putative rat model of psychosis. *J. Pharmacol. Exp. Ther.* **290**, 1393–1408.
- Anichtchik, O. V., Huotari, M., Peitsaro, N., Haycock, J. W., Mannisto, P. T., and Panula, P. (2000a). Modulation of histamine H_3 receptors in the brain of 6-hydroxydopamine-lesioned rats. *Eur. J. Neurosci.* **12**, 3823–3832.
- Anichtchik, O. V., Rinne, J. O., Kalimo, H., and Panula, P. (2000b). An altered histaminergic innervation of the substantia nigra in Parkinson's disease. *Exp. Neurol.* **163**, 20–30.
- Anichtchik, O. V., Peitsaro, N., Rinne, J. O., Kalimo, H., and Panula, P. (2001). Distribution and modulation of histamine H_3 receptors in basal ganglia and frontal cortex of healthy controls and patients with Parkinson's disease. *Neurobiol. Dis.* **8**, 707–716.

- Anis, N. A., Berry, S. C., Burton, N. R., and Lodge, D. (1983). The dissociative anaesthetics, ketamine and phencyclidine, selectively reduce excitation of central mammalian neurones by N-methyl-aspartate. *Br. J. Pharmacol.* **79**, 565–575.
- Arias-Montano, J. A., Floran, B., Garcia, M., Aceves, J., and Young, J. M. (2001). Histamine H₃ receptor-mediated inhibition of depolarization-induced, dopamine D₁ receptor-dependent release of [³H]-gamma-aminobutyric acid from rat striatal slices. *Br. J. Pharmacol.* **133**, 165–171.
- Arrang, J. M., Garbarg, M., and Schwartz, J. C. (1983). Auto-inhibition of brain histamine release mediated by a novel class (H₃) of histamine receptor. *Nature* **302**, 832–837.
- Arrang, J. M., Garbarg, M., and Schwartz, J. C. (1985). Autoregulation of histamine release in brain by presynaptic H₃-receptors. *Neuroscience* **15**, 553–562.
- Arrang, J. M., Garbarg, M., Lancelot, J. C., Lecomte, J. M., Pollard, H., Robba, M., Schunack, W., and Schwartz, J. C. (1987). Highly potent and selective ligands for histamine H₃-receptors. *Nature* **327**, 117–123.
- Arrang, J. M., Devaux, B., Chodkiewicz, J. P., and Schwartz, J. C. (1988). H₃-receptors control histamine release in human brain. *J. Neurochem.* **51**, 105–108.
- Atack, C., and Carlsson, A. (1972). *In vitro* release of endogenous histamine, together with noradrenaline and 5-hydroxytryptamine, from slices of mouse cerebral hemispheres. *J. Pharm. Pharmacol.* **24**, 990–992.
- Bajic, D., VanManh Hoang, Q., Nakajima, S., and Nakajima, Y. (2004). Dissociated histaminergic neuron cultures from the tuberomammillary nucleus of rats: Culture methods and ghrelin effects. *J. Neurosci. Methods* **132**, 177–184.
- Barbier, A. J., Berridge, C., Dugovic, C., Laposky, A. D., Wilson, S. J., Boggs, J., Aluisio, L., Lord, B., Mazur, C., Pudiak, C. M., Langlois, X., Xiao, W., *et al.* (2004). Acute wake-promoting actions of JNJ-5207852, a novel, diamine-based H₃ antagonist. *Br. J. Pharmacol.* **143**, 649–661.
- Baudry, M., Martres, M. P., and Schwartz, J. C. (1975). H₁ and H₂ receptors in the histamine-induced accumulation of cyclic AMP in guinea pig brain slices. *Nature* **253**, 362–364.
- Bayer, L., Eggermann, E., Serafin, M., Saint-Mieux, B., Machard, D., Jones, B., and Muhlethaler, M. (2001). Orexins (hypocretins) directly excite tuberomammillary neurons. *Eur. J. Neurosci.* **14**, 1571–1575.
- Bayliss, D. A., Wang, Y. M., Zahnow, C. A., Joseph, D. R., and Millhorn, D. E. (1990). Localization of histidine decarboxylase mRNA in rat brain. *Mol. Cell. Neurosci.* **1**, 3–9.
- Bouthenet, M. L., Martres, M. P., Sales, N., and Schwartz, J. C. (1987). A detailed mapping of dopamine D₂ receptors in rat central nervous system by autoradiography with [¹²⁵I]iodosulpride. *Neuroscience* **20**, 117–155.
- Browman, K. E., Komater, V. A., Curzon, P., Rueter, L. E., Hancock, A. A., Decker, M. W., and Fox, G. B. (2004). Enhancement of prepulse inhibition of startle in mice by the H₃ receptor antagonists thioperamide and ciproxifan. *Behav. Brain Res.* **153**, 69–76.
- Brown, R. E., and Reymann, K. G. (1996). Histamine H₃ receptor-mediated depression of synaptic transmission in the dentate gyrus of the rat *in vitro*. *J. Physiol.* **496**, 175–184.
- Brown, R. E., and Haas, H. L. (1999). On the mechanism of histaminergic inhibition of glutamate release in the rat dentate gyrus. *J. Physiol.* **515**, 777–786.
- Brown, R. E., Stevens, D. R., and Haas, H. L. (2001). The physiology of brain histamine. *Prog. Neurobiol.* **63**, 637–672.
- Buckland, K. F., Williams, T. J., and Conroy, D. M. (2003). Histamine induces cytoskeletal changes in human eosinophils via the H₄ receptor. *Br. J. Pharmacol.* **140**, 1117–1127.
- Carlsson, M., and Carlsson, A. (1990). Interactions between glutamatergic and monoaminergic systems within the basal ganglia—implications for schizophrenia and Parkinson's disease. *Trends Neurosci.* **13**, 272–276.

- Chemelli, R. M., Willie, J. T., Sinton, C. M., Elmquist, J. K., Scammell, T., Lee, C., Richardson, J. A., Williams, S. C., Xiong, Y., Kisanuki, Y., Fitch, T. E., Nakazato, M., *et al.* (1999). Narcolepsy in orexin knockout mice: Molecular genetics of sleep regulation. *Cell* **98**, 437–451.
- Chen, G. L., Wang, H., Wang, W., Xu, Z. H., Zhou, G., He, F., and Zhou, H. H. (2003a). Histamine N-methyltransferase gene polymorphisms in Chinese and their relationship with enzyme activity in erythrocytes. *Pharmacogenetics* **13**, 389–397.
- Chen, Z., Li, Z., Sakurai, E., Izadi Mobarakeh, J., Ohtsu, H., Watanabe, T., Iinuma, K., and Yanai, K. (2003b). Chemical kindling induced by pentylentetrazol in histamine H₁ receptor gene knockout mice (H₁KO), histidine decarboxylase-deficient mice (HDC(–/–)) and mast cell-deficient W/W(v) mice. *Brain Res.* **968**, 162–166.
- Chikai, T., Oishi, R., and Saeki, K. (1993). Microdialysis study of the effects of sedative drugs on extracellular histamine in the striatum of freely moving rats. *J. Pharmacol. Exp. Ther.* **266**, 1277–1281.
- Chikai, T., Oishi, R., and Saeki, K. (1994). Increase in the extracellular histamine concentration in the rat striatum by *mu*-opioid receptor activation. *J. Neurochem.* **62**, 724–729.
- Chou, T. C., Bjorkum, A. A., Gaus, S. E., Lu, J., Scammell, T. E., and Saper, C. B. (2002). Afferents to the ventrolateral preoptic nucleus. *J. Neurosci.* **22**, 977–990.
- Clapham, J., and Kilpatrick, G. J. (1994). Thioperamide, the selective histamine H₃ receptor antagonist, attenuates stimulant-induced locomotor activity in the mouse. *Eur. J. Pharmacol.* **259**, 107–114.
- Coge, F., Guenin, S. P., Rique, H., Boutin, J. A., and Galizzi, J. P. (2001). Structure and expression of the human histamine H₄-receptor gene. *Biochem. Biophys. Res. Commun.* **284**, 301–309.
- Cumming, P., Jakubovic, A., and Vincent, S. R. (1989). Cerebral histamine levels are unaffected by MPTP administration in the mouse. *Eur. J. Pharmacol.* **166**, 299–301.
- Cumming, P., Shaw, C., and Vincent, S. R. (1991). High affinity histamine binding site is the H₃ receptor: characterization and autoradiographic localization in rat brain. *Synapse* **8**, 144–151.
- Dam Trung Tuong, M., Garbarg, M., and Schwartz, J. C. (1980). Pharmacological specificity of brain histamine H₂-receptors differs in intact cells and cell-free preparations. *Nature* **287**, 548–551.
- De Backer, M. D., Loonen, I., Verhasselt, P., Neefs, J. M., and Luyten, W. H. (1998). Structure of the human histamine H₁ receptor gene. *Biochem. J.* **335**, 663–670.
- de Esch, I. J., Thurmond, R. L., Jongejan, A., and Leurs, R. (2005). The histamine H₄ receptor as a new therapeutic target for inflammation. *Trends Pharmacol. Sci.* **26**, 462–469.
- Dere, E., De Souza-Silva, M. A., Topic, B., Spieler, R. E., Haas, H. L., and Huston, J. P. (2003). Histidine-decarboxylase knockout mice show deficient nonreinforced episodic object memory, improved negatively reinforced water-maze performance, and increased neo- and ventro-striatal dopamine turnover. *Learn. Mem.* **10**, 510–519.
- Dere, E., De Souza-Silva, M. A., Spieler, R. E., Lin, J. S., Ohtsu, H., Haas, H. L., and Huston, J. P. (2004). Changes in motoric, exploratory and emotional behaviours and neuronal acetylcholine content and 5-HT turnover in histidine decarboxylase-KO mice. *Eur. J. Neurosci.* **20**, 1051–1058.
- D'Este, L., Wimalawansa, S. J., and Renda, T. G. (2001). Distribution of amylin-immunoreactive neurons in the monkey hypothalamus and their relationships with the histaminergic system. *Arch. Histol. Cytol.* **64**, 295–303.
- Doreulee, N., Yanovsky, Y., Flagmeyer, I., Stevens, D. R., Haas, H. L., and Brown, R. E. (2001). Histamine H₃ receptors depress synaptic transmission in the corticostriatal pathway. *Neuropharmacology* **40**, 106–113.
- Dringenberg, H. C., de Souza-Silva, M. A., Schwarting, R. K., and Huston, J. P. (1998). Increased levels of extracellular dopamine in neostriatum and nucleus accumbens after histamine H₁ receptor blockade. *Naunyn Schmiedebergs Arch. Pharmacol.* **358**, 423–429.

- Drutel, G., Peitsaro, N., Karlstedt, K., Wieland, K., Smit, M. J., Timmerman, H., Panula, P., and Leurs, R. (2001). Identification of rat H₃ receptor isoforms with different brain expression and signaling properties. *Mol. Pharmacol.* **59**, 1–8.
- Ericson, H., Watanabe, T., and Kohler, C. (1987). Morphological analysis of the tuberomammillary nucleus in the rat brain: Delineation of subgroups with antibody against L-histidine decarboxylase as a marker. *J. Comp. Neurol.* **263**, 1–24.
- Ericson, H., Blomqvist, A., and Kohler, C. (1989). Brainstem afferents to the tuberomammillary nucleus in the rat brain with special reference to monoaminergic innervation. *J. Comp. Neurol.* **281**, 169–192.
- Ericson, H., Blomqvist, A., and Kohler, C. (1991a). Origin of neuronal inputs to the region of the tuberomammillary nucleus of the rat brain. *J. Comp. Neurol.* **311**, 45–64.
- Ericson, H., Kohler, C., and Blomqvist, A. (1991b). GABA-like immunoreactivity in the tuberomammillary nucleus: An electron microscopic study in the rat. *J. Comp. Neurol.* **305**, 462–469.
- Eriksson, K. S., Stevens, D. R., and Haas, H. L. (2000). Opposite modulation of histaminergic neurons by nociceptin and morphine. *Neuropharmacology* **39**, 2492–2498.
- Eriksson, K. S., Sergeeva, O., Brown, R. E., and Haas, H. L. (2001a). Orexin/hypocretin excites the histaminergic neurons of the tuberomammillary nucleus. *J. Neurosci.* **21**, 9273–9279.
- Eriksson, K. S., Stevens, D. R., and Haas, H. L. (2001b). Serotonin excites tuberomammillary neurons by activation of Na⁺/Ca²⁺-exchange. *Neuropharmacology* **40**, 345–351.
- Esbenshade, T. A., Fox, G. B., Krueger, K. M., Miller, T. R., Kang, C. H., Denny, L. I., Witte, D. G., Yao, B. B., Pan, L., Wetter, J., Marsh, K., Bennani, Y. L., *et al.* (2005). Pharmacological properties of ABT-239 [4-(2-{2-[(2R)-2-methylpyrrolidinyl]ethyl}-benzofuran-5-yl)benzonitrile]: I. Potent and selective histamine H₃ receptor antagonist with drug-like properties. *J. Pharmacol. Exp. Ther.* **313**, 165–175.
- Faucard, R., Armand, V., Héron, A., Cochois, V., and Arrang, J. M. (2006). N-methyl-D-aspartate receptor antagonists enhance histamine neuron activity in rodent brain. *J. Neurochem.* **98**, 1487–1496.
- Fekete, C., and Liposits, Z. (2003). Histamine-immunoreactive neurons of the tuberomammillary nucleus are innervated by alpha-melanocyte stimulating hormone-containing axons. Generation of a new histamine antiserum for ultrastructural studies. *Brain Res.* **969**, 70–77.
- Fox, G. B., Pan, J. B., Esbenshade, T. A., Bennani, Y. L., Black, L. A., Faghih, R., Hancock, A. A., and Decker, M. W. (2002). Effects of histamine H₃ receptor ligands GT-2331 and ciproxifan in a repeated acquisition avoidance response in the spontaneously hypertensive rat pup. *Behav. Brain Res.* **131**, 151–161.
- Fox, G. B., Esbenshade, T. A., Pan, J. B., Radek, R. J., Krueger, K. M., Yao, B. B., Browman, K. E., Buckley, M. J., Ballard, M. E., Komater, V. A., Miner, H., Zhang, M., *et al.* (2005). Pharmacological properties of ABT-239 [4-(2-{2-[(2R)-2-methylpyrrolidinyl]ethyl}-benzofuran-5-yl)benzonitrile]: II. Neurophysiological characterization and broad preclinical efficacy in cognition and schizophrenia of a potent and selective histamine H₃ receptor antagonist. *J. Pharmacol. Exp. Ther.* **313**, 176–190.
- Fukushima, Y., Saitoh, T., Anai, M., Tsukuda, K., Onishi, Y., Sakoda, H., Inukai, K., Ogihara, T., Funaki, M., Ono, H., Fujishiro, M., Ishikawa, T., *et al.* (2001). G649, an allelic variant of the human H₂ receptor with low basal activity, is resistant to upregulation upon antagonist exposure. *Pharmacogenomics J.* **1**, 78–83.
- Fülop, A. K., Foldes, A., Buzas, E., Hegyi, K., Miklos, I. H., Romics, L., Kleiber, M., Nagy, A., Falus, A., and Kovacs, K. J. (2003). Hyperleptinemia, visceral adiposity, and decreased glucose tolerance in mice with a targeted disruption of the histidine decarboxylase gene. *Endocrinology* **144**, 4306–4314.
- Gantz, I., Munzert, G., Tashiro, T., Schaffer, M., Wang, L., DelValle, J., and Yamada, T. (1991a). Molecular cloning of the human histamine H₂ receptor. *Biochem. Biophys. Res. Commun.* **178**, 1386–1392.

- Gantz, I., Schaffer, M., DelValle, J., Logsdon, C., Campbell, V., Uhler, M., and Yamada, T. (1991b). Molecular cloning of a gene encoding the histamine H₂ receptor. *Proc. Natl. Acad. Sci. USA* **88**, 429–433.
- Garbarg, M., Barbin, G., Feger, J., and Schwartz, J. C. (1974). Histaminergic pathway in rat brain evidenced by lesions of the medial forebrain bundle. *Science* **186**, 833–835.
- Garbarg, M., Barbin, G., Rodergas, E., and Schwartz, J. C. (1980). Inhibition of histamine synthesis in brain by alpha-fluoromethylhistidine, a new irreversible inhibitor: *in vitro* and *in vivo* studies. *J. Neurochem.* **35**, 1045–1052.
- Garbarg, M., Tuong, M. D., Gros, C., and Schwartz, J. C. (1989). Effects of histamine H₃-receptor ligands on various biochemical indices of histaminergic neuron activity in rat brain. *Eur. J. Pharmacol.* **164**, 1–11.
- Garbarg, M., Traiffort, E., Ruat, M., Arrang, J. M., and Schwartz, J. C. (1992). Reversible labeling of H₁, H₂ and H₃ receptors. In "The Histamine Receptor" (J. C. Schwartz and H. L. Haas, Eds.), pp. 73–95. Wiley-Liss, New York.
- Garcia, M., Floran, B., Arias-Montano, J. A., Young, J. M., and Aceves, J. (1997). Histamine H₃ receptor activation selectively inhibits dopamine D₁ receptor-dependent [³H]GABA release from depolarization-stimulated slices of rat substantia nigra pars reticulata. *Neuroscience* **80**, 241–249.
- Gbahou, F., Rouleau, A., Morisset, S., Parmentier, R., Crochet, S., Lin, J. S., Ligneau, X., Tardivel-Lacombe, J., Stark, H., Schunack, W., Ganellin, C. R., Schwartz, J. C., *et al.* (2003). Protean agonism at histamine H₃ receptors *in vitro* and *in vivo*. *Proc. Natl. Acad. Sci. USA* **100**, 11086–11091.
- Gbahou, F., Vincent, L., Humbert-Claude, M., Tardivel-Lacombe, J., Chabret, C., and Arrang, J. M. (2006). Compared pharmacology of human histamine H₃ and H₄ receptors: Structure-activity relationships of histamine derivatives. *Br. J. Pharmacol.* **147**, 744–754.
- Gomez-Ramirez, J., Ortiz, J., and Blanco, I. (2002). Presynaptic H₃ autoreceptors modulate histamine synthesis through cAMP pathway. *Mol. Pharmacol.* **61**, 239–245.
- Goodchild, R. E., Court, J. A., Hobson, I., Piggott, M. A., Perry, R. H., Ince, P., Jaros, E., and Perry, E. K. (1999). Distribution of histamine H₃-receptor binding in the normal human basal ganglia: Comparison with Huntington's and Parkinson's disease cases. *Eur. J. Neurosci.* **11**, 449–456.
- Green, J. P., and Maayani, S. (1977). Tricyclic antidepressant drugs block histamine H₂ receptor in brain. *Nature* **269**, 163–165.
- Green, J. P., Johnson, C. L., Weinstein, H., and Maayani, S. (1977). Antagonism of histamine-activated adenylate cyclase in brain by D-lysergic acid diethylamide. *Proc. Natl. Acad. Sci. USA* **74**, 5697–5701.
- Gulat-Marnay, C., Lafitte, A., Arrang, J. M., and Schwartz, J. C. (1989a). Modulation of histamine release and synthesis in the brain mediated by *alpha* 2-adrenoceptors. *J. Neurochem.* **53**, 513–518.
- Gulat-Marnay, C., Lafitte, A., Arrang, J. M., and Schwartz, J. C. (1989b). Regulation of histamine release and synthesis in the brain by muscarinic receptors. *J. Neurochem.* **52**, 248–254.
- Haas, H. L., and Bucher, U. M. (1975). Histamine H₂-receptors on single central neurones. *Nature* **255**, 634–635.
- Haas, H. L., and Panula, P. (2003). The role of histamine and the tuberomammillary nucleus in the nervous system. *Nat. Rev. Neurosci.* **4**, 121–130.
- Hancock, A. A. (2003). H₃ receptor antagonists/inverse agonists as anti-obesity agents. *Curr. Opin. Investig. Drugs* **4**, 1190–1197.
- Hancock, A. A., and Fox, G. B. (2004). Perspectives on cognitive domains, H₃ receptor ligands and neurological disease. *Expert Opin. Investig. Drugs* **13**, 1237–1248.
- Hancock, A. A., Bennani, Y. L., Bush, E. N., Esbenshade, T. A., Faghghi, R., Fox, G. B., Jacobson, P., Knourek-Segel, V., Krueger, K. M., Nuss, M. E., Pan, J. B., Shapiro, R., *et al.* (2004). Antiobesity effects of A-331440, a novel non-imidazole histamine H₃ receptor antagonist. *Eur. J. Pharmacol.* **487**, 183–197.

- Hegstrand, L. R., Kanof, P. D., and Greengard, P. (1976). Histamine-sensitive adenylate cyclase in mammalian brain. *Nature* **260**, 163–165.
- Hill, S. J., Ganellin, C. R., Timmerman, H., Schwartz, J. C., Shankley, N. P., Young, J. M., Schunack, W., Levi, R., and Haas, H. L. (1997). International union of pharmacology. XIII. Classification of histamine receptors. *Pharmacol. Rev.* **49**, 253–278.
- Hirai, T., Okuma, C., Harada, C., Mio, M., Ohtsu, H., Watanabe, T., and Kamei, C. (2004). Development of amygdaloid kindling in histidine decarboxylase-deficient and histamine H₁ receptor-deficient mice. *Epilepsia* **45**, 309–313.
- Hong, C. J., Lin, C. H., Yu, Y. W., Chang, S. C., Wang, S. Y., and Tsai, S. J. (2002). Genetic variant of the histamine-1 receptor (glu349asp) and body weight change during clozapine treatment. *Psychiatr. Genet.* **12**, 169–171.
- Honrubia, M. A., Vilario, M. T., Palacios, J. M., and Mengod, G. (2000). Distribution of the histamine H₂ receptor in monkey brain and its mRNA localization in monkey and human brain. *Synapse* **38**, 343–354.
- Hough, L. B. (2001). Genomics meets histamine receptors: New subtypes, new receptors. *Mol. Pharmacol.* **59**, 415–419.
- Huang, Z. L., Qu, W. M., Li, W. D., Mochizuki, T., Eguchi, N., Watanabe, T., Urade, Y., and Hayaishi, O. (2001). Arousal effect of orexin A depends on activation of the histaminergic system. *Proc. Natl. Acad. Sci. USA* **98**, 9965–9970.
- Hussain, N., Flumerfelt, B. A., and Rajakumar, N. (2002). Muscarinic, adenosine A₂ and histamine H₃ receptor modulation of haloperidol-induced c-fos expression in the striatum and nucleus accumbens. *Neuroscience* **112**, 427–438.
- Imaizumi, M., and Onodera, K. (1993). The behavioral and biochemical effects of thioperamide, a histamine H₃-receptor antagonist, in a light/dark test measuring anxiety in mice. *Life Sci.* **53**, 1675–1683.
- Inoue, I., Yanai, K., Kitamura, D., Taniuchi, I., Kobayashi, T., Niimura, K., and Watanabe, T. (1996). Impaired locomotor activity and exploratory behavior in mice lacking histamine H₁ receptors. *Proc. Natl. Acad. Sci. USA* **93**, 13316–13320.
- Ito, C. (2000). The role of brain histamine in acute and chronic stresses. *Biomed. Pharmacother.* **54**, 263–267.
- Ito, C., Onodera, K., Sakurai, E., Sato, M., and Watanabe, T. (1996). Effects of dopamine antagonists on neuronal histamine release in the striatum of rats subjected to acute and chronic treatments with methamphetamine. *J. Pharmacol. Exp. Ther.* **279**, 271–276.
- Ito, C., Onodera, K., Watanabe, T., and Sato, M. (1997). Effects of histamine agents on methamphetamine-induced stereotyped behavior and behavioral sensitization in rats. *Psychopharmacology* **130**, 362–367.
- Ito, C., Morisset, S., Krebs, M. O., Olie, J. P., Loo, H., Poirier, M. F., Lannfelt, L., Schwartz, J. C., and Arrang, J. M. (2000). Histamine H₂ receptor gene variants: lack of association with schizophrenia. *Mol. Psychiatry* **5**, 159–164.
- Itoh, Y., Nishibori, M., Oishi, R., and Saeki, K. (1984). Neuronal histamine inhibits methamphetamine-induced locomotor hyperactivity in mice. *Neurosci. Lett.* **48**, 305–309.
- Itoh, Y., Oishi, R., Nishibori, M., and Saeki, K. (1985). Phencyclidine and the dynamics of mouse brain histamine. *J. Pharmacol. Exp. Ther.* **235**, 788–792.
- Itoh, Y., Oishi, R., Nishibori, M., and Saeki, K. (1986). Comparison of effects of phencyclidine and methamphetamine on body temperature in mice: A possible role for histamine neurons in thermoregulation. *Naunyn Schmiedebergs Arch. Pharmacol.* **332**, 293–296.
- Itoh, Y., Oishi, R., Nishibori, M., and Saeki, K. (1987). Involvement of opioid receptors in phencyclidine-induced enhancement of brain histamine turnover in mice. *Naunyn Schmiedebergs Arch. Pharmacol.* **335**, 285–289.

- Itoh, Y., Oishi, R., Nishibori, M., and Saeki, K. (1988). Involvement of *Mu* receptors in the opioid-induced increase in the turnover of mouse brain histamine. *J. Pharmacol. Exp. Ther.* **244**, 1021–1026.
- Itoh, Y., Oishi, R., Nishibori, M., and Saeki, K. (1991). Characterization of histamine release from the rat hypothalamus as measured by *in vivo* microdialysis. *J. Neurochem.* **56**, 769–774.
- Itoh, Y., Oishi, R., Adachi, N., and Saeki, K. (1992). A highly sensitive assay for histamine using ion-pair HPLC coupled with postcolumn fluorescent derivatization: Its application to biological specimens. *J. Neurochem.* **58**, 884–889.
- Iwabuchi, K., Kubota, Y., Ito, C., Watanabe, T., and Yanai, K. (2004). Methamphetamine and brain histamine: A study using histamine-related gene knockout mice. *Ann. NY Acad. Sci.* **1025**, 129–134.
- Iwabuchi, K., Ito, C., Tashiro, M., Kato, M., Kano, M., Itoh, M., Iwata, R., Matsuoka, H., Sato, M., and Yanai, K. (2005). Histamine H₁ receptors in schizophrenic patients measured by positron emission tomography. *Eur. Neuropsychopharmacol.* **15**, 185–191.
- Jang, I. S., Rhee, J. S., Watanabe, T., and Akaike, N. (2001). Histaminergic modulation of GABAergic transmission in rat ventromedial hypothalamic neurones. *J. Physiol.* **534**, 791–803.
- Javitt, D. C., and Zukin, S. R. (1991). Recent advances in the phencyclidine model of schizophrenia. *Am. J. Psychiatry.* **148**, 1301–1308.
- Jentsch, J. D., and Roth, R. H. (1999). The neuropsychopharmacology of phencyclidine: From NMDA receptor hypofunction to the dopamine hypothesis of schizophrenia. *Neuropsychopharmacology* **20**, 201–225.
- Jiang, X., Chen, A., and Li, H. (2005). Histaminergic modulation of excitatory synaptic transmission in the rat basolateral amygdala. *Neuroscience* **131**, 691–703.
- Jin, C. Y., and Panula, P. (2005). The laminar histamine receptor system in human prefrontal cortex suggests multiple levels of histaminergic regulation. *Neuroscience* **132**, 137–149.
- Jin, C. Y., Kalimo, H., and Panula, P. (2002). The histaminergic system in human thalamus: Correlation of innervation to receptor expression. *Eur. J. Neurosci.* **15**, 1125–1138.
- Jonker, J. W., and Schinkel, A. H. (2004). Pharmacological and physiological functions of the polyspecific organic cation transporters: OCT1, 2, and 3 (SLC22A1–3). *J. Pharmacol. Exp. Ther.* **308**, 2–9.
- Kamei, C. (2001). Involvement of central histamine in amygdaloid kindled seizures in rats. *Behav. Brain Res.* **124**, 243–250.
- Kaminsky, R., Moriarty, T. M., Bodine, J., Wolf, D. E., and Davidson, M. (1990). Effect of famotidine on deficit symptoms of schizophrenia. *Lancet* **335**, 1351–1352.
- Kanayama, H., Yasuhara, O., Matsuo, A., Tooyama, I., Aimi, Y., Bellier, J. P., Nagy, J. I., Fukui, K., and Kimura, H. (2003). Expression of a splice variant of choline acetyltransferase in magnocellular neurons of the tuberomammillary nucleus of rat. *Neuroscience* **118**, 243–251.
- Kanof, P. D., and Greengard, P. (1978). Brain histamine receptors as targets for antidepressant drugs. *Nature* **272**, 329–333.
- Kathmann, M., Schlicker, E., and Gothert, M. (1994). Intermediate affinity and potency of clozapine and low affinity of other neuroleptics and of antidepressants at H₃ receptors. *Psychopharmacology* **116**, 464–468.
- Katoh, Y., Niimi, M., Yamamoto, Y., Kawamura, T., Morimoto-Ishizuka, T., Sawada, M., Takemori, H., and Yamatodani, A. (2001). Histamine production by cultured microglial cells of the mouse. *Neurosci. Lett.* **305**, 181–184.
- Kim, S. H., Krapfenbauer, K., Cheon, M. S., Fountoulakis, M., Cairns, N. J., and Lubec, G. (2002). Human brain cytosolic histamine-N-methyltransferase is decreased in Down syndrome and increased in Pick's disease. *Neurosci. Lett.* **321**, 169–172.
- Knigge, U., and Warberg, J. (1991). The role of histamine in the neuroendocrine regulation of pituitary hormone secretion. *Acta Endocrinol.* **124**, 609–619.

- Komater, V. A., Browman, K. E., Curzon, P., Hancock, A. A., Decker, M. W., and Fox, G. B. (2003). H₃ receptor blockade by thioperamide enhances cognition in rats without inducing locomotor sensitization. *Psychopharmacology* **167**, 363–372.
- Koyama, M., Seyedi, N., Fung-Leung, W. P., Lovenberg, T. W., and Levi, R. (2003). Norepinephrine release from the ischemic heart is greatly enhanced in mice lacking histamine H₃ receptors. *Mol. Pharmacol.* **63**, 378–382.
- Kroeze, W. K., Hufeisen, S. J., Popadak, B. A., Renock, S. M., Steinberg, S., Ernsberger, P., Jayathilake, K., Meltzer, H. Y., and Roth, B. L. (2003). H₁-histamine receptor affinity predicts short-term weight gain for typical and atypical antipsychotic drugs. *Neuropsychopharmacology* **28**, 519–526.
- Laitinen, K. S., Tuomisto, L., and Laitinen, J. T. (1995a). Endogenous serotonin modulates histamine release in the rat hypothalamus as measured by *in vivo* microdialysis. *Eur. J. Pharmacol.* **285**, 159–164.
- Laitinen, K. S., Tuomisto, L., and MacDonald, E. (1995b). Effects of a selective α 2-adrenoceptor antagonist, atipamezole, on hypothalamic histamine and noradrenaline release *in vivo*. *Eur. J. Pharmacol.* **285**, 255–260.
- Laruelle, M., Abi-Dargham, A., van Dyck, C. H., Gil, R., D'Souza, C. D., Erdos, J., McCance, E., Rosenblatt, W., Fingado, C., Zoghbi, S. S., Baldwin, R. M., Seibyl, J. P., *et al.* (1996). Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug-free schizophrenic subjects. *Proc. Natl. Acad. Sci. USA* **93**, 9235–9240.
- Leopoldt, D., Harteneck, C., and Nurnberg, B. (1997). G proteins endogenously expressed in Sf 9 cells: Interactions with mammalian histamine receptors. *Naunyn Schmiedebergs Arch. Pharmacol.* **356**, 216–224.
- Leurs, R., Traiffort, E., Arrang, J. M., Tardivel-Lacombe, J., Ruat, M., and Schwartz, J. C. (1994). Guinea pig histamine H₁ receptor. II. Stable expression in Chinese hamster ovary cells reveals the interaction with three major signal transduction pathways. *J. Neurochem.* **62**, 519–527.
- Ligneau, X., Lin, J., Vanni-Mercier, G., Jouvet, M., Muir, J. L., Ganellin, C. R., Stark, H., Elz, S., Schunack, W., and Schwartz, J. (1998). Neurochemical and behavioral effects of ciproxifan, a potent histamine H₃-receptor antagonist. *J. Pharmacol. Exp. Ther.* **287**, 658–666.
- Lim, H. D., van Rijn, R. M., Ling, P., Bakker, R. A., Thurmond, R. L., and Leurs, R. (2005). Evaluation of histamine H₁, H₂, and H₃-receptor ligands at the human histamine H₄ receptor: Identification of 4-methylhistamine as the first potent and selective H₄ receptor agonist. *J. Pharmacol. Exp. Ther.* **314**, 1310–1321.
- Lin, J. S. (2000). Brain structures and mechanisms involved in the control of cortical activation and wakefulness, with emphasis on the posterior hypothalamus and histaminergic neurons. *Sleep Med. Rev.* **4**, 471–503.
- Lin, J. S., Sakai, K., Vanni-Mercier, G., and Jouvet, M. (1989). A critical role of the posterior hypothalamus in the mechanisms of wakefulness determined by microinjection of muscimol in freely moving cats. *Brain Res.* **479**, 225–240.
- Lin, L., Wisor, J., Shiba, T., Taheri, S., Yanai, K., Wurts, S., Lin, X., Vitaterna, M., Takahashi, J., Lovenberg, T. W., Koehl, M., Uhl, G., *et al.* (2002). Measurement of hypocretin/orexin content in the mouse brain using an enzyme immunoassay: The effect of circadian time, age and genetic background. *Peptides* **23**, 2203–2211.
- Liu, C., Ma, X., Jiang, X., Wilson, S. J., Hofstra, C. L., Blevitt, J., Pyati, J., Li, X., Chai, W., Carruthers, N., and Lovenberg, T. W. (2001a). Cloning and pharmacological characterization of a fourth histamine receptor (H₄) expressed in bone marrow. *Mol. Pharmacol.* **59**, 420–426.
- Liu, C., Wilson, S. J., Kuei, C., and Lovenberg, T. W. (2001b). Comparison of human, mouse, rat, and guinea pig histamine H₄ receptors reveals substantial pharmacological species variation. *J. Pharmacol. Exp. Ther.* **299**, 121–130.

- Lovenberg, T. W., Roland, B. L., Wilson, S. J., Jiang, X., Pyati, J., Huvar, A., Jackson, M. R., and Erlander, M. G. (1999). Cloning and functional expression of the human histamine H₃ receptor. *Mol. Pharmacol.* **55**, 1101–1107.
- Lovenberg, T. W., Pyati, J., Chang, H., Wilson, S. J., and Erlander, M. G. (2000). Cloning of rat histamine H₃ receptor reveals distinct species pharmacological profiles. *J. Pharmacol. Exp. Ther.* **293**, 771–778.
- Mammoto, T., Yamamoto, Y., Kagawa, K., Hayashi, Y., Mashimo, T., Yoshiya, I., and Yamatodani, A. (1997). Interactions between neuronal histamine and halothane anesthesia in rats. *J. Neurochem.* **69**, 406–411.
- Manahan-Vaughan, D., Reymann, K. G., and Brown, R. E. (1998). *In vivo* electrophysiological investigations into the role of histamine in the dentate gyrus of the rat. *Neuroscience* **84**, 783–790.
- Mancama, D., Arranz, M. J., Munro, J., Makoff, A., and Kerwin, R. (2000). The histamine 1 and histamine 2 receptor genes-candidates for schizophrenia and clozapine drug response. *GeneScreen* **1**, 29–34.
- Mancama, D., Arranz, M. J., Munro, J., Osborne, S., Makoff, A., Collier, D., and Kerwin, R. (2002). Investigation of promoter variants of the histamine 1 and 2 receptors in schizophrenia and clozapine response. *Neurosci. Lett.* **333**, 207–211.
- Marcus, J. N., Aschkenasi, C. J., Lee, C. E., Chemelli, R. M., Saper, C. B., Yanagisawa, M., and Elmquist, J. K. (2001). Differential expression of orexin receptors 1 and 2 in the rat brain. *J. Comp. Neurol.* **435**, 6–25.
- Martinez-Mir, M. I., Pollard, H., Moreau, J., Arrang, J. M., Ruat, M., Traiffort, E., Schwartz, J. C., and Palacios, J. M. (1990). Three histamine receptors (H₁, H₂ and H₃). visualized in the brain of human and non-human primates. *Brain Res.* **526**, 322–327.
- Martinez-Mir, M. I., Pollard, H., Moreau, J., Traiffort, E., Ruat, M., Schwartz, J. C., and Palacios, J. M. (1993). Loss of striatal histamine H₂ receptors in Huntington's chorea but not in Parkinson's disease: Comparison with animal models. *Synapse* **15**, 209–220.
- Masaki, T., Yoshimatsu, H., Chiba, S., Watanabe, T., and Sakata, T. (2001). Central infusion of histamine reduces fat accumulation and upregulates UCP family in leptin-resistant obese mice. *Diabetes* **50**, 376–384.
- Masaki, T., Chiba, S., Yasuda, T., Noguchi, H., Kakuma, T., Watanabe, T., Sakata, T., and Yoshimatsu, H. (2004). Involvement of hypothalamic histamine H₁ receptor in the regulation of feeding rhythm and obesity. *Diabetes* **53**, 2250–2260.
- Miklos, I. H., and Kovacs, K. J. (2003). Functional heterogeneity of the responses of histaminergic neuron subpopulations to various stress challenges. *Eur. J. Neurosci.* **18**, 3069–3079.
- Miyazaki, S., Onodera, K., Imaizumi, M., and Timmerman, H. (1997). Effects of clobenpropit (VUF-9153), a histamine H₃-receptor antagonist, on learning and memory, and on cholinergic and monoaminergic systems in mice. *Life Sci.* **61**, 355–361.
- Mobarakeh, J. I., Sakurada, S., Katsuyama, S., Kutsuwa, M., Kuramasu, A., Lin, Z. Y., Watanabe, T., Hashimoto, Y., and Yanai, K. (2000). Role of histamine H₁ receptor in pain perception: A study of the receptor gene knockout mice. *Eur. J. Pharmacol.* **391**, 81–89.
- Mochizuki, T., Yamatodani, A., Okakura, K., Takemura, M., Inagaki, N., and Wada, H. (1991). *In vivo* release of neuronal histamine in the hypothalamus of rats measured by microdialysis. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **343**, 190–195.
- Molina-Hernandez, A., Nunez, A., and Arias-Montano, J. A. (2000). Histamine H₃-receptor activation inhibits dopamine synthesis in rat striatum. *Neuroreport* **11**, 163–166.
- Molina-Hernandez, A., Nunez, A., Sierra, J. J., and Arias-Montano, J. A. (2001). Histamine H₃ receptor activation inhibits glutamate release from rat striatal synaptosomes. *Neuropharmacology* **41**, 928–934.
- Mori, T., Narita, M., Onodera, K., and Suzuki, T. (2004). Involvement of histaminergic system in the discriminative stimulus effects of morphine. *Eur. J. Pharmacol.* **491**, 169–172.

- Morimoto, T., Yamamoto, Y., Mobarakeh, J. I., Yanai, K., Watanabe, T., and Yamatodani, A. (1999). Involvement of the histaminergic system in leptin-induced suppression of food intake. *Physiol. Behav.* **67**, 679–683.
- Morimoto, T., Yamamoto, Y., and Yamatodani, A. (2000). Leptin facilitates histamine release from the hypothalamus in rats. *Brain Res.* **868**, 367–369.
- Morisset, S., Traiffort, E., and Schwartz, J. C. (1996). Inhibition of histamine versus acetylcholine metabolism as a mechanism of tacrine activity. *Eur. J. Pharmacol.* **315**, R1–R2.
- Morisset, S., Sahm, U. G., Traiffort, E., Tardivel-Lacombe, J., Arrang, J. M., and Schwartz, J. C. (1999). Atypical neuroleptics enhance histamine turnover in brain via 5-Hydroxytryptamine_{2A} receptor blockade. *J. Pharmacol. Exp. Ther.* **288**, 590–596.
- Morisset, S., Rouleau, A., Ligneau, X., Gbahou, F., Tardivel-Lacombe, J., Stark, H., Schunack, W., Ganellin, C. R., Schwartz, J. C., and Arrang, J. M. (2000). High constitutive activity of native H₃ receptors regulates histamine neurons in brain. *Nature* **408**, 860–864.
- Morisset, S., Pilon, C., Tardivel-Lacombe, J., Weinstein, D., Rostene, W., Betancur, C., Sokoloff, P., Schwartz, J. C., and Arrang, J. M. (2002). Acute and chronic effects of methamphetamine on tele-methylhistamine levels in mouse brain: Selective involvement of the D2 and not D3 receptor. *J. Pharmacol. Exp. Ther.* **300**, 621–628.
- Munzar, P., Tanda, G., Justinova, Z., and Goldberg, S. R. (2004). Histamine H₃ receptor antagonists potentiate methamphetamine self-administration and methamphetamine-induced accumbal dopamine release. *Neuropsychopharmacology* **29**, 705–717.
- Nakai, T., Kitamura, N., Hashimoto, T., Kajimoto, Y., Nishino, N., Mita, T., and Tanaka, C. (1991). Decreased histamine H₁ receptors in the frontal cortex of brains from patients with chronic schizophrenia. *Biol. Psychiatry* **30**, 349–356.
- Nakamura, S., Takemura, M., Ohnishi, K., Suenaga, T., Nishimura, M., Akiguchi, I., Kimura, J., and Kimura, T. (1993). Loss of large neurons and occurrence of neurofibrillary tangles in the tuberomammillary nucleus of patients with Alzheimer's disease. *Neurosci. Lett.* **151**, 196–199.
- Nakazato, M., Murakami, N., Date, Y., Kojima, M., Matsuo, H., Kangawa, K., and Matsukura, S. (2001). A role for ghrelin in the central regulation of feeding. *Nature* **409**, 194–198.
- Nalwalk, J. W., Koch, J. E., Barke, K. E., Bodnar, R. J., and Hough, L. B. (1995). Modulation of morphine antinociception by the brain-penetrating H₂ antagonist zolantidine: Detailed characterization in five nociceptive test systems. *Pharmacol. Biochem. Behav.* **50**, 421–429.
- Nelson, L. E., Guo, T. Z., Lu, J., Saper, C. B., Franks, N. P., and Maze, M. (2002). The sedative component of anesthesia is mediated by GABA_A receptors in an endogenous sleep pathway. *Nat. Neurosci.* **5**, 979–984.
- Nishibori, M., Oishi, R., Itoh, Y., and Saeki, K. (1985). Morphine-induced changes in histamine dynamics in mouse brain. *J. Neurochem.* **45**, 719–724.
- Nishibori, M., Tahara, A., Sawada, K., Sakiyama, J., Nakaya, N., and Saeki, K. (2000). Neuronal and vascular localization of histamine N-methyltransferase in the bovine central nervous system. *Eur. J. Neurosci.* **12**, 415–424.
- Nishino, S., Fujiki, N., Ripley, B., Sakurai, E., Kato, M., Watanabe, T., Mignot, E., and Yanai, K. (2001). Decreased brain histamine content in hypocretin/orexin receptor-2 mutated narcoleptic dogs. *Neurosci. Lett.* **313**, 125–128.
- Nowak, J. Z., Arrang, J. M., Schwartz, J. C., and Garbarg, M. (1983). Interaction between mianserin, an antidepressant drug, and central H₁- and H₂-histamine-receptors: *In vitro* and *in vivo* studies and radioreceptor assay. *Neuropharmacology* **22**, 259–266.
- Oda, T., Morikawa, N., Saito, Y., Masuho, Y., and Matsumoto, S. (2000). Molecular cloning and characterization of a novel type of histamine receptor preferentially expressed in leukocytes. *J. Biol. Chem.* **275**, 36781–36786.
- Oishi, R., Nishibori, M., Itoh, Y., and Saeki, K. (1986). Diazepam-induced decrease in histamine turnover in mouse brain. *Eur. J. Pharmacol.* **124**, 337–342.

- Oishi, R., Adachi, N., Okada, K., Muroi, N., and Saeki, K. (1990a). Regulation of histamine turnover via muscarinic and nicotinic receptors in the brain. *J. Neurochem.* **55**, 1899–1904.
- Oishi, R., Nishibori, M., Itoh, Y., Shishido, S., and Saeki, K. (1990b). Is monoamine turnover in the brain regulated by histamine H₃ receptors? *Eur. J. Pharmacol.* **184**, 135–142.
- Oishi, R., Itoh, Y., and Saeki, K. (1992). Inhibition of histamine turnover by 8-OH-DPAT, buspirone and 5-hydroxytryptophan in the mouse and rat brain. *Naunyn Schmiedebergs Arch. Pharmacol.* **345**, 495–499.
- Okakura-Mochizuki, K., Mochizuki, T., Yamamoto, Y., Horii, A., and Yamatodani, A. (1996). Endogenous GABA modulates histamine release from the anterior hypothalamus of the rat. *J. Neurochem.* **67**, 171–176.
- Orange, P. R., Heath, P. R., Wright, S. R., Ramchand, C. N., Kolkeiwicz, L., and Pearson, R. C. (1996). Individuals with schizophrenia have an increased incidence of the H2R649G allele for the histamine H₂ receptor gene. *Mol. Psychiatry* **1**, 466–469.
- Oyewumi, L. K., Vollick, D., Merskey, H., and Plumb, C. (1994). Famotidine as an adjunct treatment of resistant schizophrenia. *J. Psychiatry Neurosci.* **19**, 145–150.
- Panula, P., and Airaksinen, M. (1991). The histaminergic neuronal system as revealed with antisera against histamine. In “Histaminergic Neurons: Morphology and Function” (T. Watanabe and H. Wada, Eds.), pp. 127–144. CRC, Boca Raton, FL.
- Panula, P., Airaksinen, M. S., Pirvola, U., and Kotilainen, E. (1990). A histamine-containing neuronal system in human brain. *Neuroscience* **34**, 127–132.
- Panula, P., Rinne, J., Kuokkanen, K., Eriksson, K. S., Sallmen, T., Kalimo, H., and Relja, M. (1998). Neuronal histamine deficit in Alzheimer’s disease. *Neuroscience* **82**, 993–997.
- Panula, P., Yang, H. Y., and Costa, E. (1984). Histamine-containing neurons in the rat hypothalamus. *Proc. Natl. Acad. Sci. USA* **81**, 2572–2576.
- Panula, P., Karlstedt, K., Sallmen, T., Peitsaro, N., Kaslin, J., Michelsen, K. A., Anichtchik, O., Kukko-Lukjanov, T., and Lintunen, M. (2000). The histaminergic system in the brain: Structural characteristics and changes in hibernation. *J. Chem. Neuroanat.* **18**, 65–74.
- Parmentier, R., Ohtsu, H., Djebbara-Hannas, Z., Valatx, J. L., Watanabe, T., and Lin, J. S. (2002). Anatomical, physiological, and pharmacological characteristics of histidine decarboxylase knock-out mice: Evidence for the role of brain histamine in behavioral and sleep-wake control. *J. Neurosci.* **22**, 7695–7711.
- Patel, B. T., Tudball, N., Wada, H., and Watanabe, T. (1986). Adenosine deaminase and histidine decarboxylase coexist in certain neurons of the rat brain. *Neurosci. Lett.* **63**, 185–189.
- Peter, D., Jimenez, J., Liu, Y., Kim, J., and Edwards, R. H. (1994). The chromaffin granule and synaptic vesicle amine transporters differ in substrate recognition and sensitivity to inhibitors. *J. Biol. Chem.* **269**, 7231–7237.
- Pillot, C., Heron, A., Cochois, V., Tardivel-Lacombe, J., Ligneau, X., Schwartz, J. C., and Arrang, J. M. (2002a). A detailed mapping of the histamine H₃ receptor and its gene transcripts in rat brain. *Neuroscience* **114**, 173–193.
- Pillot, C., Ortiz, J., Heron, A., Ridray, S., Schwartz, J. C., and Arrang, J. M. (2002b). Ciproxifan, a histamine H₃-receptor antagonist/inverse agonist, potentiates neurochemical and behavioral effects of haloperidol in the rat. *J. Neurosci.* **22**, 7272–7280.
- Pillot, C., Heron, A., Schwartz, J. C., and Arrang, J. M. (2003). Ciproxifan, a histamine H₃-receptor antagonist/inverse agonist, modulates the effects of methamphetamine on neuropeptide mRNA expression in rat striatum. *Eur. J. Neurosci.* **17**, 307–314.
- Pollard, H., Bischoff, S., and Schwartz, J. C. (1973a). Decreased histamine synthesis in the rat brain by hypnotics and anaesthetics. *J. Pharm. Pharmacol.* **25**, 920–922.
- Pollard, H., Bischoff, S., and Schwartz, J. C. (1973b). Increased synthesis and release of ³H-histamine in rat brain by reserpine. *Eur. J. Pharmacol.* **24**, 399–401.

- Pollard, H., Bischoff, S., and Schwartz, J. C. (1974). Turnover of histamine in rat brain and its decrease under barbiturate anesthesia. *J. Pharmacol. Exp. Ther.* **190**, 88–99.
- Pollard, H., and Bouthenet, M. L. (1992). Autoradiographic visualization of the three histamine receptor subtypes in the brain. In "The Histamine Receptor" (J. C. Schwartz and H. L. Haas, Eds.), pp. 179–192. Wiley-Liss, New York.
- Pollard, H., Moreau, J., Arrang, J. M., and Schwartz, J. C. (1993). A detailed autoradiographic mapping of histamine H³ receptors in rat brain areas. *Neuroscience* **52**, 169–189.
- Prast, H., Fischer, H. P., Prast, M., and Philippu, A. (1994). *In vivo* modulation of histamine release by autoreceptors and muscarinic acetylcholine receptors in the rat anterior hypothalamus. *Naunyn Schmiedeberg Arch. Pharmacol.* **350**, 599–604.
- Prast, H., Heistracher, M., and Philippu, A. (1991). *In vivo* modulation of the histamine release in the hypothalamus by adrenoreceptor agonists and antagonists. *Naunyn Schmiedeberg Arch. Pharmacol.* **344**, 183–186.
- Prast, H., Heistracher, M., and Philippu, A. (1993). Modulation by dopamine receptors of the histamine release in the rat hypothalamus. *Naunyn Schmiedeberg Arch. Pharmacol.* **347**, 301–305.
- Prell, G. D., Green, J. P., Kaufmann, C. A., Khandelwal, J. K., Morrishow, A. M., Kirch, D. G., Linnoila, M., and Wyatt, R. J. (1995). Histamine metabolites in cerebrospinal fluid of patients with chronic schizophrenia: Their relationships to levels of other aminergic transmitters and ratings of symptoms. *Schizophr. Res.* **14**, 93–104.
- Prell, G. D., Khandelwal, J. K., Burns, R. S., LeWitt, P. A., and Green, J. P. (1990). Influence of age and gender on the levels of histamine metabolites and pro-methylimidazoleacetic acid in human cerebrospinal fluid. *Arch. Gerontol. Geriatr.* **11**, 85–95.
- Preuss, C. V., Wood, T. C., Szumlanski, C. L., Raftogianis, R. B., Otterness, D. M., Girard, B., Scott, M. C., and Weinshilboum, R. M. (1998). Human histamine N-methyltransferase pharmacogenetics: Common genetic polymorphisms that alter activity. *Mol. Pharmacol.* **53**, 708–717.
- Quach, T. T., Duchemin, A. M., Rose, C., and Schwartz, J. C. (1979). *In vivo* occupation of cerebral histamine H₁-receptors evaluated with ³H-mepyramine may predict sedative properties of psychotropic drugs. *Eur. J. Pharmacol.* **60**, 391–392.
- Rauscher, F. P., Nasrallah, H. A., and Wyatt, R. J. (1980). Cutaneous histamine response in schizophrenia. *J. Clin. Psychiatry* **41**, 44–51.
- Reilly, M. A., and Schayer, R. W. (1970). *In vivo* studies on histamine catabolism and its inhibition. *Br. J. Pharmacol.* **38**, 478–489.
- Richelson, E., and Souder, T. (2000). Binding of antipsychotic drugs to human brain receptors focus on newer generation compounds. *Life Sci.* **68**, 29–39.
- Rinne, J. O., Anichtchik, O. V., Eriksson, K. S., Kaslin, J., Tuomisto, L., Kalimo, H., Roytta, M., and Panula, P. (2002). Increased brain histamine levels in Parkinson's disease but not in multiple system atrophy. *J. Neurochem.* **81**, 954–960.
- Rizk, A., Curley, J., Robertson, J., and Raber, J. (2004). Anxiety and cognition in histamine H₃ receptor–/– mice. *Eur. J. Neurosci.* **19**, 1992–1996.
- Rosse, R. B., Kendrick, K., Fay-McCarthy, M., Prell, G. D., Rosenberg, P., Tsui, L. C., Wyatt, R. J., and Deutsch, S. I. (1996). An open-label study of the therapeutic efficacy of high-dose famotidine adjuvant pharmacotherapy in schizophrenia: Preliminary evidence for treatment efficacy. *Clin. Neuropharmacol.* **19**, 341–348.
- Rouleau, A., Ligneau, X., Tardivel-Lacombe, J., Morisset, S., Gbahou, F., Schwartz, J. C., and Arrang, J. M. (2002). Histamine H₃-receptor-mediated [³⁵S]GTPγ[S] binding: Evidence for constitutive activity of the recombinant and native rat and human H₃ receptors. *Br. J. Pharmacol.* **135**, 383–392.
- Ruat, M., Traffort, E., Bouthenet, M. L., Schwartz, J. C., Hirschfeld, J., Buschauer, A., and Schunack, W. (1990). Reversible and irreversible labeling and autoradiographic localization of

- the cerebral histamine H₂ receptor using [¹²⁵I]iodinated probes. *Proc. Natl. Acad. Sci. USA* **87**, 1658–1662.
- Ryu, J. H., Yanai, K., Iwata, R., Ido, T., and Watanabe, T. (1994a). Heterogeneous distributions of histamine H₃, dopamine D₁ and D₂ receptors in rat brain. *Neuroreport* **5**, 621–624.
- Ryu, J. H., Yanai, K., and Watanabe, T. (1994b). Marked increase in histamine H₃ receptors in the striatum and substantia nigra after 6-hydroxydopamine-induced denervation of dopaminergic neurons: An autoradiographic study. *Neurosci. Lett.* **178**, 19–22.
- Ryu, J. H., Yanai, K., Sakurai, E., Kim, C. Y., and Watanabe, T. (1995). Ontogenetic development of histamine receptor subtypes in rat brain demonstrated by quantitative autoradiography. *Brain Res. Dev. Brain Res.* **87**, 101–110.
- Ryu, J. H., Yanai, K., Zhao, X. L., and Watanabe, T. (1996). The effect of dopamine D₁ receptor stimulation on the up-regulation of histamine H₃-receptors following destruction of the ascending dopaminergic neurones. *Br. J. Pharmacol.* **118**, 585–592.
- Sakai, K., Yoshimoto, Y., Luppi, P. H., Fort, P., el Mansari, M., Salvert, D., and Jouvet, M. (1990). Lower brainstem afferents to the cat posterior hypothalamus: A double-labeling study. *Brain Res. Bull.* **24**, 437–455.
- Sakata, T., Fukagawa, K., Ookuma, K., Fujimoto, K., Yoshimatsu, H., Yamatodani, A., and Wada, H. (1990). Hypothalamic neuronal histamine modulates ad libitum feeding by rats. *Brain Res.* **537**, 303–306.
- Sakata, T., Yoshimatsu, H., and Kurokawa, M. (1997). Hypothalamic neuronal histamine: Implications of its homeostatic control of energy metabolism. *Nutrition* **13**, 403–411.
- Sakurada, S., Watanabe, H., Mizoguchi, H., Yonezawa, A., Orito, T., Katsuyama, S., Kuramasu, A., Sakurada, C., Yanai, K., and Sakurada, T. (2004). Involvement of the histaminergic system in the nociceptin-induced pain-related behaviors in the mouse spinal cord. *Pain* **112**, 171–182.
- Sanchez-Lemus, E., and Arias-Montano, J. A. (2004). Histamine H₃ receptor activation inhibits dopamine D₁ receptor-induced cAMP accumulation in rat striatal slices. *Neurosci. Lett.* **364**, 179–184.
- Sangalli, B. C. (1997). Role of the central histaminergic neuronal system in the CNS toxicity of the first generation H₁-antagonists. *Prog. Neurobiol.* **52**, 145–157.
- Sautel, F., Griffon, N., Sokoloff, P., Schwartz, J. C., Launay, C., Simon, P., Costentin, J., Schoenfelder, A., Garrido, F., and Mann, A. (1995). Nafadotride, a potent preferential dopamine D₃ receptor antagonist, activates locomotion in rodents. *J. Pharmacol. Exp. Ther.* **275**, 1239–1246.
- Schlicker, E., Fink, K., Detzner, M., and Gothert, M. (1993). Histamine inhibits dopamine release in the mouse striatum via presynaptic H₃ receptors. *J. Neural. Transm. Gen. Sect.* **93**, 1–10.
- Schlicker, E., Kathmann, M., Detzner, M., Exner, H. J., and Gothert, M. (1994a). H₃ receptor-mediated inhibition of noradrenaline release: an investigation into the involvement of Ca²⁺ and K⁺ ions, G protein and adenylate cyclase. *Naunyn Schmiedeberg's Arch. Pharmacol.* **350**, 34–41.
- Schlicker, E., Malinowska, B., Kathmann, M., and Gothert, M. (1994b). Modulation of neurotransmitter release via histamine H₃ heteroreceptors. *Fundam. Clin. Pharmacol.* **8**, 128–137.
- Schneider, C., Risser, D., Kirchner, L., Kitzmuller, E., Cairns, N., Prast, H., Singewald, N., and Lubec, G. (1997). Similar deficits of central histaminergic system in patients with Down syndrome and Alzheimer disease. *Neurosci. Lett.* **222**, 183–186.
- Schwartz, J. C. (1975). Histamine as a transmitter in brain. *Life Sci.* **17**, 503–517.
- Schwartz, J. C. (1977). Histaminergic mechanisms in brain. *Annu. Rev. Pharmacol. Toxicol.* **17**, 325–339.
- Schwartz, J. C., and Arrang, J. M. (2002). Histamine. In "Neuropsychopharmacology: The Fifth Generation of Progress" (K. L. Davis, D. Charney, J. T. Coyle, and C. Nemeroff, Eds.), pp. 179–190. Lippincott Williams & Wilkins, Philadelphia.
- Schwartz, J. C., Lampart, C., and Rose, C. (1970). Properties and regional distribution of histidine decarboxylase in rat brain. *J. Neurochem.* **17**, 1527–1534.

- Schwartz, J. C., Pollard, H., Bischoff, S., Rehault, M. C., and Verdiere-Sahuque, M. (1971). Catabolism of ^3H -histamine in the rat brain after intracisternal administration. *Eur. J. Pharmacol.* **16**, 326–335.
- Schwartz, J. C., Arrang, J. M., Garbarg, M., Gulat-Marnay, C., and Pollard, H. (1990). Modulation of histamine synthesis and release in brain via presynaptic autoreceptors and heteroreceptors. *Ann. NY Acad. Sci.* **604**, 40–54.
- Schwartz, J. C., Arrang, J. M., Garbarg, M., Pollard, H., and Ruat, M. (1991). Histaminergic transmission in the mammalian brain. *Physiol. Rev.* **71**, 1–51.
- Schwartz, J. C., Morisset, S., Rouleau, A., Tardivel-Lacombe, J., Gbahou, F., Ligneau, X., Heron, A., Sasse, A., Stark, H., Schunack, W., Ganellin, R. C., and Arrang, J. M. (2001). Application of genomics to drug design: The example of the histamine H_3 receptor. *Eur. Neuropsychopharmacol.* **11**, 441–448.
- Senba, E., Daddona, P. E., Watanabe, T., Wu, J. Y., and Nagy, J. I. (1985). Coexistence of adenosine deaminase, histidine decarboxylase, and glutamate decarboxylase in hypothalamic neurons of the rat. *J. Neurosci.* **5**, 3393–3402.
- Sherin, J. E., Shiromani, P. J., McCarley, R. W., and Saper, C. B. (1996). Activation of ventrolateral preoptic neurons during sleep. *Science* **271**, 216–219.
- Sherin, J. E., Elmquist, J. K., Torrealba, F., and Saper, C. B. (1998). Innervation of histaminergic tuberomammillary neurons by GABAergic and galaninergic neurons in the ventrolateral preoptic nucleus of the rat. *J. Neurosci.* **18**, 4705–4721.
- Shigemoto, Y., Fujii, Y., Shinomiya, K., and Kamei, C. (2004). Participation of histaminergic H_1 and noradrenergic $\alpha 1$ receptors in orexin A-induced wakefulness in rats. *Brain Res.* **1023**, 121–125.
- Silver, R. B., Poonwasi, K. S., Seyedi, N., Wilson, S. J., Lovenberg, T. W., and Levi, R. (2002). Decreased intracellular calcium mediates the histamine H_3 -receptor-induced attenuation of norepinephrine exocytosis from cardiac sympathetic nerve endings. *Proc. Natl. Acad. Sci. USA* **99**, 501–506.
- Sindelar, D. K., Shepperd, M. L., Pickard, R. T., Alexander-Chacko, J., Dill, M. J., Cramer, J. W., Smith, D. P., and Gadski, R. (2004). Central H_3R activation by thioperamide does not affect energy balance. *Pharmacol. Biochem. Behav.* **78**, 275–283.
- Steininger, T. L., Alam, M. N., Gong, H., Szymusiak, R., and McGinty, D. (1999). Sleep-waking discharge of neurons in the posterior lateral hypothalamus of the albino rat. *Brain Res.* **840**, 138–147.
- Stevens, D. R., Kuramasu, A., and Haas, H. L. (1999). GABA B -receptor-mediated control of GABAergic inhibition in rat histaminergic neurons in vitro. *Eur. J. Neurosci.* **11**, 1148–1154.
- Stevens, D. R., Eriksson, K. S., Brown, R. E., and Haas, H. L. (2001). The mechanism of spontaneous firing in histamine neurons. *Behav. Brain Res.* **124**, 105–112.
- Takahashi, K., Suwa, H., Ishikawa, T., and Kotani, H. (2002). Targeted disruption of H_3 receptors results in changes in brain histamine tone leading to an obese phenotype. *J. Clin. Invest.* **110**, 1791–1799.
- Takeshita, Y., Watanabe, T., Sakata, T., Munakata, M., Ishibashi, H., and Akaike, N. (1998). Histamine modulates high-voltage-activated calcium channels in neurons dissociated from the rat tuberomammillary nucleus. *Neuroscience* **87**, 797–805.
- Tardivel-Lacombe, J., Rouleau, A., Heron, A., Morisset, S., Pillot, C., Cochois, V., Schwartz, J. C., and Arrang, J. M. (2000). Cloning and cerebral expression of the guinea pig histamine H_3 receptor: Evidence for two isoforms. *Neuroreport* **11**, 755–759.
- Tashiro, M., Mochizuki, H., Iwabuchi, K., Sakurada, Y., Itoh, M., Watanabe, T., and Yanai, K. (2002). Roles of histamine in regulation of arousal and cognition: functional neuroimaging of histamine H_1 receptors in human brain. *Life Sci.* **72**, 409–414.
- Taylor, K. M., and Snyder, S. H. (1971a). Brain histamine: Rapid apparent turnover altered by restraint and cold stress. *Science* **172**, 1037–1039.

- Taylor, K. M., and Snyder, S. H. (1971b). Histamine in rat brain: sensitive assay of endogenous levels, formation in vivo and lowering by inhibitors of histidine decarboxylase. *J. Pharmacol. Exp. Ther.* **179**, 619–633.
- Taylor, K. M., and Snyder, S. H. (1973). The release of histamine from tissue slices of rat hypothalamus. *J. Neurochem.* **21**, 1215–1223.
- Thurmond, R. L., Desai, P. J., Dunford, P. J., Fung-Leung, W. P., Hofstra, C. L., Jiang, W., Nguyen, S., Riley, J. P., Sun, S., Williams, K. N., Edwards, J. P., and Karlsson, L. (2004). A potent and selective histamine H₄ receptor antagonist with anti-inflammatory properties. *J. Pharmacol. Exp. Ther.* **309**, 404–413.
- Tohyama, M., Tamiya, R., Inagaki, N., and Takagi, H. (1991). Morphology of histaminergic neurons with histidine decarboxylase as a marker. In “Histaminergic Neurons: Morphology and Function” (T. Watanabe and H. Wada, Eds.), pp. 107–126. CRC, Boca Raton, FL.
- Torrent, A., Moreno-Delgado, D., Gomez-Ramirez, J., Rodriguez-Agudo, D., Rodriguez-Caso, C., Sanchez-Jimenez, F., Blanco, I., and Ortiz, J. (2005). H₃ autoreceptors modulate histamine synthesis through calcium/calmodulin- and cAMP-dependent protein kinase pathways. *Mol. Pharmacol.* **67**, 195–203.
- Toyota, H., Dugovic, C., Koehl, M., Laposky, A. D., Weber, C., Ngo, K., Wu, Y., Lee, D. H., Yanai, K., Sakurai, E., Watanabe, T., Liu, C., *et al.* (2002). Behavioral characterization of mice lacking histamine H₃ receptors. *Mol. Pharmacol.* **62**, 389–397.
- Traiffort, E., Pollard, H., Moreau, J., Ruat, M., Schwartz, J. C., Martinez-Mir, M. I., and Palacios, J. M. (1992). Pharmacological characterization and autoradiographic localization of histamine H₂ receptors in human brain identified with [¹²⁵I]iodoaminopotentidine. *J. Neurochem.* **59**, 290–299.
- Trotter, S., Chotard, C., Traiffort, E., Unmehopa, U., Fisser, B., Swaab, D. F., and Schwartz, J. C. (2002). Co-localization of histamine with GABA but not with galanin in the human tuberomammillary nucleus. *Brain Res.* **939**, 52–64.
- Vanni-Mercier, G., Gigout, S., Debilly, G., and Lin, J. S. (2003). Waking selective neurons in the posterior hypothalamus and their response to histamine H₃-receptor ligands: an electrophysiological study in freely moving cats. *Behav. Brain Res.* **144**, 227–241.
- Verdiere, M., Rose, C., and Schwartz, J. C. (1975). Synthesis and release of histamine studied on slices from rat hypothalamus. *Eur. J. Pharmacol.* **34**, 157–168.
- Vizuete, M. L., Traiffort, E., Bouthenet, M. L., Ruat, M., Souil, E., Tardivel-Lacombe, J., and Schwartz, J. C. (1997). Detailed mapping of the histamine H₂ receptor and its gene transcripts in guinea-pig brain. *Neuroscience* **80**, 321–343.
- Wang, L., Thomae, B., Eckloff, B., Wieben, E., and Weinshilboum, R. (2002). Human histamine N-methyltransferase pharmacogenetics: Gene resequencing, promoter characterization, and functional studies of a common 5'-flanking region single nucleotide polymorphism (SNP). *Biochem. Pharmacol.* **64**, 699–710.
- Watanabe, T., Taguchi, Y., Hayashi, H., Tanaka, J., Shiosaka, S., Tohyama, M., Kubota, H., Terano, Y., and Wada, H. (1983). Evidence for the presence of a histaminergic neuron system in the rat brain: An immunohistochemical analysis. *Neurosci. Lett.* **39**, 249–254.
- Watanabe, T., and Yanai, K. (2001). Studies on functional roles of the histaminergic neuron system by using pharmacological agents, knockout mice and positron emission tomography. *Tohoku J. Exp. Med.* **195**, 197–217.
- Watanabe, T., Taguchi, Y., Shiosaka, S., Tanaka, J., Kubota, H., Terano, Y., Tohyama, M., and Wada, H. (1984). Distribution of the histaminergic neuron system in the central nervous system of rats; a fluorescent immunohistochemical analysis with histidine decarboxylase as a marker. *Brain Res.* **295**, 13–25.
- Willie, J. T., Chemelli, R. M., Sinton, C. M., Tokita, S., Williams, S. C., Kisanuki, Y. Y., Marcus, J. N., Lee, C., Elmquist, J. K., Kohlmeier, K. A., Leonard, C. S., Richardson, J. A., *et al.* (2003). Distinct

- narcolepsy syndromes in Orexin receptor-2 and Orexin null mice: Molecular genetic dissection of non-REM and REM sleep regulatory processes. *Neuron* **38**, 715–730.
- Wilson, J. R., Manning, K. A., Forestner, D. M., Counts, S. E., and Uhlich, D. J. (1999). Comparison of cholinergic and histaminergic axons in the lateral geniculate complex of the macaque monkey. *Anat. Rec.* **255**, 295–305.
- Wong, E. H., Kemp, J. A., Priestley, T., Knight, A. R., Woodruff, G. N., and Iversen, L. L. (1986). The anticonvulsant MK-801 is a potent N-methyl-D-aspartate antagonist. *Proc. Natl. Acad. Sci. USA* **83**, 7104–7108.
- Wouterlood, F., and Steinbusch, H. (1991). Afferent and efferent fiber connections of histaminergic neurons in the rat brain: Comparison with dopaminergic, noradrenergic and serotonergic systems. In “Histaminergic Neurons: Morphology and Function” (T. Watanabe and H. Wada, Eds.), pp. 145–162. CRC, Boca Raton, FL.
- Yamashita, M., Fukui, H., Sugama, K., Horio, Y., Ito, S., Mizuguchi, H., and Wada, H. (1991). Expression cloning of a cDNA encoding the bovine histamine H₁ receptor. *Proc. Natl. Acad. Sci. USA* **88**, 11515–11519.
- Yan, L., Szumlanski, C. L., Rice, S. R., Sobell, J. L., Lachman, H. M., and Weinshilboum, R. M. (2000). Histamine N-methyltransferase functional polymorphism: lack of association with schizophrenia. *Am. J. Med. Genet.* **96**, 404–406.
- Yanai, K., Son, L. Z., Endou, M., Sakurai, E., Nakagawasai, O., Tadano, T., Kisara, K., Inoue, I., and Watanabe, T. (1998). Behavioural characterization and amounts of brain monoamines and their metabolites in mice lacking histamine H₁ receptors. *Neuroscience* **87**, 479–487.
- Yang, Q. Z., and Hatton, G. I. (1997). Electrophysiology of excitatory and inhibitory afferents to rat histaminergic tuberomammillary nucleus neurons from hypothalamic and forebrain sites. *Brain Res.* **773**, 162–172.
- Yang, R., Hey, J. A., Aslanian, R., and Rizzo, C. A. (2002). Coordination of histamine H₃ receptor antagonists with human adrenal cytochrome P450 enzymes. *Pharmacology* **66**, 128–135.
- Yatsunami, K., Ohtsu, H., Tsuchikawa, M., Higuchi, T., Ishibashi, K., Shida, A., Shima, Y., Nakagawa, S., Yamauchi, K., Yamamoto, M., and *et al.* (1994). Structure of the L-histidine decarboxylase gene. *J. Biol. Chem.* **269**, 1554–1559.
- Yokoyama, H. (2001). The role of central histaminergic neuron system as an anticonvulsive mechanism in developing brain. *Brain Dev.* **23**, 542–547.
- Yoshimatsu, H., Itateyama, E., Kondou, S., Tajima, D., Himeno, K., Hidaka, S., Kurokawa, M., and Sakata, T. (1999). Hypothalamic neuronal histamine as a target of leptin in feeding behavior. *Diabetes* **48**, 2286–2291.
- Zhang, M., Ballard, M. E., Pan, L., Roberts, S., Faghieh, R., Cowart, M., Esbenshade, T. A., Fox, G. B., Decker, M. W., Hancock, A. A., and Rueter, L. E. (2005). Lack of cataleptogenic potentiation with non-imidazole H₃ receptor antagonists reveals potential drug-drug interactions between imidazole-based H₃ receptor antagonists and antipsychotic drugs. *Brain Res.* **1045**, 142–149.

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CANNABINOIDS AND PSYCHOSIS

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Recent epidemiological studies and advances in understanding of brain cannabinoid function have renewed interest in the long-recognized association between cannabinoids and psychosis. This chapter presents evidence supporting and refuting the association between cannabinoids and psychosis. Cannabinoids can induce acute transient psychotic symptoms or an acute psychosis in some individuals. What makes some individuals vulnerable to cannabinoid-related psychosis is unclear. Also clear is that cannabinoids can also exacerbate psychosis in individuals with an established psychotic disorder, and these exacerbations may last beyond the period of intoxication. Less clear is whether cannabis *causes* a persistent *de novo* psychosis. The available evidence meets many but not all the criteria for causality, including dose–response, temporality, direction, specificity, and biological plausibility. On the other hand, the large majority of individuals exposed to cannabinoids do not experience psychosis or develop schizophrenia and the rates of schizophrenia have not increased commensurate with the increase in rates of cannabis use. Similar to smoking and lung cancer, it is more likely that cannabis exposure is a component cause that interacts with other factors, for example, genetic risk, to “cause” schizophrenia. Nevertheless, in the absence of known causes of schizophrenia, the role of component causes such as cannabis

exposure (exogenous hypothesis) is important and warrants further study. There is also tantalizing evidence from postmortem, neurochemical, and genetic studies suggesting CB1 receptor dysfunction (endogenous hypothesis) in schizophrenia that warrants further investigation. Further work is necessary to identify those factors that place individuals at higher risk for cannabinoid-related psychosis, to identify the biological mechanisms underlying the risks and to further study whether CB1 receptor dysfunction contributes to the pathophysiology of psychotic disorders.

I. Introduction

... acute psychotic reactions, generally lasting but a few hours, but occasionally as long as a week; the reaction seemed dose related and its main features included paranoid ideation, illusions, hallucinations, delusions, depersonalization, confusion, restlessness and excitement. There can be delirium, disorientation and marked clouding of consciousness ...

In 'Du Haschisch et d'alimentation mentale' J.J. Moreau de Tours (1845) (Moreau, 1973).

An association between cannabis and psychosis has long been recognized. However, recent advances in the understanding of cannabinoid receptor function have renewed in the association between cannabis and psychosis. In addition to epidemiological studies, there are case reports of psychosis following cannabis use, reports of psychosis in surveys of cannabis users from community samples, and pharmacological studies with various cannabinoid compounds. These data are relevant to an exogenous hypothesis according to which the consumption of cannabinoid compounds is associated with psychotic disorders. We also suggest an endogenous hypothesis according to which brain cannabinoid receptor (CB1) dysfunction may contribute to the pathophysiology of psychosis and/or schizophrenia, and further, that the putative CB1 receptor dysfunction maybe unrelated to the consumption of cannabinoid compounds. These two hypotheses are by no means mutually exclusive, and may in fact interact.

The data supporting an association between cannabis consumption and the manifestation of psychotic symptoms in humans is now reviewed. However, a review of the literature will not be complete without a discussion about the constituents of cannabis. The principal active ingredient of cannabis is delta-9-tetrahydrocannabinol (Δ -9-THC). However, in addition to Δ -9-THC, herbal cannabis contains nearly 70 cannabinoid compounds, including cannabidiol (CBD), Δ -8-tetrahydrocannabinol, cannabinal, cannabigerol, and also terpenoids and flavonoids (Elsohly and Slade, 2005). These constituent compounds may modulate the effects of Δ -9-THC and may also have "entourage" effects (Mechoulam and Ben-Shabat, 1999; Russo and McPartland, 2003). The principal active metabolite of Δ -9-THC, 11-hydroxy-THC is more potent than Δ -9-THC. The time course of 11-hydroxy-THC blood levels correlates well with the psychological effects of

inhaled and oral Δ -9-THC reviewed in Agurell *et al.* (1986). CBD may offset some Δ -9-THC effects by its anxiolytic effects (Guimaraes *et al.*, 1994; Zuardi *et al.*, 1982), antipsychotic-like effects (Zuardi *et al.*, 1991, 1995, 2006), and may block the conversion of Δ -9-THC to the more psychoactive 11-hydroxy-THC (Bornheim *et al.*, 1995). Therefore, the net effect of herbal cannabis is a composite of its constituents. The CBD content of cannabis varies greatly and some samples of cannabis have been reported to be devoid of CBD (Pitts *et al.*, 1990, 1992). Thus, a relatively lower CBD content of cannabis has been implicated in the occurrence of psychotic and anxiety reactions with cannabis use (Solomons and Neppe, 1989; Solomons *et al.*, 1990). An example is South African cannabis, also known as dagga, which has very low levels of CBD compared to other varieties of cannabis obtained elsewhere. Naturalistic studies suggest an association between dagga consumption and high rates of psychotic symptom (Solomons and Neppe, 1989; Solomons *et al.*, 1990).

There are some data from studies with synthetic cannabinoids including dronabinol, nabilone, and levonantradol that are informative about the psychotic adverse effects of cannabinoids (Fig. 1). Dronabinol is synthetic Δ -9-THC. The 9-*trans*-ketocannabinoid Nabilone (Cesamet®) is a synthetic analogue of Δ -9-THC that was developed as an antiemetic and is available in Europe and Canada. Levonantradol, also a synthetic cannabinoid, was developed as an analgesic agent but abandoned because of a high incidence of intolerable behavioral side effects.

Evidence for an association between cannabis and psychosis comes from several sources, including case series of psychosis following cannabis use, autobiographical accounts, and surveys of cannabis users in the general population, epidemiological studies, and pharmacological studies with various cannabinoid compounds.

II. Anecdotal Reports

A. AUTOBIOGRAPHICAL ACCOUNTS

There are several exquisitely detailed autobiographical accounts of the effects of cannabinoids. In perhaps one of the first detailed accounts of cannabis effects, Moreau de Tours (1845) described acute, transient, dose-related psychotic reactions lasting hours to days following hashish use (Moreau, 1973). The reaction was characterized by paranoid ideation, illusions, hallucinations, delusions, depersonalization, confusion, restlessness, and excitement. Among other effects of cannabis, Marshall described grandiosity (“my powers became superhuman, my knowledge of the universe was infinite, and so on”) (Marshall, 1897). At higher doses he described disturbing hallucinations (“demons”). While informative, these individual accounts have limited generalizability.

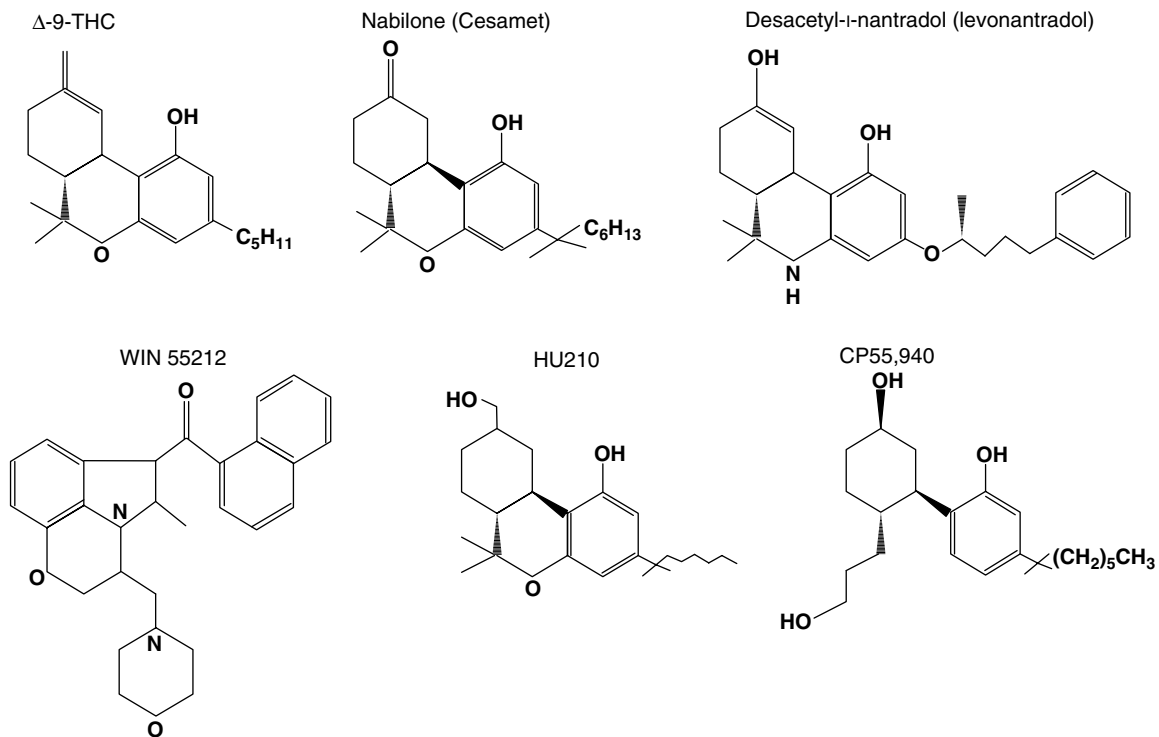


FIG. 1. Natural and synthetic cannabinoid receptor agonists.

B. SURVEYS OF CANNABIS USERS

Some of the limitations of individual accounts can be addressed in surveys of large groups of cannabis users from community samples. Thomas surveyed 1000 New Zealanders aged 18–35 years about the effects of cannabis (Thomas, 1996). Thirty-eight percent of respondents admitted to cannabis use. The most common adverse effects included anxiety and panic attacks (22%) followed by psychotic symptoms such as auditory hallucinations and persecutory ideas (15%). A significant relationship between panic attacks and psychosis was found. However, the survey, failed to find a dose–response relationship between the levels of cannabis use and the occurrence of these symptoms. As discussed later, there are challenges to accurately estimate dose–response relationships from naturalistic data. In a study of young Australian cannabis users ($n = 268$), about 21% reported negative effects that included paranoia (Reilly *et al.*, 1998). Green *et al.* (2003) reviewed pooled data ($n > 2500$) from surveys of cannabis users that used closed questions and found that 51.4% of the sample reported paranoia whereas 19.8% reported hallucinations while under the influence of cannabis. Thus, psychotic symptoms are not uncommonly experienced under the influence of cannabis. However, survey data too have their limitations, including sampling bias, reliance on self-report, lack of structured scales to assess psychosis, lack of reliable dose–response data, and so on.

At this point, it would be important to make the distinction between a psychotic disorder and psychotic symptoms, since these terms are somewhat poorly defined in the literature. Psychotic disorder refers to a condition characterized by *persistent* psychotic symptoms accompanied by functional deficits. Further, most discussions associating cannabis and psychosis have referred mainly to positive symptoms (hallucinations, perceptual alterations, delusions, paranoia, ideas of reference, disorganized speech, and disorganized behavior). However, any discussion of schizophrenia would be incomplete without reference to negative symptoms (emotional withdrawal, blunted affect, amotivation, alogia, and social withdrawal) and cognitive deficits (deficits in memory, attention, and executive function). Furthermore, the inclusion of schizophrenia-spectrum conditions such as schizotypy makes for some interesting findings. Several groups have found higher scores on measures of schizotypy, positive psychotic symptoms, and perceptual alterations in cannabis users (Dumas *et al.*, 2002; Nunn *et al.*, 2001; Skosnik *et al.*, 2001). Verdoux *et al.* (2003c) studied the association between cannabis use and psychotic dimensions in a nonclinical sample (571 college students) 18–51 years of age and found an association between cannabis use and positive and negative dimensions of psychosis and a correlation between the frequency of use and intensity of symptoms. This relationship appeared to be specific for cannabis use as alcohol use was not associated with dimensions of psychosis. Further the association with cannabis use was specific to positive and negative symptoms of psychosis but not depression.

Another limitation of cross-sectional surveys is that they are not very informative on the direction of causality. Thus, cannabis use may be a consequence rather than a cause of psychosis or cannabis use and psychosis may be independently associated with some common risk factor(s). Prospective studies have addressed some of these limitations. Verdoux *et al.* (2003a) investigated the impact of cannabis use on the onset of psychotic experiences using an experience sampling method, a self-reported structured diary technique that has been validated as a method of collecting information on psychotic experiences in daily life. Subjects were instructed to respond to randomly programmed cues from a wristwatch to describe their present substance use and psychotic experiences five times a day over 7 consecutive days. There was a clear temporal relationship between cannabis use and the acute occurrence of psychotic-like abnormal perceptions.

C. PSYCHOSIS IN CANNABIS USERS FROM COMMUNITY SAMPLES

Using data from the US Epidemiological Catchment Area (ECA) study, Tien and Anthony (1990) found that daily cannabis use doubled the risk of reporting psychotic symptoms after adjusting for baseline alcohol use and psychiatric diagnosis. Similarly, in the Australian National Survey of Mental Health and Well-Being, 17.2% of individuals diagnosed with cannabis dependence screened positive for a psychotic disorder (Degenhardt *et al.*, 2001).

D. NATURALISTIC CASE SERIES

In a review, Hall and Degenhardt (2004) found 397 cases of “cannabis psychosis” reported in the literature. In perhaps the earliest case series, Chopra and Smith (1974) described 200 patients admitted to a psychiatric hospital in India for psychosis following cannabis use (Chopra, 1973; Chopra and Smith, 1974). The psychosis was typically preceded by ingestion of large doses of cannabis and was characterized by hallucinations, paranoia, delusions, depersonalization, emotional lability, amnesia, confusion, and disorientation. One-third of the patients reportedly had no previous psychiatric history, suggesting that the heavy cannabis abuse in these subjects was not a sign of preexisting psychopathology. Further, the use of more potent forms of cannabis, for example, hashish, was associated with a quicker onset of psychosis suggesting a dose-response. Similar case series have been reported from other geographical areas, including Denmark (Arendt *et al.*, 2005), Sweden (Bernhardson and Gunne, 1972), the Caribbean (Harding and Knight, 1973), the United Kingdom (Carney *et al.*, 1984; Mathers *et al.*, 1991), the United States (Talbot and Teague, 1969b; Tennant and Groesbeck, 1972a), Scotland (Wylie *et al.*, 1995), and South Africa (Rottanburg *et al.*, 1982).

Most of these studies suggest that psychotic episodes resolved completely when cannabis use stopped but recurred with the resumption of cannabis use reviewed in Hall and Solowij (1998) and typically resolved fairly quickly in comparison with endogenous psychoses (Basu *et al.*, 1999; Carney *et al.*, 1984; Chaudry *et al.*, 1991; Cohen, 1994; Kolansky and Moore, 1971a; Rottanburg *et al.*, 1982; Talbott and Teague, 1969a; Thacore, 1973; Thacore and Shukla, 1976; Wylie *et al.*, 1995). Further, some studies suggested that patients with preexisting mental problems had a less favorable outcome (Bernhardson and Gunne, 1972; Bromberg, 1939; Chopra and Smith, 1974; Palsson *et al.*, 1982; Tennant and Groesbeck, 1972b).

In most of the studies discussed thus far, cases/patients were followed no greater than 3 months after remission of psychosis and hence, long-term outcome of these cases could not be conclusively determined. Arendt *et al.* (2005) reported the long-term outcome of a cohort of patients treated for cannabis-induced psychotic disorder extracted from the Danish Psychiatric Central Register. All patients treated for ICD-10 cannabis-induced psychotic disorder between 1994 and 1999 who had never been previously treated for psychosis ($n = 535$) were examined for the development of schizophrenia-spectrum disorder in the ensuing years (Table I). Schizophrenia-spectrum disorders were diagnosed in 44.5% of the sample. When the diagnosis was broadened to include new psychotic episodes of *any* type, the diagnosed sample increased to 77.2%. Further, only 15.9% of individuals were not in psychiatric care at the time of long-term follow-up. About half of the patients received the diagnosis of schizophrenia-spectrum disorder more than a year after seeking treatment for a cannabis-induced psychosis. The patients had an earlier age of onset of schizophrenia compared to a control group without a history of cannabis-induced psychosis. This study is the first to show that such psychotic symptoms induced by cannabis may be the first manifestations of a long-term psychotic disorder such as a schizophrenia-spectrum disorder.

These case series have some shortcomings. First, it is possible that the individuals who developed psychosis after using cannabis were not “healthy” and carried some risk for developing a psychotic disorder. Here, it is important to point out that other than family history there are no other known risks factors that are either specific and/or sensitive predictors of the future development of a psychotic disorder. Further, even though having a first-degree relative diagnosed with schizophrenia increases the risk of developing schizophrenia by 9–18 times, more than 80% of individuals diagnosed with schizophrenia do not have an affected first-degree relative and over 60% do not have an affected first- or second-degree relative (Gottesman and Shields, 1982). As discussed later, genetic factors may influence the risk of psychosis associated with cannabis exposure (Caspi *et al.*, 2005). Second, since individuals who use cannabis often co-use other drugs, it is unclear whether the use of other drugs contributed to the development

TABLE I
RISK OF MENTAL ILLNESS FOLLOWING HOSPITALIZATION FOR CANNABIS-INDUCED PSYCHOSIS PATIENTS
TREATED FOR MENTAL OR BEHAVIORAL DISORDERS AFTER INDEX POINT (N = 535)

Diagnosis ^a	Within 3 years n (%)	After 3 years n (%)	Total follow-up n (%)
Schizophrenia-spectrum disorder ^b	197 (36.8)	41 (7.7)	238 (44.5)
Persistent delusional disorder (F22)	18 (3.4)	4 (0.7)	22 (4.1)
Other or non-organic psychotic disorder (F28/F29)	5 (0.9)	3 (0.6)	8 (1.5)
Bipolar affective disorder	12 (2.2)	6 (1.1)	18 (3.4)
Acute and transient psychotic disorder	28 (5.2)	7 (1.3)	35 (6.5)
Cannabis-induced psychosis	78 (14.6)	1 (0.2)	79 (14.8)
Other drug-induced psychosis	11 (2.1)	2 (0.4)	13 (2.4)
Depression, anxiety or personality disorder	29 (5.4)	8 (1.5)	37 (6.9)
No treatment			85 (15.9)
Total			535 (100)

^aPatients are entered only once, in a hierarchical manner as described in the method section.

^bSchizophrenia (ICD-10 code F20), schizotypal disorder (F21) or schizoaffective disorder (F25).

of psychosis. Third, only a minority of case series studies employed standardized measures of psychosis. Fourth, the amount of cannabis use preceding the psychotic episode was not quantified limiting any speculation about dose-response relationships. In contrast, the temporal relationship between cannabis use and psychosis, the fact that psychosis resolved with abstinence from the drug and recurred with renewed use, lends support to the notion that the relationship between cannabis exposure and the development of psychosis is not merely coincidental.

On the basis of some similarities between the phenomenology of psychosis associated with cannabis use and the psychosis of schizophrenia, some (Chaudry *et al.*, 1991; Ghodse, 1986; Mathers *et al.*, 1991; Rolfe *et al.*, 1993; Rottanburg *et al.*, 1982; Thacore, 1973; Thacore and Shukla, 1976) but not others (Hall and Degenhardt, 2004; Imade and Ebie, 1991; McGuire *et al.*, 1994, 1995; Thomas, 1993; Thornicroft, 1990) have argued for the inclusion of “cannabis psychosis” as a distinct nosological entity. Arguments challenging the validity of “cannabis psychosis” as a distinct diagnostic entity should not be confused with the debate on the association between cannabis and psychosis.

As mentioned earlier, it is difficult to derive dose-response relationships from naturalistic data for a number of reasons. The reliability of self-reported, long-term, retrospective estimates of cannabis use is unclear. Individuals who use cannabis will often share a “joint” with one or more individuals, thus estimating

the dose consumed by the individual may be difficult. Cannabis is consumed in many different ways, for example, “joints,” “bongs,” and so on, which are not equivalent. The estimates of the number of lifetime exposures cannot accurately reflect the actual dose of Δ -9-THC that is consumed. Finally, the Δ -9-THC content of cannabis varies greatly. The Potency Monitoring program, a collaboration between the University of Mississippi and the National Institute on Drug Abuse (NIDA), provides analytical data about the potency of confiscated marijuana seized in the United States. In the most recent report covering the last 10 years on \sim 30,000 cannabis samples, 207 hashish samples, and 86 hash oil samples there appears to be an upward trend in the average THC content of confiscated cannabis (Mehmedic *et al.*, 2005). The Δ -9-THC content of cannabis doubled from 3.48% in 1994 to 7.08% in 2004. While there was no consistent increase in Δ -9-THC content in hashish samples from 1994 to 1999, the average potency of hashish samples increased from 4.16% Δ -9-THC in 2000 to 11.2% in 2004. No potency trends were observed for hash oil samples. Finally, there was no change in the average levels of the other cannabinoids (CBD, CBC, CBG, and CBN) in the cannabis samples over the reported time frame. Similarly, the average Δ -9-THC content of Dutch cannabis, Dutch hashish, and imported hashish has significantly increased between 1999 and 2005 (Niesink *et al.*, 2005). For example, in 2005, the average Δ -9-THC content of Dutch home-grown cannabis (Nederwiet) was 17.73%, and was nearly three times higher than that of imported cannabis (6.7% Δ -9-THC). Dutch hashish (Nederhasj) contained 26% Δ -9-THC in 2005, compared with 16.9% THC in imported hashish. In summary, deriving accurate dose-response from naturalistic data may have significant limitations.

III. Epidemiological Studies

Epidemiological studies have provided *the* major contribution to the evidence supporting an association between cannabis and psychosis (Table II). The study that first brought significant attention to the topic was a large historical, longitudinal cohort study of all Swedes conscripted between 1969 and 1970 (Andreasson *et al.*, 1987). Since Sweden mandates military service, 97% of males aged 18–20 years were included. The relationship between self-reported cannabis use at the time of conscription and psychiatric hospitalization for schizophrenia in the ensuing 15 years was examined. A dose-response relationship was observed between cannabis use at conscription (age 18 years) and schizophrenia diagnosis in the following 15 years. Individuals who reported having used cannabis more than 50 times were 6 times more likely than nonusers to have been diagnosed with schizophrenia 15 years later. Adjusting for other relevant risk factors including

TABLE II
EPIDEMIOLOGICAL STUDIES

References	<i>N</i>	Design	Instrument	Age range at f/u (years)	Outcome		Adjusted Risk
Arendt <i>et al.</i> 2005	535	Longitudinal follow up of cannabis-induced psychosis (Denmark)	Registry	Not specified (estimated as 32 years)	New psychotic episode of any type diagnosed in 77.2%. Schizophrenia-spectrum disorders diagnosed in 44.5%	Earlier onset of schizophrenia	–
Ferdinand <i>et al.</i> 2005	1580	Prospective cohort (The Netherlands)	CIDI CBCL	18–30	Psychotic symptoms	Lifetime CB use	2.8
Henquet <i>et al.</i> 2005	2437	Prospective cohort (Germany)	M-CIDI	18.3–21.8	Any psychotic symptom	Lifetime CB use	1.7
Stefanis <i>et al.</i> 2004	3500	Cross-sectional cohort of adolescents (Greece)	CAPE PDI	18	Positive and negative symptoms	Lifetime CB use until age 19 years	4.3
Fergusson, 2003	1011	Birth cohort (Christchurch)	SCL90	21	Psychotic symptoms	DSM-IV CB dependence at 21 years	1.8
Arseneault <i>et al.</i> 2002	759	Longitudinal, prospective, birth cohort (Dunedin)	DSM-IV	26	Schizophrenia and schizophreniform disorders	Lifetime CB use until age 15 and 18 years	3.12
Weiser <i>et al.</i> 2002	50,413	Longitudinal conscript cohort (Israel)	registry	4–15	Hospitalization for schizophrenia	Lifetime drug use at 16–17 years	2
van Os <i>et al.</i> 2002	4095	Longitudinal population-based 3 year follow-up (The Netherlands)	CIDI, SCID	18–64	Any psychotic symptoms	Lifetime CB use at 16–17 years	2.76
Zammit <i>et al.</i> 2002	50,053	Longitudinal conscript cohort (Sweden)	Registry	45–47	Hospitalization for schizophrenia	>50 exposures to CB at age 18 years	3.1
Andreasson <i>et al.</i> 1987	45,570	Longitudinal conscript cohort (Sweden)	Registry	33–35	Hospitalization for schizophrenia	>50 exposures to CB at age 18 years	2.3

psychiatric diagnosis other than psychosis at conscription reduced but did not eliminate the higher risk (odds ratio = 2.3) of schizophrenia conferred by cannabis use.

A reanalysis and extension of the same Swedish conscript cohort reconfirmed that heavy cannabis users by the age of 18 years were 6.7 times more likely than nonusers to be hospitalized for schizophrenia in the following 27 years (Zammit *et al.*, 2002). This study addressed the confounding effects of concomitant use of other drugs of abuse, premorbid personality traits, and cannabis use as a form of self-medication of schizophrenia. The adjusted odds ratio for cannabis use and schizophrenia remained significant (1.2), despite adjusting for a number of confounds, including low IQ, urbanicity, cigarette smoking, poor social integration, function, and stimulant use. Further, after controlling for the possibility that cannabis use is a consequence of prodromal manifestations of psychosis by excluding subjects who developed schizophrenia within 5 years of conscription, the finding of an increased risk of schizophrenia conferred by cannabis use persisted. The authors concluded that cannabis use was associated in a causal way with an increased risk of developing schizophrenia and that 13% of cases of schizophrenia would be averted if cannabis use were prevented.

The historical studies have been complemented by a number of recent prospective cohort studies. In a general population birth cohort study of 1037 people born in Dunedin, New Zealand, and followed through until age 26 years, cannabis use conferred a higher risk for the subsequent development of schizophrenia (Arseneault *et al.*, 2002). One of the strengths of this study was that it collected data on self-reported psychotic symptoms at age 11 years, that is, to address whether psychosis preceded cannabis use. Self-reported cannabis at both ages 15 and 18 years use was also collected. Further, the entire sample was assessed at age 26 years using a standardized psychiatric interview that allowed the determination of both schizophrenia symptoms and disorder. Compared to nonusers, individuals using cannabis at ages 15 and 18 years had higher rates of psychotic symptoms and schizophreniform disorder at age 26 years, even after controlling for psychotic symptoms pre-dating the onset of cannabis use. Cannabis users at age 15 years had a higher rate (OR = 3.1) of developing schizophreniform disorder at age 26 years, even after controlling for psychotic symptoms pre-dating the onset of cannabis use (Fig. 2).

In The Netherlands Mental Health Survey and Incidence Study (NEMESIS), 4045 psychosis-free individuals and 59 individuals with a psychotic disorder were assessed at baseline, 1 and 3 years (van Os *et al.*, 2002) using a measure of psychosis. Individuals using cannabis at baseline were nearly three times more likely to manifest psychotic symptoms at follow-up even after adjustment for a range of factors. Further, a dose-response relationship was established with the highest risk (odds ratio = 6.8) for the highest level of cannabis use. The relationship between cannabis use and psychotic symptoms was stronger for cases with

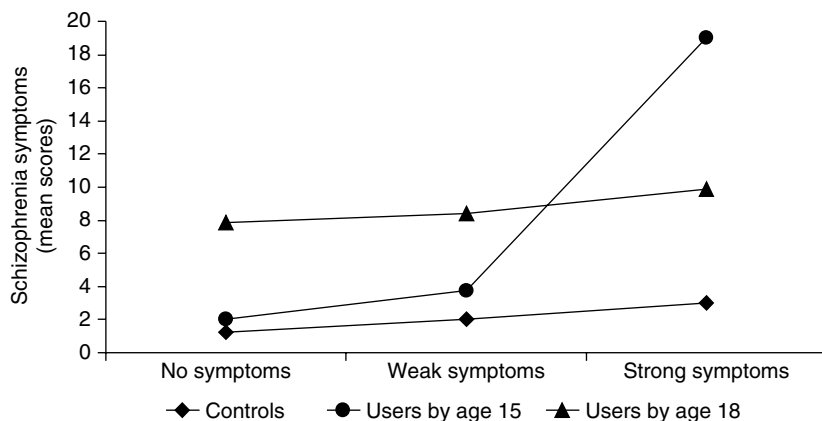


Fig. 2. Interaction between cannabis use at age 18 and psychotic symptoms at age 11 in predicting schizophrenia symptoms.

more severe psychotic symptoms. Individuals who reported psychotic symptoms at baseline were also more likely to develop schizophrenia if they used cannabis than were individuals who did not. The attributable risk of cannabis to psychosis was estimated at 13% for psychotic symptoms and 50% for cases with psychotic disorders that required psychiatric treatment.

Henquet *et al.* (2005) studied the relation between cannabis use and psychotic symptoms in individuals at risk for psychosis who first used cannabis during adolescence.

They tracked 2437 subjects (14–24 years) with and without risk for psychosis from the general population for 4 years and found a dose-dependent increased risk of psychosis in subjects exposed to cannabis (Henquet *et al.*, 2005). Interestingly, predisposition to psychosis was not found to be a predictor of future cannabis use at 4-year follow up. Adding to these studies, Stefanis *et al.* (2004) reported that both positive and negative symptoms can be induced by cannabis consumption and are independent of each other. Finally, a number of cohort studies have reported a dose-response relationship in the increased risk of psychosis with cannabis exposure (Ferdinand *et al.*, 2005; Fergusson *et al.*, 2003; Henquet *et al.*, 2005; Stefanis *et al.*, 2004; van Os *et al.*, 2002; Weiser *et al.*, 2002).

Collectively, these epidemiological studies suggest that cannabis use may confer a nearly twofold higher risk for developing schizophrenia. This increased risk is comparable to the better known associations, such as the risk conferred by cigarette smoking in the development of lung cancer and the risk of heart disease from hypercholesterolemia.

Temporal relationship between cannabis use and the onset of schizophrenia: The onset of cannabis use may precede, follow, or co-occur with the onset of schizophrenia. However, schizophrenia begins insidiously, and evolves through several identifiable stages with the emergence of psychotic symptoms as the final step in the evolution of the disorder. As a result, while it may be easy to pinpoint the emergence of positive psychotic symptoms in retrospective studies, pinpointing the onset of the less obvious prodromal symptoms is extremely challenging. Thus, while there is evidence suggesting a temporal association between cannabis use and the onset of positive psychotic symptoms, the temporal relationship between cannabis use and the less obvious symptoms has not been studied. Further, if as the neurodevelopmental hypothesis posits, that the pathophysiological processes underlying the illness precede the clinical manifestations by years or even decades and that these processes may even begin *in utero*, then, the argument about a temporal relationship is no longer relevant.

Nevertheless, there are a few studies that have systematically investigated the temporal order of cannabis use and the onset of schizophrenia. Allebeck and colleagues reported that in 69% of a schizophrenic patient sample from a Swedish case registry ($n = 112$), cannabis abuse preceded the onset of psychotic symptoms by at least 1 year (Allebeck *et al.*, 1993b). Further, in only 11% did the onset of psychotic symptoms precede the onset of cannabis abuse. Similarly, Linszen *et al.* (1994) found that cannabis abuse preceded the onset of psychotic symptoms by at least 1 year in 23 of 24 cannabis-abusing recent onset schizophrenic patients. Hambrecht and Hafner (1996, 2000) in their study of first-episode schizophrenic patients found that 14.2% of the sample had a lifetime history of drug abuse with cannabis being the most frequently abused drug (88%). Furthermore, drug abuse preceded the first sign of schizophrenia by more than a year but typically by more than 5 years in 27.5% of patients. In 37.9% of individuals, drug abuse followed the first sign of schizophrenia, and in 34.6% of individuals the first sign of schizophrenia and drug abuse started within the same month.

Effects of cannabis in individuals at high risk for developing schizophrenia: Another way to assess the risk of psychosis conferred by cannabis exposure is by studying the effects of cannabis in individuals at high risk for developing schizophrenia. In the Edinburgh High Risk study, individuals with a high genetic risk of schizophrenia, as evidenced by two or more affected relatives, cannabis use increased the risk for psychosis (Miller *et al.*, 2001). Furthermore, frequent cannabis use conferred a sixfold higher risk of schizophrenia in individuals with a family history of schizophrenia. In contrast, cannabis use or dependence in the previous year was not associated with a heightened risk of developing psychosis over the following 12-month period in a group of individuals at ultrahigh risk for developing schizophrenia (Phillips *et al.*, 2002). While the authors concluded that cannabis use did not appear to contribute to the onset of psychosis, they acknowledged

several limitations to the study design, including a low level of cannabis use in the sample and the lack of monitoring of cannabis use.

Cannabis is associated with an earlier onset of psychotic symptoms in schizophrenia: Some studies suggest that cannabis and other substance use is associated with an earlier age of and more abrupt onset of psychotic symptoms in schizophrenic patients (Addington and Addington, 1998; Allebeck *et al.*, 1993a; Andreasson *et al.*, 1987, 1989; Cleghorn *et al.*, 1991; Green *et al.*, 2004; Hambrecht and Hafner, 1996; Linszen *et al.*, 1994; McGuire *et al.*, 1994; Van Mastrigt *et al.*, 2004; Veen *et al.*, 2004).

Parallels in the association between amphetamines and psychosis and cannabis and psychosis: At this juncture, it would be illustrative to review an older but relevant story about the association between cannabis and psychosis. As early as 1938, Young and Scoville first reported an association between amphetamine use and psychosis. Nearly 20 years later, in a seminal report of 42 cases, Connell (1958) reported that high-dose amphetamine use by amphetamine addicts was associated with a florid psychosis. Despite some supporting data (dose-response, temporal association, and phenomenological similarities), there was considerable skepticism about the suggested association between amphetamines and psychosis as reviewed in (D'Souza *et al.*, 1999). First, it was not possible to determine whether amphetamine precipitated latent psychosis or *de novo* psychosis. Second, at the time "thought disorder," which was not a prominent feature described in early reports of amphetamine psychosis, was believed to be fundamental to and diagnostic of schizophrenia in American Psychiatry. Third, the prevailing diagnostic criteria of schizophrenia were poorly defined. Fourth, the role of sleep deprivation induced by amphetamines, in the development of amphetamine psychosis could not be excluded. Finally, it was unclear whether other drugs (sedative hypnotics, marijuana, and hallucinogens) taken along with amphetamines may have contributed to the development of psychosis in amphetamine abusers reviewed in (D'Souza *et al.*, 1999).

Prospective, controlled pharmacological studies with amphetamine provided critical support for the early DA hypothesis by addressing the limitations of naturalistic studies. In a series of studies, amphetamine loading was shown to induce psychosis in nonschizophrenic volunteers that spontaneously resolved (Angrist and Gershon, 1970; Angrist *et al.*, 1971; Bell, 1965, 1973; Griffith *et al.*, 1972). Drawing a parallel, pharmacological studies with cannabinoids address some of the limitations of naturalistic data similar to pharmacological studies with amphetamines. In particular, pharmacological studies offer the advantages of providing more accurate dose-response data, a sample carefully screened for preexisting illness, a more precise estimation of temporality and control of various confounds. While there are several reports of pharmacological studies with cannabinoids in humans, most of the studies were not specifically designed to study psychosis.

IV. Pharmacological Studies

In order to better interpret the pharmacological studies, it would be essential to understand some of the pharmacokinetic issues relevant to cannabis and Δ -9-THC. The pharmacokinetics and effects of Δ -9-THC vary as a function of route of administration. Herbal cannabis and cannabinoid compounds are typically consumed during recreational use by the inhaled or oral route. However, cannabinoids have also been administered for therapeutic or experimental purposes by the intravenous, rectal, sublingual, transdermal, topical (eye drops), and aerosolized route. Δ -9-THC administered by inhalation results in peak plasma concentrations within minutes, with psychotropic effects starting within seconds to a few minutes, reaching a peak after 15–30 min, and then tapering off within 2–3 h (Fig. 3). In attempting to quantify the dose of Δ -9-THC extracted from a typical cannabis joint several factors need to be considered including, but not limited to, the weight of a cannabis joint, the potency of Δ -9-THC in the herbal cannabis preparation, and the presence of other cannabinoids in herbal cannabis that might contribute to the effects of cannabis and/or alter the effects of Δ -9-THC (Karniol and Carlini, 1973; Karniol *et al.*, 1974, 1975; Turner *et al.*, 1980). Further, the amount of THC delivered is influenced by several factors, including the rate of inhalation, depth of puffs, duration of puffs, volume inhaled, extent of breath-holding after inhalation,

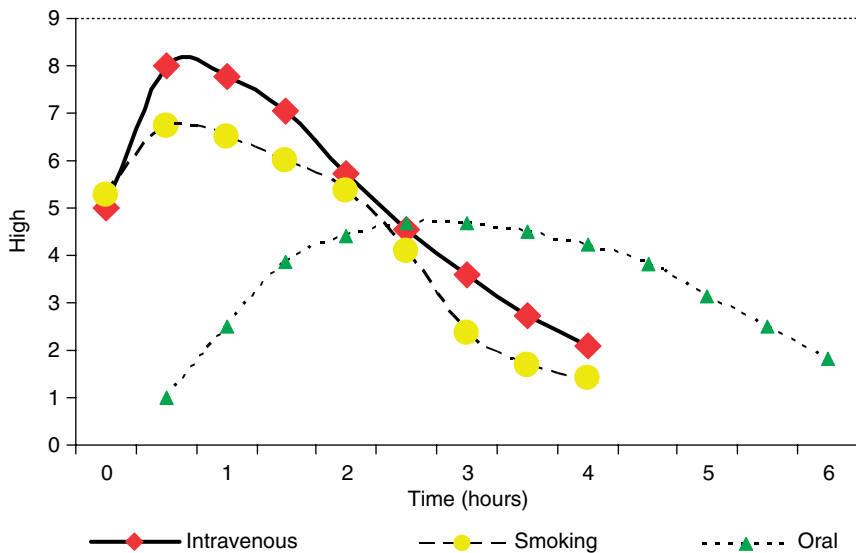


FIG. 3. Time course of subjective effects according to route of administration.

amount lost by smoke escaping into the air or respiratory dead space, vital capacity, length of cigarette smoked, adeptness of smoking, and subject's overall experience in titrating the dose. In fact, only 10–25% of the Δ -9-THC content of a cannabis joint enters the circulation when smoked (Adams and Martin, 1996). Thus, quantifying the typical dose of Δ -9-THC that a typical cannabis joint delivers is not without challenges. Intravenous dosing follows the pharmacokinetics of the inhaled route, though blood levels tend to be higher. Following oral ingestion, psychotropic effects set in with a delay of 30–90 min, reach their maximum after 2–3 h, and last for about 4–12 h (Hollister *et al.*, 1981; Ohlsson *et al.*, 1980, 1981). Nabilone is administered by oral route while levonantradol is administered by intramuscular route.

Psychosis associated with the consumption of medicinal cannabinoids: Cannabinoids including cannabis, natural and synthetic Δ -9-THC, nabilone, and levonantradol have been used in the treatment of chemotherapy-induced nausea, spasticity in multiple sclerosis, pain syndromes, glaucoma, stimulation of appetite, Tourette's syndrome, parkinsonism, dyskinesia, and traumatic brain injury. Adverse events causally linked to Marinol that occurred at >1% in the clinical trials included hallucinations, abnormal thinking, paranoid reaction, amnesia, and so on, all of which are symptoms of psychosis (Marinol Product monograph). Further, the incidence of "disturbing" psychiatric reactions increased with dose escalation. Similarly, studies with oral and intramuscular levonantradol have reported "loss of control," hallucinations, other perceptual alterations, thought disturbance, feelings of unreality, fear and paranoia, apprehension, difficulty concentrating, dissociation, depersonalization, dysphoria, anxiety, and panic (Citron *et al.*, 1985; Cronin *et al.*, 1981; Diasio *et al.*, 1981; Heim *et al.*, 1981, 1984; Jain *et al.*, 1981; Kenny and Wilkinson, 1982; Laszlo *et al.*, 1981; Sheidler *et al.*, 1984; Stuart-Harris *et al.*, 1983). Psychotropic adverse effects increased both with increasing dose and with repeated dosing (Citron *et al.*, 1985; Stambaugh *et al.*, 1984). Further, some subjects refused further testing because of the disturbing psychotropic effects. Nabilone (CESAMETTM) was developed by Eli Lilly and marketed in Europe as an analgesic agent. A "toxic psychosis" has been reported as one of its side effects.

In a systematic review of randomized controlled trials comparing the antiemetic effects of cannabinoids with placebo or other antiemetics, 6% of patients receiving cannabinoids presented with hallucinations and 5% with "paranoia," while no patient treated with control drugs presented with such side effects (Tramer *et al.*, 2001). The same group conducted a systematic review of randomized controlled trials comparing the analgesic effects cannabinoids with placebo or other analgesics (codeine and benzopyranoperidine), but they did not specifically mention psychotic symptoms (Campbell *et al.*, 2001).

Experimental studies: There are a small number of pharmacological studies that were specifically designed to examine the behavioral and/or cognitive effects of

cannabinoids. As far back as the 1940s, pharmacological investigations were conducted under the direction of the "LaGuardia Committee on Marihuana" (Mayor's, 1944). With cannabis doses of about 30–50 mg (oral) and 8–30 mg (smoked), 12.5% of subjects experienced psychotic reactions. However, these subjects were prisoners and their mental status cannot be presumed to be healthy. Ames (1958) studied the effects of unassayed oral doses of cannabis extract (about 50–70 g Δ -9-THC) in 12 medical house staff who were presumably healthy. Subjects reported immediate recall deficits, thought fragmentation, dissociation between thoughts and action, disturbed temporal and spatial perception, visual illusions and hallucinations, derealization and depersonalization, mood alterations, and anxiety. Some subjects reported delusions of the presence of hidden recorders, fear of being hypnotized, fear of ECT, and fear of developing schizophrenia. One subject refused to answer questions for fear of being certified as insane. Isbell and colleagues (1967) studied the effects of varying doses of Δ -9-THC (120–480 μ g/kg orally and 50–250 μ g/kg smoked) in 40 former opiate addicts. At Δ -9-THC 120 μ g/kg orally and 50 μ g/kg smoking, in addition to recognizing the effects as being similar to marijuana, the subjects reported alterations in visual, auditory, and time perception. However, at Δ -9-THC doses of 300–480 μ g/kg orally and 200–250 μ g/kg by smoking there were marked auditory and visual distortions, depersonalization, derealization, and hallucinations. Of note, "occasional" individuals experienced psychosis even at low doses of Δ -9-THC. In a related study, Isbell and Jasinski (1969) compared the effects of Δ -9-THC (75–225 μ g/kg, smoked) and LSD (0.5–1.5) in 10 "normal" controls. Both drugs produced perceptual distortions, mood changes, and at higher doses hallucinations. Of note, two subjects dropped out from the study after experiencing psychotic "reactions" from Δ -9-THC. However, Hollister showed that Δ -9-THC was not associated with as prominent psychotomimetic effects as LSD (reviewed in Hollister, 1986).

Melges *et al.* (1970) in a double-blind, placebo-controlled study with high and low dose Δ -9-THC reported that cannabis users were noted to have core symptoms of psychosis, including thought disorder and paranoia. The authors specifically described "tracking difficulties" that subjects reported, including racing thoughts, thought blocking, losing their train of thought, and so on. Jones and Stone (1970) did not observe robust psychotomimetic effects in studies of "normal" controls with Δ -9-THC (20 mg smoked or 40 mg orally). However, a few subjects reported ideas of reference and delusions that the researcher was using secret (unexplained) tests and hidden recording devices. At doses higher than 20 mg smoked or 40 mg orally, psychotomimetic effects including delusions, loosening of associations, and marked illusions began to emerge. In a ^{18}F -2-fluoro-2-deoxyglucose positron emission tomography (FDG-PET) study of intravenous Δ -9-THC (2 mg) on regional brain metabolism, two of eight healthy subjects who occasionally used cannabis, experienced paranoid-anxious reactions (Volkow *et al.*, 1991).

The pharmacological studies discussed thus far had several limitations, including the absence of placebo/control, lack of a double-blind, inclusion of psychiatrically ill individuals, and lack of standardized measures of psychosis. Recently, there have been a few laboratory studies examining the psychotogenic effects of cannabinoids that address some of these limitations.

Leweke *et al.* (1999b) reported the effects of synthetic Δ -9-THC (120 μ g/kg) by oral route in 17 healthy individuals under controlled laboratory conditions. The study included subjects with past experience but no recent consumption of cannabinoids. The overall lifetime consumption of cannabinoids was limited to 10 times to exclude the long-term effects of cannabis use. Subjects with a history of recurrent abuse of illicit drugs other than cannabinoids or other psychiatric disorders were excluded. The primary outcome measure was binocular depth perception which has been described as a model of illusionary perception. While the study was not placebo controlled, subjects were told that they might receive a placebo or active drug, but in fact they always received active drug. Subjective reactions ranged from mild euphoria to more pronounced reactions, including feelings of loss of self-control and body distortion suggestive of psychotic-like symptoms. One subject experienced a transient psychotic episode described as "a paranoid psychotic state with persecutory delusions, delusions of thought insertion, attentional irritability, fear, and—to some extent—verbal aggressive behavior." These symptoms resolved spontaneously within minutes to hours. Leweke *et al.* (2000) repeated the study with nabilone, a synthetic analogue of Δ -9-THC, and observed effects on binocular depth inversion similar to that of Δ -9-THC (Leweke *et al.*, 2000).

D'Souza *et al.* (2004) characterized the behavioral and cognitive effects of Δ -9-THC in a double-blind, placebo-controlled study of healthy controls ($n = 22$). Only subjects with past cannabis experience but without lifetime cannabis abuse or dependence were included. Healthy subjects also underwent a Structured Clinical Interview for DSM-IV (healthy) and an unstructured psychiatric evaluation. Subjects were excluded if they had any significant psychiatric disorder and/or a family history of any DSM Axis I disorder. Subjects received in random order 5 or 2.5 mg of Δ -9-THC, or vehicle by intravenous route over 2 min. Positive and negative symptoms of psychosis were measured using the Positive and Negative Syndrome Scale (PANSS). Perceptual alterations that did not quite meet the threshold of psychosis were measured using the Clinician Administered Dissociative Symptoms Scale (CADSS). Cognitive symptoms were measured using tests of immediate recall (learning) and delayed recall, verbal fluency, working memory, and vigilance and distractibility. Δ -9-THC produced transient positive symptoms (Fig. 4), perceptual alterations, negative symptoms, euphoria, anxiety, and deficits in working memory and verbal recall, and the executive control of attention without altering general orientation.

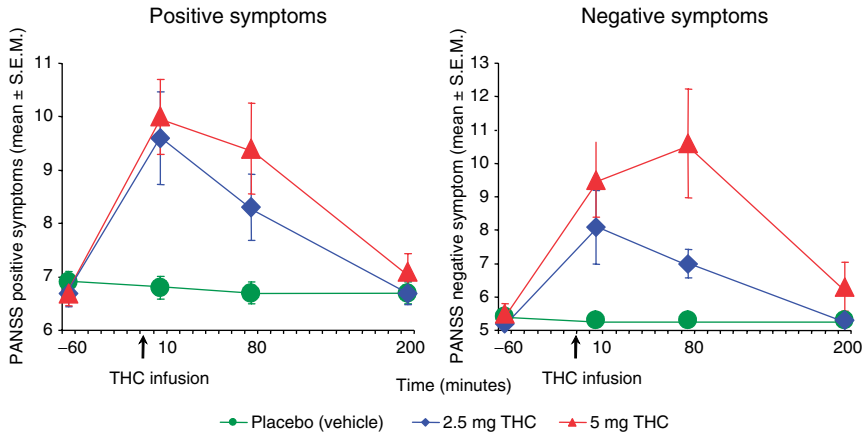


FIG. 4. Δ -9-THC induces positive and negative symptoms in healthy individuals; Positive and Negative Syndrome Scale.

Positive symptoms and perceptual alterations: The positive symptoms induced by Δ -9-THC included suspiciousness, paranoid and grandiose delusions, conceptual disorganization, and illusions. For example, healthy controls reported suspiciousness such as “I thought you all were *trying to trick me* by changing the rules of the tests to make me fail. I thought you were turning the clock back to confuse me,” or “I thought that this was real . . . I was convinced this wasn’t an experiment,” or “I thought you all were giving me THC thru the BP (blood pressure) machine and the sheets.” Healthy controls also reported conceptual disorganization such as “I couldn’t keep track of my thoughts . . . they’d suddenly disappear,” or “It seemed as if all the questions were coming to me at once . . . everything was happening in staccato,” or “my thoughts were fragmented. . . the past present and future all seemed to happening at once.” Healthy subjects also reported unusual thoughts such as “I thought you could read my mind, that’s why I didn’t answer . . . I felt as if my mind was nude,” or “I felt I could see into the future . . . I thought I was God.” These effects reported by carefully screened healthy subjects appear to be remarkably similar to the kinds of psychotic symptoms reported by patients with schizophrenia.

An identical study was conducted in parallel in medicated schizophrenic patients. Δ -9-THC *transiently* exacerbated a range of positive and negative symptoms, perceptual alterations, cognitive deficits, and medication side effects associated with schizophrenia without producing any obvious “beneficial” effects. The increases in psychosis were brief, modest, and occurred even though subjects were clinically stable, medication-responsive, and were receiving therapeutic

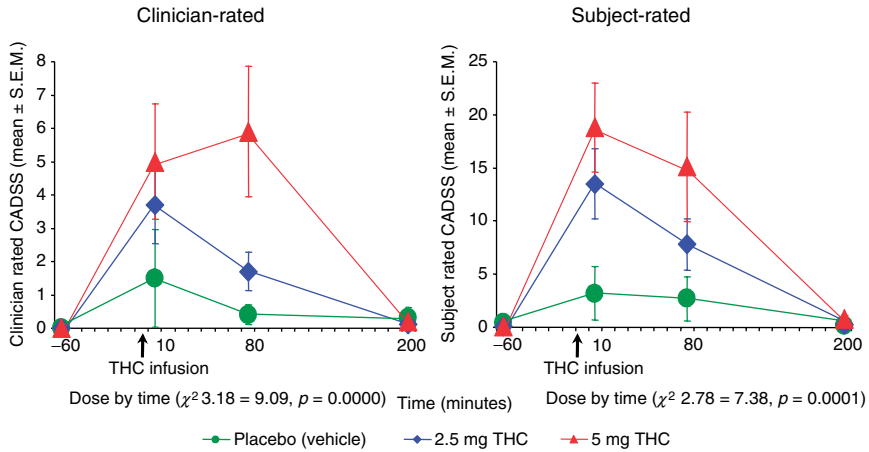


FIG. 6. Δ -9-THC induces transient perceptual alterations in healthy individuals; Clinician Administered Dissociative Symptoms Scale.

psychomotor retardation, and emotional withdrawal (Fig. 4). How much sedation and/or being inwardly preoccupied contributed to the increased ratings of negative symptoms was unclear. These schizophrenia-like negative symptoms may have been confounded by the known cataleptic and sedating effects of Δ -9-THC. Besides, acute pharmacological studies may have limitations in their capacity to “model” negative symptoms. Finally, Δ -9-THC transiently increased negative symptoms in schizophrenic patients.

Of note, a persistent “amotivational syndrome” has been described in chronic heavy cannabis users by some (Halikas *et al.*, 1982; Hall and Solowij, 1998; Kolansky and Moore, 1971b; Millman and Sbriglio, 1986; Tennant and Groesbeck, 1972b) but not others (Carter *et al.*, 1980; Hollister, 1988; Rubin and Comitas, 1975). This so-called “amotivational syndrome” is characterized by apathy, amotivation, social withdrawal, narrowing of interests, lethargy, impaired memory, impaired concentration, disturbed judgment, and impaired occupational achievement. The syndrome has a striking phenomenological similarity with the negative dimension of psychosis and appears to be dose related. However, other drug use, poverty, low socio-economic status, or preexisting psychiatric disorders existing data may confound the interpretation of the existing literature.

Cognitive deficits: The most consistent acute cognitive effects of cannabinoids in humans include deficits in learning, short-term memory, working memory, and attention (Hart *et al.*, 2001; Heishman *et al.*, 1990; Hooker and Jones, 1987; Leweke *et al.*, 1998; Marks and MacAvoy, 1989; Miller *et al.*, 1977). These are also the cognitive deficits observed in schizophrenia (Heinrichs and Zakzanis, 1998). Of note, the most robust effects of cannabinoids are on verbal memory

reviewed in Ranganathan and D'Souza (2006), the latter is also the most robust cognitive deficit observed in schizophrenia (Heinrichs and Zakzanis, 1998).

In healthy subjects (hatched lines), Δ -9-THC significantly impaired immediate free recall in a dose-dependent manner across all three trials of immediate recall (Fig. 7). Δ -9-THC also impaired delayed (+30 min) free recall and delayed cued recall in a significant, dose-dependent manner. The effect on delayed recognition recall trended toward significance. Finally, Δ -9-THC increased the number of false positives and intrusions with a trend toward significance. Relative to controls, schizophrenia patients were specifically more vulnerable to the dose-related learning impairments produced by Δ -9-THC (D'Souza *et al.*, 2005). Under the influence of 5 mg Δ -9-THC schizophrenia patients (solid lines) showed no learning whatsoever (Fig. 7). Δ -9-THC also increased the number of intrusions and false positives generated during recall. Further, 5 mg Δ -9-THC reduced learning and recall in healthy controls to the level of schizophrenia patients on the placebo condition.

While the acute transient effects of cannabinoids on memory are quite clear, whether cannabinoids produce impairments in memory that persist beyond the period of intoxication, remains inconclusive. Heavy and prolonged cannabis exposure may be associated with deficits in memory, sustained attention, and executive functioning (Bolla *et al.*, 2002; Pope and Yurgelun-Todd, 1996; Pope *et al.*, 1995; Solowij, 1995; Solowij *et al.*, 1995, 2002). Eleven of 22 (50%) studies

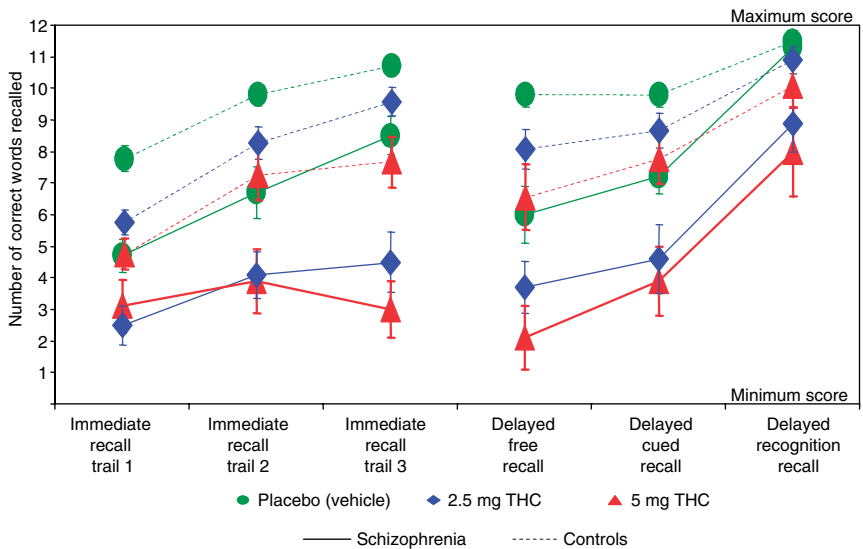


FIG. 7. Δ -9-THC induces memory impairments; Hopkins Verbal Learning Test.

involving clinical samples (cannabis abusers) and 5 of 7 (75%) population-based samples support an association between cannabis use (excluding intoxication) and cognitive deficits. With a few exceptions, the literature has limitations including (1) the lack of any measures of cognitive functioning prior to onset of cannabis use, (2) small samples (range 9–63; median = 26), (3) selection bias, (4) confound of other drug and/or alcohol use, and (5) sensitivity of the cognitive measures, and so on.

In a meta-analysis of 15 studies, Gonzalez *et al.* (2002) concluded that a majority of studies found evidence for persistent but subtle cognitive deficits associated with nonacute (remote) cannabis use. However, whether these cognitive deficits are reversible and persist despite cessation of cannabis use remains an open question. In the first published report examining whether the residual cognitive effects of cannabis persist beyond a period of abstinence longer than 12–72 h, Pope *et al.* (2001) found that deficits in cognitive test performance in cannabis abusers that were present at 7 days normalized by day 28 (Pope *et al.*, 2001). In contrast, Bolla *et al.* (2002) found that cognitive test performance in cannabis abusers with a history of very heavy use of cannabis showed persistent decrements in learning and recall, executive function, psychomotor speed, complex reaction time, and manual dexterity even after 28 days of abstinence. The magnitude of the difference in mean performance between the heavy and light users was between 1.0 and 3.3 S.D. units (Bolla *et al.*, 2002). Similarly, the magnitude of the association between cannabis use and decreasing Wisconsin Card Test Performance (executive functioning) was large (4.1–4.2 S.D. units).

The question of enduring deficits associated with cannabis abuse are only beginning to be investigated with more sensitive measures such as brain imaging and electrophysiology. Cannabis abusers have altered regional cerebral blood flow (rCBF) at rest in several brain regions even after 26 h of monitored abstinence (Block *et al.*, 2000). Similarly, Block *et al.*, have shown decreases in memory-related blood flow in prefrontal cortex in frequent cannabis users in the unintoxicated state relative to nonusers, increases in memory-relevant regions of cerebellum, and altered lateralization in hippocampus. The speed of information processing, measured by the latency of parietal P300, was delayed significantly with increasing frequency of use (Solowij *et al.*, 1995). In summary, there is some overlap between the cognitive effects of cannabinoids and the cognitive deficits observed in schizophrenia.

Δ -9-THC produces a plethora of effects including euphoria, a calm and relaxed feeling state, psychotomimetic symptoms, tachycardia, etc. in the absence of any change in orientation. While some but not other studies reviewed suggest that cannabis can induce a broad range of transient effects in healthy individuals that share some similarities with some, though not all, of the symptoms of schizophrenia, the data from these acute studies do not, however, address the

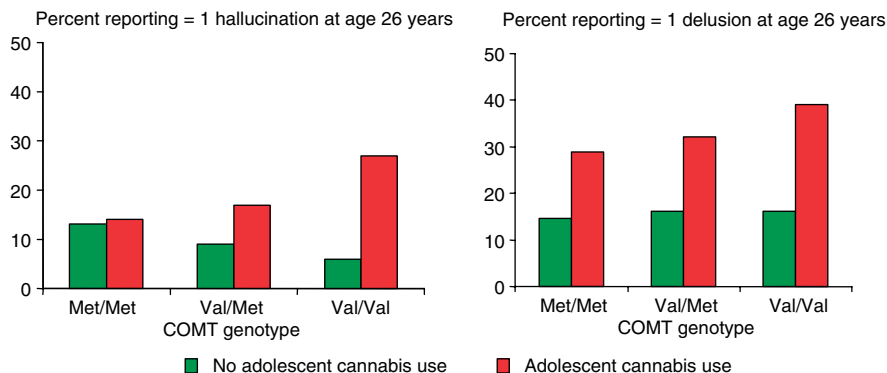


FIG. 8. COMT gene polymorphism moderates influence of adolescent cannabis use on developing psychosis in adulthood.

question of whether cannabis can cause a chronic psychotic disorder such as schizophrenia, that persists through life.

What puts some individuals at higher risk for experiencing psychotic symptoms following exposure to cannabinoids? Individuals with a vulnerability to psychosis as estimated by a psychosis proneness scale were more likely to report psychotic symptoms with cannabis use (Verdoux *et al.*, 2003b). In a preliminary report, no structural mutations in CB1R were found in individuals who developed acute psychotic symptoms after cannabis intake (Hoehe *et al.*, 2000). Caspi *et al.* (2005) reported that a polymorphism of the catechol-*O*-methyl transferase (COMT) gene modulates the risk of schizophrenia conferred by cannabis (Fig. 8). After adolescent exposure to cannabis, individuals homozygous for the COMT valine158 allele were most likely to exhibit psychotic symptoms and to develop schizophreniform disorder later. The effects of cannabinoids on dopamine function may be involved in this gene by cannabis interaction.

V. Cannabis and Psychosis: Causality

Some of the criteria that have been used to establish disease causality include temporality, strength of the association, direction, dose-response or biological gradient, consistency, specificity, coherence, strength of the relationship, experimental evidence, and biologic plausibility (Aiello and Larson, 2002; Hill, 1965).

Most studies provide evidence of direction by showing that the association between cannabis use and psychosis persists even after controlling for many potential confounding variables such as gender, age, ethnicity, low IQ, level of

education, urbanicity, disturbed behavior, cigarette smoking, poor social integration, unemployment, single marital status, and previous psychotic symptoms. While there is a strong association between cigarette smoking and schizophrenia, there is little evidence to support the notion that cigarette smoking “causes” schizophrenia. In contrast, the high rates of cigarette smoking in schizophrenia may reflect an attempt by schizophrenia patients to self-medicate deficits in information processing. Several studies reviewed here provide evidence of a dose–response relationship between cannabis exposure and the risk of psychosis. With regard to temporality, some, though not all, studies suggest that cannabis use precedes or coincides with the onset of psychosis in about two-third of patients with schizophrenia who use cannabis. Related to this, cannabis use may be associated with an earlier age of onset of schizophrenia. Further, there is also evidence that the earlier the onset of cannabis use, the stronger the effect on schizophrenia outcomes. There is also some evidence for both the relative specificity of exposure (i.e., cannabis) and specificity of the outcome (i.e., psychosis). Experimental evidence from laboratory studies suggests that cannabinoids can induce a wide range of transient schizophrenia-like symptoms in healthy individuals and relative to controls, schizophrenia patients are more vulnerable to the psychotomimetic effects of Δ -9-THC. Whether exposure to cannabis can result in a chronic psychotic state that persists beyond the period of intoxication is unclear.

However, not all patients with psychosis have been exposed to cannabis and not all cannabis users develop psychosis. Furthermore, there is a disparity in the incidence and prevalence of cannabis abuse (7–12%) and that of schizophrenia (1–2%), and despite different rates of cannabis consumption across the globe, there is relative uniformity in the incidence of schizophrenia. Further, the increase in cannabis use and the use of more potent forms of cannabis in certain geographical areas has not been accompanied or followed by a commensurate increase in the rates of schizophrenia (Degenhardt *et al.*, 2003). Similarly, if cannabis use is associated with an earlier age of onset, then the increase rates of cannabis use should result in a trend toward a lower age of onset of schizophrenia. This does not seem to be the case.

Taken collectively, it appears that cannabis is neither a necessary nor a sufficient cause of schizophrenia. Similarly, cigarette smoking is neither a necessary nor a sufficient cause of lung cancer. More likely, cannabis exposure is a component or contributing cause which interacts with other known, for example, genetic and heretofore unknown factors, leading to schizophrenia. In terms of strength, cannabis confers about a two- to threefold increase in the relative risk for schizophrenia. Arsenault *et al.* (2004) have suggested that the number of cases of schizophrenia in a population that could be eliminated by removal of cannabis use, the population attributable fraction (PAF), is about 8% (Arsenault *et al.*, 2004). In the absence of known causes of schizophrenia, the role of component causes such as cannabinoid exposure remains important and warrants further study.

As reviewed by Piomelli, advances in the understanding of brain cannabinoid receptor function now offer several biologically plausible mechanisms by which cannabis exposure might induce psychosis. While it is out of the scope of this chapter, the interactions between cannabinoid receptor function and dopamine, glutamate and GABA receptor function provide potential mechanisms by which cannabis contribute to the pathophysiology of psychosis reviewed in D'Souza *et al.* (2004, 2005).

VI. Cannabinoid Receptor Dysfunction and Psychotic Disorders

Emerging findings from postmortem (Dean *et al.*, 2001; Zavitsanou *et al.*, 2004), neurochemical (Giuffrida *et al.*, 2004; Leweke *et al.*, 1999a), and genetic (Ujike *et al.*, 2002) studies suggest that cannabinoid receptor system dysfunction that may contribute to the pathophysiology of schizophrenia.

Leweke *et al.* (1999a) and colleagues were the first to suggest altered cannabinoid receptor function in schizophrenia. Levels of anandamide, 2-AG, palmitoylethanolamide (PEA), and a noncannabinoid acylethanolamide, oleylethanolamide (OEA) as a positive control, were measured in cerebrospinal fluid (CSF) sampled from 10 schizophrenia patients and 11 healthy controls (Fig. 9). Mean anandamide and PEA levels were approximately twofold higher in the schizophrenia patients. These differences could not be attributable to drug use, and neither age, gender nor medication correlated with CSF endocannabinoid levels. However, since some subjects were neuroleptic naïve and others were not, it was not possible to fully exclude an effect of antipsychotic drugs.

These data were replicated in a larger sample and the confound of medication status was also addressed (Giuffrida *et al.*, 2004). CSF anandamide levels were eightfold higher in antipsychotic-naïve first-episode paranoid schizophrenics than in healthy and psychiatric controls (Fig. 10). The elevations in CSF anandamide seen in antipsychotic naïve schizophrenic patients were absent in schizophrenics treated with "typical" antipsychotics but not in those treated with "atypical" antipsychotics ($n = 34$). Finally, CSF anandamide levels negatively correlated with psychotic symptoms in unmedicated patients. Since anandamide release may serve as an inhibitory feedback signal countering dopamine activation, the increase in CSF anandamide levels in unmedicated acutely psychotic patients was interpreted as a compensatory increase in endocannabinoids secondary to psychosis-related hyperdopaminergia. While CSF studies of endocannabinoids have served to draw attention to possible endocannabinoid dysfunction in schizophrenia, they are not without limitations. Endocannabinoids are very challenging to assay. Anandamide has a very short half-life and therefore differences in collection and processing of samples might explain the group differences. Finally, CSF

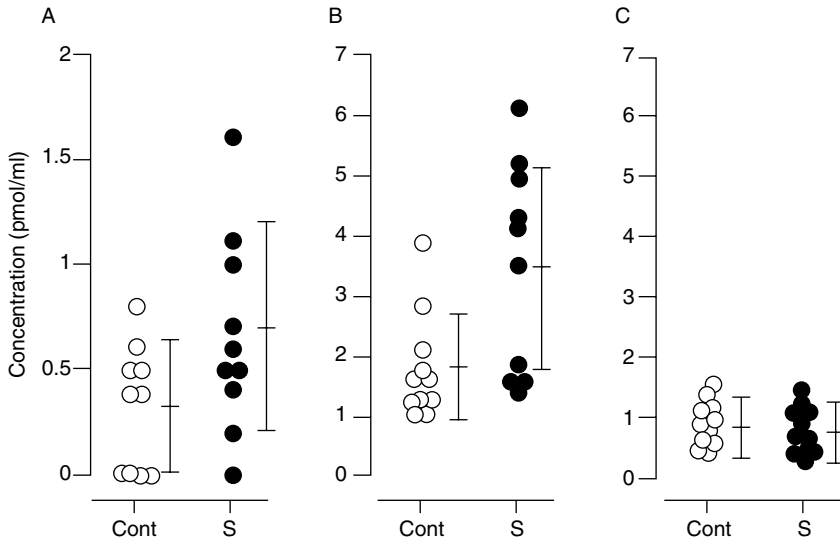


FIG. 9. Cerebrospinal fluid endocannabinoid levels in schizophrenia. Concentrations of anandamide (A), palmitylethanolamide (B), and OEA, oleylethanolamide (C) in CSF of schizophrenic (S) and nonschizophrenic (Cont) subjects. Bars indicate mean \pm S.D. for each group [(A) and (B), $p < 0.05$; Student's t -test]; Lewke *et al.* (1999).

endocannabinoid levels may reflect global rather than regional changes. Studies directly examining CB1 receptor may address some of these limitations.

There are two postmortem studies of CB1 receptor changes in schizophrenia in the literature. Compared with control subjects, schizophrenia patients showed a 19% increase ($p < 0.05$) in CB1 receptor density only in the dorsolateral prefrontal cortex (DLPFC) (Dean *et al.*, 2001). These differences could not be attributable to postmortem interval, brain pH, age, or gender. Further, there were no significant correlations between CB-1R binding and duration of illness or antipsychotic drug dose in those with schizophrenia. Of note, chronic antipsychotic drug treatment study in rats does not result in changes in CB1-receptor binding in the cerebral cortex, caudate-putamen (CP), or hippocampus (Sundram *et al.*, 2004). There were no significant differences between the groups in the CP or hippocampal formation. In subjects who had recently consumed cannabis, there was a significant ($p < 0.05$) 23% increase in CB1 receptor density in the CP compared to nonusers independent of schizophrenia. Zavitsanou *et al.* (2004) compared CB1 receptor in the anterior cingulate cortex (ACC) taken postmortem from patients with schizophrenia ($n = 10$) and matched control subjects ($n = 9$) using [^3H]SR141716A. Compared to the control group schizophrenia patients had a significant 64% increase in CB1 receptor density in the ACC. The effects of

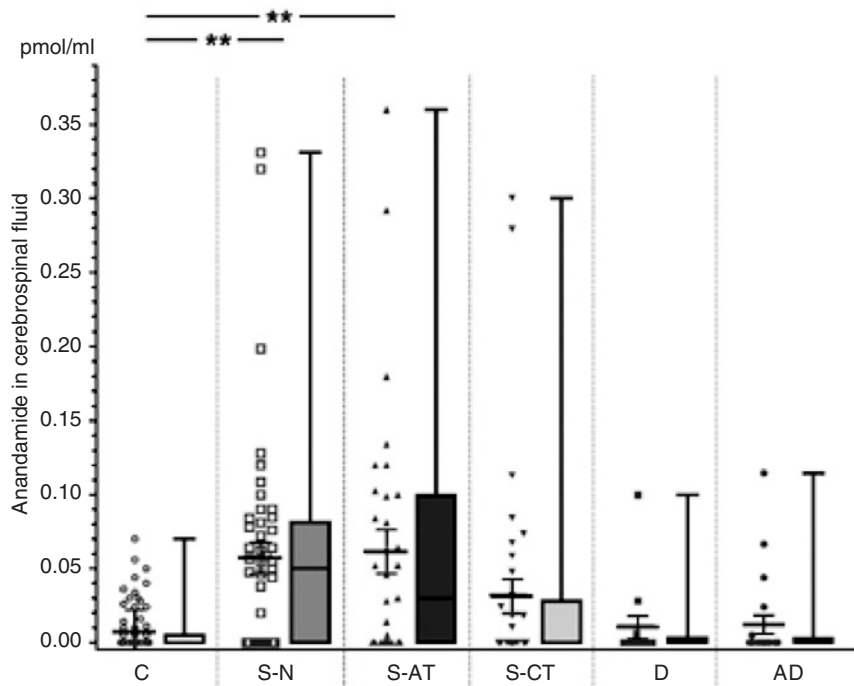


FIG. 10. Elevated anandamide in the CSF of antipsychotic-free first-episode schizophrenic patients. CSF anandamide levels in healthy volunteers (C); antipsychotic-free schizophrenics with paranoid schizophrenia (S-N); acute schizophrenics (paranoid type) on “atypical” (S-AT) or “typical” (S-CT) antipsychotic drugs; and patients with dementia (D) or affective disorders (AD) without psychotic symptoms. Single values with mean \pm SEM as well as corresponding ox-plots illustrating median, range, and quartiles for each group. Statistically significant differences between groups are shown (* $p < 0.01$; ** $p < 0.001$; Kruskal-Wallis test followed by Mann–Whitney test; a Bonferroni correction (5 tests) of the α -level was applied: $\alpha = 0.05/5 = 0.01$).

antipsychotic treatment or premorbid cannabis use explaining the group differences could not be ruled out. Together the postmortem studies provide some preliminary evidence suggestive of endocannabinoids dysfunction in schizophrenia. The development of good radioligands that permit *in vivo* imaging of the CB1 receptor in schizophrenia patients would help confirm the limited postmortem findings.

Finally, several groups have looked for associations between schizophrenia and genes relevant to cannabinoid receptor function. The CB1 receptor gene is mapped to chromosome 6q14–15, and linkage studies have produced evidence for a schizophrenia-susceptibility locus in this region. Two polymorphisms for the

CB1 receptor gene have been identified, a triplet repeat (AAT) n in the 3'-flanking region (Dawson *et al.*, 1995) and a biallelic silent mutation of 1359 G-to-A at the 453 codon in the coding exon (Gadzicki *et al.*, 1999). Dawson failed to show a significant association between the (AAT) n triplet repeat polymorphism of the CB1 receptor gene and schizophrenia in 135 schizophrenic subjects compared to 101 control subjects (Dawson, 1995). Similarly, Tsai replicated these findings in a study comparing 127 subjects with schizophrenia and 146 control subjects in a Han Chinese population (Tsai *et al.*, 2000). They concluded the (AAT) n triplet repeat in the promoter region of the CB1 receptor gene is not directly involved in the pathogenesis of schizophrenia in Chinese populations. In contrast, Ujike *et al.* (2002) reported that the (AAT) n triplet repeat polymorphism of the CB1 receptor gene was significantly associated with patients with schizophrenia, especially the hebephrenic subtype. The functional effect of this triplet repeat on the CB1 receptor gene transcription rate remains unclear. Leroy studied the 1359 A/G polymorphism in a French Caucasian sample of 102 subjects with schizophrenia and 63 healthy controls (Leroy *et al.*, 2001). Overall there were no significant differences between the two groups either in allele frequency or genotype distribution. However, when the patient group was divided into a substance using ($n = 42$) and nonusing, they found a significant decrease in homozygosity for the G allele in nonusers compared to users ($p < 0.04$). Fatty acid amide hydrolase (FAAH) is the primary catabolic enzyme of endocannabinoids. Morita *et al.* (2005) found no significant association of a nonsynonymous polymorphism (Pro129Thr) of the FAAH gene with schizophrenia.

VII. Summary and Conclusions

Cannabinoids can induce acute transient psychotic symptoms or an acute psychosis in some individuals. What makes some individuals vulnerable to cannabinoid-related psychosis is unclear. Cannabinoids can also exacerbate psychosis in individuals with an established psychotic disorder, and these exacerbations may last beyond the period of intoxication. Less clear is whether cannabis causes a persistent *de novo* psychosis. The available evidence meets many but not all the criteria for causality, including dose-response, temporality, direction, specificity, and biological plausibility. On the other hand, the large majority of individuals exposed to cannabinoids do not experience psychosis or develop schizophrenia. It is more likely that cannabis exposure is a component cause that interacts with other factors, for example, genetic risk, to "cause" schizophrenia. Nevertheless, in the absence of known causes of schizophrenia, the role of component causes such as cannabis exposure (exogenous hypothesis), is important and warrants further study. There is also tantalizing evidence from postmortem,

neurochemical, and genetic studies, suggesting CB1 receptor dysfunction (endogenous hypothesis) in schizophrenia that warrants further investigation. Further work is necessary to identify those factors that place individuals at higher risk cannabinoid-related psychosis, to identify the biological mechanisms underlying the risks and to further study whether CB1 receptor dysfunction contributes to the pathophysiology of psychotic disorders. Finally, from a treatment perspective, given the negative impact of cannabis use on the course and expression of schizophrenia, and the potential for precipitating psychosis in individuals at risk for schizophrenia, efforts need to be directed toward developing effective treatments for cannabis use disorders.

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References

- Adams, I. B., and Martin, B. R. (1996). Cannabis: Pharmacology and toxicology in animals and humans. *Addiction* **91**, 1585-1614.
- Addington, J., and Addington, D. (1998). Effect of substance misuse in early psychosis. *Br. J. Psychiatry Suppl.* **172**, 134-136.
- Agurell, S., Halldin, M., Lindgren, J. E., Ohlsson, A., Widman, M., Gillespie, H., and Hollister, L. (1986). Pharmacokinetics and metabolism of delta 1-tetrahydrocannabinol and other cannabinoids with emphasis on man. *Pharmacol. Rev.* **38**, 21-43.
- Aiello, A. E., and Larson, E. L. (2002). Causal inference: The case of hygiene and health. *Am. J. Infect. Control* **30**, 503-511.
- Allebeck, P., Adamsson, C., Engstrom, A., and Rydberg, U. (1993a). Cannabis and schizophrenia: A longitudinal study of cases treated in Stockholm County. *Acta Psychiatr. Scand.* **88**, 21-24.
- Allebeck, P., Adamsson, C., Engstrom, A., and Rydberg, U. (1993b). Cannabis and schizophrenia: A longitudinal study of cases treated in Stockholm County. [erratum appears in *Acta. Psychiatr. Scand.* (1993) **88**(4), 304]. *Acta Psychiatr. Scand.* **88**, 21-24.
- Ames, F. (1958). A clinical and metabolic study of acute intoxication with cannabis sativa and its role in model psychoses. *J. Ment. Sci.* **104**, 972-999.
- Andreasson, S., Allebeck, P., Engstrom, A., and Rydberg, U. (1987). Cannabis and schizophrenia. A longitudinal study of Swedish conscripts. *Lancet* **2**, 1483-1486.
- Andreasson, S., Allebeck, P., and Rydberg, U. (1989). Schizophrenia in users and nonusers of cannabis. A longitudinal study in Stockholm County. *Acta Psychiatr. Scand.* **79**, 505-510.
- Angrist, B. M., and Gershon, S. (1970). The phenomenology of experimentally induced amphetamine psychosis: Preliminary observations. *Biol. Psychiatry* **2**, 95-107.

- Angrist, B. M., Shopsin, B., and Gershon, S. (1971). Comparative psychotomimetic effects of stereoisomers of amphetamine. *Nature* **234**, 152–153.
- Arendt, M., Rosenberg, R., Foldager, L., Perto, G., and Munk-Jorgensen, P. (2005). Cannabis-induced psychosis and subsequent schizophrenia-spectrum disorders: Follow-up study of 535 incident cases. *Br. J. Psychiatry* **187**, 510–515.
- Arseneault, L., Cannon, M., Poulton, R., Murray, R., Caspi, A., and Moffitt, T. E. (2002). Cannabis use in adolescence and risk for adult psychosis: Longitudinal prospective study. *BMJ* **325**, 1212–1213.
- Arseneault, L., Cannon, M., Witton, J., and Murray, R. (2004). “Cannabis as a Potential Causal Factor in Schizophrenia.” Cambridge University Press, New York.
- Basu, D., Malhotra, A., Bhagat, A., and Varma, V. K. (1999). Cannabis psychosis and acute schizophrenia: A case-control study from India. *Eur. Addict. Res.* **5**, 71–73.
- Bell, D. S. (1965). Comparison of amphetamine psychosis and schizophrenia. *Br. J. Psychiatry* **111**, 701–707.
- Bell, D. S. (1973). The experimental reproduction of amphetamine psychosis. *Arch. Gen. Psychiatry* **29**, 35–40.
- Bernhardson, G., and Gunne, L. M. (1972). Forty-six cases of psychosis in cannabis abusers. *Int. J. Addict.* **7**, 9–16.
- Block, R. I., O’Leary, D. S., Hichwa, R. D., Augustinack, J. C., Ponto, L. L., Ghoneim, M. M., Arndt, S., Ehrhardt, J. C., Hurtig, R. R., Watkins, G. L., Hall, J. A., Nathan, P. E., et al. (2000). Cerebellar hypoactivity in frequent marijuana users. *Neuroreport* **11**, 749–753.
- Bolla, K. I., Brown, K., Eldreth, D., Tate, K., and Cadet, J. L. (2002). Dose-related neurocognitive effects of marijuana use. *Neurology* **59**, 1337–1343.
- Bornheim, L. M., Kim, K. Y., Li, J., Perotti, B. Y., and Benet, L. Z. (1995). Effect of cannabidiol pretreatment on the kinetics of tetrahydrocannabinol metabolites in mouse brain. *Drug Metab. Dispos.* **23**, 825–831.
- Bromberg, W. (1939). Marihuana: A psychiatric study. *JAMA* **113**, 4–12.
- Campbell, F. A., Tramer, M. R., Carroll, D., Reynolds, D. J., Moore, R. A., and McQuay, H. J. (2001). Are cannabinoids an effective and safe treatment option in the management of pain? A qualitative systematic review. *BMJ* **323**, 13–16.
- Carney, M. W., Bacelle, L., and Robinson, B. (1984). Psychosis after cannabis abuse. *Br. Med. J. Clin. Res. Ed.* **288**, 1047.
- Carter, W. E., Coggins, W., and Doughty, P. L. (1980). Cannabis in Costa Rica: A study of chronic marijuana use.
- Caspi, A., Moffitt, T. E., Cannon, M., McClay, J., Murray, R., Harrington, H., Taylor, A., Arseneault, L., Williams, B., Braithwaite, A., Poulton, R., and Craig, I. W. (2005). Moderation of the effect of adolescent-onset cannabis use on adult psychosis by a functional polymorphism in the catechol-O-methyltransferase gene: Longitudinal evidence of a gene X environment interaction. *Biol. Psychiatry* **57**, 1117–1127.
- Chaudry, H. R., Moss, H. B., Bashir, A., and Suliman, T. (1991). Cannabis psychosis following bhang ingestion. *Br. J. Addict.* **86**, 1075–1081.
- Chopra, G. S. (1973). Studies on psycho-clinical aspects of long-term marihuana use in 124 cases. *Int. J. Addict.* **8**, 1015–1026.
- Chopra, G. S., and Smith, J. W. (1974). Psychotic reactions following cannabis use in East Indians. *Arch. Gen. Psychiatry* **30**, 24–27.
- Citron, M. L., Herman, T. S., Vreeland, F., Krasnow, S. H., Fossieck, B. E., Jr., Harwood, S., Franklin, R., and Cohen, M. H. (1985). Antiemetic efficacy of levonantradol compared to delta-9-tetrahydrocannabinol for chemotherapy-induced nausea and vomiting. *Cancer Treat. Rep.* **69**, 109–112.

- Cleghorn, J. M., Kaplan, R. D., Szechtman, B., Szechtman, H., Brown, G. M., and Franco, S. (1991). Substance abuse and schizophrenia: Effect on symptoms but not on neurocognitive function. *J. Clin. Psychiatry* **52**, 26–30.
- Cohen, S. I. (1994). Cannabis consumption and schizophrenia. *Br. J. Psychiatry* **165**, 410–411.
- Connel, P. H. (1958). *Amphetamine Psychoses*. Oxford University Press. London.
- Cronin, C. M., Sallan, S. E., Gelber, R., Lucas, V. S., and Laszlo, J. (1981). Antiemetic effect of intramuscular levonantradol in patients receiving anticancer chemotherapy. *J. Clin. Pharmacol.* **21**, 43S–50S.
- D'Souza, D. C., Berman, R. M., Krystal, J. H., and Charney, D. S. (1999). Symptom provocation studies in psychiatric disorders: Scientific value, risks, and future. *Biol. Psychiatry* **46**, 1060–1080.
- D'Souza, D. C., Perry, E., MacDougall, L., Ammerman, Y., Cooper, T., Wu, Y. T., Braley, G., Gueorguieva, R., and Krystal, J. H. (2004). The psychotomimetic effects of intravenous delta-9-tetrahydrocannabinol in healthy individuals: Implications for psychosis. *Neuropsychopharmacology* **29**, 1558–1572.
- D'Souza, D. C., Abi-Saab, W. M., Madonick, S., Forselius-Bielen, K., Doersch, A., Braley, G., Gueorguieva, R., Cooper, T. B., and Krystal, J. H. (2005). Delta-9-tetrahydrocannabinol effects in schizophrenia: Implications for cognition, psychosis, and addiction. *Biol. Psychiatry* **57**, 594–608.
- Dawson, E., Powell, J. F., Sham, P., Shaikh, S., Taylor, C., Clements, A., Asherson, P., Sargeant, M., Collier, D., and Nanko, S. (1995). Systematic search for major genes in schizophrenia: Methodological issues and results from chromosome 12. *Am. J. Med. Genet.* **60**, 424–433.
- Dean, B., Sundram, S., Bradbury, R., Scarr, E., and Copolov, D. (2001). Studies on [3H]CP-55940 binding in the human central nervous system: Regional specific changes in density of cannabinoid-1 receptors associated with schizophrenia and cannabis use. *Neuroscience* **103**, 9–15.
- Degenhardt, L., Hall, W., and Lynskey, M. (2001). Alcohol, cannabis and tobacco use among Australians: A comparison of their associations with other drug use and use disorders, affective and anxiety disorders, and psychosis. *Addiction* **96**, 1603–1614.
- Degenhardt, L., Hall, W., and Lynskey, M. (2003). Testing hypotheses about the relationship between cannabis use and psychosis. *Drug Alcohol. Depend.* **71**, 37–48.
- Diasio, R. B., Ettinger, D. S., and Satterwhite, B. E. (1981). Oral levonantradol in the treatment of chemotherapy-induced emesis: Preliminary observations. *J. Clin. Pharmacol.* **21**, 81S–85S.
- Dumas, P., Saoud, M., Bouafia, S., Gutknecht, C., Ecohard, R., Dalery, J., Rochet, T., and d'Amato, T. (2002). Cannabis use correlates with schizotypal personality traits in healthy students. *Psychiatry Res.* **109**, 27–35.
- Elsohly, M. A., and Slade, D. (2005). Chemical constituents of marijuana: The complex mixture of natural cannabinoids. *Life Sci.* **78**, 539–548.
- Ferdinand, R. F., Sondeijker, F., van der Ende, J., Selten, J. P., Huizink, A., and Verhulst, F. C. (2005). Cannabis use predicts future psychotic symptoms, and vice versa. *Addiction* **100**, 612–618.
- Fergusson, D. M., Horwood, L. J., and Swain-Campbell, N. R. (2003). Cannabis dependence and psychotic symptoms in young people. *Psychol. Med.* **33**, 15–21.
- Gadzicki, D., Muller-Vahl, K., and Stuhmann, M. (1999). A frequent polymorphism in the coding exon of the human cannabinoid receptor (CNR1) gene. *Mol. Cell. Probes* **13**, 321–323.
- Ghose, A. H. (1986). Cannabis psychosis. *Br. J. Addict.* **81**, 473–478.
- Giuffrida, A., Leweke, F. M., Gerth, C. W., Schreiber, D., Koethe, D., Faulhaber, J., Klosterkotter, J., and Piomelli, D. (2004). Cerebrospinal anandamide levels are elevated in acute schizophrenia and are inversely correlated with psychotic symptoms. *Neuropsychopharmacology* **29**, 2108–2114.
- Gonzalez, R., Carey, C., and Grant, I. (2002). Nonacute (residual) neuropsychological effects of cannabis use: A qualitative analysis and systematic review. *J. Clin. Pharmacol.* **42**, 48S–57S.
- Gottesman, I. I., and Shields, J. (1982). "Schizophrenia: The Epigenetic Puzzle." Cambridge University Press, Cambridge.

- Green, A. I., Tohen, M. F., Hamer, R. M., Strakowski, S. M., Lieberman, J. A., Glick, I., Clark, W. S., and Group, H. R. (2004). First episode schizophrenia-related psychosis and substance use disorders: Acute response to olanzapine and haloperidol. [see comment]. *Schizophr. Res.* **66**, 125–135.
- Green, B., Kavanagh, D., and Young, R. (2003). Being stoned: A review of self-reported cannabis effects. *Drug Alcohol Rev.* **22**, 453–460.
- Griffith, J. D., Cavanaugh, J., Held, J., and Oates, J. A. (1972). Dextroamphetamine: Evaluation of psychomimetic properties in man. *Arch. Gen. Psychiatry* **26**, 97–100.
- Guimaraes, F. S., de Aguiar, J. C., Mechoulam, R., and Breuer, A. (1994). Anxiolytic effect of cannabidiol derivatives in the elevated plus-maze. *Gen. Pharmacol.* **25**, 161–164.
- Halikas, J. A., Weller, R. A., Morse, C., and Shapiro, T. (1982). Incidence and characteristics of a motivational syndrome, including associated findings, among chronic marijuana users. In “Marijuana and Youth: Clinical Observations on Motivation and Learning” (N. I. O. D. Abuse, Ed.), pp. 11–23. National Institute on Drug Abuse, Rockville, Maryland.
- Hall, W., and Degenhardt, L. (2004). “Is There a Specific ‘Cannabis Psychosis?’” Cambridge University Press, Cambridge.
- Hall, W., and Solowij, N. (1998). Adverse effects of cannabis. *Lancet* **352**, 1611–1616.
- Hambrecht, M., and Hafner, H. (1996). Substance abuse and the onset of schizophrenia. *Biol. Psychiatry* **40**, 1155–1163.
- Hambrecht, M., and Hafner, H. (2000). Cannabis, vulnerability, and the onset of schizophrenia: An epidemiological perspective. *Aust. N.Z. J. Psychiatry* **34**, 468–475.
- Harding, T., and Knight, F. (1973). Marijuana-modified mania. *Arch. Gen. Psychiatry* **29**, 635–637.
- Hart, C. L., van Gorp, W., Haney, M., Foltin, R. W., and Fischman, M. W. (2001). Effects of acute smoked marijuana on complex cognitive performance. *Neuropsychopharmacology* **25**, 757–765.
- Heim, M. E., Romer, W., and Queisser, W. (1981). Clinical experience with levonantradol hydrochloride in the prevention of cancer chemotherapy-induced nausea and vomiting. *J. Clin. Pharmacol.* **21**, 86S–89S.
- Heim, M. E., Queisser, W., and Altenburg, H. P. (1984). Randomized crossover study of the antiemetic activity of levonantradol and metoclopramide in cancer patients receiving chemotherapy. *Cancer Chemother. Pharmacol.* **13**, 123–125.
- Heinrichs, R. W., and Zakzanis, K. K. (1998). Neurocognitive deficit in schizophrenia: A quantitative review of the evidence. *Neuropsychology* **12**, 426–445.
- Heishman, S. J., Huestis, M. A., Henningfield, J. E., and Cone, E. J. (1990). Acute and residual effects of marijuana: Profiles of plasma THC levels, physiological, subjective, and performance measures. *Pharmacol. Biochem. Behav.* **37**, 561–565.
- Henquet, C., Krabbendam, L., Spauwen, J., Kaplan, C., Lieb, R., Wittchen, H. U., and van Os, J. (2005). Prospective cohort study of cannabis use, predisposition for psychosis, and psychotic symptoms in young people. *BMJ* **330**, 11.
- Hill, A. B. (1965). The environment and disease: Association or causation? *Proc. R. Soc. Med.* **58**, 295–300.
- Hoehle, M. R., Rinn, T., Flachmeier, C., Heere, P., Kunert, H. J., Timmermann, B., Kopke, K., and Ehrenreich, H. (2000). Comparative sequencing of the human CB1 cannabinoid receptor gene coding exon: No structural mutations in individuals exhibiting extreme responses to cannabis. *Psychiatr. Genet.* **10**, 173–177.
- Hollister, L. E. (1986). Health aspects of cannabis. *Pharmacol. Rev.* **38**, 1–20.
- Hollister, L. E. (1988). Cannabis 1988. *Acta Psychiatr. Scand. Suppl.* **345**, 108–118.
- Hollister, L. E., Gillespie, H. K., Ohlsson, A., Lindgren, J. E., Wahlen, A., and Agurell, S. (1981). Do plasma concentrations of delta 9-tetrahydrocannabinol reflect the degree of intoxication? *J. Clin. Pharmacol.* **21**, 171S–177S.

- Hooker, W. D., and Jones, R. T. (1987). Increased susceptibility to memory intrusions and the Stroop interference effect during acute marijuana intoxication. *Psychopharmacology* **91**, 20–24.
- Imade, A. G., and Ebbe, J. C. (1991). A retrospective study of symptom patterns of cannabis-induced psychosis. *Acta Psychiatr. Scand.* **83**, 134–136.
- Isbell, H., and Jasinski, D. R. (1969). A comparison of LSD-25 with (–)-delta-9-trans-tetrahydrocannabinol (THC) and attempted cross tolerance between LSD and THC. *Psychopharmacologia* **14**(20), 115–123.
- Isbell, H., Gorodetzky, C. W., Jasinski, D., Claussen, U., Von Spulak, F., and Korte, F. (1967). Effects of (–)-delta-9-trans-tetrahydrocannabinol in man. *Psychopharmacologia* **11**, 184–188.
- Jain, A. K., Ryan, J. R., McMahon, F. G., and Smith, G. (1981). Evaluation of intramuscular levonantradol and placebo in acute postoperative pain. *J. Clin. Pharmacol.* **21**, 320S–326S.
- Jones, R. T., and Stone, G. C. (1970). Psychological studies of marijuana and alcohol in man. *Psychopharmacologia* **18**, 108–117.
- Karniol, I. G., and Carlini, E. A. (1973). Pharmacological interaction between cannabidiol and delta 9-tetrahydrocannabinol. *Psychopharmacologia* **33**, 53–70.
- Karniol, I. G., Shirakawa, I., Kasinski, N., Pfeferman, A., and Carlini, E. A. (1974). Cannabidiol interferes with the effects of delta 9-tetrahydrocannabinol in man. *Eur. J. Pharmacol.* **28**, 172–177.
- Karniol, I. G., Shirakawa, I., Takahashi, R. N., Knobel, E., and Musty, R. E. (1975). Effects of delta-9-tetrahydrocannabinol and cannabiniol in man. *Pharmacology* **13**, 502–512.
- Kenny, J. B., and Wilkinson, P. M. (1982). Levonantradol effectiveness in cancer patients resistant to conventional anti-emetics. *Clin. Oncol.* **8**, 335–339.
- Kolansky, H., and Moore, W. T. (1971a). Effects of marihuana on adolescents and young adults. *JAMA* **216**, 486–492.
- Kolansky, H., and Moore, W. T. (1971b). Effects of marihuana on adolescents and young adults. *J. Psychiatr. Nurs. Ment. Health Serv.* **9**, 9–16.
- Laszlo, J., Lucas, V. S., Jr., Hanson, D. C., Cronin, C. M., and Sallan, S. E. (1981). Levonantradol for chemotherapy-induced emesis: Phase I-II oral administration. *J. Clin. Pharmacol.* **21**, 51S–56S.
- Leroy, S., Griffon, N., Bourdel, M. C., Olie, J. P., Poirier, M. F., and Krebs, M. O. (2001). Schizophrenia and the cannabinoid receptor type 1 (CB1): Association study using a single-base polymorphism in coding exon 1. *Am. J. Med. Genet.* **105**, 749–752.
- Leweke, M., Kampmann, C., Radwan, M., Dietrich, D. E., Johannes, S., Emrich, H. M., and Munte, T. F. (1998). The effects of tetrahydrocannabinol on the recognition of emotionally charged words: An analysis using event-related brain potentials. *Neuropsychobiology* **37**, 104–111.
- Leweke, F. M., Giuffrida, A., Wurster, U., Emrich, H. M., and Piomelli, D. (1999a). Elevated endogenous cannabinoids in schizophrenia. *Neuroreport* **10**, 1665–1669.
- Leweke, F. M., Schneider, U., Thies, M., Munte, T. F., and Emrich, H. M. (1999b). Effects of synthetic delta-9-tetrahydrocannabinol on binocular depth inversion of natural and artificial objects in man. *Psychopharmacology (Berl.)* **142**, 230–235.
- Leweke, F. M., Schneider, U., Radwan, M., Schmidt, E., and Emrich, H. M. (2000). Different effects of nabilone and cannabidiol on binocular depth inversion in Man. *Pharmacol. Biochem. Behav.* **66**, 175–181.
- Lindemann, E., and Malamud, W. (1934). Experimental analysis of the psychopathological effects of intoxicating drugs. *Am. J. Psychiatry* **90**, 853–881.
- Linszen, D. H., Dingemans, P. M., and Lenior, M. E. (1994). Cannabis abuse and the course of recent-onset schizophrenic disorders. *Arch. Gen. Psychiatry* **51**, 273–279.
- Marks, D. F., and MacAvoy, M. G. (1989). Divided attention performance in cannabis users and non-users following alcohol and cannabis separately and in combination. *Psychopharmacology* **99**, 397–401.

- Marshall, C. R. (1897). The active principle of Indian hemp: A preliminary communication. *Lancet* 235–238.
- Mathers, D. C., Ghodse, A. H., Caan, A. W., and Scott, S. A. (1991). Cannabis use in a large sample of acute psychiatric admissions. *Br. J. Addict.* **86**, 779–784.
- Mayor's, C. (1944). "The Marijuana Problem in the City of New York." Jacques Catell Press, Lancaster, PA.
- McGuire, P. K., Jones, P., Harvey, I., Bebbington, P., Toone, B., Lewis, S., and Murray, R. M. (1994). Cannabis and acute psychosis. *Schizophr. Res.* **13**, 161–167.
- McGuire, P. K., Jones, P., Harvey, I., Williams, M., McGuffin, P., and Murray, R. M. (1995). Morbid risk of schizophrenia for relatives of patients with cannabis-associated psychosis. *Schizophr. Res.* **15**, 277–281.
- Mechoulam, R., and Ben-Shabat, S. (1999). From gan-zi-gun-nu to anandamide and 2-arachidonoylglycerol: The ongoing story of cannabis. *Nat. Prod. Rep.* **16**, 131–143.
- Mehmedic, Z., Martin, J., Foster, S., and ElSohly, M. A. (2005). Δ 9-THC and other cannabinoids content of confiscated marijuana: Potency trends, 1994–2004. In "International Association for Cannabis as Medicine (IACM) 3rd Conference on Cannabinoids in Medicine." Leiden, The Netherlands.
- Melges, F. T., Tinklenberg, J. R., Hollister, L. E., and Gillespie, H. K. (1970). Marihuana and temporal disintegration. *Science* **168**, 1118–1120.
- Miller, L. L., McFarland, D., Cornett, T. L., and Brightwell, D. (1977). Marijuana and memory impairment: Effect on free recall and recognition memory. *Pharmacol. Biochem. Behav.* **7**, 99–103.
- Miller, P., Lawrie, S. M., Hodges, A., Clafferty, R., Cosway, R., and Johnstone, E. C. (2001). Genetic liability, illicit drug use, life stress and psychotic symptoms: Preliminary findings from the Edinburgh study of people at high risk for schizophrenia. *Soc. Psychiatry Psychiatr. Epidemiol.* **36**, 338–342.
- Millman, R. B., and Sbriglio, R. (1986). Patterns of use and psychopathology in chronic marijuana users. *Psychiatr. Clin. North Am.* **9**, 533–545.
- Moreau, J. (1973). "Hashish and Mental Illness." Raven Press, New York.
- Morita, Y., Ujike, H., Tanaka, Y., Uchida, N., Nomura, A., Ohtani, K., Kishimoto, M., Morio, A., Imamura, T., Sakai, A., Inada, T., Harano, M., *et al.* (2005). A nonsynonymous polymorphism in the human fatty acid amide hydrolase gene did not associate with either methamphetamine dependence or schizophrenia. *Neurosci. Lett.* **376**, 182–187.
- Niesink, J. M., Rigter, S. M., Pijlman, F. T. A., and Bossong, M. G. (2005). Increase in total delta-9-THC in Nederweert as sold in Dutch Coffee Shops. In "International Association for Cannabis as Medicine (IACM) 3rd Conference on Cannabinoids in Medicine." Leiden, The Netherlands.
- Nunn, J. A., Rizza, F., and Peters, E. R. (2001). The incidence of schizotypy among cannabis and alcohol users. *J. Nerv. Ment. Dis.* **189**, 741–748.
- Ohlsson, A., Lindgren, J. E., Wahlen, A., Agurell, S., Hollister, L. E., and Gillespie, H. K. (1980). Plasma delta-9 tetrahydrocannabinol concentrations and clinical effects after oral and intravenous administration and smoking. *Clin. Pharmacol. Ther.* **28**, 409–416.
- Ohlsson, A., Lindgren, J. E., Wahlen, A., Agurell, S., Hollister, L. E., and Gillespie, H. K. (1981). Plasma levels of delta 9-tetrahydrocannabinol after intravenous, oral, and smoke administration. *NIDA Res. Monogr.* **34**, 250–256.
- Palsson, A., Thulin, S. O., and Tunving, K. (1982). Cannabis psychoses in south Sweden. *Acta Psychiatr. Scand.* **66**, 311–321.
- Phillips, L. J., Curry, C., Yung, A. R., Yuen, H. P., Adlard, S., and McGorry, P. D. (2002). Cannabis use is not associated with the development of psychosis in an 'ultra' high-risk group. *Aust. N. Z. J. Psychiatry* **36**, 800–806.
- Pitts, J. E., O'Neil, P. J., and Leggo, K. P. (1990). Variation in the THC content of illicitly imported Cannabis products: 1984–1989. *J. Pharm. Pharmacol.* **42**, 817–820.

- Pitts, J. E., Neal, J. D., and Gough, T. A. (1992). Some features of Cannabis plants grown in the United Kingdom from seeds of known origin. *J. Pharm. Pharmacol.* **44**, 947–951.
- Pope, H. G., Jr., Gruber, A. J., and Yurgelun-Todd, D. (1995). The residual neuropsychological effects of cannabis: The current status of research. *Drug Alcohol Depend.* **38**, 25–34.
- Pope, H. G., Jr., and Yurgelun-Todd, D. (1996). The residual cognitive effects of heavy marijuana use in college students. *JAMA* **275**, 521–527.
- Pope, H. G., Jr., Gruber, A. J., Hudson, J. I., Huestis, M. A., and Yurgelun-Todd, D. (2001). Neuropsychological performance in long-term cannabis users. *Arch. Gen. Psychiatry* **58**, 909–915.
- Ranganathan, M., and D'Souza, D. (2006). The Acute effects of cannabinoids on memory in humans: A review. *Psychopharmacology*. **188**(4), 425–444. EPub.
- Reilly, D., Didcott, P., Swift, W., and Hall, W. (1998). Long-term cannabis use: Characteristics of users in an Australian rural area. *Addiction* **93**, 837–846.
- Rolfe, M., Tang, C. M., Sabally, S., Todd, J. E., Sam, E. B., and Hatib N'Jie, A. B. (1993). Psychosis and cannabis abuse in The Gambia. A case-control study [see comment]. *Br. J. Psychiatry* **163**, 798–801.
- Rottanburg, D., Robins, A. H., Ben-Arie, O., Teggin, A., and Elk, R. (1982). Cannabis-associated psychosis with hypomanic features. *Lancet* **2**, 1364–1366.
- Rubin, V., and Comitas, L. (1975). “Ganja in Jamaica: A Medical Anthropological Study of Chronic Marihuana Use.” Mouton Publishers, The Hague.
- Russo, E. B., and McPartland, J. M. (2003). Cannabis is more than simply delta(9)-tetrahydrocannabinol. *Psychopharmacology (Berl.)* **165**, 431–432; author reply 433–434.
- Sheidler, V. R., Ettinger, D. S., Diasio, R. B., Enterline, J. P., and Brown, M. D. (1984). Double-blind multiple-dose crossover study of the antiemetic effect of intramuscular levonantradol compared to prochlorperazine. *J. Clin. Pharmacol.* **24**, 155–159.
- Skosnik, P. D., Spatz-Glenn, L., and Park, S. (2001). Cannabis use is associated with schizotypy and attentional disinhibition. *Schizophr. Res.* **48**, 83–92.
- Solomons, K., and Neppe, V. M. (1989). Cannabis: Its clinical effects. *S. Afr. Med. J.* **76**, 102–104.
- Solomons, K., Neppe, V. M., and Kuyl, J. M. (1990). Toxic cannabis psychosis is a valid entity. *S. Afr. Med. J.* **78**, 476–481.
- Solowij, N. (1995). Do cognitive impairments recover following cessation of cannabis use? *Life Sci.* **56**, 2119–2126.
- Solowij, N., Michie, P. T., and Fox, A. M. (1995). Differential impairments of selective attention due to frequency and duration of cannabis use. *Biol. Psychiatry* **37**, 731–739.
- Solowij, N., Stephens, R. S., Roffman, R. A., Babor, T., Kadden, R., Miller, M., Christiansen, K., McRee, B., Vendetti, J., and Marijuana Treatment Project Research, G. (2002). Cognitive functioning of long-term heavy cannabis users seeking treatment.[comment][erratum appears in *JAMA* (2002) April 3 **287**(13), 1651]. *JAMA* **287**, 1123–1131.
- Stambaugh, J. E., Jr., McAdams, J., and Vreeland, F. (1984). Dose ranging evaluation of the antiemetic efficacy and toxicity of intramuscular levonantradol in cancer subjects with chemotherapy-induced emesis. *J. Clin. Pharmacol.* **24**, 480–485.
- Stefanis, N. C., Delespaul, P., Henquet, C., Bakoula, C., Stefanis, C. N., and Van Os, J. (2004). Early adolescent cannabis exposure and positive and negative dimensions of psychosis. *Addiction* **99**, 1333–1341.
- Stuart-Harris, R. C., Mooney, C. A., and Smith, I. E. (1983). Levonantradol: A synthetic cannabinoid in the treatment of severe chemotherapy-induced nausea and vomiting resistant to conventional anti-emetic therapy. *Clin. Oncol.* **9**, 143–146.
- Sundram, S., Dean, B., and Copolov, D. (2004). “The Endogenous Cannabinoid System in Schizophrenia.” Cambridge University Press, New York.
- Talbott, J. A., and Teague, J. W. (1969a). Marihuana psychosis. Acute toxic psychosis associated with the use of Cannabis derivatives. *JAMA* **210**, 299–302.

- Talbot, J. A., and Teague, J. W. (1969b). Marihuana psychosis: Acute toxic psychosis associated with the use of cannabis derivatives. *JAMA* **210**, 299–302.
- Tennant, F. S., and Groesbeck, C. J. (1972a). Psychiatric effects of hashish. *Arch. Gen. Psychiatry* **27**, 133–136.
- Tennant, F. S., Jr., and Groesbeck, C. J. (1972b). Psychiatric effects of hashish. *Arch. Gen. Psychiatry* **27**, 133–136.
- Thacore, V. R. (1973). Bhang psychosis. *Br. J. Psychiatry* **123**, 225–229.
- Thacore, V. R., and Shukla, S. R. (1976). Cannabis psychosis and paranoid schizophrenia. *Arch. Gen. Psychiatry* **33**, 383–386.
- Thomas, H. (1993). Cannabis and the PSE. *Br. J. Psychiatry* **162**, 271–272.
- Thomas, H. (1996). A community survey of adverse effects of cannabis use. *Drug Alcohol Depend.* **42**, 201–207.
- Thornicroft, G. (1990). Cannabis and psychosis. Is there epidemiological evidence for an association? *Br. J. Psychiatry* **157**, 25–33.
- Tien, A. Y., and Anthony, J. C. (1990). Epidemiological analysis of alcohol and drug use as risk factors for psychotic experiences. *J. Nerv. Ment. Dis.* **178**, 473–480.
- Tramer, M. R., Carroll, D., Campbell, F. A., Reynolds, D. J., Moore, R. A., and McQuay, H. J. (2001). Cannabinoids for control of chemotherapy induced nausea and vomiting: Quantitative systematic review. *BMJ* **323**, 16–21.
- Tsai, S. J., Wang, Y. C., and Hong, C. J. (2000). Association study of a cannabinoid receptor gene (CNR1) polymorphism and schizophrenia. *Psychiatr. Genet.* **10**, 149–151.
- Turner, C. E., Elsohly, M. A., and Boeren, E. G. (1980). Constituents of Cannabis sativa L. XVII. A review of the natural constituents. *J. Nat. Prod.* **43**, 169–234.
- Ujike, H., Takaki, M., Nakata, K., Tanaka, Y., Takeda, T., Kodama, M., Fujiwara, Y., Sakai, A., and Kuroda, S. (2002). CNR1, central cannabinoid receptor gene, associated with susceptibility to hebephrenic schizophrenia. *Mol. Psychiatry* **7**, 515–518.
- Van Mastrigt, S., Addington, J., and Addington, D. (2004). Substance misuse at presentation to an early psychosis program. *Soc. Psychiatry Psychiatr. Epidemiol.* **39**, 69–72.
- van Os, J., Bak, M., Hanssen, M., Bijl, R. V., de Graaf, R., and Verdoux, H. (2002). Cannabis use and psychosis: A longitudinal population-based study. *Am. J. Epidemiol.* **156**, 319–327.
- Veen, N. D., Selten, J. P., van der Tweel, I., Feller, W. G., Hoek, H. W., and Kahn, R. S. (2004). Cannabis use and age at onset of schizophrenia. *Am. J. Psychiatry* **161**, 501–506.
- Verdoux, H., Gindre, C., Sorbara, F., Tournier, M., and Swendsen, J. D. (2003a). Effects of cannabis and psychosis vulnerability in daily life: An experience sampling test study. *Psychol. Med.* **33**, 23–32.
- Verdoux, H., Gindre, C., Sorbara, F., Tournier, M., and Swendsen, J. D. (2003b). Effects of cannabis and psychosis vulnerability in daily life: An experience sampling test study. [comment]. *Psychol. Med.* **33**, 23–32.
- Verdoux, H., Sorbara, F., Gindre, C., Swendsen, J. D., and van Os, J. (2003c). Cannabis use and dimensions of psychosis in a nonclinical population of female subjects. *Schizophr. Res.* **59**, 77–84.
- Volkow, N. D., Fowler, J. S., Wolf, A. P., and Gillespi, H. (1991). Metabolic studies of drugs of abuse. *NIDA Res. Monogr.* **105**, 47–53.
- Weiser, M., Knobler, H. Y., Noy, S., and Kaplan, Z. (2002). Clinical characteristics of adolescents later hospitalized for schizophrenia. *Am. J. Med. Genet.* **114**, 949–955.
- Wylie, A. S., Scott, R. T., and Burnett, S. J. (1995). Psychosis due to “skunk.” *BMJ* **311**, 125.
- Young, D., and Scoville, W. B. (1938). Paranoid Psychosis in narcolepsy and the possible dosages of benzedrine treatment. *Medical clinics of North America* **22**, 673.
- Zammit, S., Allebeck, P., Andreasson, S., Lundberg, I., and Lewis, G. (2002). Self reported cannabis use as a risk factor for schizophrenia in Swedish conscripts of 1969: Historical cohort study. [comment]. *BMJ* **325**, 1199.

- Zavitsanou, K., Garrick, T., and Huang, X. F. (2004). Selective antagonist [3H]SR141716A binding to cannabinoid CB1 receptors is increased in the anterior cingulate cortex in schizophrenia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **28**, 355–360.
- Zuardi, A. W., Shirakawa, I., Finkelfarb, E., and Karniol, I. G. (1982). Action of cannabidiol on the anxiety and other effects produced by delta 9-THC in normal subjects. *Psychopharmacology* **76**, 245–250.
- Zuardi, A. W., Rodrigues, J. A., and Cunha, J. M. (1991). Effects of cannabidiol in animal models predictive of antipsychotic activity. *Psychopharmacology* **104**, 260–264.
- Zuardi, A. W., Morais, S. L., Guimaraes, F. S., and Mechoulam, R. (1995). Antipsychotic effect of cannabidiol. *J. Clin. Psychiatry* **56**, 485–486.
- Zuardi, A. W., Hallak, J. E., Dursun, S. M., Morais, S. L., Faria Sanches, R., Musty, R. E., and Crippa, J. A. (2006). Cannabidiol monotherapy for treatment-resistant schizophrenia. *J. Psychopharmacol.* **20**(5), 683–686. Epub.

INVOLVEMENT OF NEUROPEPTIDE SYSTEMS IN SCHIZOPHRENIA: HUMAN STUDIES

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Neuropeptides are heterogeneously distributed throughout the digestive, circulatory, and nervous systems and serve as neurotransmitters, neuromodulators, and hormones. Neuropeptides are phylogenetically conserved and have been demonstrated to regulate numerous behaviors. They have been hypothesized to be pathologically involved in several psychiatric disorders, including schizophrenia. On the basis of preclinical data, numerous studies have sought to examine the role of neuropeptide systems in schizophrenia. This chapter reviews the clinical data, linking alterations in neuropeptide systems to the etiology, pathophysiology, and treatment of schizophrenia. Data for the following neuropeptide systems are included: arginine-vasopressin, cholecystokinin (CCK), corticotropin-releasing factor (CRF), interleukins, neuregulin 1 (NRG1), neurotensin (NT), neuropeptide Y (NPY), opioids, secretin, somatostatin, tachykinins, thyrotropin-releasing hormone (TRH), and vasoactive intestinal peptide (VIP). Data from cerebrospinal fluid (CSF), postmortem and genetic studies, as well

as clinical trials are described. Despite the inherent difficulties associated with human studies (including small sample size, variable duration of illness, medication status, the presence of comorbid psychiatric disorders, and diagnostic heterogeneity), several findings are noteworthy. Postmortem studies support disease-related alterations in several neuropeptide systems in the frontal and temporal cortices. The strongest genetic evidence supporting a role for neuropeptides in schizophrenia are those studies linking polymorphisms in *NRG1* and the *CCK_A* receptor with schizophrenia. Finally, the only compounds that act directly on neuropeptide systems that have demonstrated therapeutic efficacy in schizophrenia are neurokinin receptor antagonists. Clearly, additional investigation into the role of neuropeptide systems in the etiology, pathophysiology, and treatment of schizophrenia is warranted.

I. Introduction

Elucidation of the etiology and pathophysiology of schizophrenia with the goal of developing novel prevention and treatment approaches has included examination of the role of neuropeptide systems. Neuropeptides, often referred to as gut-brain peptides because of their high concentrations in both tissues, regulate numerous behaviors (Bennett *et al.*, 1997; Ramirez *et al.*, 2004) and have been implicated in the pathophysiology of several major psychiatric diseases (Holsboer, 2003; Inui, 2003; Kinkead and Nemeroff, 2004; Nemeroff and Vale, 2005). Although neuropeptides clearly function as neurotransmitters (directly modifying the electrical state of neurons), they also function as neuromodulators (modifying the effects of other, primary neurotransmitters such as dopamine, glutamate, serotonin, and GABA) and as neurohormones. Elimination of a neuropeptide or neuropeptide receptor has a wide impact on normal animal behavior, ranging from no effect in the absence of an additional pharmacological or environmental challenge (Coste *et al.*, 2000; Ragnauth *et al.*, 2005; Sharpe *et al.*, 2005; Weninger *et al.*, 1999a,b) to a frankly abnormal phenotype (Bielsky *et al.*, 2005; Colwell *et al.*, 2003; Dauge *et al.*, 2001; Nishimori *et al.*, 1996; Yamada *et al.*, 2001). Despite the relatively subtle behavioral effects of neuropeptides, considerable preclinical data have implicated neuropeptide systems in schizophrenia, not the least of which is their widespread CNS distribution in brain regions implicated in this disorder and the well-documented effects of antipsychotic drugs on these circuits.

Overall, the clinical data implicating a preeminent role for one or another neuropeptide in schizophrenia is not strong, although the overall database is far smaller than the multitude of studies on dopamine and other monoamines. Factors contributing to this include the heterogeneity of schizophrenic patients, including sex, age, substance abuse, comorbid disorders, and chronic exposure to

antipsychotic drugs. However, several clinical studies have consistently associated dysregulation of neuropeptide systems with specific patient subgroups characterized by common symptomatology or treatment response (Breslin *et al.*, 1994; Garver *et al.*, 1991; Tachikawa *et al.*, 2000; Zhang *et al.*, 2000).

Moreover, inadequate methods for antemortem measurement of indices of the activity of neuropeptidergic systems in the human brain in patients contribute to our paucity of information on neuropeptides in schizophrenia. For example, the virtual absence of ligands to measure peptide receptor density with positron emission tomography (PET) or single photon emission computed tomography (SPECT) has limited antemortem studies to measurements of neuropeptides in the cerebrospinal fluid (CSF). However, although neuropeptides measured in the CSF are believed to originate largely but not completely from the CNS, the specific brain regions of origin and their relative contribution remain obscure. Although postmortem studies allow for a high degree of neuroanatomical resolution, results are subject to a number of potential confounds, including postmortem delay interval and chronic antipsychotic (and other) drug exposure, as well as effects of agonal state. An additional consideration in CSF and postmortem studies is the difficulty of obtaining samples from antipsychotic drug-free subjects.

In this chapter, the results of clinical studies examining the role of neuropeptide systems in schizophrenia are reviewed. The peptides discussed include arginine-vasopressin (AVP), cholecystokinin (CCK), corticotropin-releasing factor (CRF), interleukins, neurotensin (NT), neuropeptide Y (NPY), opioids, secretin, somatostatin, tachykinins, thyrotropin-releasing hormone (TRH) and vasoactive intestinal peptide (VIP). Table I summarizes the neuropeptides discussed in this chapter, identified receptor subtypes, and the available receptor agonists and antagonists. Data from CSF, postmortem and genetic studies, as well as clinical trials will be described.

II. Cholecystokinin

CCK was first isolated from the gastrointestinal tract in 1928 (Ivy and Oldberg, 1928) and was shown to stimulate pancreatic secretion and gall bladder contraction. CCK is heterogeneously distributed in the gastrointestinal, central, and peripheral nervous systems (for review see Miyasaka and Funakoshi, 2003). Several biologically active forms of CCK have been identified and share a common N-terminal sequence (CCK-4, CCK-8, CCK-22, CCK-33, CCK-39, and CCK-58), with CCK-8 and CCK-58 being the most abundant forms in the brain. The peptide gastrin (encoded on human chromosome 17) and CCK (encoded on human chromosome 3) have identical 6 N-terminal amino acids but exhibit different sulphation sites (Lund *et al.*, 1986). In the CNS, CCK is

TABLE I
GENERAL PROPERTIES OF NEUROPEPTIDES IMPLICATED IN SCHIZOPHRENIA

Peptide	Related peptides (number of aa)	Identified receptors	Agonists	Antagonists
CCK	Prepro-CCK (4, 8, 22, 33, 39, 58)	CCK _A CCK _B	Cerulein (CCK _A ,CCK _B) Gastrin (CCK _B)	Proglumide (CCK _A) Dexloxiglumide (CCK _A) Devazepide (CCK _A) L365,260 (CCK _B)
CRF	CRF (41) Urocortin I, II, III (40, 38, 38) Urotensin I, II, III (41, 11, 29) Sauvagine (40)	CRF ₁ CRF ₂ ,(sCRF _{2α} , sCRF _{2β} , sCRF _{2γ})	Urocortins (CRF ₂)	GSK 876008 (CRF ₁) R121919 (CRF ₁)
Dynorphins	Prodynorphin Dynorphin A (8, 17) Dynorphin B (13)	κ Receptor	Ethylketocyclazocine (κ) U50,488H (κ) U69,593 (κ)	Norbinaltorphimine (κ)
Endorphins	POMC -β-endorphin (30) -ACTH (39) -α-MSH (13) -α, -β, -γ endorphins	μ Receptor	Morphine (μ) Fentanyl (μ) Levorphanol (μ) Dermorphin (μ)	Naloxone (μ, κ, δ) Naltrexone (μ, κ, δ) Cyprodime (μ)
Enkephalins	Proenkephalin -Leu-enkephalin (5) -Met-enkephalin (5)	δ Receptor	Deltorphin (δ)	Naloxone (μ, δ, κ) Naltrexone (μ, δ, κ) ICI 174864 (δ)

Gastrin NT	Gastrin (6) Prepro-NT/Neuromedin N -Neurotensin (13) -Neuromedin N (6)	CCK _B NT ₁ -NT ₄	JB93242 (CCK _B) NT69L (NT ₁) Eisai compound (NT ₁) JMV 449 (NT ₁) PD149163 (NT ₁)	JB93182 (CCK _B) SR 48692 (NT ₁) SR 142948A (NT ₁ , NT ₂)
NPY	Pro-NPY -NPY(36)	NPY ₁ -NPY ₅	Pancreatic polypeptide Peptide YY	H-409/22 (NPY ₁) 1229U91 (NPY ₁)
Somatostatin	Prosomatostatin -Somatostatin (14, 28)	SST ₁ -SST ₅	Octeotride (SST ₂) Seglitide (SST ₂) Lanreotide (SST ₂) Cortistatin (SST ₁ -SST ₅) GR73632 (NK ₁)	CYN 154806 (SST ₂) PRL-2903 (SST ₂) BIM-23056 (SST ₅)
Tachykinins	Preprotachykinin I -Substance P (11) -Neurokinin A (10) -Neurokinin B (10) Preprotachykinin II -Neurokinin B (10)	NK ₁ NK ₂ NK ₃		Vestipitant (NK ₁) Saredutant (NK ₂) Osanetant (NK ₃) Talnetant (NK ₃) SSR 146977 (NK ₃)
TRH	Pro-TRH -TRH (3)	THR-R1 THR-R2	TA-0910 DN-1417	—
VIP	Prepro-VIP -VIP (28) -PHI (27) -PHV (27) -PACAP (27, 38)	VPAC ₁ VPAC ₂	Ro 25-1553 (VPAC ₂)	PG 97-269 (VPAC ₁) VIP 6-28

TABLE II
STUDIES OF NEUROPEPTIDES IN CEREBROSPINAL FLUID IN PATIENTS WITH SCHIZOPHRENIA

Peptide	APD therapy	Compared to controls	Associated features
AVP	Drug-free	↔ ^{1, 2}	—
CCK	Drug-free	↓ ³⁻⁷	Inversely associated with latency to APD response ^{6,7}
CRF	Maintained	↓ ⁸	Modestly increased compared to depression ⁸
	Drug-free	↑	
	Drug-free	↔ ⁹⁻¹²	Trend to CRF increase ¹⁰
	Withdrawn	↔ ¹³	Trend for relapsers to have higher CRF ¹³
Dynorphin	Drug-free	↑ ¹⁴⁻¹⁵	Associated with severity of symptoms ¹⁴ and poor outcome ¹⁵
Endorphin	Drug-free	↓ ¹⁶	—
	Drug-free	↓ ¹⁷⁻¹⁸	—
	Drug-free	↑ ¹⁹⁻²³	Decreased after APD treatment ²⁰⁻²³
	Drug-free	↔ ²⁴⁻²⁷	—
Enkephalin	Drug-free	↓ ^{26, 28}	—
Gastrin	Drug-free	↔ ^{29, 30}	—
Hypocretin	Drug free	↔ ³¹	—
IL-2 and -6	Drug free	↑ ^{32, 33}	—
	Drug-free	↔ ³⁴⁻³⁷	Trend to IL-6 increase ³⁴
	Maintained	↔ ³⁶	—
	Withdrawn	↑ ³⁸	Associated with relapse ³⁸
NT	Drug-free	↓ ³⁹⁻⁴³	Associated with severity of positive and negative of symptoms ⁴²⁻⁴⁴
			Associated with latency to APD response ⁴³
			Normalization after successful APD treatment ^{40, 42}
NPY	Drug-free	↔ ^{45, 46}	Increased PYY ⁴⁶ . Not modified by APD treatment ⁴⁶
	Drug-free	↑ ⁴⁷	Associated with duration of illness, brain abnormalities and severity of symptoms ⁴⁷
Oxytocin	Withdrawn	↓ ⁴⁷	—
	Maintained	↔ ⁴⁸	—
	Withdrawn	↔ ⁴⁸	—
SOM	Drug-free	↔ ^{5, 10}	Not modified by APD treatment ⁴⁹
	Drug-free	↓ ⁴⁹⁻⁵¹	—
	Drug-free	NA ^{52, 53, 54}	Reduced by APD treatment
SP	Drug-free	↔ ^{14, 55, 56}	—
TRH	Drug-free	↔ ^{2, 10, 57}	—
VIP	Drug-free	↔ ^{29, 30}	—

AVP, arginine-vasopressin; CCK, cholecystokinin; CRF, corticotropin-releasing factor; IL, interleukin; NPY, neuropeptide Y; NT, neurotensin; SOM, somatostatin; SP, substance P; TRH, thyrotropin-releasing hormone; VIP, vasoactive intestinal peptide; ↑, increase; ↓, decrease; ↔, no changes; APD, antipsychotic drug.

located in the ventral mesencephalon, medial nucleus accumbens (NAcc), septum, hypothalamus, and solitary complex (Vanderhaeghen *et al.*, 1981). Two CCK receptors have been identified, CCK_A (CCK₁) and CCK_B (CCK₂), both G-protein-coupled receptors (GPCRs). The CCK_A receptor has a higher affinity for CCK than gastrin, whereas the CCK_B receptor has similar affinity for CCK and gastrin.

CCK-8 is the predominant form of CCK in the CNS and CSF, and the CNS is the main source of CSF CCK (Hökfelt *et al.*, 1994; Rehfeld and Kruse-Larsen, 1978). CSF CCK concentrations in schizophrenic patients (drug-free and treated with antipsychotic drugs) have been reported to be reduced compared to controls in some studies (Beinfeld and Garver, 1991; Garver *et al.*, 1990; Lotstra *et al.*, 1985; Verbanck *et al.*, 1984) or not different in others (Gerner and Yamada, 1982; Gerner *et al.*, 1985; Rafaelsen and Gjerris, 1985; Tamminga *et al.*, 1986). An association between CSF concentrations of CCK and the latency to antipsychotic drug response has been reported (Beinfeld and Garver, 1991; Garver *et al.*, 1990). See Table II for a summary of the available CSF studies.

The postmortem data suggest disruption of CCK neurotransmission in the temporal lobe, possibly involving the limbic system in schizophrenia (for summary see Table III). CCK-immunoreactivity (IR), CCK mRNA expression, and CCK receptor binding are decreased in the temporal cortex, parahippocampal cortex, hippocampus, and frontal cortex in schizophrenic patients compared to normal controls (Bachus *et al.*, 1997; Farmery *et al.*, 1985; Gabriel *et al.*, 1996; Kerwin *et al.*, 1992; Roberts *et al.*, 1983; Virgo *et al.*, 1995), but discordant results have also been obtained (Bachus *et al.*, 1997; Roberts *et al.*, 1983). Additionally, increased CCK mRNA expression in the substantia nigra of antipsychotic drug-treated schizophrenic patients has been reported (Schalling *et al.*, 1990).

In humans, the gene encoding CCK is located on chromosome 3 (Takahashi *et al.*, 1986). The CCK_A receptor is encoded on chromosome 4, in proximity to the gene encoding the dopamine (DA) D₅ receptor, whereas the CCK_B receptor

-, No positive or negative associations; NA, not assessed.

¹(Sorensen *et al.*, 1985), ²(Gjerris *et al.*, 1985), ³(Lotstra *et al.*, 1985), ⁴(Verbanck *et al.*, 1984), ⁵(Gerner and Yamada, 1982), ⁶(Beinfeld and Garver, 1991), ⁷(Garver *et al.*, 1990), ⁸(Banki *et al.*, 1987), ⁹(Nemeroff *et al.*, 1984), ¹⁰(Banki *et al.*, 1992b), ¹¹(Risch *et al.*, 1992), ¹²(Nishino *et al.*, 1998), ¹³(Forman *et al.*, 1994), ¹⁴(Heikkilä *et al.*, 1990), ¹⁵(Lindström, 1996), ¹⁶(Zhang *et al.*, 1985), ¹⁷(Naber *et al.*, 1981), ¹⁸(Pickar *et al.*, 1981), ¹⁹(Domschke *et al.*, 1979), ²⁰(Rimon *et al.*, 1980), ²¹(Lindström *et al.*, 1978), ²²(Lindström *et al.*, 1986), ²³(Lindström *et al.*, 1992), ²⁴(Emrich *et al.*, 1979), ²⁵(Gerner and Sharp, 1982), ²⁶(Burbach *et al.*, 1979), ²⁷(Nakao *et al.*, 1980), ²⁸(Wen *et al.*, 1983), ²⁹(Gjerris *et al.*, 1984), ³⁰(Rafaelsen and Gjerris, 1985), ³¹(Nishino *et al.*, 2002), ³²(Licinio *et al.*, 1993), ³³(Garver *et al.*, 2003), ³⁴(van Kammen *et al.*, 1999), ³⁵(Barak *et al.*, 1995), ³⁶(Katila *et al.*, 1994), ³⁷(el-Mallakh *et al.*, 1993), ³⁸(McAllister *et al.*, 1995), ³⁹(Widerlöv *et al.*, 1982), ⁴⁰(Lindström *et al.*, 1988), ⁴¹(Nemeroff *et al.*, 1989), ⁴²(Breslin *et al.*, 1994), ⁴³(Garver *et al.*, 1991), ⁴⁴(Sharma *et al.*, 1997), ⁴⁵(Berrettini *et al.*, 1987), ⁴⁶(Widerlöv *et al.*, 1988), ⁴⁷(Peters *et al.*, 1990), ⁴⁸(Glovinsky *et al.*, 1994), ⁴⁹(Heikkilä, 1993), ⁵⁰(Rubinow, 1986), ⁵¹(Sharma *et al.*, 1995), ⁵²(Bissette *et al.*, 1986), ⁵³(Gerner *et al.*, 1985), ⁵⁴(Doran *et al.*, 1989), ⁵⁵(Rimon *et al.*, 1984), ⁵⁶(Miller *et al.*, 1996), ⁵⁷(Sharma *et al.*, 2001).

TABLE III

SUMMARY OF RESULTS FROM STUDIES EXAMINING NEUROPEPTIDE SYSTEMS IN POSTMORTEM BRAIN TISSUE FROM SUBJECTS WITH SCHIZOPHRENIA [DATA REPRESENT CHANGES IN IMMUNOREACTIVITY (IR), mRNA EXPRESSION OR BINDING FOR EACH PEPTIDE UNLESS OTHERWISE SPECIFIED]

Peptide	Frontal cortex	Temporal cortex	Entorhinal cortex	Cingulate cortex	Nucleus accumbens	Caudate/ Putamen	Amygdala	Hippocampus	Substantia nigra
AVP	NA	↓IR ¹	NA	NA	NA	NA	NA	NA	NA
CCK	↓ and ↔ mRNA ^{2,3}	↓mRNA ²	↓ mRNA ^{2,3}	↔ mRNA ³				↓ mRNA ⁶⁻⁸	
	↓ and ↔ IR ^{5,6}	↓ and ↔ IR ⁵⁻⁸	↔ IR ⁹	↔ IR ⁸	↔ IR ⁵	↔ IR ⁵	↓ and ↔ IR ^{5,7,8}	↓ and ↔ IR ⁵⁻⁸	↑ IR ¹¹
	↓ binding ⁴	↓ binding ⁴				↔ binding ⁴	↔ binding ⁴	↓ binding ^{4,10}	
Dyn	↔ proDyn mRNA ¹²	NA	NA	↔ proDyn mRNA ¹²	NA	NA	↔ proDyn mRNA ¹³	NA	↔ Dyn-IR ¹⁴
	↔κ receptor mRNA ¹²			↔κ receptor mRNA ¹²					
Enk	↑ IR ¹⁵	NA	NA	NA		↓ IR ⁵ ↔ proEnk mRNA ¹⁶	↔ IR ¹⁷	NA	↑ IR ¹⁵
CRF	↔ IR ⁶	↔ IR ⁶	NA	↓ IR ⁶	NA	NA	NA	NA	NA
Gal	↔ IR ¹⁸	↓ and ↔ IR ^{1,18}	NA	NA	NA	NA	NA	NA	NA
NT	↑ IR ¹⁹	↔ IR ⁸		↔ IR ⁸	↔ IR ^{5,8}	↔ IR ⁸	↔ IR ^{8,17}	↔ IR ⁸	
	↓ binding ²⁰		↓ binding ^{21, 22}	↓ binding ²⁰		↓ binding ^{20,23}			↑ binding ²⁴

NPY	Altered distribution ²⁵ ↓ and ↔ mRNA ^{26,27} ↔ NPY ₁ mRNA ²⁸ ↔ NPY ₂ mRNA ²⁸	↓ IR ^{1,6} ↓ PYY-IR ¹	NA	↓ IR ⁶	NA	NA	↔ IR ^{17,29}	Altered morphology in NPY fibers in CA4 ³⁰	NA
SOM	↓ IR ^{6,19}	↓ IR ⁶	NA	↓ IR ⁶	NA	↔ IR ⁸	↔ IR ^{8,29}	↓ IR ⁶⁻⁸	NA
SP	↔ mRNA ³¹ ↑ and ↔ IR ^{5,15} ↑ NK ₁ -IR ³¹	↔ mRNA ³² ↑ IR ¹⁵ ↔ binding ³²	NA	↔ IR ⁸	↔ IR ^{5,15}	↔ mRNA ¹⁶ ↔ IR ^{5,15}	↓ mRNA ³² ↔ IR ⁸ ↔ binding ³²	↑ IR ^{8,15}	↑ IR ¹⁵ ↔ IR ^{14,15}
TRH	↓ IR ¹⁹	NA	NA	NA	↔ IR ¹⁹	↔ IR ¹⁹	↔ IR ¹⁹	NA	NA
VIP	↔ IR ^{6,8}	↔ IR ^{6,8}	↔ IR ⁹	↔ IR ⁶	NA	↔ IR ⁸	↑ IR ^{8,17}	↔ IR ^{6, 8}	NA

AVP, arginine-vasopressin; CCK, cholecystokinin; CRF, corticotropin-releasing factor; Dyn, dynorphin; Enk, enkephalin; Gal, galanin; NPY, neuropeptide Y; NT, neurotensin; PYY, peptide YY; SOM, somatostatin; SP, substance P; TRH, thyrotropin-releasing hormone; VIP, vasoactive intestinal peptide; ↑, increased; ↓, decreased; ↔, no change; NA, not assessed; IR: immunoreactivity.

¹(Frederiksen *et al.*, 1991), ²(Virgo *et al.*, 1995), ³(Bachus *et al.*, 1997), ⁴(Farmery *et al.*, 1985), ⁵(Kleinman *et al.*, 1985), ⁶(Gabriel *et al.*, 1996), ⁷(Ferrier *et al.*, 1983), ⁸(Roberts *et al.*, 1983), ⁹(Perry *et al.*, 1981), ¹⁰(Kerwin *et al.*, 1992), ¹¹(Schalling *et al.*, 1990), ¹²(Peckys and Hurd, 2001), ¹³(Hurd, 2002), ¹⁴(Iadarola *et al.*, 1991), ¹⁵(Toru *et al.*, 1988), ¹⁶(Harrington *et al.*, 1995), ¹⁷(Zech *et al.*, 1986), ¹⁸(Gabriel *et al.*, 1994), ¹⁹(Nemeroff *et al.*, 1983), ²⁰(Lahti *et al.*, 1998), ²¹(Hamid *et al.*, 2002), ²²(Wolf *et al.*, 1995), ²³(Palacios *et al.*, 1991), ²⁴(Uhl and Kuhar, 1984), ²⁵(Ikeda *et al.*, 2004), ²⁶(Kuromitsu *et al.*, 2001), ²⁷(Caberlotto and Hurd, 1999), ²⁸(Caberlotto and Hurd, 2001), ²⁹(Beal *et al.*, 1987), ³⁰(Iritani *et al.*, 2000), ³¹(Tooney *et al.*, 2001), ³²(Carletti *et al.*, 2005).

is encoded on chromosome 11, in proximity to the gene encoding the DA D₄ receptor (Huppi *et al.*, 1995). On the basis of the similarity in expression patterns, biological effects, and their proximity in the human genome, it has been proposed that receptors for CCK and DA may possibly interact with one another, perhaps via coregulation (Huppi *et al.*, 1995). Two studies of polymorphisms in the CCK gene found no association with schizophrenia (Bowen *et al.*, 1998; Hattori *et al.*, 2001a). In contrast, a significant association was found between a polymorphism in the promoter region of the CCK gene and schizophrenia in a family-based analysis (Wang *et al.*, 2002). Polymorphisms in the CCK_A receptor gene and its promoter have also been associated with schizophrenia (Tachikawa *et al.*, 2000, 2001), specifically with the paranoid type of schizophrenia (Tachikawa *et al.*, 2000, 2001) and positive symptom severity (Lu *et al.*, 2004; Zhang *et al.*, 2000). Two studies have failed to find any association between polymorphisms in the CCK_B receptor gene and schizophrenia (Hattori *et al.*, 2001b; Tachikawa *et al.*, 1999).

On the basis of biochemical and behavioral data demonstrating antidopaminergic and antipsychotic-like effects of central CCK administration (for review see Nair *et al.*, 1986), the efficacies of CCK-8, CCK-33, and the decapeptide caerulein (a mixed CCK_A and CCK_B receptor agonist) have been tested in schizophrenic patients in more than 20 clinical trials (for a summary see Table IV). Most of these trials involved i.v. administration of CCK or caerulein (single to ten doses), alone or in combination with antipsychotic drug maintenance therapy, to chronic schizophrenic patients resistant to neuroleptic treatment. Initial open or single-blind studies reported promising findings of symptom relief that lasted up to several weeks in at least a subset of patients (for review see Montgomery and Green, 1988 and Nair *et al.*, 1986). However, 7 out of 10 double-blind studies reported no difference from placebo (Albus *et al.*, 1986; Hommer *et al.*, 1984; Itoh *et al.*, 1986; Lotstra *et al.*, 1984, 1985; Mattes *et al.*, 1985; Nair *et al.*, 1984, 1985; Peselow *et al.*, 1987; Tamminga *et al.*, 1986; Verhoeven *et al.*, 1986). Moreover, caerulein monotherapy was found to be ineffective in two small clinical trials in schizophrenia (Lotstra *et al.*, 1984; Tamminga *et al.*, 1986). The last clinical trial with a CCK receptor agonist was nearly 20 years ago. The lack of efficacy of CCK in these trials could be related to the small number of patients, inadequate dosing, poor brain penetration, or insufficient treatment duration. The possibility that CCK receptor agonists may represent a novel treatment option for a subset of patients with schizophrenia probably merits additional study.

On the basis of the observation that CCK and CCK analogs stimulate midbrain DA cell firing (Skirboll *et al.*, 1981), the antipsychotic potential of the CCK receptor antagonist proglumide was examined. Three clinical trials failed to demonstrate efficacy of proglumide in the treatment of schizophrenia (Hicks *et al.*, 1989; Innis *et al.*, 1986; Whiteford *et al.*, 1992).

The few clinical studies evaluating gastrin found no evidence of a role for this peptide in the pathophysiology of schizophrenia (Detera-Wadleigh *et al.*, 1987; Gjerris *et al.*, 1984; Rafaelsen and Gjerris, 1985).

III. Corticotropin-Releasing Factor

The importance of stress on the course of schizophrenia is well established. Stressful life events are associated with the onset of schizophrenia (Gruen and Baron, 1984; Leff and Vaughn, 1980; Lukoff *et al.*, 1984), relapse frequency, and psychotic decompensation (Altamura *et al.*, 1999; Gispen-de Wied, 2000; Howes *et al.*, 2004; Norman *et al.*, 2002). Additionally, schizophrenic patients are more vulnerable to stress associated with minor life events (Cotter and Pariante, 2002; Gispen-de Wied, 2000). The 41 amino acid peptide CRF coordinates the mammalian stress response in part via regulation of the hypothalamic-pituitary-adrenal (HPA) axis and the autonomic nervous system (ANS). In the CNS, CRF-containing neurons are found in the hypothalamus, amygdala, cerebral cortex, septum, bed nucleus of the stria terminalis (BNST), cerebellum, brain stem, and spinal cord (Fischman and Moldow, 1982). Following exposure to stress, CRF is released from nerve terminals in the paraventricular nucleus of the hypothalamus (PVN) into the portal circulation, binds to CRF receptors located in the anterior pituitary, and induces the release of adrenocorticotrophic hormone (ACTH) into the general circulation. ACTH stimulates the release of cortisol from the adrenal cortex. Cortisol is the final mediator of the stress response, modulating glucose metabolism, blood pressure, and immune function. Finally, cortisol feeds back negatively on the HPA axis, binding to glucocorticoid receptors at the level of the hypothalamus, hippocampus, and anterior pituitary. Other members of the CRF family including the urocortins and perhaps urotensin and sauvagine also participate in regulation of the stress response (Skelton *et al.*, 2000). CRF has two known receptors, CRF₁ and CRF₂ (also known as the urocortin receptor). Although still under debate, anxiogenic and anxiolytic effects are attributed to activation of CRF₁ and CRF₂ receptors, respectively (Nemeroff and Vale, 2005).

Some, but not all studies report a modest increase in CSF CRF concentrations in schizophrenia, although of lesser magnitude than those found in depression (Banki *et al.*, 1987, 1992a; Nishino *et al.*, 1998). Further, discontinuation of haloperidol maintenance therapy in chronic stable schizophrenic patients is associated with increased CSF CRF concentrations (Forman *et al.*, 1994). In postmortem tissue, CRF-IR is decreased in the cingulate gyrus but not in the frontal, temporal, or occipital cortices of schizophrenic patients with cognitive impairment (Frederiksen *et al.*, 1991; Gabriel *et al.*, 1996). To the best of our

TABLE IV
THE RESULTS OF CLINICAL TRIALS WITH NEUROPEPTIDE RECEPTOR AGONISTS AND ANTAGONISTS IN SCHIZOPHRENIC PATIENTS

Compound	Class	Drug administration	Study design	Schizophrenic patient population	APD therapy	Outcome	References
CCK-33	CCK _A , CCK _B agonist	Single	Open	6 Chronic paranoid	Maintained	Improvement	Nair <i>et al.</i> (1982)
		Single	Open	21 Chronic resistant	Maintained	Improvement	Nair <i>et al.</i> (1983)
CCK-8	CCK _A , CCK _B agonist	Single	Open	8 Chronic resistant	Maintained	Improvement	Bloom <i>et al.</i> (1983)
		4 d, b.i.d.	Double-blind cross-over	4 Chronic resistant	Maintained	Not different from placebo	Hommer <i>et al.</i> (1984)
		1/wk for 8 wk	Double-blind w/placebo	9 Chronic	Maintained	Improvement in cognition and delusions	Nair <i>et al.</i> (1984)
		1/wk for 8 wk	Double-blind w/placebo	14 Chronic	Maintained	Improvement	Nair <i>et al.</i> (1985)
		Single	Double-blind cross-over	30 Chronic resistant	Maintained	Not different from placebo	Peselow <i>et al.</i> (1987)
		Single	Open	20 Chronic	Maintained	Improvement	Moroji <i>et al.</i> (1982)
Caerulein	CCK _A , CCK _B agonist	1–2 wk, q.d.	Open	58 Chronic resistant	Maintained	Improvement in 23/58 cases	Itoh <i>et al.</i> (1982)
		2 wk, 1/wk	Open	6 Chronic	Maintained	Improvement	Albus <i>et al.</i> (1984)
		3 wk, 1/wk	Double-blind w/placebo	10 Chronic	Maintained	Not different from placebo	Albus <i>et al.</i> (1984)
		Single	Double-blind cross-over	9 Chronic	Drug-free	Not different from placebo	Lotstra <i>et al.</i> (1984)
		4 wk, b.i.d.	Double-blind cross-over	8 Chronic resistant	Maintained	Not different from placebo	Hommer <i>et al.</i> (1984)
		1/wk for 2 wk	Single blind	6 Chronic	Maintained	Improvement	van Ree <i>et al.</i> (1984)

			Single	Double-blind cross-over	17 Chronic	Maintained	Not different from placebo	Mattes <i>et al.</i> (1985)
			Single	Open	9 Chronic resistant	Maintained	No improvement	Boza and Rotondo (1985)
			5 d, q.d.	Double-blind cross-over	5 Chronic	Drug-free	Not different from placebo	Tamminga <i>et al.</i> (1986)
			7 d, q.d.	Double-blind w/placebo	135 Chronic	Maintained	Not different from placebo	Itoh <i>et al.</i> (1986)
			1 wk, 3 wk	Double-blind w/placebo	10 Chronic	Maintained	Not different from placebo	Albus <i>et al.</i> (1986)
			7 × in 3 wk	Double-blind w/placebo	15 Chronic	Maintained	Improvement, especially in patients with less negative symptoms	Verhoeven <i>et al.</i> (1986)
	Proglumide	CCK _A , CCK _B	4–7 d, q.d.	Double-blind cross-over	4 Chronic	Maintained	No improvement	Innis <i>et al.</i> (1986)
		antagonist	4 wk, q.d.	Open	4 Chronic resistant	Maintained	No improvement	Hicks <i>et al.</i> (1989)
			4 wk, b.i.d.	Single-blind	7 Chronic resistant	Maintained	No improvement, worsening in some cases	Hicks <i>et al.</i> (1989)
			4 wk, q.d.	Double-blind w/placebo	14 Chronic resistant	Maintained	Not different from placebo	Whiteford <i>et al.</i> (1992)
	DTγE	μ, κ Agonist	7 d, q.d.	Open	6 Chronic resistant	Drug-free	Improvement of psychotic symptoms	Verhoeven <i>et al.</i> (1979)
			8 d, q.d.	Double-blind cross-over	8 Chronic resistant	Maintained (6)	Improvement of psychotic symptoms	Verhoeven <i>et al.</i> (1979)
			4 d, q.d.	Double-blind cross-over	13 Chronic resistant	Maintained	Not different from placebo	Emrich <i>et al.</i> (1980)
			3 d, q.d.	Open	10 Chronic	Maintained	No improvement	Casey <i>et al.</i> (1981)
			10 d, q.d.	Open	11 Chronic	Maintained	No improvement, euphoria in 3 cases	Manchanda and Hirsch (1981)
			10 d, q.d.	Double-blind w/placebo	17 Chronic	Maintained	Improvement	Verhoeven <i>et al.</i> (1982)

(Continued)

TABLE IV (Continued)
THE RESULTS OF CLINICAL TRIALS WITH NEUROPEPTIDE RECEPTOR AGONISTS AND ANTAGONISTS IN SCHIZOPHRENIC PATIENTS

Compound	Class	Drug administration	Study design	Schizophrenic patient population	APD therapy	Outcome	References
DE γ E	μ , κ Agonist	12 d, q.d.	Open	8 Mixed	Drug-free	Improvement in 6	Meltzer <i>et al.</i> (1982)
		Single	Open	5 Chronic	Drug-free	No improvement	Tamminga <i>et al.</i> (1981)
		Single	Open	4 Chronic	Maintained	No improvement	Korsgaard <i>et al.</i> (1982)
		1/d, 8 d	Double-blind cross-over	9 Chronic	Maintained	Transient but modest improvement	Volavka <i>et al.</i> (1983)
		14 d, q.d.	Open	6 Chronic	Maintained	No improvement	Mizuki <i>et al.</i> (1986)
		10 d, q.d.	Double-blind cross-over	18 Chronic	Drug-free	Improvement in subset (hebephrenic, paranoid types with less negative symptoms)	Verhoeven <i>et al.</i> (1984b)
		7 \times in 3wk	Double-blind w/placebo	15 Chronic	Maintained	Improvement in subset (less negative symptoms)	Verhoeven <i>et al.</i> (1986)
		4 wk, q.d.	Double-blind var. doses	93 Chronic	Maintained	Not different from placebo	Azorin <i>et al.</i> (1990)
Naltrexone	μ , κ , δ Antagonist	4 wk, q.d.	Double-blind w/placebo	31 Chronic	Drug-free	Not different from placebo	Montgomery <i>et al.</i> (1992)
		6 wk, q.d. to t.i.d.	Open	5 Chronic	Maintained	No improvement	Mielke and Gallant (1977)
		7 wk, q.d.	Double-blind cross-over	8 Chronic	Drug-free	Not different from placebo	Gitlin <i>et al.</i> (1981)
		14 wk, b.i.d.	Double-blind w/placebo	4 Chronic negative	Maintained	Not different from placebo	Marchesi <i>et al.</i> (1992)
		2 wk, b.i.d.	Double-blind w/placebo	9 Chronic	Maintained	Improvement in positive and negative symptoms	Marchesi <i>et al.</i> (1995)

Naloxone	μ, κ, δ Antagonist	Single	Single blind	6 Chronic	Maintained	Decreased hallucinations in 4 patients	Gunne <i>et al.</i> (1977)
		1 d, b.i.d.	Double-blind w/placebo	12 Chronic	Maintained	Not different from placebo	Kurland <i>et al.</i> (1977)
		2 d, q.d.	Double-blind cross-over	9 Chronic	Maintained	Decreased hallucinations	Watson <i>et al.</i> (1978)
		Single	Double-blind cross-over	9 Chronic	Maintained	No improvement	Lipinski <i>et al.</i> (1979)
		2 d, q.d.	Double-blind cross-over	14 Chronic	Maintained	Decreased hallucinations	Berger <i>et al.</i> (1981)
		2 d, q.d.	Double-blind cross-over	11 Chronic	Drug free	Not different from placebo	Sethi and Prakash (1981)
		Single	Double-blind cross-over	13 Chronic hallucinating	Maintained		Freeman and Fairburn, (1981)
		2 d, q.d.	Double-blind cross-over	32 Chronic	Drug free (13) Maintained (19)	Drug-free: not different from placebo APD- treated: improvement particularly in hallucinations	Pickar <i>et al.</i> (1982)
		2 d, q.d.	Double-blind cross-over	4 Chronic	Maintained	Improvement in noncatatonics	Cohen <i>et al.</i> (1985)
		4 d, q.d.	Double-blind cross-over	6 Chronic	Maintained	Not different from placebo	Naber and Leibl (1983)
		5 d, q.d.	Double-blind cross-over	12 Chronic	Maintained	Not different from placebo	Naber <i>et al.</i> (1983)
		4 d, q.d.	Double-blind cross-over	43 Chronic	Maintained	Not different from placebo	Pickar <i>et al.</i> (1989)
		4 d, q.d.	Double-blind cross-over	10	Maintained	Placebo showed a slightly better effect	Verhoeven <i>et al.</i> (1984a)
		Single	Open	44 Nonpsychiatric	Drug free	Induced psychotic symptoms	Denicoff <i>et al.</i> (1987)

(Continued)

TABLE IV (Continued)
THE RESULTS OF CLINICAL TRIALS WITH NEUROPEPTIDE RECEPTOR AGONISTS AND ANTAGONISTS IN SCHIZOPHRENIC PATIENTS

Compound	Class	Drug administration	Study design	Schizophrenic patient population	APD therapy	Outcome	References
Oxytocin		6–10 d, q.d.	Open	Number not reported	Not reported	Improvement, particularly in acute cases	Bujanow (1972); Bujanow (1974)
SR 48692	Neurotensin receptor (NT ₁) antagonist	6 wk, q.d.	Double-blind multi-arm	63 Schizophrenics and schizoaffective	Drug-free	Not different from placebo	Meltzer <i>et al.</i> (2004)
Secretin		Single	Double-blind w/placebo	11 Severely ill, resistant	Maintained	Not different from placebo, improvement in a subgroup	Sheitman <i>et al.</i> (2004)
Osanetant	NK ₃ antagonist	6 wk, q.d.	Double-blind multi-arm	71 Schizophrenics and schizoaffective	Drug-free	Improvement in positive and cognitive symptoms, Very mild side effects	Meltzer <i>et al.</i> , 2004)
TRH		1/wk for 2 wk	Open	10 Chronic	Drug-free	Improvement in affect and thought	Wilson <i>et al.</i> (1973)
		5 d, q.d.	Double-blind cross-over	3 Chronic resistant	Drug-free	Worsening of symptoms in 2 cases	Bigelow <i>et al.</i> (1975)
		14 d, t.i.d.	Open	9 Chronic	Drug-free	Worsening of symptoms	Davis <i>et al.</i> (1975)
		3 wk, q.d.	Double-blind cross-over	5 Chronic	Maintained	Not different from placebo	Clark <i>et al.</i> (1975)

		4 d, q.d.	Double-blind cross-over	10 Chronic	Drug-free	Not different from placebo, Increase in TSH	Lindström <i>et al.</i> (1977)
		14 d, q.d.	Double-blind w/placebo	70 Chronic	Maintained	Global improvement, onset within a week	Inanaga <i>et al.</i> (1978)
		Single	Single blind	5 Chronic	Drug-free	Improvement	Prange <i>et al.</i> (1979)
		Single	Double-blind w/placebo	12 Chronic	Drug-free	Transient improvement in psychotic symptoms	Prange <i>et al.</i> (1979)
		14 d, q.d.	Double-blind cross-over	11 Chronic	Maintained	Not different from placebo, improvement in affect in some cases	Kobayashi <i>et al.</i> (1980)
		15× in 30 d	Open	10 Chronic	Maintained	Improvement of negative symptoms, Transient borderline hyperthyroidism	Brambilla <i>et al.</i> (1986)
DN-1417	TRH analogue	14 d, q.d.	Open	6 Chronic	Maintained	Improvement in hallucinations and thought content, Increase in frontal EEG activity	Mizuki <i>et al.</i> (1986)

CCK, cholecystokinin; DT γ E, des-tyrosine-gamma-endorphin; DE γ E, des-enkephalin-gamma-endorphin; TRH, thyrotropin-releasing hormone; APD, antipsychotic drug; wk, week; d, day; q.d., once daily; b.i.d., twice daily; t.i.d., three times a day.

knowledge, association studies between CRF system genes and schizophrenia or the clinical efficacy of CRF-related compounds in schizophrenia have not been examined.

Several groups have reported increased baseline plasma cortisol concentrations in schizophrenic patients, although of a lesser magnitude than in depressed patients (for review see Altamura *et al.*, 1999; Gispén-de Wied, 2000; Lieberman and Koren, 1993). This hypercortisolemia has been postulated to be associated with an increase in inflammatory cytokines (Altamura *et al.*, 1999). In addition, a subset of schizophrenic patients displays cortisol hyposecretion following pharmacological challenge of the HPA axis (i.e., dexamethasone, apomorphine, and DA receptor antagonists) (Altamura *et al.*, 1989; Coryell and Tsuang, 1992; McGauley *et al.*, 1989; Meltzer *et al.*, 2001; Tandon *et al.*, 2000, 1991; Yeragani, 1990). In the dexamethasone suppression test (DST), the best studied test of HPA axis function in schizophrenia, rates of nonsuppression vary from 0% to 73%, and are higher in drug-free than in medicated patients (Tandon *et al.*, 1991). DST nonsuppression, particularly in the drug-free state, is associated with negative symptoms and is a predictor of poor outcome (Altamura *et al.*, 1999; Coryell and Tsuang, 1992; Tandon *et al.*, 1991, 2000). An abnormal stress response in schizophrenia is further supported by alterations in the regulation of the HPA axis and the ANS. Alterations of ANS function in schizophrenia include increased basal heart rate and abnormalities in temperature regulation and skin conductance (Gispén-de Wied, 2000).

IV. Interleukins

Cytokines are small peptides produced by immune cells that serve as an important component of the immune response. The effects of cytokines on the CNS have received considerable attention. Cytokines have both direct and indirect access to the brain via the so called “leaky” regions in the blood-brain barrier (i.e., the circumventricular organs), via cytokine-specific transporters in endothelial cells and via vagal afferents. Moreover, interleukin 2 (IL-2) and its receptor are widely distributed in the brain (Lapchak *et al.*, 1991). Of particular interest to schizophrenia are studies demonstrating cytokine regulation of the mesolimbic DA system. IL-2 enhances DA release in striatal slice preparations (Lapchak, 1992; Petitto *et al.*, 1997) and *in vivo*, systemic IL-2 administration enhances norepinephrine (NE) and DA turnover in the hypothalamus and prefrontal cortex, respectively (Zalcman *et al.*, 1994) and induces hyperlocomotion (Petitto *et al.*, 1997). Further support for a possible role of cytokines in the pathogenesis of schizophrenia was provided by the demonstration that prenatal lipopolysaccharide exposure produces disruptions in sensorimotor gating (one of

the core features of schizophrenia), increased serum IL-2 and IL-6 concentrations, and distinct abnormalities in the DA system and glial cells in mesolimbic regions in adult rats. The effect of prenatal lipopolysaccharide exposure on sensorimotor gating was reversed by antipsychotic drug administration (Borrell *et al.*, 2002).

Several additional lines of evidence suggest that cytokines may be involved in the pathogenesis of schizophrenia. First, IL-2 administration produced psychotic-like symptoms (paranoia and perceptual abnormalities) in nonpsychiatric patients (Denicoff *et al.*, 1987). Second, epidemiological studies have repeatedly demonstrated associations between pre- and perinatal (see Pearce, 2001 for review) as well as childhood (Koponen *et al.*, 2004) viral infections and schizophrenia. Moreover, several studies have reported immune abnormalities in schizophrenia (reviewed in Muller *et al.*, 2000; Rothermundt *et al.*, 1998). These abnormalities range from alterations in white blood cell counts and activation, to changes in serum and CSF cytokine concentrations, and cytokine production by activated lymphocytes. The most consistent cytokine findings in schizophrenia are decreased mitogen stimulated IL-2 and IFN- γ production by CD4 lymphocytes, increased CSF and serum concentrations of IL-2 and IL-6, and increased serum soluble IL-2 receptors (for review see Smith, 1992). It has been suggested that abnormally activated T lymphocytes oversecrete IL-2 leading to both increased serum IL-2 concentrations and depletion of IL-2 in lymphocytes themselves (Smith, 1992; Smith and Maes, 1995). These abnormalities are more pronounced in patients with treatment-resistant schizophrenia, and have been associated with the severity of positive symptoms and poor treatment outcome (Arolt *et al.*, 2000; McAllister *et al.*, 1995; Zhang *et al.*, 2002, 2005). In addition, antipsychotic drugs seem to have an inhibitory effect on inflammatory cytokines (Arolt *et al.*, 2000; Cazzullo *et al.*, 2002; Rothermundt *et al.*, 2000; Sirota *et al.*, 2005; Song *et al.*, 2000; Zhang *et al.*, 2005 but see also Kim *et al.*, 1995; Muller *et al.*, 1997). In addition to the variability commonly observed in clinical studies, cytokine studies exhibit differences in bioassays, cell preparation techniques, ongoing infectious diseases, production of counterregulatory cytokines, stress, and circadian variation. It has been suggested that abnormally activated T lymphocytes oversecrete IL-2 leading to both increased serum IL-2 concentrations and depletion of IL-2 in lymphocytes themselves (Smith, 1992; Smith and Maes, 1995).

A study with 230 schizophrenic patients found an association between a single nucleotide polymorphism in the IL-2 gene and schizophrenia (Schwarz *et al.*, 2006). It is interesting to note that a double-blind, placebo-controlled study revealed that addition of the cyclo-oxygenase 2 inhibitor celecoxib (an anti-inflammatory drug used in the treatment of rheumatoid arthritis) to risperidone markedly improved psychotic symptoms in schizophrenic patients (Muller *et al.*, 2004). Many of these immunologic abnormalities in schizophrenia resemble the natural history (including age of onset, relapsing course, and differential gender

distribution) of a number of well-known immune diseases such as rheumatoid arthritis and systemic lupus erythematosus. Additionally, proinflammatory cytokine concentrations are increased by stress and have been postulated to play a key role in depression (Raison *et al.*, 2006). Overall, it is tempting to posit a role for proinflammatory cytokines, particularly IL-2, in the pathogenesis of schizophrenia, mostly in mediating the increase in schizophrenia associated with birth season and perinatal insult.

V. Neurotensin

The 13 amino acid-containing peptide NT was first isolated by Carraway and Leeman 1973 and is encoded on human chromosome 12 (Marondel *et al.*, 1996). NT is widely distributed in the gastrointestinal tract, circulatory system, and in the central and peripheral nervous systems. In the CNS, NT is involved in regulation of reward, pain and body temperature, and has been hypothesized to play a role in the pathophysiology of schizophrenia, in the mechanism of action of antipsychotic drugs and in drug abuse. There are three identified NT receptors (NT₁–NT₃) and a fourth putative receptor (NT₄). NT₁ and NT₂ are GPCRs, whereas the NT₃ and putative NT₄ receptors are members of the vacuolar sorting and LDL receptor families (Dobner, 2005; Vincent *et al.*, 1999).

NT was first proposed by our group to be an endogenous antipsychotic in 1980 based on the close anatomical and neurochemical interactions between NT and the DA system and the striking behavioral similarities between the effects of antipsychotic drugs and centrally administered NT (Nemeroff, 1980). Since then, copious data supporting this hypothesis have been published (for review see Cáceda *et al.*, 2003; Dobner, 2005; Kinkead and Nemeroff, 2004).

The strongest clinical research evidence for the involvement of NT in schizophrenia has been provided by the measurement of NT concentrations in the CSF of drug-free and antipsychotic drug-treated schizophrenic patients. CSF NT concentrations are independent of serum concentrations and are believed to largely reflect CNS NTergic activity (Widerlöv *et al.*, 1982). CSF NT concentrations did not vary with patient age, duration of disease, or previous antipsychotic drug treatment (Lindström *et al.*, 1988). Low CSF NT concentrations have consistently been found in a subset of drug-free schizophrenic patients relative to normal volunteers and patients with other psychiatric disorders (Breslin *et al.*, 1994; Lindström *et al.*, 1988; Manberg *et al.*, 1985; Nemeroff *et al.*, 1989; Sharma *et al.*, 1994; Widerlöv *et al.*, 1982). In this subset of patients, clinical improvement (especially in negative symptoms) was associated with normalization of CSF NT concentrations (Breslin *et al.*, 1994; Garver *et al.*, 1991; Sharma *et al.*, 1997).

Low CSF NT concentrations were positively correlated with the severity of psychopathology, including thought disorder, deficit symptoms, disorganized behavior, and impaired functioning (Garver *et al.*, 1991; Sharma *et al.*, 1997). There is considerable specificity to schizophrenic patients, as CSF NT concentrations were unchanged in patients with other neuropsychiatric conditions, including anorexia/bulimia, depression, premenstrual syndrome, and Huntington's disease when compared to age-matched control subjects (Nemeroff *et al.*, 1989). Until technological advances to assess CNS NTergic neurotransmission such as the use of PET or SPECT to measure NT receptor subtype ligands are available, CSF studies represent the best *in vivo* evidence of NT dysfunction in schizophrenia. Taken together, these studies reveal a subset of schizophrenic patients with reduced NT neurotransmission.

Examination of the NT system in human postmortem tissue studies has generated variable results with several negative studies (Manberg *et al.*, 1982; Palacios *et al.*, 1991; Zech *et al.*, 1986). Among the positive findings are increased NT-IR in the frontal cortex (Manberg *et al.*, 1985; Nemeroff *et al.*, 1983), decreased NT receptor binding in layer II of the entorhinal cortex, caudate/putamen, and cingulate cortex (Hamid *et al.*, 2002; Lahti *et al.*, 1998; Wolf *et al.*, 1995), and increased NT receptor binding in the substantia nigra of medicated patients (Uhl and Kuhar, 1984). See Table III for summary of results.

Several polymorphisms have been identified in the noncoding region of the human NT₁ receptor gene (Austin *et al.*, 2000a; Huezio-Diaz *et al.*, 2004; Le *et al.*, 1997a,b; Watson *et al.*, 1993). However, no associations have been found between genetic variations in the NT or NT₁ receptor genes and schizophrenia (Austin *et al.*, 2000a,b; Huezio-Diaz *et al.*, 2004).

In contrast to the hypothesis that NT serves as an endogenous antipsychotic, is evidence that NT receptor antagonists may exhibit antipsychotic properties. Chronic administration of an NT receptor antagonist, like antipsychotic drugs, produces depolarization block in the ventral tegmental area (Santucci *et al.*, 1997). Additionally, similar to the effects of both NT receptor agonists (Feifel *et al.*, 1999) and antipsychotic drugs (Bakshi and Geyer, 1995; Geyer *et al.*, 2001), acute administration of an NT receptor antagonist prevents amphetamine- and dizocilpine-induced disruption of prepulse inhibition (PPI) of the startle response in rats (Cáceda *et al.*, submitted for publication). These seemingly contradictory findings are most likely explained by the different roles played by distinct anatomical NT circuits. NT has antipsychotic-like behavioral effects in the NAcc but stimulant-like behavioral effects in the VTA (Cáceda *et al.*, 2005; Feifel *et al.*, 1997; Kalivas *et al.*, 1981, 1984). The antipsychotic-like effects of systemic NT receptor antagonists may be due to blockade of NT neurotransmission in regions other than the NAcc, such as the VTA or subiculum. Despite this promissory preclinical evidence that NT receptor antagonists may have antipsychotic drug

properties, an NT receptor antagonist was ineffective in the treatment of refractory schizophrenic patients (Meltzer *et al.*, 2004). The patient population used in this study (refractory to antipsychotic drug treatment) and the testing of only a single (possibly suboptimal) dose of the NT receptor antagonist limits any firm conclusions to be drawn. Although NT receptor agonists have repeatedly been shown to possess antipsychotic-like behavioral effects in laboratory animals, no clinical trials using an NT receptor agonist have been conducted. This delay is due to the lack of a specific and potent small molecule NT receptor agonist.

VI. Neuropeptide Y

The pancreatic polypeptide family is composed of NPY, pancreatic polypeptide, peptide YY (PYY), and polypeptide Y. NPY is 36 amino acids long and is encoded on human chromosome 7. The five known receptors for NPY (NPY₁–NPY₅) are all GPCRs. Central NPY systems have been implicated in anxiety, major depression, bipolar disorder, suicide, and schizophrenia (for review see Obuchowicz *et al.*, 2004).

Of the three studies in which NPY concentrations were measured in the CSF of schizophrenic patients (Berrettini *et al.*, 1987; Peters *et al.*, 1990; Widerlöv *et al.*, 1988), only one found an increase in schizophrenic patients (drug free and after haloperidol withdrawal) compared to normal controls (Peters *et al.*, 1990). In this study, CSF concentrations of NPY positively correlated with duration of illness, the presence of abnormalities on brain CT scans and severity of clinical symptomatology in stable patients (Peters *et al.*, 1990).

Abnormal distribution of NPY positive interneurons (Ikeda *et al.*, 2004) and decreased NPY mRNA expression (Kuromitsu *et al.*, 2001) were observed in the dorsal prefrontal cortex of schizophrenic patients in postmortem studies, especially in the disorganized and paranoid type (but see Caberlotto and Hurd, 1999, 2001). Additionally, decreased NPY-IR was reported in the cingulate and temporal cortices of schizophrenic patients (Frederiksen *et al.*, 1991; Gabriel *et al.*, 1996). Finally, morphological alterations in NPY positive fibers have been reported in the CA4 region of the hippocampus in schizophrenia (Iritani *et al.*, 2000; Table III).

An association between a polymorphism in the promoter region of NPY and schizophrenia was reported (Buckland *et al.*, 2004; Itokawa *et al.*, 2003) but not replicated (Duan *et al.*, 2005; Lindberg *et al.*, 2006). In addition, two studies found no association between polymorphisms in the NPY gene and schizophrenia (Detera-Wadleigh *et al.*, 1987; Duan *et al.*, 2005). There are no published clinical trials in schizophrenia with compounds that increase or decrease NPY neurotransmission.

VII. Opioid Peptides

Three families of opioid peptides are known, each arising from different genes and precursor molecules: enkephalin A, proopiomelanocortin (POMC), and proenkephalin B or prodynorphin. Met-enkephalin, β -endorphin, and dynorphin are the best known molecules (Table I). The precursor molecules undergo extensive posttranslational modifications in the Golgi apparatus, including cleavage and acetylation. Opioid peptides interact with the μ , κ , σ , and δ GPCRs. The opioid receptors are associated with, and display differential selectivity for, each of the opioid peptides.

A. ENDORPHINS

Endorphins, relatively selective for the μ receptor, are widely expressed in the brain and spinal cord, particularly in the median eminence, periaqueductal gray matter, and substantia nigra (Zamir *et al.*, 1984). A large body of research has investigated whether endorphins play a preeminent role in pain regulation, reward, and drug addiction, as well as in schizophrenia and mood disorders. The hypothesized role of endorphins in schizophrenia is based on the antipsychotic-like effects of γ -endorphin in rodents and the reports of elevated concentrations of nonbiologically active α and γ -endorphins in the hypothalamus of schizophrenic patients (Wiegant *et al.*, 1988). However, the existent clinical research data do not support a role for endorphins in the pathophysiology of schizophrenia (for review see De Wied and Sigling, 2002; Wiegant *et al.*, 1992). CSF β -endorphin concentrations in unmedicated schizophrenic patients have been reported to be decreased (Naber *et al.*, 1981; Pickar *et al.*, 1981), unchanged (Burbach *et al.*, 1979; Emrich *et al.*, 1979; Gerner and Sharp, 1982), or increased (Domschke *et al.*, 1979; Lindström *et al.*, 1986, 1992) and decreased after antipsychotic drug treatment (Lindström *et al.*, 1986, 1992; Rimon *et al.*, 1980).

Initial open and double-blind studies with β -endorphin or [Des-Tyr1]-gamma-endorphin (DT γ E) alone or in combination with antipsychotic drugs in schizophrenic patients produced some positive results (see Wiegant *et al.*, 1992 for review), especially in the hebephrenic and paranoid subtypes (Verhoeven *et al.*, 1979, 1984). However, double-blind, placebo-controlled studies with more than 90 patients failed to demonstrate efficacy compared to placebo (Azorin *et al.*, 1990; Montgomery *et al.*, 1992). Similarly, although case reports and small pilot studies reported that the opiate antagonists, naltrexone and naloxone, demonstrate clinical efficacy (particularly against hallucinations) double-blind placebo-controlled studies failed to replicate these findings (see Welch and Thompson, 1994 for review).

B. DYNORPHIN

κ Opiate receptors have a high affinity and are relatively selective for dynorphin. Dynorphin is found in high concentrations in the neo- and allocortices, caudate/putamen, NAcc, amygdala, BNST, hypothalamus, medial preteccal area, nucleus of the optic tract, periaqueductal gray, raphe nuclei, and in brain stem nuclei involved in pain and nociception (Fallon and Leslie, 1986). Increased CSF concentrations of dynorphin were reported in unmedicated schizophrenic patients compared to healthy controls or other psychiatric patients (Heikkilä *et al.*, 1990; Lindström, 1996), with no decrease after antipsychotic treatment. CSF dynorphin concentrations were associated with symptom severity, as well as poor clinical outcome (Heikkilä *et al.*, 1990; Lindström, 1996 but see also Zhang *et al.*, 1985). Postmortem studies have revealed no alterations in dynorphin-IR or κ receptor expression in schizophrenic patients (Hurd, 2002; Iadarola *et al.*, 1991; Peckys and Hurd, 2001).

Although a direct association between polymorphisms in the prodynorphin gene promoter and schizophrenia has not been observed, an increased risk of susceptibility to schizophrenia was associated with the Ser9Gly polymorphism in the DA D₃ receptor, particularly in individuals carrying allele 3 of the prodynorphin gene. It was suggested that prodynorphin and DA D₃ receptor genes cooperatively contribute to a background of susceptibility to the development of schizophrenia (Ventriglia *et al.*, 2002).

C. ENKEPHALINS

δ Receptors are relatively selective for the enkephalins. The anatomical distribution of enkephalins is similar to dynorphin (Fallon and Leslie, 1986). Few studies examining the role of enkephalins in schizophrenia have been published. Decreased met-enkephalin concentrations were reported in the CSF (Burbach *et al.*, 1979; Wen *et al.*, 1983) and caudate/putamen (Kleinman *et al.*, 1985) of schizophrenic patients. Increased met-enkephalin concentrations were reported in the frontal cortex and substantia nigra, with no changes in the thalamus, or parietal and occipital cortices (Toru *et al.*, 1988).

A single mutation in the promoter region of the proenkephalin A gene was found in one schizophrenic patient, but in a larger study no more subjects with this mutation were found and its functional significance remains obscure (Mikesell *et al.*, 1997, 1996). Enkephalin-related compounds have not proven efficacious in the treatment of schizophrenia (Azorin *et al.*, 1990; de Jongh *et al.*, 1982; Jorgensen *et al.*, 1993).

VIII. Secretin

In 1902, Bayliss and Starling identified the first peptide ever discovered, secretin. Secretin, a member of the secretin/somatostatin/VIP superfamily, is composed of 27 amino acids (for review see Chey and Chang, 2003). In the brain, secretin is most abundant in the hippocampus and hypothalamus. Peripheral secretin administration increases fos protein expression in the central nucleus of the amygdala in rats (Goulet *et al.*, 2003), induces cAMP formation in the hippocampus and hypothalamus (Karelson *et al.*, 1995), and antagonizes Phencyclidine (PCP)-induced PPI disruption in rats (Myers *et al.*, 2005). In a double-blind clinical trial in which patients with refractory schizophrenia received a single intravenous injection of porcine secretin or placebo, a number of patients exhibited a transient (up to 4 days) but significant improvement in symptoms, although the overall effect was not significant (Alamy *et al.*, 2004; Sheitman *et al.*, 2004). Data from human and rodent studies suggest that the antipsychotic-like behavioral effects of secretin may be related to the effects of secretin in the limbic system (amygdala or hippocampus). Further investigation into the clinical potential of secretin in the treatment of schizophrenia is warranted.

IX. Somatostatin

Somatostatin, also known as growth hormone release-inhibiting hormone, was discovered in 1968 and five receptors have been identified, all of which are GPCRs (see Panteris and Karamanolis, 2005 for review). Two cyclic splice variants of somatostatin exist, 14 and 28 amino acids, each displaying unique tissue distributions. Somatostatin 28 is the most abundant form in the nervous system, with highest concentrations found in the olfactory tubercles, superior and inferior colliculi, and cerebellum.

The majority of CSF studies found no significant differences in CSF somatostatin concentrations in schizophrenic patients compared to controls (Banki *et al.*, 1992a,b; Gerner and Yamada, 1982; Heikkilä, 1993; Rubinow, 1986). However, reports of both increased and decreased CSF somatostatin concentrations in drug free schizophrenic patients have also been reported (Bissette *et al.*, 1986; Gerner *et al.*, 1985). Variable results have also been obtained after antipsychotic drug administration (Doran *et al.*, 1989; Sharma *et al.*, 1994). Low concentrations of CSF somatostatin were reported in schizophrenic, as well as in other psychiatric patients who were dexamethasone nonsuppressors, suggesting a functional relationship between HPA axis hyperactivity and reduced CSF somatostatin concentrations (Doran *et al.*, 1986; Rubinow, 1986). Both plasma somatostatin

concentrations (Saiz-Ruiz *et al.*, 1992) and serum somatostatin autoantibodies (Rogaeva *et al.*, 1990; Roy *et al.*, 1994) are repeatedly found to be increased in schizophrenic patients.

Postmortem studies of somatostatin in schizophrenia consistently demonstrate decreased somatostatin concentrations in the hippocampus of patients with predominant negative symptoms (Ferrier *et al.*, 1983; Roberts *et al.*, 1983). Decreased somatostatin concentrations have also been reported in cerebral cortex and lateral thalamus in schizophrenia (Gabriel *et al.*, 1996; Nemeroff *et al.*, 1983; Roberts *et al.*, 1983).

To date, no associations between somatostatin system genes and schizophrenia have been reported (Detera-Wadleigh *et al.*, 1987). Several somatostatin agonists have been approved by the FDA for the treatment of cancer, diabetes, and normalization of digestive function; however, no clinical trials in schizophrenia have been conducted.

X. Vasoactive Intestinal Peptide

The 28 amino acid peptide VIP is a potent vasodilator with a wide distribution in the CNS and in the periphery (for review see Delgado *et al.*, 2004). VIP acts on two identified receptor subtypes (VPAC₁ and VPAC₂) and historically is associated with regulation of digestive function (Harmar *et al.*, 1998). There are few studies exploring the role of VIP in schizophrenia.

No alterations in VIP concentrations have been found in the CSF of schizophrenic patients (Gjerris *et al.*, 1984; Rafaelsen and Gjerris, 1985). Postmortem studies report increased VIP concentrations in the amygdala, particularly in the central nucleus (Roberts *et al.*, 1983; Zech *et al.*, 1986). The most consistent results have been decreased VIP concentrations in the lymphocytes of schizophrenic patients, which are not altered by haloperidol treatment (Mauri *et al.*, 1998; Panerai and Sacerdote, 1993; Panza *et al.*, 1992). This observation has been postulated to be associated with the low prevalence of appendicitis in schizophrenic patients (Lauerma, 1999).

XI. Tachykinins

Mammalian tachykinins include the closely related peptides substance P (SP), neurokinin A, neurokinin B, neuropeptide K, neuropeptide γ , and hemokinin 1. The neurokinins share the common C-terminal sequence Phe-X-Gly-Leu-Met-NH₂ and are encoded by two preprotachykinin genes. All of the identified

neurokinin receptors (NK₁-NK₃) are GPCRs (see Almeida *et al.*, 2004 and Page, 2005 for review). Neurokinins and their receptors are widely and heterogeneously distributed in the CNS, particularly in cerebral cortex, NAcc, amygdala, and hypothalamus (Gale *et al.*, 1978). NK₃ is the dominant neurokinin receptor in the rat brain, whereas in the human, NK₁ is the most prevalent.

Substance P concentrations in CSF (Heikkilä *et al.*, 1990; Miller *et al.*, 1996; Rimón *et al.*, 1984) and plasma (Kaiya *et al.*, 1981) of schizophrenic patients do not differ from controls. In postmortem studies, NK₁ receptor-IR is increased in the prefrontal cortex (Tooney *et al.*, 2001). SP concentrations are increased in the prefrontal cortex, thalamus, hippocampus, and substantia nigra (Roberts *et al.*, 1983; Toru *et al.*, 1988) and decreased in the amygdala (Carletti *et al.*, 2005; Toru *et al.*, 1988). Additionally, decreased preprotachykinin A mRNA expression was reported in the amygdala (Carletti *et al.*, 2005). Two studies failed to find associations between SP, or angiotensin converting enzyme (likely the enzyme largely responsible for SP degradation in the CNS) and susceptibility to schizophrenia (Arinami *et al.*, 1996; Detera-Wadleigh *et al.*, 1987).

Because activation of NK₃ receptors increases the firing rate of DA cells in the VTA and subsequent DA release in the ventral striatum, it has been hypothesized that NK₃ receptor antagonists may possess antipsychotic-like behavioral effects (Marco *et al.*, 1998; Nalivaiko *et al.*, 1997; Seabrook *et al.*, 1995) and two clinical trials have demonstrated clinical efficacy of NK₃ receptor antagonists. The nonpeptide NK₃ receptor antagonist Talnetant (GlaxoSmithKline) was effective at improving positive and possibly cognitive symptoms in schizophrenic patients in a phase II clinical trial (Spooren *et al.*, 2005). Similarly, the nonpeptide NK₃ receptor antagonist Osanetant (SR 142801) improved total BPRS scores and positive symptoms in patients with schizophrenia and schizoaffective disorder in a multiarm clinical trial (Meltzer *et al.*, 2004). Both drugs were well tolerated and their side effects did not differ from those of placebo. Additional studies with both agents are currently underway.

XII. Thyrotropin-Releasing Hormone

The best characterized function of TRH is regulation of the hypothalamus-pituitary-thyroid (HPT) axis. TRH is associated with two GPCR subtypes; TRH-R1 and TRH-R2. TRH is found in the olfactory bulbs, piriform and entorhinal cortices, hippocampus, amygdala, NAcc, olfactory tubercle, parvocellular portion of the PVN and the periaqueductal central gray. In addition to a central role in the regulation of the HPT axis, TRH has been hypothesized to play a role in regulation of autonomic functions, arousal and cognition, locomotion, and water intake (for review see Prokai, 2002).

The TRH stimulation test assesses HPT axis function by measurement of thyroid stimulating hormone (TSH) secretion after intravenous administration of a standard dose of TRH. The TRH-induced TSH response is blunted in a significant proportion (25–33%) of depressed, alcoholic, and personality disorder patients (for review see Loosen, 1985). The frequency of a blunted TSH response to TRH in schizophrenic patients is lower than that observed in other psychiatric groups and has been associated with favorable antipsychotic drug response (Braddock and Blake, 1981; Garver, 1988; Langer *et al.*, 1986; Yazici *et al.*, 2002). No difference has been found in CSF TRH concentrations between schizophrenic patients and controls (Banki *et al.*, 1992a,b; Gjerris *et al.*, 1985; Sharma *et al.*, 2001). The only positive finding in postmortem studies of schizophrenic patients is decreased TRH-IR in the frontal cortex (Biggins *et al.*, 1983; Nemeroff *et al.*, 1983). Currently, there are no genetic studies of TRH polymorphisms in schizophrenia.

Clinical trials with TRH or its analogues have produced variable results, including exacerbation of symptoms (Bigelow *et al.*, 1975; Davis *et al.*, 1975; Wilson *et al.*, 1973), absence of therapeutic effects (Clark *et al.*, 1975; Lindström *et al.*, 1977) and improvement of positive and negative symptoms and emotionality (Brambilla *et al.*, 1986; Inanaga *et al.*, 1978; Kobayashi *et al.*, 1980; Mizuki *et al.*, 1985, 1986; Prange *et al.*, 1979).

XIII. Other Peptides

On the basis of animal data, clinical studies have investigated the role of galanin, hypocretin/orexin, arginine-vasopressin, oxytocin, and LHRH in schizophrenia. Galanin-IR is reportedly reduced in the temporal (Frederiksen *et al.*, 1991) but not frontal or occipital cortex (Sharma *et al.*, 1994) of schizophrenic patients.

The close anatomical association between hypocretin/orexin and the meso-limbic DA system was the basis for exploration of this peptide system in schizophrenia. Whereas no differences in CSF hypocretin concentrations were found between schizophrenic patients and control subjects, CSF hypocretin concentrations were significantly correlated with sleep latency in schizophrenics, one of the most consistent sleep abnormalities in schizophrenia (Nishino *et al.*, 2002). A study found decreased CSF hypocretin in patients with schizophrenia treated with haloperidol, but not atypical antipsychotic drugs, compared to unmedicated subjects (Dalal *et al.*, 2003). Additionally, a single nucleotide polymorphism in the hypocretin 1 receptor gene was associated with polydipsia-hyponatremia in schizophrenia, a not uncommon condition that appears after years of antipsychotic

drug treatment (Meerabux *et al.*, 2005). Overall, these findings suggest that hypocretin may be associated with antipsychotic drug response.

The nonapeptide arginine-vasopressin (AVP) plays a crucial role in the control of water balance in humans. Considerable attention has been given to AVP in relation to the common prevalence of hyponatremia in schizophrenic patients (3–5%) who develop potentially fatal episodes of water intoxication associated with impaired water secretion (de Leon *et al.*, 1994). Schizophrenic patients, particularly the subset that develops hyponatremia, display increased AVP plasma concentrations (Delva *et al.*, 1990; Ryan *et al.*, 2004) and alterations in AVP regulation (Goldman *et al.*, 1996; Kishimoto *et al.*, 1989; Ohsawa *et al.*, 1993). Additionally, water intoxication crises and enhanced AVP release often coincide with psychotic exacerbations (Goldman *et al.*, 1997). CSF studies have found no difference in AVP between schizophrenic patients and controls (Gjerris *et al.*, 1985; Glovinsky *et al.*, 1994; Sorensen *et al.*, 1985). Decreased AVP content was reported in the temporal cortex, but not the hypothalamus, of schizophrenic patients (Frederiksen *et al.*, 1991).

The neurohypophyseal peptide oxytocin, best known for its role in parturition, lactation, and regulation of social behavior, has also been studied in schizophrenic patients. CSF oxytocin levels were increased in drug naïve patients and increased with antipsychotic drug treatment (Beckmann *et al.*, 1985 but also see Glovinsky *et al.*, 1994). One investigator has reported improvement of psychotic symptoms in open clinical trials with oxytocin (Bujanow, 1972).

Luteinizing hormone-releasing hormone (LHRH), (gonadotropin-releasing hormone (GnRH) or gonadorelin, controls sex hormones and regulates reproductive behavior in humans and several mammals. Normal basal secretion of gonadorelin has been reported in men, women, and adolescent schizophrenic patients (Apter *et al.*, 1983; Brown *et al.*, 1995; Gil-Ad *et al.*, 1981). However, increased response of growth hormone (Gil-Ad *et al.*, 1981) and LH (Brambilla *et al.*, 1976 but also see Brown *et al.*, 1995), to LHRH challenge has also been described. The effect of antipsychotic drug treatment on the response to LHRH is unclear (Apter *et al.*, 1983; Brambilla *et al.*, 1976; Gil-Ad *et al.*, 1981; Naber *et al.*, 1980). No postmortem studies or clinical trials involving LHRH have been published.

Neuregulin 1 (NRG1) is part of epithelial growth factor family and is associated with the erbB receptors. Neuregulin has crucial roles in neurodevelopmental processes, including neuronal migration, myelination, hormonal control of puberty, synaptic plasticity, and regulation of neurotransmitter expression and signaling (Corfas *et al.*, 2004; Owen *et al.*, 2005). An initial report from a genome-wide scan identified a haplotype in the 5'-end of the NRG1 gene with a highly significant association with schizophrenia in an Icelandic population (Stefansson *et al.*, 2002). Subsequently, NRG1 has been repeatedly found to be a susceptibility

gene for schizophrenia in subjects of European (Bakker *et al.*, 2004; Corvin *et al.*, 2004; Green *et al.*, 2005; Norton *et al.*, 2006; Petryshen *et al.*, 2005; Stefansson *et al.*, 2003; Williams *et al.*, 2003), Asian (Fukui *et al.*, 2006; Li *et al.*, 2004; Liu *et al.*, 2005; Tang *et al.*, 2004; Yang *et al.*, 2003; Zhao *et al.*, 2004), and African descent (Lachman *et al.*, 2006) (but also see (Hong *et al.*, 2004; Iwata *et al.*, 2004; Kampman *et al.*, 2004; Thiselton *et al.*, 2004). Additionally, the NRG1 gene has been associated with poor response to antipsychotic drugs (Kampman *et al.*, 2004) and susceptibility to bipolar disorder (Green *et al.*, 2005; Tkachev *et al.*, 2003). Furthermore, NRG1 gene is suspected to interact with erB4 to increase susceptibility to schizophrenia (Norton *et al.*, 2006). In agreement with these genetic studies are reports of decreased expression or altered expression of NRG1 (Hashimoto *et al.*, 2004) and its receptor erB3 (Hakak *et al.*, 2001; Tkachev *et al.*, 2003) and erB4 (Silberberg *et al.*, 2006) in the prefrontal cortex of schizophrenic patients. Overall, NRG1 despite its relatively recent identification, is the one with the strongest genetic data in schizophrenia and its study promises to greatly expand our understanding of this disease.

XIV. Conclusions

A clear role for neuropeptides in the etiology, pathophysiology, and treatment response of schizophrenia has not been consistently demonstrated. Factors contributing to the negative clinical data include the lack of identifiable subgroups of schizophrenic patients, the inherent heterogeneity associated with human studies (including low subject number and heterogeneity of the patient population), and inadequate methods for evaluation of peptide systems antemortem.

In postmortem studies, the frontal and temporal cortices show the most consistent abnormalities in neuropeptides and neuropeptide receptors in schizophrenic patients. These abnormalities are primarily of a quantitative (variation in the concentration of peptides or their receptors) rather than qualitative (variation in the distribution of peptides or their receptors) nature. Despite substantial preclinical evidence suggesting that neuropeptide systems in the mesolimbic pathways (specifically in the NAcc and VTA) may be involved in the pathophysiology of schizophrenia, postmortem data do not support this hypothesis.

Overall, genetic evidence of neuropeptide abnormalities in schizophrenia is weak, with the possible exception of the association between polymorphisms in the NRG1 gene and schizophrenia, as well as in the CCK_A receptor gene and positive symptoms. Ultimately, polymorphisms in the promoter regions, rather than in processing regions (i.e., exons and introns), of neuropeptide genes may prove more likely to be associated with the degree of gene expression and with the pathogenesis of schizophrenia (Tachikawa *et al.*, 2000, 2001).

Numerous agonist and antagonist ligands for neuropeptide receptors have been developed and tested for clinical efficacy in the treatment of schizophrenia. Although the majority of clinical trials are negative (possibly due to low sample size, short treatment duration, and the use of treatment-resistant populations) data support further development and testing of NK₃ receptor antagonists, and possibly secretin, TRH, and CCK_A and NT receptor agonists. In addition to larger controlled clinical trials to replicate and extend these findings, the use of psychopharmacogenetics would allow identification of specific subgroups of patients that would benefit from specific treatments. Additionally, although data support the testing of peptide agonists within several peptide systems, neuropeptide receptor antagonists seem to be the prime pharmacological targets to pursue, due to the greater availability of small molecule nonpeptide antagonists (with greater stability and ability to cross the blood-brain barrier) and reduced likelihood of developing receptor downregulation and tachyphylaxis.

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References

- Alamy, S. S., Jarskog, L. F., Sheitman, B. B., and Lieberman, J. A. (2004). Secretin in a patient with treatment-resistant schizophrenia and prominent autistic features. *Schizophr. Res.* **66**, 183–186.
- Albus, M., Ackenheil, M., Munch, U., and Naber, D. (1984). Ceruletide: A new drug for the treatment of schizophrenic patients? *Arch. Gen. Psychiatry* **41**, 528.
- Albus, M., von Gellhorn, K., Munch, U., Naber, D., and Ackenheil, M. (1986). A double-blind study with ceruletide in chronic schizophrenic patients: Biochemical and clinical results. *Psychiatry Res.* **19**, 1–7.

- Almeida, T. A., Rojo, J., Nieto, P. M., Pinto, F. M., Hernandez, M., Martin, J. D., and Candenas, M. L. (2004). Tachykinins and tachykinin receptors: Structure and activity relationships. *Curr. Med. Chem.* **11**, 2045–2081.
- Altamura, A. C., Boin, F., and Maes, M. (1999). HPA axis and cytokines dysregulation in schizophrenia: Potential implications for the antipsychotic treatment. *Eur. Neuropsychopharmacol.* **10**, 1–4.
- Altamura, C., Guercetti, G., and Percudani, M. (1989). Dexamethasone suppression test in positive and negative schizophrenia. *Psychiatry Res.* **30**, 69–75.
- Apter, A., Dickerman, Z., Gonen, N., Assa, S., Prager-Lewin, R., Kaufman, H., Tyano, S., and Laron, Z. (1983). Effect of chlorpromazine on hypothalamic-pituitary-gonadal function in 10 adolescent schizophrenic boys. *Am. J. Psychiatry* **140**, 1588–1591.
- Arinami, T., Li, L., Mitsushio, H., Itokawa, M., Hamaguchi, H., and Toru, M. (1996). An insertion/deletion polymorphism in the angiotensin converting enzyme gene is associated with both brain substance P contents and affective disorders. *Biol. Psychiatry* **40**, 1122–1127.
- Arolt, V., Rothermundt, M., Wandering, K. P., and Kirchner, H. (2000). Decreased *in vitro* production of interferon-gamma and interleukin-2 in whole blood of patients with schizophrenia during treatment. *Mol. Psychiatry* **5**, 150–158.
- Austin, J., Buckland, P., Cardno, A. G., Williams, N., Spurlock, G., Hoogendoorn, B., Zammit, S., Jones, G., Sanders, R., Jones, L., McCarthy, G., Jones, S., *et al.* (2000a). The high affinity neurotensin receptor gene (NTSR1): Comparative sequencing and association studies in schizophrenia. *Mol. Psychiatry* **5**, 552–557.
- Austin, J., Hoogendoorn, B., Buckland, P., Speight, G., Cardno, A., Bowen, T., Williams, N., Spurlock, G., Sanders, R., Jones, L., Murphy, K., McCarthy, G., *et al.* (2000b). Comparative sequencing of the proneurotensin gene and association studies in schizophrenia. *Mol. Psychiatry* **5**, 208–212.
- Azorin, J. M., Blum, A., Charbaut, J., Escande, M., Granier, F., Huber, J. P., Metzger, J. Y., Richou, H., Sitsen, A., Van Amerongen, P., *et al.* (1990). Des-enkephalin-gamma-endorphin in the treatment of schizophrenia. *Int. Clin. Psychopharmacol.* **5**, 205–215.
- Bachus, S. E., Hyde, T. M., Herman, M. M., Egan, M. F., and Kleinman, J. E. (1997). Abnormal cholecystokinin mRNA levels in entorhinal cortex of schizophrenics. *J. Psychiatr. Res.* **31**, 233–256.
- Bakker, S. C., Hoogendoorn, M. L., Selten, J. P., Verduijn, W., Pearson, P. L., Sinke, R. J., and Kahn, R. S. (2004). Neuregulin1: Genetic support for schizophrenia subtypes. *Mol. Psychiatry* **9**, 1061–1063.
- Bakshi, V. P., and Geyer, M. A. (1995). Antagonism of phencyclidine-induced deficits in prepulse inhibition by the putative atypical antipsychotic olanzapine. *Psychopharmacology* **122**, 198–201.
- Banki, C. M., Bissette, G., Arato, M., O'Connor, L., and Nemeroff, C. B. (1987). CSF corticotropin-releasing factor-like immunoreactivity in depression and schizophrenia. *Am. J. Psychiatry* **144**, 873–877.
- Banki, C. M., Karmacsi, L., Bissette, G., and Nemeroff, C. B. (1992a). Cerebrospinal-fluid neuropeptides: A biochemical subgrouping approach. *Neuropsychobiology* **26**, 37–42.
- Banki, C. M., Karmacsi, L., Bissette, G., and Nemeroff, C. B. (1992b). CSF corticotropin releasing hormone, somatostatin, and thyrotropin releasing hormone in schizophrenia. *Psychiatry Res.* **43**, 13–21.
- Barak, V., Barak, Y., Levine, J., Nisman, B., and Roisman, I. (1995). Changes in interleukin-1 beta and soluble interleukin-2 receptor levels in CSF and serum of schizophrenic patients. *J. Basic Clin. Physiol. Pharmacol.* **6**, 61–69.
- Bayliss, H. P., and Starling, E. H. (1902). Mechanism of pancreatic secretion. *J. Physiol. Lond* **28**, 325–353.
- Beal, M. F., Svendsen, C. N., Bird, E. D., and Martin, J. B. (1987). Somatostatin and neuropeptide Y are unaltered in the amygdala in schizophrenia. *Neurochem. Pathol.* **6**, 169–176.

- Beckmann, H., Lang, R. E., and Gattaz, W. F. (1985). Vasopressin-oxytocin in cerebrospinal fluid of schizophrenic patients and normal controls. *Psychoneuroendocrinology* **10**, 187-191.
- Beinfeld, M. C., and Garver, D. L. (1991). Concentration of cholecystokinin in cerebrospinal fluid is decreased in psychosis: Relationship to symptoms and drug response. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **15**, 601-609.
- Bennett, G. W., Ballard, T. M., Watson, C. D., and Fone, K. C. (1997). Effect of neuropeptides on cognitive function. *Exp. Gerontol.* **32**, 451-469.
- Berger, P. A., Watson, S. J., Akil, H., and Barchas, J. D. (1981). The effects of naloxone in chronic schizophrenia. *Am. J. Psychiatry* **138**, 913-918.
- Berrettini, W. H., Doran, A. R., Kelsoe, J., Roy, A., and Pickar, D. (1987). Cerebrospinal fluid neuropeptide Y in depression and schizophrenia. *Neuropsychopharmacology* **1**, 81-83.
- Bielsky, I. F., Hu, S. B., and Young, L. J. (2005). Sexual dimorphism in the vasopressin system: lack of an altered behavioral phenotype in female V1a receptor knockout mice. *Behav. Brain Res.* **164**, 132-136.
- Bigelow, L. B., Gillin, J. C., Semal, C., and Wyatt, R. J. (1975). Letter: Thyrotropin-releasing hormone in chronic schizophrenia. *Lancet* **2**, 869-870.
- Biggins, J. A., Perry, E. K., McDermott, J. R., Smith, A. I., Perry, R. H., and Edwardson, J. A. (1983). Post mortem levels of thyrotropin-releasing hormone and neurotensin in the amygdala in Alzheimer's disease, schizophrenia and depression. *J. Neurol. Sci.* **58**, 117-122.
- Bissette, G., Widerlöv, E., Walleus, H., Karlsson, I., Eklund, K., Forsman, A., and Nemeroff, C. B. (1986). Alterations in cerebrospinal fluid concentrations of somatostatin-like immunoreactivity in neuropsychiatric disorders. *Arch. Gen. Psychiatry* **43**, 1148-1154.
- Bloom, D., Nair, N. P., and Schwartz, G. (1983). CCK-8 in the treatment of chronic schizophrenia. *Psychopharmacol. Bull.* **19**, 361-363.
- Borrell, J., Vela, J. M., Arevalo-Martin, A., Molina-Holgado, E., and Guaza, C. (2002). Prenatal immune challenge disrupts sensorimotor gating in adult rats: Implications for the etiopathogenesis of schizophrenia. *Neuropsychopharmacology* **26**, 204-215.
- Bowen, T., Norton, N., Jacobsen, N. J., Guy, C., Daniels, J. K., Sanders, R. D., Cardno, A. G., Jones, L. A., Murphy, K. C., McGuffin, P., Craddock, N., O'Donovan, M. C., *et al.* (1998). Linked polymorphisms upstream of exons 1 and 2 of the human cholecystokinin gene are not associated with schizophrenia or bipolar disorder. *Mol. Psychiatry* **3**, 67-71.
- Boza, R. A., and Rotondo, D. J. (1985). Is cholecystokinin therapeutic in chronic schizophrenia? *J. Clin. Psychiatry* **46**, 485-486.
- Braddock, L. E., and Blake, I. M. (1981). Neuroendocrine tests during treatment with neuroleptic drugs. II. The TRH test. *Br. J. Psychiatry* **139**, 404-407.
- Brambilla, F., Rovere, C., Guastalla, A., Guerrini, A., and Riggi, F. (1976). Gonadotropin response to synthetic gonadotropin hormone-releasing hormone (GnRH) in chronic schizophrenia. *Acta Psychiatr. Scand.* **54**, 131-145.
- Brambilla, F., Aguglia, E., Massironi, R., Maggioni, M., Grillo, W., Castiglioni, R., Catalano, M., and Drago, F. (1986). Neuropeptide therapies in chronic schizophrenia: TRH and vasopressin administration. *Neuropsychobiology* **15**, 114-121.
- Breslin, N. A., Suddath, R. L., Bissette, G., Nemeroff, C. B., Lowrimore, P., and Weinberger, D. R. (1994). CSF concentrations of neurotensin in schizophrenia: An investigation of clinical and biochemical correlates. *Schizophr. Res.* **12**, 35-41.
- Brown, A. S., Hembree, W. C., Friedman, J. H., Kaufmann, C. A., and Gorman, J. M. (1995). The gonadal axis in men with schizophrenia. *Psychiatry Res.* **57**, 231-239.
- Buckland, P. R., Hoogendoorn, B., Guy, C. A., Coleman, S. L., Smith, S. K., Buxbaum, J. D., Haroutunian, V., and O'Donovan, M. C. (2004). A high proportion of polymorphisms in the promoters of brain expressed genes influences transcriptional activity. *Biochem. Biophys. Acta* **1690**, 238-249.

- Bujanow, W. (1972). Hormones in the treatment of psychoses. *Br. Med. J.* **4**, 298.
- Bujanow, W. (1974). Is oxytocin an anti-schizophrenic hormone? *Can. Psychiatr. assoc. J.* **19**, 323.
- Burbach, J. P., Loeber, J. G., Verhoef, J., de Kloet, E. R., van Ree, J. M., and de Wied, D. (1979). Schizophrenia and degradation of endorphins in cerebrospinal fluid [letter]. *Lancet* **2**, 480–481.
- Caberlotto, L., and Hurd, Y. L. (1999). Reduced neuropeptide Y mRNA expression in the prefrontal cortex of subjects with bipolar disorder. *Neuroreport* **10**, 1747–1750.
- Caberlotto, L., and Hurd, Y. L. (2001). Neuropeptide Y Y(1) and Y(2) receptor mRNA expression in the prefrontal cortex of psychiatric subjects: Relationship of Y(2) subtype to suicidal behavior. *Neuropsychopharmacology* **25**, 91–97.
- Cáceda, R., Kinkead, B., and Nemeroff, C. B. (2003). Do neurotensin receptor agonists represent a novel class of antipsychotic drugs? *Sem. Clin. Neuropsychiatry* **8**, 94–108.
- Cáceda, R., Kinkead, B., Owens, M. J., and Nemeroff, C. B. (2005). Virally mediated increased neurotensin 1 receptor in the nucleus accumbens decreases behavioral effects of mesolimbic system activation. *J. Neurosci.* **25**, 11748–11756.
- Carletti, R., Corsi, M., Melotto, S., and Caberlotto, L. (2005). Down-regulation of amygdala preprotachykinin A mRNA but not 3H-SP receptor binding sites in subjects affected by mood disorders and schizophrenia. *Eur. J. Neurosci.* **21**, 1712–1718.
- Carraway, R. E., and Leeman, S. E. (1973). The isolation of a new hypotensive peptide, neurotensin, from bovine hypothalamus. *J. Biol. Chem.* **248**, 6854–6861.
- Casey, D. E., Korsgaard, S., Gerlach, J., Jorgensen, A., and Simmelsgaard, H. (1981). Effect of des-tyrosine-gamma-endorphin in tardive dyskinesia. *Arch. Gen. Psychiatry* **38**, 158–160.
- Cazzullo, C. L., Sacchetti, E., Galluzzo, A., Panariello, A., Adorni, A., Pegoraro, M., Bosis, S., Colombo, F., Trabattoni, D., Zagliani, A., and Clerici, M. (2002). Cytokine profiles in schizophrenic patients treated with risperidone: A 3-month follow-up study. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **26**, 33–39.
- Chey, W. Y., and Chang, T. M. (2003). Secretin, 100 years later. *J. Gastroenterol.* **38**, 1025–1035.
- Clark, M. L., Paredes, A., Costiloe, J. P., and Wood, F. (1975). Synthetic thyroid releasing hormone (TRH) administered orally to chronic schizophrenic patients. *Psychopharmacol. Commun.* **1**, 191–200.
- Cohen, M. R., Pickar, D., and Cohen, R. M. (1985). High-dose naloxone administration in chronic schizophrenia. *Biol. Psychiatry* **20**, 573–575.
- Colwell, C. S., Michel, S., Itri, J., Rodriguez, W., Tam, J., Lelievre, V., Hu, Z., Liu, X., and Waschek, J. A. (2003). Disrupted circadian rhythms in VIP- and PHI-deficient mice. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **285**, R939–R949.
- Corfas, G., Roy, K., and Buxbaum, J. D. (2004). Neuregulin 1-erbB signaling and the molecular/cellular basis of schizophrenia. *Nat. Neurosci.* **7**, 575–580.
- Corvin, A. P., Morris, D. W., McGhee, K., Schwaiger, S., Scully, P., Quinn, J., Meagher, D., Clair, D. S., Waddington, J. L., and Gill, M. (2004). Confirmation and refinement of an 'at-risk' haplotype for schizophrenia suggests the EST cluster, Hs.97362, as a potential susceptibility gene at the Neuregulin-1 locus. *Mol. Psychiatry* **9**, 208–213.
- Coryell, W., and Tsuang, D. (1992). Hypothalamic-pituitary-adrenal axis hyperactivity and psychosis: Recovery during an 8-year follow-up. *Am. J. Psychiatry* **149**, 1033–1039.
- Coste, S. C., Kesterson, R. A., Heldwein, K. A., Stevens, S. L., Heard, A. D., Hollis, J. H., Murray, S. E., Hill, J. K., Pantely, G. A., Hohimer, A. R., Hatton, D. C., Phillips, T. J., *et al.* (2000). Abnormal adaptations to stress and impaired cardiovascular function in mice lacking corticotropin-releasing hormone receptor-2. *Nat. Gen.* **24**, 403–409.
- Cotter, D., and Pariante, C. M. (2002). Stress and the progression of the developmental hypothesis of schizophrenia. *Br. J. Psychiatry* **181**, 363–365.
- Dalal, M. A., Schulz, A., and Pollmacher, T. (2003). Lower CSF orexin A (hypocretin-1) levels in patients with schizophrenia treated with haloperidol compared to unmedicated subjects. *Mol. Psychiatry* **8**, 836–837.

- Dauge, V., Sebret, A., Beslot, F., Matsui, T., and Roques, B. P. (2001). Behavioral profile of CCK2 receptor-deficient mice. *Neuropsychopharmacology* **25**, 690–698.
- Davis, K., Hossister, L. E., and Berger, P. A. (1975). Thyrotropin-releasing hormone in schizophrenia. *Am. J. Psychiatry* **132**, 951–953.
- de Jongh, B. M., Verhoeven, W. M., van Ree, J. M., de Wied, D., and van Rood, J. J. (1982). Hla, and the response to treatment with gamma-type endorphins in schizophrenia. *J. Immunogenet.* **9**, 381–388.
- de Leon, J., Verghese, C., Tracy, J. I., Josiassen, R. C., and Simpson, G. M. (1994). Polydipsia and water intoxication in psychiatric patients: A review of the epidemiological literature. *Biol. Psychiatry* **35**, 408–419.
- De Wied, D., and Sigling, H. O. (Wied 2002). Neuropeptides involved in the pathophysiology of schizophrenia and major depression. *Neurotoxicol. Res.* **4**, 453–468.
- Delgado, M., Pozo, D., and Ganea, D. (2004). The significance of vasoactive intestinal peptide in immunomodulation. *Pharmacol. Rev.* **56**, 249–290.
- Delva, N. J., Crammer, J. L., Lawson, J. S., Lightman, S. L., Sribney, M., and Weier, B. J. (1990). Vasopressin in chronic psychiatric patients with primary polydipsia. *Br. J. Psychiatry* **157**, 703–712.
- Denicoff, K. D., Rubinow, D. R., Papa, M. Z., Simpson, C., Seipp, C. A., Lotze, M. T., Chang, A. E., Rosenstein, D., and Rosenberg, S. A. (1987). The neuropsychiatric effects of treatment with interleukin-2 and lymphokine-activated killer cells. *Ann. Intern. Med.* **107**, 293–300.
- Detera-Wadleigh, S. D., de Miguel, C., Berrettini, W. H., DeLisi, L. E., Goldin, L. R., and Gershon, E. S. (1987). Neuropeptide gene polymorphisms in affective disorder and schizophrenia. *J. Psychiatr. Res.* **21**, 581–587.
- Dobner, P. R. (2005). Multitasking with neurotensin in the central nervous system. *Cell. Mol. Life Sci.* **62**, 1946–1963.
- Domschke, W., Dickschas, A., and Mitznegg, P. (1979). C.S.F. beta-endorphin in schizophrenia [letter]. *Lancet* **1**, 1024.
- Doran, A. R., Rubinow, D. R., Roy, A., and Pickar, D. (1986). CSF somatostatin and abnormal response to dexamethasone administration in schizophrenic and depressed patients. *Arch. Gen. Psychiatry* **43**, 365–369.
- Doran, A. R., Rubinow, D. R., Wolkowitz, O. M., Roy, A., Breier, A., and Pickar, D. (1989). Fluphenazine treatment reduces CSF somatostatin in patients with schizophrenia: Correlations with CSF HVA. *Biol. Psychiatry* **25**, 431–439.
- Duan, S., Gao, R., Xing, Q., Du, J., Liu, Z., Chen, Q., Wang, H., Feng, G., and He, L. (2005). A family-based association study of schizophrenia with polymorphisms at three candidate genes. *Neurosci. Lett.* **379**, 32–36.
- el-Mallakh, R. S., Suddath, R. L., and Wyatt, R. J. (1993). Interleukin-1 alpha and interleukin-2 in cerebrospinal fluid of schizophrenic subjects. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **17**, 383–391.
- Emrich, H. M., Holtt, V., Kissling, W., Fischler, M., Laspe, H., Heinemann, H., von Zerssen, D., and Herz, A. (1979). Beta-endorphin-like immunoreactivity in cerebrospinal fluid and plasma of patients with schizophrenia and other neuropsychiatric disorders. *Pharmakopsychiatrie Neuro-Psychopharmakologie* **12**, 269–276.
- Emrich, H. M., Zaudig, M., Kissling, W., Dirlich, G., von Zerssen, D., and Herz, A. (1980). Des-tyrosyl-gamma-endorphin in schizophrenia: A double-blind trial in 13 patients. *Pharmakopsychiatrie Neuro-Psychopharmakologie* **13**, 290–298.
- Fallon, J. H., and Leslie, F. M. (1986). Distribution of dynorphin and enkephalin peptides in the rat brain. *J. Comp. Neurol.* **249**, 293–336.
- Farmary, S. M., Owen, F., Poulter, M., and Crow, T. J. (1985). Reduced high affinity cholecystokinin binding in hippocampus and frontal cortex of schizophrenic patients. *Life Sci.* **36**, 473–477.

- Feifel, D., Minor, K. L., Dulawa, S., and Swerdlow, N. R. (1997). The effects of intra-accumbens neurotensin on sensorimotor gating. *Brain Res.* **760**, 80–84.
- Feifel, D., Reza, T. L., Wustrow, D. J., and Davis, M. D. (1999). Novel antipsychotic-like effects on prepulse inhibition of startle produced by a neurotensin agonist. *J. Pharmacol. Exp. Ther.* **288**, 710–713.
- Ferrier, I. N., Roberts, G. W., Crow, T. J., Johnstone, E. C., Owens, D. G., Lee, Y. C., O'Shaughnessy, D., Adrian, T. E., Polak, J. M., and Bloom, S. R. (1983). Reduced cholecystokinin-like and somatostatin-like immunoreactivity in limbic lobe is associated with negative symptoms in schizophrenia. *Life Sci.* **33**, 475–482.
- Fischman, A. J., and Moldow, R. L. (1982). Extrahypothalamic distribution of CRF-like immunoreactivity in the rat brain. *Peptides* **3**, 149–153.
- Forman, S. D., Bisette, G., Yao, J., Nemeroff, C. B., and van Kammen, D. P. (1994). Cerebrospinal fluid corticotropin-releasing factor increases following haloperidol withdrawal in chronic schizophrenia. *Schizophr. Res.* **12**, 43–51.
- Frederiksen, S. O., Ekman, R., Gottfries, C. G., Widerlöv, E., and Jonsson, S. (1991). Reduced concentrations of galanin, arginine vasopressin, neuropeptide Y and peptide YY in the temporal cortex but not in the hypothalamus of brains from schizophrenics. *Acta Psychiatr. Scand.* **83**, 273–277.
- Freeman, C. P., and Fairburn, C. G. (1981). Lack of effect of naloxone and schizophrenic auditory hallucinations. *Psychol. Med.* **11**, 405–407.
- Fukui, N., Muratake, T., Kaneko, N., Amagane, H., and Someya, T. (2006). Supportive evidence for neuregulin 1 as a susceptibility gene for schizophrenia in a Japanese population. *Neurosci. Lett.* **396**, 117–120.
- Gabriel, S. M., Bierer, L. M., Davidson, M., Purohit, D. P., Perl, D. P., and Haroutunian, V. (1994). Galanin-like immunoreactivity is increased in the postmortem cerebral cortex from patients with Alzheimer's disease. *J. Neurochem.* **62**, 1516–1523.
- Gabriel, S. M., Davidson, M., Haroutunian, V., Powchik, P., Bierer, L. M., Purohit, D. P., Perl, D. P., and Davis, K. L. (1996). Neuropeptide deficits in schizophrenia vs. Alzheimer's disease cerebral cortex. *Biol. Psychiatry* **39**, 82–91.
- Gale, J. S., Bird, E. D., Spoke, E. G., Iveysen, L. L., and Jessel, T. (1978). Human brain substance P: Distribution in controls and Huntington's chorea. *J. Neurochem.* **30**, 633–634.
- Garver, D. L. (1988). Neuroendocrine findings in the schizophrenias. *Endocrinol. Metab. Clin. North Am.* **17**, 103–109.
- Garver, D. L., Beinfeld, M. C., and Yao, J. K. (1990). Cholecystokinin, dopamine and schizophrenia. *Psychopharmacol. Bull.* **26**, 377–380.
- Garver, D. L., Bisette, G., Yao, J. K., and Nemeroff, C. B. (1991). Relation of CSF neurotensin concentrations to symptoms and drug response of psychotic patients. *Am. J. Psychiatry* **148**, 484–488.
- Garver, D. L., Tamas, R. L., and Holcomb, J. A. (2003). Elevated interleukin-6 in the cerebrospinal fluid of a previously delineated schizophrenia subtype. *Neuropsychopharmacology* **28**, 1515–1520.
- Gerner, R. H., and Sharp, B. (1982). CSF beta-endorphin-immunoreactivity in normal, schizophrenic, depressed, manic and anorexic subjects. *Brain Res.* **237**, 244–247.
- Gerner, R. H., van Kammen, D. P., and Ninan, P. T. (1985). Cerebrospinal fluid cholecystokinin, bombesin and somatostatin in schizophrenia and normals. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **9**, 73–82.
- Gerner, R. H., and Yamada, T. (1982). Altered neuropeptide concentrations in cerebrospinal fluid of psychiatric patients. *Brain Res.* **238**, 298–302.
- Geyer, M. A., Krebs-Thomson, K., Braff, D. L., and Swerdlow, N. R. (2001). Pharmacological studies of prepulse inhibition models of sensorimotor gating deficits in schizophrenia: A decade in review. *Psychopharmacology* **156**, 117–154.

- Gil-Ad, I., Dickerman, Z., Weizman, R., Weizman, A., Tyano, S., and Laron, Z. (1981). Abnormal growth hormone response to LRH and TRH in adolescent schizophrenic boys. *Am. J. Psychiatry* **138**, 357–360.
- Gispén-de Wied, C. C. (2000). Stress in schizophrenia: An integrative view. *Eur. J. Pharmacol.* **405**, 375–384.
- Gitlin, M. J., Gerner, R. H., and Rosenblatt, M. (1981). Assessment of naltrexone in the treatment of schizophrenia. *Psychopharmacology* **74**, 51–53.
- Gjerris, A., Rafaelsen, O. J., Vendsborg, P., Fahrenkrug, J., and Rehfeld, J. F. (1984). Vasoactive intestinal polypeptide decreased in cerebrospinal fluid (CSF) in atypical depression. Vasoactive intestinal polypeptide, cholecystokinin and gastrin in CSF in psychiatric disorders. *J. Affect. Disord.* **7**, 325–337.
- Gjerris, A., Hammer, M., Vendsborg, P., Christensen, N. J., and Rafaelsen, O. J. (1985). Cerebrospinal fluid vasopressin—changes in depression. *Br. J. Psychiatry* **147**, 696–701.
- Glovinsky, D., Kalogeras, K. T., Kirch, D. G., Suddath, R., and Wyatt, R. J. (1994). Cerebrospinal fluid oxytocin concentration in schizophrenic patients does not differ from control subjects and is not changed by neuroleptic medication. *Schizophr. Res.* **11**, 273–276.
- Goldman, M. B., Robertson, G. L., and Hedeker, D. (1996). Oropharyngeal regulation of water balance in polydipsic schizophrenics. *Clin. Endocrinol. (Oxf.)* **44**, 31–37.
- Goldman, M. B., Robertson, G. L., Luchins, D. J., Hedeker, D., and Pandey, G. N. (1997). Psychotic exacerbations and enhanced vasopressin secretion in schizophrenic patients with hyponatremia and polydipsia. *Arch. Gen. Psychiatry* **54**, 443–449.
- Goulet, M., Shiromani, P. J., Ware, C. M., Strong, R. A., Boismenu, R., and Rusche, J. R. (2003). A secretin i.v. infusion activates gene expression in the central amygdala of rats. *Neuroscience* **118**, 881–888.
- Green, E. K., Raybould, R., Macgregor, S., Gordon-Smith, K., Heron, J., Hyde, S., Grozeva, D., Hamshere, M., Williams, N., Owen, M. J., O'Donovan, M. C., Jones, L., *et al.* (2005). Operation of the schizophrenia susceptibility gene, neuregulin 1, across traditional diagnostic boundaries to increase risk for bipolar disorder. *Arch. Gen. Psychiatry* **62**, 642–648.
- Gruen, R., and Baron, M. (1984). Stressful life events and schizophrenia: Relation to illness onset and family history. *Neuropsychobiology* **12**, 206–208.
- Gunne, L. M., Lindström, L., and Terenius, L. (1977). Naloxone-induced reversal of schizophrenic hallucinations. *J. Neural. Transm.* **40**, 13–19.
- Hakak, Y., Walker, J. R., Li, C., Wong, W. H., Davis, K. L., Buxbaum, J. D., Haroutunian, V., and Fienberg, A. A. (2001). Genome-wide expression analysis reveals dysregulation of myelination-related genes in chronic schizophrenia. *Proc. Natl. Acad. Sci. USA* **98**, 4746–4751.
- Hamid, E. H., Hyde, T. M., Egan, M. F., Wolf, S. S., Herman, M. M., Nemeroff, C. B., and Kleinman, J. E. (2002). Neurotensin receptor binding abnormalities in the entorhinal cortex in schizophrenia and affective disorders. *Biol. Psychiatry* **51**, 795–800.
- Harmar, A. J., Arimura, A., Gozes, I., Journot, L., Laburthe, M., Pisegna, J. R., Rawlings, S. R., Robberecht, P., Said, S. I., Sreedharan, S. P., Wank, S. A., and Waschek, J. A. (1998). International Union of Pharmacology. XVIII. Nomenclature of receptors for vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide. *Pharmacol. Rev.* **50**, 265–270.
- Harrington, K. A., Augood, S. J., Faull, R. L., McKenna, P. J., and Emson, P. C. (1995). Dopamine D1 receptor, D2 receptor, proenkephalin A and substance P gene expression in the caudate nucleus of control and schizophrenic tissue: A quantitative cellular *in situ* hybridisation study. *Mol. Brain Res.* **33**, 333–342.
- Hashimoto, R., Straub, R. E., Weickert, C. S., Hyde, T. M., Kleinman, J. E., and Weinberger, D. R. (2004). Expression analysis of neuregulin-1 in the dorsolateral prefrontal cortex in schizophrenia. *Mol. Psychiatry* **9**, 299–307.

- Hattori, E., Ebihara, M., Yamada, K., Ohba, H., Shibuya, H., and Yoshikawa, T. (2001a). Identification of a compound short tandem repeat stretch in the 5'-upstream region of the cholecystokinin gene, and its association with panic disorder but not with schizophrenia. *Mol. Psychiatry* **6**, 465–470.
- Hattori, E., Yamada, K., Toyota, T., Yoshitsugu, K., Toru, M., Shibuya, H., and Yoshikawa, T. (2001b). Association studies of the CT repeat polymorphism in the 5' upstream region of the cholecystokinin B receptor gene with panic disorder and schizophrenia in Japanese subjects. *Am. J. Med. Genet.* **105**, 779–782.
- Heikkilä, L. (1993). Somatostatin in the cerebrospinal fluid of schizophrenic patients before and after neuroleptic drug treatment. *Schizophr. Res.* **8**, 273–277.
- Heikkilä, L., Rimón, R., and Terenius, L. (1990). Dynorphin A and Substance P in the cerebrospinal fluid of schizophrenic patients. *Psychiatry Res.* **34**, 229–236.
- Hicks, P. B., Vinogradov, S., Riney, S. J., Su, K., and Csernansky, J. G. (1989). A preliminary dose-ranging trial of proglumide for the treatment of refractory schizophrenics. *J. Clin. Psychopharmacol.* **9**, 209–212.
- Hökfelt, T., Morino, P., Verge, V., Castel, M. N., Broberger, C., Zhang, X., Herrera-Marschitz, M., Meana, J. J., Ungerstedt, U., and Xu, X. J. (1994). CCK in cerebral cortex and at the spinal level. *Ann. N. Y. Acad. Sci.* **713**, 157–163.
- Holsboer, F. (2003). The role of peptides in treatment of psychiatric disorders. *J. Neural Transm. Suppl.* **110**, 17–34.
- Hommer, D. W., Pickar, D., Roy, A., Ninan, P., Boronow, J., and Paul, S. M. (1984). The effects of ceruletide in schizophrenia. *Arch. Gen. Psychiatry* **41**, 617–619.
- Hong, C. J., Huo, S. J., Liao, D. L., Lee, K., Wu, J. Y., and Tsai, S. J. (2004). Case-control and family-based association studies between the neuregulin 1 (Arg38Gln) polymorphism and schizophrenia. *Neurosci. Lett.* **366**, 158–161.
- Howes, O. D., McDonald, C., Cannon, M., Arseneault, L., Boydell, J., and Murray, R. M. (2004). Pathways to schizophrenia: The impact of environmental factors. *Int. J. Neuropsychopharmacol.* **7**(Suppl. 1), S7–S13.
- Huezo-Diaz, P., Arranz, M. J., Munro, J., Osborne, S., Makoff, A., Kerwin, R. W., Austin, J., and O'Donovan, M. (2004). An association study of the neurotensin receptor gene with schizophrenia and clozapine response. *Schizophr. Res.* **66**, 193–195.
- Huppi, K., Siwarski, D., Pisegna, J. R., and Wank, S. (1995). Chromosomal localization of the gastric and brain receptors for cholecystokinin (CCKAR and CCKBR) in human and mouse. *Genomics* **25**, 727–729.
- Hurd, Y. L. (2002). Subjects with major depression or bipolar disorder show reduction of prodynorphin mRNA expression in discrete nuclei of the amygdaloid complex. *Mol. Psychiatry* **7**, 75–81.
- Iadarola, M. J., Ofri, D., and Kleinman, J. E. (1991). Enkephalin, dynorphin and substance P in postmortem substantia nigra from normals and schizophrenic patients. *Life Sci.* **48**, 1919–1930.
- Ikeda, K., Iritani, S., Ueno, H., and Niizato, K. (2004). Distribution of neuropeptide Y interneurons in the dorsal prefrontal cortex of schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **28**, 379–383.
- Inanaga, K., Nakano, T., Nagata, T., Tanaka, M., and Ogawa, N. (1978). Behavioral effects of protirelin in schizophrenia. *Arch. Gen. Psychiatry* **35**, 1011–1014.
- Innis, R. B., Bunney, B. S., Charney, D. S., Price, L. H., Glazer, W. M., Sternberg, D. E., Rubin, A. L., and Heninger, G. R. (1986). Does the cholecystokinin antagonist proglumide possess antipsychotic activity? *Psychiatry Res.* **18**, 1–7.
- Inui, A. (2003). Neuropeptide gene polymorphisms and human behavioural disorders. *Nat. Rev. Drug Discov.* **2**, 986–998.

- Iritani, S., Kuroki, N., Niizato, K., and Ikeda, K. (2000). Morphological changes in neuropeptide Y-positive fiber in the hippocampal formation of schizophrenics. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **24**, 241–249.
- Itoh, H., Tanoue, S., Yagi, G., Tateyama, M., Kamisada, M., Fujii, Y., Takamiya, M., and Nakajima, S. (1982). Clinical study on the psychotropic effects of caerulein: An open clinical trial in chronic schizophrenic patients. *Keio J. Med.* **31**, 71–95.
- Itoh, H., Shimazono, Y., Kawakita, Y., Kudo, Y., Satoh, Y., and Takahashi, R. (1986). Clinical evaluation of ceruletide in schizophrenia: A multi-institutional cooperative double-blind controlled study. *Psychopharmacol. Bull.* **22**, 123–128.
- Itokawa, M., Arai, M., Kato, S., Ogata, Y., Furukawa, A., Haga, S., Ujike, H., Sora, I., Ikeda, K., and Yoshikawa, T. (2003). Association between a novel polymorphism in the promoter region of the neuropeptide Y gene and schizophrenia in humans. *Neurosci. Lett.* **347**, 202–204.
- Ivy, A. C., and Oldberg, E. (1928). A hormone mechanism for gall bladder contraction and evacuation. *Am. J. Physiol.* **66**, 196–202.
- Iwata, N., Suzuki, T., Ikeda, M., Kitajima, T., Yamanouchi, Y., Inada, T., and Ozaki, N. (2004). No association with the neuregulin 1 haplotype to Japanese schizophrenia. *Mol. Psychiatry* **9**, 126–127.
- Jorgensen, L. S., Bach, F. W., Christiansen, P., Raundahl, U., Ostgaard, S., and Ekman, R. (1993). Decreased cerebrospinal fluid beta-endorphin and increased pain sensitivity in patients with functional abdominal pain. *Scand. J. Gastroenterol.* **28**, 763–766.
- Kaiya, H., Tamura, Y., Adachi, S., Moriuchi, I., Namba, M., Tanaka, M., Yoshida, H., Yanaihara, N., and Yanaihara, C. (1981). Substance P-like immunoreactivity in plasma of psychotic patients and effects of neuroleptics and electroconvulsive therapy. *Psychiatry Res.* **5**, 11–21.
- Kalivas, P. W., Nemeroff, C. B., and Prange, A. J., Jr. (1981). Increase in spontaneous motor activity following infusion of neurotensin into the ventral tegmental area. *Brain Res.* **229**, 525–529.
- Kalivas, P. W., Nemeroff, C. B., and Prange, A. J., Jr. (1984). Neurotensin microinjection into the nucleus accumbens antagonizes dopamine-induced increase in locomotion and rearing. *Neuroscience* **11**, 919–930.
- Kampman, O., Anttila, S., Illi, A., Saarela, M., Rontu, R., Mattila, K. M., Leinonen, E., and Lehtimäki, T. (2004). Neuregulin genotype and medication response in Finnish patients with schizophrenia. *Neuroreport* **15**, 2517–2520.
- Karelson, E., Laasik, J., and Sillard, R. (1995). Regulation of adenylate cyclase by galanin, neuropeptide Y, secretin and vasoactive intestinal polypeptide in rat frontal cortex, hippocampus and hypothalamus. *Neuropeptides* **28**, 21–28.
- Katila, H., Hurme, M., Wahlbeck, K., Appelberg, B., and Rimön, R. (1994). Plasma and cerebrospinal fluid interleukin-1 beta and interleukin-6 in hospitalized schizophrenic patients. *Neuropsychobiology* **30**, 20–23.
- Kerwin, R., Robinson, P., and Stephenson, J. (1992). Distribution of CCK binding sites in the human hippocampal formation and their alteration in schizophrenia: A post-mortem autoradiographic study. *Psychol. Med.* **22**, 37–43.
- Kim, C. E., Kang, D. Y., Ha, K. S., Jeong, S. H., and Kim, Y. S. (1995). Haloperidol does not affect the level of serum-soluble interleukin-2 receptor in drug-free male schizophrenics. *Biol Psychiatry* **38**, 843–845.
- Kinkead, B., and Nemeroff, C. B. (2004). Neurotensin, schizophrenia and antipsychotic drug actions. In “Disorders of Synaptic Plasticity” (J. Smythies, Ed.), Vol. 59, pp. 328–342. Elsevier, London.
- Kishimoto, T., Hirai, M., Ohsawa, H., Terada, M., Matsuoka, I., and Ikawa, G. (1989). Manners of arginine vasopressin secretion in schizophrenic patients—with reference to the mechanism of water intoxication. *Jpn. J. Psychiatry Neurol.* **43**, 161–169.
- Kleinman, J. E., Hong, J., Iadarola, M., Govoni, S., and Gillin, C. J. (1985). Neuropeptides in human brain—postmortem studies. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **9**, 91–95.

- Kobayashi, K., Nakaoka, K., Tsuji, H., and Shohmori, T. (1980). Effects of thyrotropin-releasing hormone in chronic schizophrenic patients. *Acta Med. Okayama* **34**, 263–273.
- Koponen, H., Rantakallio, P., Veijola, J., Jones, P., Jokelainen, J., and Isohanni, M. (2004). Childhood central nervous system infections and risk for schizophrenia. *Eur. Arch. Psychiatry Clin. Neurosci.* **254**, 9–13.
- Korsgaard, S., Casey, D. E., and Gerlach, J. (1982). High-dose destyrosine-gamma-endorphin in tardive dyskinesia. *Psychopharmacology* **78**, 285–286.
- Kurland, A. A., McCabe, O. L., Hanlon, T. E., and Sullivan, D. (1977). The treatment of perceptual disturbances in schizophrenia with naloxone hydrochloride. *Am. J. Psychiatry* **134**, 1408–1410.
- Kuromitsu, J., Yokoi, A., Kawai, T., Nagasu, T., Aizawa, T., Haga, S., and Ikeda, K. (2001). Reduced neuropeptide Y mRNA levels in the frontal cortex of people with schizophrenia and bipolar disorder. *Brain Res. Gene Expr. Patterns* **1**, 17–21.
- Lachman, H. M., Pedrosa, E., Nolan, K. A., Glass, M., Ye, K., and Saito, T. (2006). Analysis of polymorphisms in AT-rich domains of neuregulin 1 gene in schizophrenia. *Am. J. Med. Gen. B, Neuropsychiatr. Genet.* **141**, 102–109.
- Lahti, R. A., Cochrane, E. V., Roberts, R. C., Conley, R. R., and Tamminga, C. A. (1998). [³H] Neurotensin receptor densities in human postmortem brain tissue obtained from normal and schizophrenic persons: An autoradiographic study. *J. Neural. Transm.* **105**, 507–516.
- Langer, G., Koinig, G., Hatzinger, R., Schonbeck, G., Resch, F., Aschauer, H., Keshavan, M. S., and Sieghart, W. (1986). Response of thyrotropin to thyrotropin-releasing hormone as predictor of treatment outcome. Prediction of recovery and relapse in treatment with antidepressants and neuroleptics. *Arch. Gen. Psychiatry* **43**, 861–868.
- Lapchak, P. A. (1992). A role for interleukin-2 in the regulation of striatal dopaminergic function. *Neuroreport* **3**, 165–168.
- Lapchak, P. A., Araujo, D. M., Quirion, R., and Beaudet, A. (1991). Immunoautoradiographic localization of interleukin 2-like immunoreactivity and interleukin 2 receptors (Tac antigen-like immunoreactivity) in the rat brain. *Neuroscience* **44**, 173–184.
- Lauerma, H. (1999). Neuroimmune appendicitis, peptides, and schizophrenia. *Lancet* **354**, 1648.
- Le, F., Groshan, K., Zeng, X. P., and Richelson, E. (1997a). Characterization of the genomic structure, promoter region, and a tetranucleotide repeat polymorphism of the human neurotensin receptor gene. *J. Biol. Chem.* **272**, 1315–1322.
- Le, F., Zeng, X. P., and Richelson, E. (1997b). Tetranucleotide repeat polymorphism in the human neurotensin receptor (NT-R) gene. *Clin. Genet.* **51**, 76–77.
- Leff, J., and Vaughn, C. (1980). The interaction of life events and relatives' expressed emotion in schizophrenia and depressive neurosis. *Br. J. Psychiatry* **136**, 146–153.
- Li, T., Stefansson, H., Gudfinnsson, E., Cai, G., Liu, X., Murray, R. M., Steinthorsdottir, V., Januel, D., Gudnadottir, V. G., Petursson, H., Ingason, A., Gulcher, J. R., *et al.* (2004). Identification of a novel neuregulin 1 at-risk haplotype in Han schizophrenia Chinese patients, but no association with the Icelandic/Scottish risk haplotype. *Mol. Psychiatry* **9**, 698–704.
- Licinio, J., Seibyl, J. P., Altemus, M., Charney, D. S., and Krystal, J. H. (1993). Elevated CSF levels of interleukin-2 in neuroleptic-free schizophrenic patients. *Am. J. Psychiatry* **150**, 1408–1410.
- Lieberman, J. A., and Koreen, A. R. (1993). Neurochemistry and Neuroendocrinology of schizophrenia: A selective review. *Schizophr. Bull.* **19**, 371–429.
- Lindberg, C., Koefoed, P., Hansen, E. S., Bolwig, T. G., Rehfeld, J. F., Møllerup, E., Jørgensen, O. S., Kessing, L. V., Werge, T., Haugbol, S., Wang, A. G., and Woldbye, D. P. (2006). No association between the -399 C>T polymorphism of the neuropeptide Y gene and schizophrenia, unipolar depression or panic disorder in a Danish population. *Acta Psychiatr. Scand.* **113**, 54–58.

- Lindström, L. H. (1996). Clinical and biological markers for outcome in schizophrenia: A review of a longitudinal follow-up study in Uppsala schizophrenia research project. *Neuropsychopharmacology* **14**, 23S–26S.
- Lindström, L. H., Gunne, L. M., Ost, L. G., and Persson, E. (1977). Thyrotropin-releasing hormone (TRH) in chronic schizophrenia. A controlled study. *Acta Psychiatr. Scand.* **55**, 74–80.
- Lindström, L. H., Widerlöv, E., Gunne, L. M., Wahlström, A., and Terenius, L. (1978). Endorphins in human cerebrospinal fluid: Clinical correlations to some psychotic states. *Acta Psychiatr. Scand.* **57**, 153–164.
- Lindström, L. H., Besev, G., Gunne, L. M., and Terenius, L. (1986). CSF levels of receptor-active endorphins in schizophrenic patients: Correlations with symptomatology and monoamine metabolites. *Psychiatry Res.* **19**, 93–100.
- Lindström, L. H., Widerlöv, E., Bissette, G., and Nemeroff, C. B. (1988). Reduced CSF neurotensin concentration in drug-free schizophrenic patients. *Schizophr. Res.* **1**, 55–59.
- Lindström, L. H., Terenius, L., and Nyberg, F. (1992). Opioid peptides in psychiatric disorders. *Clin. Neuropharmacol.* **15**(Suppl. 1, Pt. A), 58A–59A.
- Lipinski, J., Meyer, R., Kornetsky, C., and Cohen, B. M. (1979). Naloxone in schizophrenia: Negative result. *Lancet* **1**, 1292–1293.
- Liu, C. M., Hwu, H. G., Fann, C. S., Lin, C. Y., Liu, Y. L., Ou-Yang, W. C., and Lee, S. F. (2005). Linkage evidence of schizophrenia to loci near neuregulin 1 gene on chromosome 8p21 in Taiwanese families. *Am. J. Med. Gen. B, Neuropsychiatr. Genet.* **134**, 79–83.
- Loosen, P. T. (1985). The TRH-induced TSH response in psychiatric patients: A possible neuroendocrine marker. *Psychoneuroendocrinology* **10**, 237–260.
- Lotstra, F., Verbanck, P., Mendlewicz, J., and Vanderhaeghen, J. J. (1984). No evidence of anti-psychotic effect of caerulein in schizophrenic patients free of neuroleptics: A double-blind cross-over study. *Biol. Psychiatry* **19**, 877–882.
- Lotstra, F., Verbanck, P. M., Gilles, C., Mendlewicz, J., and Vanderhaeghen, J. J. (1985). Reduced cholecystokinin levels in cerebrospinal fluid of parkinsonian and schizophrenic patients: Effect of ceruletide in schizophrenia. *Ann. N. Y. Acad. Sci.* **448**, 507–517.
- Lu, W. T., Zhang, X., Zhang, M., Gong, S. L., and Wei, J. (2004). Association analysis of the cholecystokinin type A receptor gene in schizophrenia. *Chin. Med. J.* **117**, 627–629.
- Lukoff, D., Snyder, K., Ventura, J., and Nuechterlein, K. H. (1984). Life events, familial stress, and coping in the developmental course of schizophrenia. *Schizophr. Bull.* **10**, 258–292.
- Lund, T., Geurts van Kessel, A. H., Haun, S., and Dixon, J. E. (1986). The genes for human gastrin and cholecystokinin are located on different chromosomes. *Hum. Genet.* **73**, 77–80.
- Manberg, P. J., Nemeroff, C. B., Iversen, L. L., Rosser, M. N., Kizer, J. S., and Prange, A. J., Jr. (1982). Human brain distribution of neurotensin in normals, schizophrenics, and Huntington's choreics. *Ann. N. Y. Acad. Sci.* **400**, 354–367.
- Manberg, P. J., Nemeroff, C. B., Bissette, G., Widerlöv, E., Youngblood, W. W., Kizer, J. S., and Prange, A. J., Jr. (1985). Neuropeptides in CSF and post-mortem brain tissue of normal controls, schizophrenics and Huntington's choreics. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **9**, 97–108.
- Manchanda, R., and Hirsch, S. R. (1981). (Des-Tyr)-gamma-endorphin in the treatment of schizophrenia. *Psychol. Med.* **11**, 401–404.
- Marchesi, G. F., Santone, G., Cotani, P., Giordano, A., and Chelli, F. (1992). Naltrexone in chronic negative schizophrenia. *Clin. Neuropharmacol.* **15**(Suppl. 1, Pt. A), 56A–57A.
- Marchesi, G. F., Santone, G., Cotani, P., Giordano, A., and Chelli, F. (1995). The therapeutic role of naltrexone in negative symptom schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **19**, 1239–1249.
- Marco, N., Thirion, A., Mons, G., Bougault, I., Le Fur, G., Soubrie, P., and Steinberg, R. (1998). Activation of dopaminergic and cholinergic neurotransmission by tachykinin NK3 receptor stimulation: An *in vivo* microdialysis approach in guinea pig. *Neuropeptides* **32**, 481–488.

- Marondel, I., Renault, B., Lieman, J., Ward, D., and Kucherlapati, R. (1996). Physical mapping of the human neurotensin gene (NTS) between markers D12S1444 and D12S81 on chromosome 12q21. *Genomics* **38**, 243–245.
- Mattes, J. A., Hom, W., Rochford, J. M., and Orlosky, M. (1985). Ceruletide for schizophrenia: A double-blind study. *Biol. Psychiatry* **20**, 533–538.
- Mauri, M. C., Rudelli, R., Vanni, S., Panza, G., Sicaro, A., Audisio, D., Sacerdote, P., and Panerai, A. E. (1998). Cholecystokinin, beta-endorphin and vasoactive intestinal peptide in peripheral blood mononuclear cells of drug-naïve schizophrenic patients treated with haloperidol compared to healthy controls. *Psychiatry Res.* **78**, 45–50.
- McAllister, C. G., van Kammen, D. P., Rehn, T. J., Miller, A. L., Gurklis, J., Kelley, M. E., Yao, J., and Peters, J. L. (1995). Increases in CSF levels of interleukin-2 in schizophrenia: Effects of recurrence of psychosis and medication status. *Am. J. Psychiatry* **152**, 1291–1297.
- McGauley, G. A., Aldridge, C. R., Fahy, T. A., and Eastment, C. (1989). The dexamethasone suppression test and negative symptoms of schizophrenia. *Acta Psychiatr. Scand.* **80**, 548–553.
- Meerabux, J., Iwayama, Y., Sakurai, T., Ohba, H., Toyota, T., Yamada, K., Nagata, R., Irukayama-Tomobe, Y., Shimizu, H., Yoshitsugu, K., Ohta, K., and Yoshikawa, T. (2005). Association of an orexin 1 receptor 408Val variant with polydipsia-hyponatremia in schizophrenic subjects. *Biol. Psychiatry* **58**, 401–407.
- Meltzer, H. Y., Arvanitis, L., Bauer, D., Rein, W., and Meta-Trial Study, W. (2004). Placebo-controlled evaluation of four novel compounds for the treatment of schizophrenia and schizoaffective disorder. *Am. J. Psychiatry* **161**, 975–984.
- Meltzer, H. Y., Busch, D. A., Tricou, B. J., and Robertson, A. (1982). Effect of (Des-Tyr)-gamma-endorphin in schizophrenia. *Psychiatry Res.* **6**, 313–326.
- Meltzer, H. Y., Lee, M. A., and Jayatilake, K. (2001). The blunted plasma cortisol response to apomorphine and its relationship to treatment response in patients with schizophrenia. *Neuropsychopharmacology* **24**, 278–290.
- Mielke, D. H., and Gallant, D. M. (1977). An oral opiate antagonist in chronic schizophrenia: A pilot study. *Am. J. Psychiatry* **134**, 1430–1431.
- Mikesell, M. J., Sobell, J. L., Sommer, S. S., and McMurray, C. T. (1996). Identification of a missense mutation and several polymorphisms in the proenkephalin A gene of schizophrenic patients. *Am. J. Med. Genet.* **67**, 459–467.
- Mikesell, M. J., Barron, Y. D., Nimgaonkar, V. L., Sobell, J. L., Sommer, S. S., and McMurray, C. T. (1997). Gly(247)– > Asp proenkephalin A mutation is rare in schizophrenia populations. *Am. J. Med. Genet.* **74**, 213–215.
- Miller, C., Kirchmair, R., Troger, J., Saria, A., Fleischhacker, W. W., Fischer-Colbrie, R., Benzer, A., and Winkler, H. (1996). CSF of neuroleptic-naïve first-episode schizophrenic patients: Levels of biogenic amines, substance P, and peptides derived from chromogranin A (GE-25) and secretogranin II (secretoneurin). *Biol. Psychiatry* **39**, 911–918.
- Miyasaka, K., and Funakoshi, A. (2003). Cholecystokinin and cholecystokinin receptors. *J. Gastroenterol.* **38**, 1–13.
- Mizuki, Y., Nishikori, S., Kajimura, N., Imaizumi, J., Yamada, M., and Inanaga, K. (1985). A treatment trial with an analog of thyrotropin-releasing hormone (DN-1417) in schizophrenia. *Biol. Psychiatry* **20**, 1030–1035.
- Mizuki, Y., Ushijima, I., Yamada, M., Tanaka, M., and Inanaga, K. (1986). A treatment trial with an analog of thyrotropin-releasing hormone (DN-1417) and des-tyrosine-gamma-endorphin in schizophrenia. *Int. Clin. Psychopharmacol.* **1**, 303–313.
- Montgomery, S. A., and Green, M. C. (1988). The use of cholecystokinin in schizophrenia: A review. *Psychol. Med.* **18**, 593–603.
- Montgomery, S. A., Green, M., Rimón, R., Heikkilä, L., Forsström, R., Hirsch, S. R., Hallström, C., Hippus, H., Naber, R., and Khan, M. C. (1992). Inadequate treatment response to

- des-enkephalin-gamma-endorphin compared with thioridazine and placebo in schizophrenia. *Acta Psychiatr. Scand.* **86**, 97–103.
- Moroji, T., Watanabe, N., Aoki, N., and Itoh, S. (1982). Antipsychotic effects of caerulein, a decapeptide chemically related to cholecystokinin octapeptide, on schizophrenia. *Int. Pharmacopsychiatry* **17**, 255–273.
- Muller, N., Empl, M., Riedel, M., Schwarz, M., and Ackenheil, M. (1997). Neuroleptic treatment increases soluble IL-2 receptors and decreases soluble IL-6 receptors in schizophrenia. *Eur. Arch. Psychiatry Clin. Neurosci.* **247**, 308–313.
- Muller, N., Riedel, M., Gruber, R., Ackenheil, M., and Schwarz, M. J. (2000). The immune system and schizophrenia. An integrative view. *Ann. N. Y. Acad. Sci.* **917**, 456–467.
- Muller, N., Ulmschneider, M., Scheppach, C., Schwarz, M. J., Ackenheil, M., Moller, H. J., Gruber, R., and Riedel, M. (2004). COX-2 inhibition as a treatment approach in schizophrenia: Immunological considerations and clinical effects of celecoxib add-on therapy. *Eur. Arch. Psychiatry Clin. Neurosci.* **254**, 14–22.
- Myers, K. M., Goulet, M., Rusche, J., Boismenu, R., and Davis, M. (2005). Partial reversal of phencyclidine-induced impairment of prepulse inhibition by secretin. *Biol. Psychiatry* **58**, 67–73.
- Naber, D., Ackenheil, M., Laakman, G., Fischer, H., and von Werder, K. (1980). Basal and stimulated levels of prolactin, TSH and LH in serum of chronic schizophrenic patients, long-term treated with neuroleptics. *Pharmacopsychiatr. Neuropsychopharmacol.* **13**, 325–330.
- Naber, D., Pickar, D., Post, R. M., Van Kammen, D. P., Waters, R. N., Ballenger, J. C., Goodwin, F. K., and Bunney, W. E., Jr. (1981). Endogenous opioid activity and beta-endorphin immunoreactivity in CSF of psychiatric patients and normal volunteers. *Am. J. Psychiatry* **138**, 1457–1462.
- Naber, D., and Leibl, K. (1983). Repeated high dosage naloxone treatment without therapeutic efficacy in schizophrenic patients. *Pharmacopsychiatria* **16**, 43–45.
- Naber, D., Munch, U., Wissmann, J., Grosse, R., Ritt, R., and Welter, D. (1983). Naloxone treatment for five days ineffective in schizophrenia: Neuroendocrine actions of the opiate antagonist. *Acta Psychiatr. Scand.* **67**, 265–271.
- Nair, N. P., Bloom, D. M., and Nestoros, J. N. (1982). Cholecystokinin appears to have antipsychotic properties. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **6**, 509–512.
- Nair, N. P., Bloom, D., Nestoros, J. N., and Schwartz, G. (1983). Therapeutic efficacy of cholecystokinin in neuroleptic-resistant schizophrenic subjects. *Psychopharmacol. Bull.* **19**, 134–136.
- Nair, N. P., Bloom, D. M., Debonnel, G., Schwartz, G., and Mosticyn, S. (1984). Cholecystokinin-octapeptide in chronic schizophrenia: A double-blind placebo-controlled study. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **8**, 711–714.
- Nair, N. P., Bloom, D., Lal, S., Debonnel, G., and Schwartz, G. (1985). Clinical and neuroendocrine studies with cholecystokinin peptides. *Ann. N. Y. Acad. Sci.* **448**, 535–541.
- Nair, N. P., Lal, S., and Bloom, D. M. (1986). Cholecystokinin and schizophrenia. *Prog. Brain Res.* **65**, 237–258.
- Nakao, K., Oki, S., Tanaka, I., Horil, K., Nakai, Y., Furui, T., Fukushima, M., Kuwayama, A., Kageyama, N., and Imura, H. (1980). Immunoreactive beta-endorphin and adrenocorticotropin in human cerebrospinal fluid. *J. Clin. Invest.* **66**, 1383–1690.
- Nalivaiko, E., Michaud, J. C., Soubrie, P., Le Fur, G., and Feltz, P. (1997). Tachykinin neurokinin-1 and neurokinin-3 receptor-mediated responses in guinea-pig substantia nigra: An *in vitro* electrophysiological study. *Neuroscience* **78**, 745–757.
- Nemeroff, C. B. (1980). Neurotensin: Perchance an endogenous neuroleptic? *Biol. Psychiatry* **15**, 283–302.
- Nemeroff, C. B., Youngblood, W. W., Manberg, P. J., Prange, A. J., Jr., and Kizer, J. S. (1983). Regional brain concentrations of neuropeptides in Huntington's chorea and schizophrenia. *Science* **221**, 972–975.

- Nemeroff, C. B., Widerlöv, E., Bissette, G., Walleus, H., Karlsson, I., Eklund, K., Kilts, C. D., Loosen, P. T., and Vale, W. (1984). Elevated concentrations of CSF corticotropin-releasing factor-like immunoreactivity in depressed patients. *Science* **226**, 1342–1344.
- Nemeroff, C. B., Bissette, G., Widerlöv, E., Beckmann, H., Gerner, R., Manberg, P. J., Lindström, L., Prange, A. J., Jr., and Gattaz, W. F. (1989). Neurotensin-like immunoreactivity in cerebrospinal fluid of patients with schizophrenia, depression, anorexia nervosa-bulimia, and premenstrual syndrome. *J. Neuropsychiatry Clin. Neurosci.* **1**, 16–20.
- Nemeroff, C. B., and Vale, W. W. (2005). The neurobiology of depression: Inroads to treatment and new drug discovery. *J. Clin. Psychiatry* **66**(Suppl. 7), 5–13.
- Nishimori, K., Young, L. J., Guo, Q., Wang, Z., Insel, T. R., and Matzuk, M. M. (1996). Oxytocin is required for nursing but is not essential for parturition or reproductive behavior. *Proc. Natl. Acad. Sci. USA* **93**, 11699–11704.
- Nishino, S., Mignot, E., Benson, K. L., and Zarcone, V. P., Jr. (1998). Cerebrospinal fluid prostaglandins and corticotropin releasing factor in schizophrenics and controls: Relationship to sleep architecture. *Psychiatry Res.* **78**, 141–150.
- Nishino, S., Ripley, B., Mignot, E., Benson, K. L., and Zarcone, V. P. (2002). CSF hypocretin-1 levels in schizophrenics and controls: Relationship to sleep architecture. *Psychiatry Res.* **110**, 1–7.
- Norman, R. M., Malla, A. K., McLean, T. S., McIntosh, E. M., Neufeld, R. W., Voruganti, L. P., and Cortese, L. (2002). An evaluation of a stress management program for individuals with schizophrenia. *Schizophr. Res.* **58**, 293–303.
- Norton, N., Moskvina, V., Morris, D. W., Bray, N. J., Zammit, S., Williams, N. M., Williams, H. J., Preece, A. C., Dwyer, S., Wilkinson, J. C., Spurlock, G., Kirov, G., *et al.* (2006). Evidence that interaction between neuregulin 1 and its receptor erbB4 increases susceptibility to schizophrenia. *Am. J. Med. Gen. B., Neuropsychiatr. Genet.* **141**, 96–101.
- Obuchowicz, Z., Krysiak, R., and Herman, Z. S. (2004). Does neuropeptide Y (NPY) mediate the effects of psychotropic drugs? *Neurosci. Biobehav. Rev.* **28**, 595–610.
- Ohsawa, H., Kishimoto, T., Shimayoshi, N., Matsumura, K., Tahara, K., Kitera, K., Higashiura, N., Noriyama, Y., Matsumoto, H., Hirai, M., *et al.* (1993). Atrial natriuretic peptide and arginine vasopressin secretion in schizophrenic patients. *Acta Psychiatr. Scand.* **88**, 130–134.
- Owen, M. J., Craddock, N., and O'Donovan, M. C. (2005). Schizophrenia: Genes at last? [Review] [83 refs]. *Trends Genet.* **21**, 518–525.
- Page, N. M. (2005). New challenges in the study of the mammalian tachykinins. *Peptides* **26**, 135613–135668.
- Palacios, J. M., Chinaglia, G., Rigo, M., Ulrich, J., and Probst, A. (1991). Neurotensin receptor binding levels in basal ganglia are not altered in Huntington's chorea or schizophrenia. *Synapse* **7**, 114–122.
- Panerai, A. E., and Sacerdote, P. (1993). Brain and gut neuropeptides in peripheral blood mononuclear cells. *J. Physiol. Paris* **87**, 283–289.
- Panteris, V., and Karamanolis, D. G. (2005). The puzzle of somatostatin: Action, receptors, analogues and therapy. *Hepatogastroenterology* **52**, 1771–1781.
- Panza, G., Monzani, E., Sacerdote, P., Penati, G., and Panerai, A. E. (1992). Beta-endorphin, vasoactive intestinal peptide and cholecystokinin in peripheral blood mononuclear cells from healthy subjects and from drug-free and haloperidol-treated schizophrenic patients. *Acta Psychiatr. Scand.* **85**, 207–210.
- Pearce, B. D. (2001). Schizophrenia and viral infection during neurodevelopment: A focus on mechanisms. *Mol. Psychiatry* **6**, 634–646.
- Peckys, D., and Hurd, Y. L. (2001). Prodynorphin and kappa opioid receptor mRNA expression in the cingulate and prefrontal cortices of subjects diagnosed with schizophrenia or affective disorders. *Brain Res. Bull.* **55**, 619–624.

- Perry, R. H., Dockray, G. J., Dimaline, R., Perry, E. K., Blessed, G., and Tomlinson, B. E. (1981). Neuropeptides in Alzheimer's disease, depression and schizophrenia: A post mortem analysis of vasoactive intestinal peptide and cholecystokinin in cerebral cortex. *J. Neurol. Sci.* **51**, 465–472.
- Peselow, E., Angrist, B., Sudilovsky, A., Corwin, J., Siekierski, J., Trent, F., and Rotrosen, J. (1987). Double blind controlled trials of cholecystokinin octapeptide in neuroleptic-refractory schizophrenia. *Psychopharmacology* **91**, 80–84.
- Peters, J., Van Kammen, D., Gelernter, J., Yao, J., and Shaw, D. (1990). Neuropeptide Y-like immunoreactivity in schizophrenia: Relationships with clinical measures. *Schizophr. Res.* **3**, 287–294.
- Petito, J. M., McCarthy, D. B., Rinker, C. M., Huang, Z., and Getty, T. (1997). Modulation of behavioral and neurochemical measures of forebrain dopamine function in mice by species-specific interleukin-2. *J. Neuroimmunol.* **73**, 183–190.
- Petryshen, T. L., Middleton, F. A., Kirby, A., Aldinger, K. A., Purcell, S., Tahl, A. R., Morley, C. P., McGann, L., Gentile, K. L., Rockwell, G. N., Medeiros, H. M., Carvalho, C., *et al.* (2005). Support for involvement of neuregulin 1 in schizophrenia pathophysiology. *Mol. Psychiatry* **10**, 366–374.
- Pickar, D., Naber, D., Post, R. M., van Kammen, D. P., Ballenger, J., Kalin, N., and Bunney, W. E., Jr. (1981). Measurement of endorphins in CSF. Relationship to psychiatric diagnosis. *Mod. Probl. Pharmacopsychiatry* **17**, 246–262.
- Pickar, D., Vartanian, F., Bunney, W. E., Jr., Maier, H. P., Gastpar, M. T., Prakash, R., Sethi, B. B., Lideman, R., Belyaev, B. S., Tsutsulkovskaja, M. V., Jungkunz, G., Nedopil, N., *et al.* (1982). Short-term naloxone administration in schizophrenic and manic patients. A World Health Organization Collaborative Study. *Arch. Gen. Psychiatry* **39**, 313–319.
- Pickar, D., Bunney, W. E., Jr., Douillet, P., Sethi, B. B., Sharma, M., Vartanian, M. E., Lideman, R. P., Naber, D., Leibl, K., Yamashita, I., Koyama, T., Verhoeven, W. M. A., *et al.* (1989). Repeated naloxone administration in schizophrenia: A phase II World Health Organization Study. *Biol. Psychiatry* **25**, 440–448.
- Prange, A. J., Jr., Loosen, P. T., Wilson, I. C., Meltzer, H. Y., and Fang, V. S. (1979). Behavioral and endocrine responses of schizophrenic patients to TRH (protirelin). *Arch. Gen. Psychiatry* **36**, 1086–1093.
- Prokai, L. (2002). Central nervous system effects of thyrotropin-releasing hormone and its analogues: Opportunities and perspectives for drug discovery and development. *Prog. Drug Res.* **59**, 133–169.
- Rafaelsen, O. J., and Gjerris, A. (1985). Neuropeptides in the cerebrospinal fluid (CSF) in psychiatric disorders. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **9**, 533–538.
- Ragnauth, A. K., Devidze, N., Moy, V., Finley, K., Goodwillie, A., Kow, L. M., Muglia, L. J., and Pfaff, D. W. (2005). Female oxytocin gene-knockout mice, in a semi-natural environment, display exaggerated aggressive behavior. *Genes Brain Behav.* **4**, 229–239.
- Raison, C. L., Capuron, L., and Miller, A. H. (2006). Cytokines sing the blues: Inflammation and the pathogenesis of depression. *Trends Immunol.* **27**, 24–31.
- Ramirez, M., Prieto, I., Vives, F., de Gasparo, M., and Alba, F. (2004). Neuropeptides, neuropeptidases and brain asymmetry. *Curr. Protein Pept. Sci.* **5**, 497–506.
- Rehfeld, J. F., and Kruse-Larsen, C. (1978). Gastrin and cholecystokinin in human cerebrospinal fluid. Immunochemical determination of concentrations and molecular heterogeneity. *Brain Res.* **155**, 19–26.
- Rimon, R., Terenius, L., and Kampman, R. (1980). Cerebrospinal fluid endorphins in schizophrenia. *Acta Psychiatr. Scand.* **61**, 395–403.
- Rimon, R., Le Greves, P., Nyberg, F., Heikkilä, L., Salmela, L., and Terenius, L. (1984). Elevation of substance P-like peptides in the CSF of psychiatric patients. *Biol. Psychiatry* **19**, 509–516.

- Risch, S. C., Lewine, R. J., Kalin, N. H., Jewart, R. D., Risby, E. D., Caudle, J. M., Stipetic, M., Turner, J., Eccard, M. B., and Pollard, W. E. (1992). Limbic-hypothalamic-pituitary-adrenal axis activity and ventricular-to-brain ratio studies in affective illness and schizophrenia. *Neuropsychopharmacology* **6**, 95–100.
- Roberts, G. W., Ferrier, I. N., Lee, Y., Crow, T. J., Johnstone, E. C., Owens, D. G., Bacarese-Hamilton, A. J., McGregor, G., O'Shaughnessey, D., Polak, J. M., and Bloom, S. R. (1983). Peptides, the limbic lobe and schizophrenia. *Brain Res.* **288**, 199–211.
- Rogaeva, E. A., Chumakova, M. M., Dergunova, N. N., Pozharitskaia, D. A., Kopeiko, G. I., Tsutsul'kovskaia, M., and Tsibezov, V. V. (1990). Anti-somatostatin autoantibodies in the blood serum of patients with schizophrenia. *Zh. Nevropatol. Psikiatr. Im S S Korsakova* **90**, 82–84.
- Rothermundt, M., Arolt, V., Weitzsch, C., Eckhoff, D., and Kirchner, H. (1998). Immunological dysfunction in schizophrenia: A systematic approach. *Neuropsychobiology* **37**, 186–193.
- Rothermundt, M., Arolt, V., Leadbeater, J., Peters, M., Rudolf, S., and Kirchner, H. (2000). Cytokine production in unmedicated and treated schizophrenic patients. *Neuroreport* **11**, 3385–3388.
- Roy, B. F., Benkelfat, C., Hill, J. L., Pierce, P. F., Dauphin, M. M., Kelly, T. M., Sunderland, T., Weinberger, D. R., and Breslin, N. (1994). Serum antibody for somatostatin-14 and prodynorphin 209–240 in patients with obsessive-compulsive disorder, schizophrenia, Alzheimer's disease, multiple sclerosis, and advanced HIV infection. *Biol. Psychiatry* **35**, 335–344.
- Rubinow, D. R. (1986). Cerebrospinal fluid somatostatin and psychiatric illness. *Biol. Psychiatry* **21**, 341–365.
- Ryan, M. C., Sharifi, N., Condren, R., and Thakore, J. H. (2004). Evidence of basal pituitary-adrenal overactivity in first episode, drug naive patients with schizophrenia. *Psychoneuroendocrinology* **29**, 1065–1070.
- Saiz-Ruiz, J., Carrasco, J. L., Martin, M., Manzanares, J., and Hernanz, A. (1992). Plasmatic somatostatin as a marker of positive symptoms of schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **16**, 203–210.
- Santucci, V., Gueudet, C., Steinberg, R., Le Fur, G., and Soubrié, P. (1997). Involvement of cortical neurotensin in the regulation of rat meso-cortico-limbic dopamine neurons: Evidence from changes in the number of spontaneously active A10 cells after neurotensin receptor blockade. *Synapse* **26**, 370–380.
- Schalling, M., Friberg, K., Seroogy, K., Riederer, P., Bird, E., Schiffmann, S. N., Mailleux, P., Vanderhaeghen, J. J., Kuga, S., and Goldstein, M. (1990). Analysis of expression of cholecystokinin in dopamine cells in the ventral mesencephalon of several species and in humans with schizophrenia. *PNAS* **87**, 8427–8431.
- Schwarz, M. J., Kronig, H., Riedel, M., Dehning, S., Douhet, A., Spellmann, I., Ackenheil, M., Moller, H. J., and Muller, N. (2006). IL-2 and IL-4 polymorphisms as candidate genes in schizophrenia. *Eur. Arch. Psychiatry Clin. Neurosci.* **256**, 72–76.
- Seabrook, G. R., Bowery, B. J., and Hill, R. G. (1995). Pharmacology of tachykinin receptors on neurones in the ventral tegmental area of rat brain slices. *Eur. J. Pharmacol.* **273**, 113–119.
- Sethi, B. B., and Prakash, R. (1981). A study of naloxone with schizophrenic and manic patients. *Br. J. Psychiatry* **138**, 501–503.
- Sharma, R. P., Bissette, G., Janicak, P., Davis, J. M., and Nemeroff, C. B. (1994). Cerebrospinal fluid somatostatin concentrations in schizophrenia and schizoaffective disorder: The effects of antipsychotic treatment. *Schizophr. Res.* **13**, 173–177.
- Sharma, R. P., Bissette, G., Janicak, P. G., Davis, J. M., and Nemeroff, C. B. (1995). Elevation of CSF somatostatin concentrations in mania. *Am. J. Psychiatry* **152**, 1807–1809.
- Sharma, R. P., Janicak, P. G., Bissette, G., and Nemeroff, C. B. (1997). CSF neurotensin concentrations and antipsychotic treatment in schizophrenia and schizoaffective disorders. *Am. J. Psychiatry* **154**, 1019–1021.

- Sharma, R. P., Martis, B., Rosen, C., Jonalagadda, J., Nemeroff, C. B., and Bissette, G. (2001). CSF thyrotropin-releasing hormone concentrations differ in patients with schizoaffective disorder from patients with schizophrenia or mood disorders. *J. Psychiatr. Res.* **35**, 287–291.
- Sharpe, A. L., Coste, S. C., Burkhart-Kasch, S., Li, N., Stenzel-Poore, M. P., and Phillips, T. J. (2005). Mice deficient in corticotropin-releasing factor receptor type 2 exhibit normal ethanol-associated behaviors. *Alcohol Clin. Exp. Res.* **29**, 1601–1609.
- Sheitman, B. B., Knable, M. B., Jarskog, L. F., Chakos, M., Boyce, L. H., Early, J., and Lieberman, J. A. (2004). Secretin for refractory schizophrenia. *Schizophr. Res.* **66**, 177–181.
- Silberberg, G., Darvasi, A., Pinkas-Kramarski, R., and Navon, R. (2006). The involvement of ErbB4 with schizophrenia: Association and expression studies. *Am. J. Med. Gen. B Neuropsychiatr. Genet.* **141**, 142–148.
- Sirota, P., Meiman, M., Herschko, R., and Bessler, H. (2005). Effect of neuroleptic administration on serum levels of soluble IL-2 receptor-alpha and IL-1 receptor antagonist in schizophrenic patients. *Psychiatry Res.* **134**, 151–159.
- Skeldon, K. H., Owens, M. J., and Nemeroff, C. B. (2000). The neurobiology of urocortin. *Reg. Peptides* **93**, 85–92.
- Skirboll, L. R., Grace, A. A., Hommer, D. W., Rehfeld, J., Goldstein, M., Hokfelt, T., and Bunney, B. S. (1981). Peptide-monoamine coexistence: Studies of the actions of cholecystokinin-like peptide on the electrical activity of midbrain dopamine neurons. *Neuroscience* **6**, 2111–2124.
- Smith, R. S. (1992). A comprehensive macrophage-T-lymphocyte theory of schizophrenia. *Med. Hypotheses* **39**, 248–257.
- Smith, R. S., and Maes, M. (1995). The macrophage-T-lymphocyte theory of schizophrenia: Additional evidence. *Med. Hypotheses* **45**, 135–141.
- Song, C., Lin, A., Kenis, G., Bosmans, E., and Maes, M. (2000). Immunosuppressive effects of clozapine and haloperidol: Enhanced production of the interleukin-1 receptor antagonist. *Schizophr. Res.* **42**, 157–164.
- Sorensen, P. S., Gjerris, A., and Hammer, M. (1985). Cerebrospinal fluid vasopressin in neurological and psychiatric disorders. *J. Neurol. Neurosurg. Psychiatry* **48**, 50–57.
- Spooren, W., Riemer, C., and Meltzer, H. (2005). Opinion: NK3 receptor antagonists: The next generation of antipsychotics? *Nat. Rev. Drug Discov.* **4**, 967–975.
- Stefansson, H., Sigurdsson, E., Steinthorsdottir, V., Bjornsdottir, S., Sigmundsson, T., Ghosh, S., Brynjolfsson, J., Gunnarsdottir, S., Ivarsson, O., Chou, T. T., Hjaltason, O., Birgisdottir, B., *et al.* (2002). Neuregulin 1 and susceptibility to schizophrenia. *Am. J. Hum. Genet.* **71**, 4.
- Stefansson, H., Sarginson, J., Kong, A., Yates, P., Steinthorsdottir, V., Gudfinnsson, E., Gunnarsdottir, S., Walker, N., Petursson, H., Crombie, C., Ingason, A., Gulcher, A., *et al.* (2003). Association of neuregulin 1 with schizophrenia confirmed in a Scottish population. *Am. J. Hum. Genet.* **72**, 83–87.
- Tachikawa, H., Harada, S., Kawanishi, Y., Okubo, T., and Shiraishi, H. (1999). Novel polymorphism in the promoter and coding regions of the human cholecystokinin B receptor gene: An association analysis with schizophrenia. *Am. J. Med. Genet.* **88**, 700–704.
- Tachikawa, H., Harada, S., Kawanishi, Y., Okubo, T., and Suzuki, T. (2001). Linked polymorphisms (-333G > T and -286A > G) in the promoter region of the CCK-A receptor gene may be associated with schizophrenia. *Psychiatry Res.* **103**, 147–155.
- Tachikawa, H., Harada, S., Kawanishi, Y., Okubo, T., and Shiraishi, H. (2000). Novel polymorphisms of the human cholecystokinin A receptor gene: An association analysis with schizophrenia. *Am. J. Med. Genet.* **96**, 141–145.
- Tamminga, C. A., Tighe, P. J., Chase, T. N., DeFraites, E. G., and Schaffer, M. H. (1981). Des-tyrosine-gamma-endorphin administration in chronic schizophrenics. A preliminary report. *Arch. Gen. Psychiatry* **38**, 167–168.

- Takahashi, Y., Fukushima, S., Murotsu, T., and Matsubara, K. (1986). Structure of human cholecystokinin gene and its chromosomal location. *Gene* **50**, 353–360.
- Tamminga, C. A., Littman, R. L., Alphas, L. D., Chase, T. N., Thaker, G. K., and Wagman, A. M. (1986). Neuronal cholecystokinin and schizophrenia: Pathogenic and therapeutic studies. *Psychopharmacology* **88**, 387–391.
- Tandon, R., Mazzara, C., DeQuardo, J., Craig, K. A., Meador-Woodruff, J. H., Goldman, R., and Greden, J. F. (1991). Dexamethasone suppression test in schizophrenia: Relationship to symptomatology, ventricular enlargement, and outcome. *Biol. Psychiatry* **29**, 953–964.
- Tandon, R., DeQuardo, J. R., Taylor, S. F., McGrath, M., Jibson, M., Eiser, A., and Goldman, M. (2000). Phasic and enduring negative symptoms in schizophrenia: Biological markers and relationship to outcome. *Schizophr. Res.* **45**, 191–201.
- Tang, J. X., Chen, W. Y., He, G., Zhou, J., Gu, N. F., Feng, G. Y., and He, L. (2004). Polymorphisms within 5' end of the Neuregulin 1 gene are genetically associated with schizophrenia in the Chinese population. *Mol. Psychiatry* **9**, 11–12.
- Thiselton, D. L., Webb, B. T., Neale, B. M., Ribble, R. C., O'Neill, F. A., Walsh, D., Riley, B. P., and Kendler, K. S. (2004). No evidence for linkage or association of neuregulin-1 (NRG1) with disease in the Irish study of high-density schizophrenia families (ISHDSE). *Mol. Psychiatry* **9**, 777–783.
- Tkachev, D., Mimmack, M. L., Ryan, M. M., Wayland, M., Freeman, T., Jones, P. B., Starkey, M., Webster, M. J., Yolken, R. H., and Bahn, S. (2003). Oligodendrocyte dysfunction in schizophrenia and bipolar disorder. *Lancet* **362**, 798–805.
- Tooney, P. A., Crawter, V. C., and Chahl, L. A. (2001). Increased tachykinin NK(1) receptor immunoreactivity in the prefrontal cortex in schizophrenia. *Biol. Psychiatry* **49**, 523–527.
- Toru, M., Watanabe, S., Shibuya, H., Nishikawa, T., Noda, K., Mitsushio, H., Ichikawa, H., Kurumaji, A., Takashima, M., Mataga, M., *et al.* (1988). Neurotransmitters, receptors and neuropeptides in post-mortem brains of chronic schizophrenic patients. *Acta Psychiatr. Scand.* **78**, 121–137.
- Uhl, G. R., and Kuhar, M. J. (1984). Chronic neuroleptic treatment enhances neurotensin receptor binding in human and rat substantia nigra. *Nature* **309**, 350–352.
- van Kammen, D. P., McAllister-Sistilli, C. G., Kelley, M. E., Gurklis, J. A., and Yao, J. K. (1999). Elevated interleukin-6 in schizophrenia. *Psychiatry Res.* **87**, 129–136.
- van Ree, J. M., Verhoeven, W. M., Brouwer, G. J., and de Wied, D. (1984). Ceruletide resembles antipsychotics in rats and schizophrenic patients. Preliminary report. *Neuropsychobiology* **12**, 4–8.
- Vanderhaeghen, J. J., Lotstra, F., Vierendeels, G., Gilles, C., Deschepper, C., and Verbanck, P. (1981). Cholecystokinins in the central nervous system and neurohypophysis. *Peptides* **2**(Suppl. 2), 81–88.
- Ventriglia, M., Bocchio Chiavetto, L., Bonvicini, C., Tura, G. B., Bignotti, S., Racagni, G., and Gennarelli, M. (2002). Allelic variation in the human prodynorphin gene promoter and schizophrenia. *Neuropsychobiology* **46**, 17–21.
- Verbanck, P. M., Lotstra, F., Gilles, C., Linkowski, P., Mendlewicz, J., and Vanderhaeghen, J. J. (1984). Reduced cholecystokinin immunoreactivity in the cerebrospinal fluid of patients with psychiatric disorders. *Life Sci.* **34**, 67–72.
- Verhoeven, W. M., van Praag, H. M., van Ree, J. M., and de Wied, D. (1979). Improvement of schizophrenic patients treated with [des-Tyr¹]-gamma-endorphin (DTgammaE). *Arch. Gen. Psychiatry* **36**, 294–298.
- Verhoeven, W. M., van Ree, J. M., Heezius-van Bentum, A., de Wied, D., and van Praag, H. M. (1982). Antipsychotic properties of Des-enkephalin-gamma-endorphin in treatment of schizophrenic patients. *Arch. Gen. Psychiatry* **39**, 648–654.
- Verhoeven, W. M., van Praag, H. M., and van Ree, J. M. (1984a). Repeated naloxone administration in schizophrenia. *Psychiatry Res.* **12**, 297–312.

- Verhoeven, W. M., van Ree, J. M., Westenberg, H. G., Krul, J. M., Brouwer, G. J., Thijssen, J. H., de Wied, D., van Praag, H. M., Ceulemans, D. L., and Kahn, R. S. (1984b). Clinical, biochemical, and hormonal aspects of treatment with Des-tyr¹-gamma-endorphin in schizophrenia. *Psychiatry Res.* **11**, 329–346.
- Verhoeven, W. M., Westenberg, H. G., and van Ree, J. M. (1986). A comparative study on the antipsychotic properties of des enkephalin-gamma-endorphin and ceruletide in schizophrenic patients. *Acta Psychiatr. Scand.* **73**, 372–382.
- Vincent, J. P., Mazella, J., and Kitabgi, P. (1999). Neurotensin and neurotensin receptors. *Trends Pharmacol. Sci.* **20**, 302–309.
- Virgo, L., Humphries, C., Mortimer, A., Barnes, T., Hirsch, S., and de Belleruche, J. (1995). Cholecystokinin messenger RNA deficit in frontal and temporal cerebral cortex in schizophrenia. *Biol. Psychiatry* **37**, 694–701.
- Volavka, J., Hui, K. S., Anderson, B., Nemes, Z., O'Donnell, J., and Lajtha, A. (1983). Short-lived effect of (Des-Tyr)-gamma-endorphin in schizophrenia. *Psychiatry Res.* **10**, 243–252.
- Wang, Z., Wassink, T., Andreasen, N. C., and Crowe, R. R. (2002). Possible association of a cholecystokinin promoter variant to schizophrenia. *Am. J. Med. Genet.* **114**, 479–482.
- Watson, S. J., Berger, P. A., Akil, H., Mills, M. J., and Barchas, J. D. (1978). Effects of naloxone on schizophrenia: Reduction in hallucinations in a subpopulation of subjects. *Science* **201**, 73–76.
- Watson, M., Isackson, P. J., Makker, M., Yamada, M. S., Yamada, M., Cusack, B., and Richelson, E. (1993). Identification of a polymorphism in the human neurotensin receptor gene. *Mayo Clinic Proc.* **68**, 1043–1048.
- Welch, E. B., and Thompson, D. F. (1994). Opiate antagonists for the treatment of schizophrenia. *J. Clin. Pharmacy Ther.* **19**, 279–283.
- Wen, H. L., Lo, C. W., and Ho, W. K. (1983). Met-enkephalin level in the cerebrospinal fluid of schizophrenic patients. *Clin. Chim. Acta* **128**, 367–371.
- Weninger, S. C., Dunn, A. J., Muglia, L. J., Dikkes, P., Miczek, K. A., Swiergiel, A. H., Berridge, C. W., and Majzoub, J. A. (1999a). Stress-induced behaviors require the corticotropin-releasing hormone (CRH) receptor, but not CRH. *PNAS* **96**, 8283–8288.
- Weninger, S. C., Muglia, L. J., Jacobson, L., and Majzoub, J. A. (1999b). CRH-deficient mice have a normal anorectic response to chronic stress. *Regul. Pept.* **84**, 69–74.
- Whiteford, H. A., Stedman, T. J., Welham, J., Csernansky, J. G., and Pond, S. M. (1992). Placebo-controlled, double-blind study of the effects of proglumide in the treatment of schizophrenia. *J. Clin. Psychopharmacol.* **12**, 337–340.
- Widerlöv, E., Lindström, L. H., Besev, G., Manberg, P. J., Nemeroff, C. B., Breese, G. R., Kizer, J. S., and Prange, A. J., Jr. (1982). Subnormal CSF levels of neurotensin in a subgroup of schizophrenic patients: Normalization after neuroleptic treatment. *Am. J. Psychiatry* **139**, 1122–1126.
- Widerlöv, E., Lindström, L. H., Wahlestedt, C., and Ekman, R. (1988). Neuropeptide Y and peptide YY as possible cerebrospinal fluid markers for major depression and schizophrenia, respectively. *J. Psychiatr. Res.* **22**, 69–79.
- Wiegant, V. M., Ronken, E., Kovacs, G., and De Wied, D. (1992). Endorphins and schizophrenia. *Prog. Brain Res.* **93**, 433–453.
- Wiegant, V. M., Verhoef, C. J., Burbach, J. P., and de Wied, D. (1988). Increased concentration of alpha- and gamma-endorphin in post mortem hypothalamic tissue of schizophrenic patients. *Life Sci.* **42**, 1733–1742.
- Williams, N. M., Preece, A., Spurlock, G., Norton, N., Williams, H. J., Zammit, S., O'Donovan, M. C., and Owen, M. J. (2003). Support for genetic variation in neuregulin 1 and susceptibility to schizophrenia. *Mol. Psychiatry* **8**, 485–487.
- Wilson, I. C., Lara, P. P., and Prange, A. J., Jr. (1973). Thyrotrophin-releasing hormone in schizophrenia. *Lancet* **2**, 43–44.

- Wolf, S. S., Hyde, T. M., Saunders, R. C., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (1995). Autoradiographic characterization of neurotensin receptors in the entorhinal cortex of schizophrenic patients and control subjects. *J. Neural. Transm. Gen. Sect.* **102**, 55–65.
- Yamada, K., Wada, E., and Wada, K. (2001). Female gastrin-releasing peptide receptor (GRP-R)-deficient mice exhibit altered social preference for male conspecifics: Implications for GRP/GRP-R modulation of GABAergic function. *Brain Res.* **894**, 281–287.
- Yang, J. Z., Si, T. M., Ruan, Y., Ling, Y. S., Han, Y. H., Wang, X. L., Zhou, M., Zhang, H. Y., Kong, Q. M., Liu, C., Zhang, D. R., Yu, D. R., *et al.* (2003). Association study of neuregulin 1 gene with schizophrenia. *Mol. Psychiatry* **8**, 706–709.
- Yazici, K., Yazici, A. E., and Taneli, B. (2002). Different neuroendocrine profiles of remitted and nonremitted schizophrenic patients. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **26**, 579–584.
- Yeragani, V. K. (1990). The incidence of abnormal dexamethasone suppression in schizophrenia: A review and a meta-analytic comparison with the incidence in normal controls. *Can. J. Psychiatry* **35**, 128–132.
- Zalcman, S., Green-Johnson, J. M., Murray, L., Nance, D. M., Dyck, D., Anisman, H., and Greenberg, A. H. (1994). Cytokine-specific central monoamine alterations induced by interleukin-1, -2 and -6. *Brain Res.* **643**, 40–49.
- Zamir, N., Palkovits, M., and Brownstein, M. J. (1984). Distribution of immunoreactive beta-neo-endorphin in discrete areas of the rat brain and pituitary gland: Comparison with alpha-neo-endorphin. *J. Neurosci.* **4**, 1248–1252.
- Zech, M., Roberts, G. W., Bogerts, B., Crow, T. J., and Polak, J. M. (1986). Neuropeptides in the amygdala of controls, schizophrenics and patients suffering from Huntington's chorea: An immunohistochemical study. *Acta Neuropathol.* **71**, 259–266.
- Zhang, A. Z., Zhou, G. Z., Xi, G. F., Gu, N. F., Xia, Z. Y., Yao, J. L., Chang, J. K., Webber, R., and Potkin, S. (1985). Lower CSF level of dynorphin(1–8) immunoreactivity in schizophrenic patients. *Neuropeptides* **5**, 553–556.
- Zhang, X. Y., Zhou, D. F., Zhang, P. Y., and Wei, J. (2000). The CCK-A receptor gene possibly associated with positive symptoms of schizophrenia. *Mol. Psychiatry* **5**, 239–240.
- Zhang, X. Y., Zhou, D. F., Zhang, P. Y., Wu, G. Y., Cao, L. Y., and Shen, Y. C. (2002). Elevated interleukin-2, interleukin-6 and interleukin-8 serum levels in neuroleptic-free schizophrenia: Association with psychopathology. *Schizophr. Res.* **57**, 247–258.
- Zhang, X. Y., Zhou, D. F., Cao, L. Y., Wu, G. Y., and Shen, Y. C. (2005). Cortisol and cytokines in chronic and treatment-resistant patients with schizophrenia: Association with psychopathology and response to antipsychotics. *Neuropsychopharmacology* **30**, 1532–1538.
- Zhao, X., Shi, Y., Tang, J., Tang, R., Yu, L., Gu, N., Feng, G., Zhu, S., Liu, H., Xing, Y., Zhao, S., Sang, S., *et al.* (2004). A case control and family based association study of the neuregulin 1 gene and schizophrenia. *J. Med. Genet.* **41**, 31–34.

BRAIN-DERIVED NEUROTROPHIC FACTOR IN SCHIZOPHRENIA AND ITS RELATION WITH DOPAMINE

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The brain-derived neurotrophic factor (BDNF) belongs to the neurotrophins family and has a role in proliferation, differentiation of neurons but also as a neurotransmitter. This neurotrophin has received much attention during the last year in regard of the pathophysiology of schizophrenia. Results of genetic studies conducted in schizophrenia support a role for BDNF in schizophrenia and in brain function associated with the disorder. The changes of BDNF observed in the brain and in the plasma of patients with schizophrenia have generated results that can be interpreted either as a hallmark of the disease or a consequence of antipsychotic drugs. Antipsychotic drugs act by blocking the dopamine transmission at the dopamine D2-like receptors. BDNF controls the expression of one of these D2-like receptors, the dopamine D3 receptor. This raises the hypothesis of a link between cortical area, via BDNF, and the dopamine neurotransmission pathway in schizophrenia and its treatment.

I. Introduction

The brain-derived neurotrophic factor (BDNF) belongs to the neurotrophins family, which comprises the prototypical member nerve growth factor (NGF), neurotrophine-3 (NT-3), and neurotrophine-4/5 (NT-4/5). NGF was initially

identified as being responsible for proliferation, differentiation, and function of sympathetic nerve cells (Levi-Montalcini and Cohen, 1960). Neurotrophins can act after their neuronal uptake and retrograde transport to the soma, through their high-affinity tropomyosine-related tyrosine (Trk) receptors and the low-affinity $p75^{\text{NTR}}$ receptor, a member of the tumor necrosis factor (TNF) receptor family (Thoenen, 1995). Trks display specificity for the neurotrophins: NGF only binds with high affinity to TrkA, NT-3 to TrkC and TrkB, BDNF, and NT-4/5 to TrkB. These receptors possess an intracellular tyrosine kinase domain, which transduce the neurotrophins signal by autophosphorylation and subsequent recruitment of enzymes such as phosphatidylinositol-3 kinase or adaptor proteins such as ShC linked to various serine/threonine kinases (Thoenen, 1995).

It was admitted that, whereas interaction with the Trk receptor promotes cell survival, interaction with $p75^{\text{NTR}}$ promotes cell death. Data have challenged this overly simplistic dual scheme (Kalb, 2005; Lu *et al.*, 2005). Neurotrophins are synthesized in a precursor form (proneurotrophins), which generate the mature neurotrophins through proteolytic cleavage. Proneurotrophins bind with higher affinity to $p75^{\text{NTR}}$ than do mature neurotrophins, but they do not bind to Trk receptors (Lee *et al.*, 2001). Moreover, complexes of $p75^{\text{NTR}}$, or homologues of this receptor (Kanning *et al.*, 2003) with either sortilin or Trk receptor modulate the affinity of the neurotrophins (Kalb, 2005). It results from these considerations that life–death decisions in neurons depend not only on neurotrophic supply but also on the pro- and mature-neurotrophin balance (Lu *et al.*, 2005).

A more diverse role for BDNF as an extracellular transmitter has, nevertheless, been inferred from observations that it is anterogradely transported (Altar *et al.*, 1997; von Bartheld *et al.*, 1996), released on neuron depolarization, and triggers rapid intracellular signals (Altar and DiStefano, 1998; Thoenen, 1995) and action potentials in central neurons (Kafitz *et al.*, 1999) via intracellular transduction of its high-affinity membrane receptor TrkB (Blum *et al.*, 2001). BDNF can alter fast synaptic transmission by speeding up the development of excitatory and inhibitory synapses (Vicario-Abejon *et al.*, 1998) but also by modulating synaptic efficacy (Huang *et al.*, 1999; Lohof *et al.*, 1993). In particular, BDNF is necessary for the induction and maintenance of hippocampal long-term potentiation (Barco *et al.*, 2005; Figurov *et al.*, 1996; Korte *et al.*, 1995; Kovalchuk *et al.*, 2002; Patterson *et al.*, 1996). ProBDNF demonstrate opposite function in activating $p75^{\text{NTR}}$ which facilitates hippocampal long-term depression (Woo *et al.*, 2005). As suggested in life–death decision, a bidirectional regulation of synaptic plasticity by proBDNF and mature BDNF might exist. Although some observations suggest a role of BDNF in nociception (Kerr *et al.*, 1999), mechanosensation (Carroll *et al.*, 1998), and learning (Du and Poo, 2004; Egan *et al.*, 2003; Lee *et al.*, 2004; Linnarsson *et al.*, 1997; Minichiello *et al.*, 1999).

For more than 40 years, dopamine has been consistently implicated in the pathophysiology of schizophrenia and its treatment (Carlsson, 1995). Evidences

have emerged showing BDNF-induced synaptic plasticity and modulating the physiological functions of the dopamine transmission pathway (Berton *et al.*, 2006; Goggi *et al.*, 2003; Grimm *et al.*, 2003; Guillin *et al.*, 2001; Horger *et al.*, 1999; Monteggia *et al.*, 2004).

In this chapter, we propose to review evidences generated by genetic, plasma, and brain concentration of BDNF studies in the human affected by schizophrenia and the relationship between dopamine and BDNF.

II. Genetic Studies

The BDNF gene is localized on the reverse strand of chromosome 11p13 and encodes a precursor peptide (proBDNF), with a strong conservation of the coding sequence across species. The gene consists of four short 5' exons with separate promoters and one 3' exon encoding mature BDNF protein (Metsis *et al.*, 1993; Timmusk *et al.*, 1993).

In schizophrenia, no significant linkage has been reported in this region in the large genome-wide scan studies. In bipolar disorders, a linkage study has generated an LOD score of 1.89 in bipolar families (Detera-Wadleigh *et al.*, 1999). Genetic association studies have been carried out in order to determine and define the association between polymorphisms located in the BDNF gene and schizophrenia.

In 1992, the first study by Pröschel *et al.* (1992) described a microsatellite: GT dinucleotide repeat polymorphism (166–174 bp) in the human BDNF gene, upstream from the transcription start site of the 1.6-kb BDNF mRNA transcript (Pröschel *et al.*, 1992). Many studies have been conducted in order to find a possible association between this polymorphism and schizophrenia (Hawi *et al.*, 1998; Krebs *et al.*, 2000; Sasaki *et al.*, 1997; Virgos *et al.*, 2001; Wassink *et al.*, 1999). No association was found between this polymorphism and schizophrenia, except for the Italian family study (Muglia *et al.*, 2003). A French study found no statistical difference in allele or genotype distribution of this polymorphism between schizophrenic patients and controls (Krebs *et al.*, 2000). However and interestingly, the authors revealed an excess of the 172- to 176-bp alleles in late onset, neuroleptic-responding patients ($n = 68$ patients) and in nonsubstance abuse patients ($n = 52$ patients).

BDNF plays an important role in activity-dependent hippocampal neuroplasticity and hippocampal-dependent memory. The valine-to-methionine variation at codon 66 of BDNF coding sequence, located in the 5' proregion of the BDNF protein, is a frequent single nucleotide missense and functional polymorphism in the human BDNF gene (Egan *et al.*, 2003). The presence of the

Val66Met has been associated with abnormal intracellular trafficking and activity-dependent secretion of BDNF in cultured hippocampal neurons (Egan *et al.*, 2003). In fact, both lower depolarization-induced secretion and failure to localize to secretory granules or synapses were demonstrated for 66Met BDNF-transfected neurons using virus-mediated transfection in cultured hippocampal neurons, compared with 66Val BDNF (Egan *et al.*, 2003).

In human subjects, the Met allele is associated with impaired episodic memory assessed with the Wechsler Memory Scale (WMS) (Egan *et al.*, 2003; Tan *et al.*, 2005a). However, these data have only been partially replicated, since schizophrenic patients and relatives were analyzed separately: 66Met was associated with a lower score at the WMS but only in relatives (Dempster *et al.*, 2005). The Met carriers (healthy subjects) exhibit relatively diminished hippocampal activity in comparison with the Val/Val subjects during both encoding and retrieval processes. This impairment was directly linked to abnormal hippocampal activation: Val/Met individuals have an abnormal pattern of increased BOLD fMRI signal activation of bilateral caudal hippocampus. In contrast, Val/Val subjects show a characteristic hippocampal deactivation pattern (Egan *et al.*, 2003). A direct effect of BDNF alleles on hippocampal processing of memory was then demonstrated when the same group showed that the interaction between the BDNF Val66Met genotype and the hippocampal response during encoding accounted for 25% of the total variation in recognition memory performance (Hariri *et al.*, 2003). Moreover, this polymorphism in the BDNF gene affects the morphology of the brain: Met-carriers have a reduction in hippocampal gray matter and in the dorsolateral prefrontal cortex compared to the Val-carrier (Pezawas *et al.*, 2004). Significant interaction between prefrontal cognitive performance (the N-back test) and Val/Val genotype implicates that working memory could also be affected by the BDNF Val66Met polymorphism (Rybakowski *et al.*, 2006). Moreover, Val66Met seems to be associated with age-related change in reasoning skills. In a cohort of healthy subjects, aged 79 years at the time of the study whom reasoning was assessed by the Raven's progressive, Met homozygotes scored significantly higher than heterozygotes and Val homozygotes (Harris *et al.*, 2006).

A study has examined a large schizophrenia sample ($n = 321$) in comparison with bipolar patients ($n = 321$) and controls ($n = 350$), testing haplotype frequencies for the BDNF GT dinucleotide-repeat and the Val66Met polymorphisms (Neves-Pereira *et al.*, 2005). The authors underlie a significant excess of the Val haplotype in schizophrenics without stratification bias and suggest that Met or Met combined with the 174-bp haplotypes may be a protective factor against schizophrenia. In a large sample of 94 families, a transmission disequilibrium test (TDT) showed a preferential transmission of the Val allele from the heterozygous parents (Val/Met) to their affected schizophrenic offspring (Rosa *et al.*, 2006).

However, no association was found between the prefrontal tests assessed (WMS) and the BDNF Val/Met polymorphism in this sample (Rosa *et al.*, 2006).

Three single nucleotide polymorphisms (rs3750934, rs6265, and rs10707) have been investigated in an association study in the Chinese population (Chen *et al.*, 2006). No significant differences were found for either the genotype or allele distribution of analyzed polymorphisms.

BDNF and dopamine D3 receptor (DRD3) interplay might be of special interest in schizophrenia (Guillin *et al.*, 2004). Interestingly, an interaction between BDNF Met-containing haplotypes and DRD3 receptor ser/ser haplotypes has been found in early age at onset of schizophrenia (Gourion *et al.*, 2005).

Association between BDNF polymorphisms and response to antipsychotic treatment or tardive dyskinesia have been also investigated. An association between the BDNF gene Val66Met polymorphism and a good response to clozapine treatment in schizophrenia was found (Hong *et al.*, 2003). Association and gene-gene interaction between the DRD3 ser⁹gly and BDNF Val66Met polymorphisms have been investigated in a cohort of patients with schizophrenia who had significantly high abnormal involuntary movements (Liou *et al.*, 2004). Heterozygosity for the BDNF genotype was associated with abnormal orofacial movement scores but neither DRD3 nor BDNF genotypes were clearly associated with tardive dyskinesia and no gene interaction was found.

A single nucleotide substitution (C270T) in the 5' noncoding BDNF region was described and a significant association with late-onset Alzheimer disease and T270 was found (Kunugi *et al.*, 2001). Comparing schizophrenic patients ($n = 178$) and controls ($n = 332$), the frequency of this nucleotide substitution was significantly increased in schizophrenia (Nanko *et al.*, 2003). The association was replicated in another sample. The C/T genotype was overrepresented in schizophrenics ($n = 101$, 25.7%) compared to controls ($n = 68$, 5.9%), despite a heterogeneity between populations (Szekeres *et al.*, 2003). However, another study has found no association between schizophrenia and the C270T polymorphism in the BDNF gene for genotype and allelic distribution (Szczechankiewicz *et al.*, 2005). Galderisi *et al.* (2005) have studied two polymorphisms: COMT Val¹⁵⁸Met and BDNF C270T in patients with schizophrenia versus controls. This case control association study does not report evidence for association between these polymorphisms and schizophrenia. The functional significance of C270T substitution in the promoter region of BDNF is not clear and there is no evidence that the C270T is involved in alterations of protein expression or function.

In conclusion, BDNF gene is a relatively new and promising target in genetic studies of mental disorders. Concerning schizophrenia, the initial results need replication. However, considering a polygenic model of schizophrenia transmission, BDNF can be involved in a genetic modulation pathway.

III. BDNF in the Serum of Patients with Schizophrenia

BDNF can be detected in the plasma of patients with schizophrenia and controls. To date, seven studies (summarized in Table I) have investigated BDNF plasma levels in patients with schizophrenia, drug free (Pirildar *et al.*, 2004; Shimizu *et al.*, 2003), drug naïve, with substance abuse (Jockers-Scherubl *et al.*, 2004; Pirildar *et al.*, 2004), chronically treated with antipsychotics drugs (Jockers-Scherubl *et al.*, 2004; Pirildar *et al.*, 2004; Shimizu *et al.*, 2003; Tan *et al.*, 2005b,c; Toyooka *et al.*, 2002) and the relationship with the presence of tardive dyskinesia (Tan *et al.*, 2005c). In drug-naïve patients no change in BDNF plasma levels was found (Jockers-Scherubl *et al.*, 2004; Shimizu *et al.*, 2003) whereas conflicting results have emerged from studies with drug-free patients: in one study BDNF plasma levels was decreased (Pirildar *et al.*, 2004) and there was no change in the second one (Huang and Lee, 2005). In patients with schizophrenia treated with antipsychotic drugs at the time BDNF plasma levels were determined, decreased (Grillo *et al.*, 2007; Tan *et al.*, 2005b,c; Toyooka *et al.*, 2002), or normal (Shimizu *et al.*, 2003) plasma levels compared to controls were observed. Interestingly, tardive dyskinesia (Tan *et al.*, 2005c) and substance abuses (Jockers-Scherubl *et al.*, 2004) have been found to be associated with decreased BDNF plasma levels in patients with schizophrenia.

Altogether these results seem to indicate that BDNF plasma levels are decreased in medicated patients with schizophrenia. However, the source of BDNF found in the plasma is still unknown as the relationship between plasma and brain levels. Nevertheless, brain- and plasma-level changes are correlated during aging in rats (Karege *et al.*, 2002). One way to explain the conflicting results between plasma levels found in patients chronically treated with antipsychotic drugs and drug-naïve patients might be the possible ability of antipsychotic drugs to decrease BDNF expression (Section IV).

IV. BDNF and TrkB Receptor in the Brain of Patients with Schizophrenia

Several studies have been conducted in order to determine BDNF protein or mRNA and TrkB mRNA levels in the postmortem brain tissue from patients with schizophrenia.

In the first study to be published, a significant increase of BDNF protein, determined by immunoassay, was found in the anterior cingulate and the hippocampus (Takahashi *et al.*, 2000). This was confirmed in another collection of brain tissue in the cortical areas but not in the hippocampus (Durany *et al.*, 2001).

TABLE I
BDNF PLASMA LEVELS IN SCHIZOPHRENIA

Author	<i>N</i> controls	<i>N</i> patients	Treatment Status	Controls (<i>n.</i> mean ± SD) ^a	Patients (<i>n.</i> mean ± SD) ^a	Patients subtype (<i>n.</i> mean ± SD) ^a	<i>p</i>	Effect size ^b
Tan <i>et al.</i> (2005b)	45	81	M	100 ± 43	74 ± 26		<0.001	0.6
Tan <i>et al.</i> (2005c)	45	125	M	100 ± 43	66 ± 26 ³	55 ± 17 ⁴	³ vs controls = 0.002	³ 0.8
Jockers-Scherubl <i>et al.</i> (2004)	72	157	DN	100 ± 40	99 ± 45	134 ± 55 ⁵ 129 ± 45 ⁶	⁴ vs controls <0.01 ⁵ vs controls = 0.0062 ⁶ vs controls = 0.0131	⁴ 1.05 ⁵ 0.85 ⁶ 0.725
Pirildar <i>et al.</i> (2004)	22	22	DF ¹	100 ± 35	53 ± 30		<0.001	1.34
Shimizu <i>et al.</i> (2003)	40	40	15 DN 25 M	100 ± 32	84 ± 28 ⁷	98 ± 43 ⁸	NS	0.5
Toyooka <i>et al.</i> (2002)	62	2 independent groups of 34	M	100 ± 67	55 ± 30		0.004	0.67
Huang and Lee (2005)	96	126	DF ²	100 ± 48	100 ± 49		NS	

^aMean normalized to mean of control subjects.
^bEffect size calculated as (mean patients – mean controls)/SD controls.
M, patients taking antipsychotics drugs at the time of the study; DN, drug naïve; DF, drug free for at least 2 weeks¹ or 1 week².
Patients with schizophrenia without³ and with tardive dyskinesia⁴; with cannabis intake⁵ or cannabis and additional substances⁶; drug naïve,⁷ or medicated⁸.
NS, nonsignificant.

A very comprehensive paper using three different methods to determine BDNF levels, RNase protection assay, Western blotting, and *in situ* hybridization was published 3 years later (Weickert *et al.*, 2003). In this study, BDNF mRNA and protein was decreased in the lateral prefrontal cortex of patients with schizophrenia, especially in the layers III, V, and VI. This result was confirmed by another group in two independent cohorts (Hashimoto *et al.*, 2005). Combining the two independent cohorts, the authors found in the prefrontal cortex a 35% decrease of BDNF mRNA in layers II, III, and V/VI. In the same sample of post-mortem brain tissue, TrkB mRNA was found to be decreased in the prefrontal cortex by 23% (Hashimoto *et al.*, 2005). The same result was found in the brain collection of the NIMH in the dorsolateral prefrontal cortex (Weickert *et al.*, 2005).

To date, two from four studies indicate that in patients with schizophrenia BDNF and TrkB are downregulated. Could decreased BDNF in some of the studies be attributable to antipsychotic drugs? First, in one of the studies showing a decrease of BDNF, any correlation between lifetime antipsychotic exposures and BDNF levels was found (Weickert *et al.*, 2003). Moreover, in the other one, some patients were free of medication for more than 1 month at the time of death and did not demonstrate significant higher expression of BDNF in the prefrontal cortex than patient under treatment (Hashimoto *et al.*, 2005). However and interestingly, in the study by Takahashi *et al.* (2000) showing an increase of BDNF in the cingulate cortex, patients were treated at the time of death with very low doses of antipsychotics as reflected by an average equivalent chlorpromazine of the sample of 72 mg/day.

Antipsychotic drugs' effects on BDNF expression in the brain of normal rats and nonhuman primates have been investigated. Haloperidol, a first-generation antipsychotic drug, given for more than 9 months in monkeys do not induce significant changes in BDNF mRNA in the prefrontal cortex (Hashimoto *et al.*, 2005). Other studies, conducted in normal rats have found no change or decreased BDNF mRNA and protein in cortical regions after chronic exposure to antipsychotic drugs (Angelucci *et al.*, 2000; Dawson *et al.*, 2001; Linden *et al.*, 2000; Lipska *et al.*, 2001; Nibuya *et al.*, 1995; Takahashi *et al.*, 2000).

Dysfunction of glutamatergic neurotransmission has been proposed to play an important role in the pathophysiology of schizophrenia (Goff and Coyle, 2001). In rats, glutamate hypofunction induced by the administration of MK-801 induces an increase of BDNF mRNA in the cortex which is circumvented by clozapine or haloperidol pretreatment (Linden *et al.*, 2000; Sokoloff *et al.*, 2006).

At this time, the more conservative conclusion of postmortem studies on BDNF expression in the prefrontal cortex is that there is discrepancy between studies and that the role of treatment on the results is still under debate.

V. Dopamine–BDNF Interactions

Since the 1960s, a crucial role for dopamine in schizophrenia and its treatment was suspected in schizophrenia (Carlsson and Lindqvist, 1963). It has been confirmed by imaging studies (Abi-Dargham and Laruelle, 2005; Laruelle, 2000, 2005). Some recent evidences link BDNF and dopamine neurotransmission.

A. BDNF SUPPORTS THE SURVIVAL AND THE DIFFERENTIATION OF DOPAMINERGIC NEURONS

BDNF is expressed in the adult dopaminergic neurons of the midbrain and destruction of dopaminergic cells by 6-hydroxydopamine result in a loss of BDNF mRNA (Seroogy *et al.*, 1994). In the same line of evidences, in disease with a loss of dopaminergic neurons like Huntington disease and Parkinson disease postmortem studies have shown a decrease of BDNF in the substantia nigra and the striatum (Chauhan *et al.*, 2001; Ferrer *et al.*, 2000; Howells *et al.*, 2000; Mogi *et al.*, 1999; Parain *et al.*, 1999). Using neurons in culture, BDNF was found to promote the survival of the dopaminergic neurons of the developing substantia nigra (Hyman *et al.*, 1991) and to elicit an increase in the depolarization-induced release of dopamine (Feng *et al.*, 1999). In mice lacking selectively the BDNF gene in the midbrain, there is a significant but not complete reduction in non-expressing calbindin and calcineurin dopaminergic neurons during development (Baquet *et al.*, 2005). However, this has not been confirmed in a report (Berton *et al.*, 2006). Moreover, BDNF protects dopaminergic neurons from the toxic effect of MPTP, 6-hydroxydopamine, oxidative stress, and hypoglycemic injury (Hung and Lee, 1996; Levivier *et al.*, 1995; Nakao *et al.*, 1995; Petersen *et al.*, 2001; Shults *et al.*, 1995). These functions are likely to complement and overlap with those of other neurotrophic factor, including glial cell line-derived neurotrophic factor and bone morphogenetic proteins, which are also known to enhance differentiation and survival of dopaminergic neurons (Brederleau *et al.*, 2002; Feng *et al.*, 1999; Gratacos *et al.*, 2001; Zuch *et al.*, 2004).

B. BDNF IN THE GABA-CONTAINING LOCAL CIRCUIT NEURONS OF THE PREFRONTAL CORTEX

Alteration in the circuitry of the dorsal prefrontal cortex appears to contribute to the working memory impairments in schizophrenia (Lewis and Lieberman, 2000), more particularly in the GABA-containing local circuit neurons of the

prefrontal cortex (Volk and Lewis, 2002), which BDNF regulates the maturation in the developing cortex (Huang *et al.*, 1999). Several lines of evidences have emerged from the Lewis's group in Pittsburgh involving BDNF in the alteration of GABA neurons in schizophrenia. These authors have shown in the postmortem brain tissue of patients with schizophrenia that the decrease in BDNF and TrkB mRNA was correlated with the decrease in GAD₆₇ mRNA, a marker of the integrity of GABA neurons (Hashimoto *et al.*, 2005). However, the presence of the BDNF Met66 allele, a polymorphism which reduces the trafficking and secretion of BDNF protein (Egan *et al.*, 2003) did not contribute to the decreased level of GAD₆₇ mRNA expression of the same sample of brain tissue of patients with schizophrenia (Hashimoto and Lewis, 2006).

C. FUNCTIONAL INTERPLAY BETWEEN BDNF AND DOPAMINE

The first line of evidences indicating a role of BDNF in modulating dopaminergic functions came from studies in rats infused with recombinant BDNF in the substantia nigra. In these animals, an increase of locomotor activity and dopamine agonists-induced rotational behavior is found suggesting that BDNF increases dopamine function. This was also suggested by the fact that supranigral infusion of BDNF elicits dopamine turnover in the striatum and increases the electrical activity of dopaminergic neurons via an activation of the PI3K and Ras-MEK pathways (Altar *et al.*, 1992; Goggi *et al.*, 2003; Shen *et al.*, 1994).

Chronic exposure to drug of abuse elicits long-lasting changes in the ventral tegmental area, a brain region involved in drug addiction (Koob, 1992). Rats chronically treated with morphine have a reduction in their ventral tegmental area of dopaminergic neurons that BDNF infusion prevents (Skclair-Tavron *et al.*, 1996). Chronic infusions of BDNF into the ventral tegmental area in the nucleus accumbens result in increased locomotor activity and enhanced locomotor sensitization to cocaine (Horger *et al.*, 1999).

In mice experiencing repeated aggression, a local deletion of the BDNF gene in the dopaminergic neurons of the ventral tegmental area does not permit the development of social avoidance like it does in animals with a normal expression of BDNF (Berton *et al.*, 2006b).

Altogether, these results are consistent with the idea of BDNF as a modulator of dopaminergic function.

D. BDNF CONTROLS THE EXPRESSION OF THE DOPAMINE D3 RECEPTOR

The DRD3 belongs to the dopamine D2-like receptors (Sokoloff *et al.*, 1990). From the beginning, attention has been attracted to the restricted distribution of the DRD3 in the brain, seemingly related to functions of dopamine associated

with the limbic brain (Bouthenet *et al.*, 1991). Hence, the hypothesis has been put forward that the DRD3 receptor could be involved in the pathophysiology of schizophrenia. Several other findings might implicate the DRD3 in the pathophysiology of schizophrenia. For instance, antipsychotic drugs display affinity at recombinant dopamine D2 receptor and DRD3 in the same magnitude (Sokoloff *et al.*, 1990). Moreover, in spite of controversy (Sabate *et al.*, 1994), the meta-analysis of the ser-gly polymorphism of the DRD3 have been found to be associated with schizophrenia (Dubertret *et al.*, 1998). However, the most direct evidence of a role of the DRD3 in schizophrenia has come from a postmortem study which, at this date, has never been refuted, neither replicated. DRD3 levels have been found elevated in the brain of drug-free schizophrenic patients, but not in patients under medication with antipsychotics at the time of death (Gurevich *et al.*, 1997). This suggests that increased DRD3 expression is a hallmark of the disease and that antipsychotic medications normalize this receptor expression. DRD3 overexpression in the etiology of schizophrenia raises the question of mechanisms governing this receptor expression during development.

In adults, the expression of the DRD3 in medium-sized neurons of the nucleus accumbens, but not in granule cells of the islands of Calleja, is highly dependent on the dopaminergic innervation: ablation of the afferent neurons by unilateral 6-hydroxydopamine results in a dramatic decrease in DRD3 density in the ipsilateral nucleus accumbens (Levesque *et al.*, 1995). This paradoxical change (the dopamine D2 receptor is upregulated under these circumstances) was shown to depend on the lack of an anterogradely transported factor from dopaminergic neurons, distinct from dopamine itself and its known peptide cotransmitters, and which is released on dopamine neuron activation (Levesque *et al.*, 1995). Among the candidate factors for regulating DRD3 expression, BDNF was particularly attractive, since it is expressed by dopamine neurons (Seroogy *et al.*, 1994). BDNF immunoreactivity is prominent in the shell of the nucleus accumbens of normal rats (Conner *et al.*, 1997), and its receptor, the TrkB, colocalizes with the DRD3 (Guillin *et al.*, 2001).

Several lines of evidences have lead to the conclusion that BDNF controls DRD3 expression (Guillin *et al.*, 2001). In mice with a BDNF-null mutation, DRD3 binding and mRNA are low at postnatal days 9–14 and do not increase at later stages as it does in their normal littermates. These results show that BDNF is required for the normal development of DRD3 expression in the shell of the nucleus accumbens. In unilaterally 6-hydroxydopamine-lesioned rats, repeated administration of levodopa, leading to extraneuronal dopamine formation, triggers DRD3 overexpression, not only in the shell of the nucleus accumbens but also in the denervated striatum, a brain structure in which DRD3 expression is hardly detectable (Bordet *et al.*, 1997). During levodopa treatment of 6-hydroxydopamine-lesioned rats, infusion into the denervated striatum of a selective BDNF antagonist impairs induction of both DRD3 mRNA and protein

expression (Guillin *et al.*, 2001). This overexpression of the DRD3 in the denervated striatum triggers the development of behavioral sensitization to levodopa (Bordet *et al.*, 1997). Infusion of the selective TrkB antagonist dose dependently inhibits behavioral sensitization, indicating that behavioral sensitization is triggered by BDNF (Guillin *et al.*, 2001). Striatal BDNF in fact originates mainly from cortical neurons (Altar *et al.*, 1997). In agreement, cortical ablation partially impairs the induction of DRD3 overexpression in the striatum and behavioral sensitization, indicating that both processes require the participation of corticostriatal neurons (Guillin *et al.*, 2001). Levodopa also induces BDNF mRNA on the frontal cortex in the 6-hydroxydopamine-lesioned side, mainly in cortical layer V, containing pyramidal cell bodies, and in layer VI, which sends projections to various subcortical areas, notably striatal and accumbal areas. This effect critically depends on activation of the dopamine D1/D5 receptors (Guillin *et al.*, 2001).

Altogether, these results demonstrate that BDNF triggers behavioral sensitization by controlling DRD3 expression and, more generally controls dopamine tone in the limbic forebrain.

VI. Conclusions

Behavioral sensitization of the dopamine system might be involved in the early stage of schizophrenia and more likely in the pathophysiology of positive symptoms (Laruelle, 2005; Lewis and Lieberman, 2000). In schizophrenia, DRD3 protein is elevated in nontreated patients. Therefore, BDNF should be found elevated in cortical regions of nontreated patients with schizophrenia. As discussed before, these data are not available at this time as all postmortem studies were performed on brain of patients treated by antipsychotic at the time of death. Moreover, the effect of antipsychotic drugs on BDNF expression is still controversial. Thus, the view that hypo- or hyperfunction of BDNF in the prefrontal cortex participate to the emergence of symptoms of schizophrenia could be supported (Hashimoto *et al.*, 2005; Sokoloff *et al.*, 2006; Weickert *et al.*, 2003).

However, results show that subchronic blockade of glutamatergic transmission induces an increase of BDNF in the frontal cortex and a striatal DRD3 overexpression in mice, that is corrected by antipsychotic drugs (Sokoloff *et al.*, 2006). In sensitized animals, DRD3 expression is under the control of prefrontal cortex BDNF and this expression is under the control of the dopamine D1 receptor stimulation (Guillin *et al.*, 2001). Dopamine D1 expression have been found to be elevated in the dorsolateral prefrontal cortex of patients with schizophrenia and associated to working performance impairment (Abi-Dargham *et al.*, 2002).

Thus, we can hypothesized that, by an unknown neurodevelopmental process, dopamine D1 function is enhanced in the dorsolateral prefrontal cortex, leading to an increase of BDNF expression, that, in turns, elicits an overexpression of the DRD3 in the striatum that participates to the expression of positive symptoms. Antipsychotic drugs might decrease BDNF levels and, therefore, normalize subcortical dopaminergic hyperfunction mediated by the DRD3.

References

- Abi-Dargham, A., and Laruelle, M. (2005). Mechanisms of action of second generation antipsychotic drugs in schizophrenia: Insights from brain imaging studies. *Eur. Psychiatry* **20**, 15–27.
- Abi-Dargham, A., Mawlawi, O., Lombardo, I., Gil, R., Martinez, D., Huang, Y., Hwang, D. R., Keilp, J., Kochan, L., Van Heertum, R., Gorman, J. M., and Laruelle, M. (2002). Prefrontal dopamine D1 receptors and working memory in schizophrenia. *J. Neurosci.* **22**, 3708–3719.
- Altar, C. A., and DiStefano, P. S. (1998). Neurotrophin trafficking by anterograde transport. *Trends Neurosci.* **21**, 433–437.
- Altar, C. A., Boylan, C., Jackson, C., Hershenson, S., Miller, J., Wiegand, S. J., Lindsay, R. M., and Hyman, C. (1992). Brain-derived neurotrophic factor augment rotational behavior and nigrostriatal dopamine turnover *in vivo*. *Proc. Natl. Acad. Sci. USA* **89**, 11347–11351.
- Altar, C. A., Cai, N., Bliven, T., Juhasz, M., Conner, J. M., Acheson, A. L., Lindsay, R. M., and Wiegand, S. J. (1997). Anterograde transport of brain-derived neurotrophic factor and its role in the brain. *Nature* **389**, 856–860.
- Angelucci, F., Mathe, A. A., and Aloe, L. (2000). Brain-derived neurotrophic factor and tyrosine kinase receptor TrkB in rat brain are significantly altered after haloperidol and risperidone administration. *J. Neurosci. Res.* **60**, 783–794.
- Baquet, Z., Bickford, P., and Jones, K. (2005). Brain-derived neurotrophic factor is required for the establishment of the proper number of dopaminergic neurins in the substantia nigra pars compacta. *J. Neurosci.* **25**, 6251–6259.
- Barco, A., Patterson, S., Alarcon, J. M., Gromova, P., Mata-Roig, M., Morozov, A., and Kandel, E. R. (2005). Gene expression profiling of facilitated L-LTP in VP16-CREB mice reveals that BDNF is critical for the maintenance of LTP and its synaptic capture. *Neuron* **48**, 123–137.
- Berton, O., McClung, C., DiLeone, R., Krishnan, V., Renthall, W., Russo, S., Graham, D., Tsankova, N., Bolanos, C. A., Rios, M., Monteggia, L. M., Self, D. W., *et al.* (2006). Essential role of BDNF in the mesolimbic dopamine pathway in social defeat stress. *Science* **311**, 864–868.
- Blum, M. W., Siegel, A. M., Meier, R., and Hess, K. (2001). Neuroleptic malignant-like syndrome and acute hepatitis during tolcapone and clozapine medication. *Eur. Neurol.* **46**, 158–160.
- Bordet, R., Ridray, S., Carboni, S., Diaz, J., Sokoloff, P., and Schwartz, J. C. (1997). Induction of dopamine D3 receptor expression as a mechanism of behavioral sensitization to levodopa. *Proc. Natl. Acad. Sci. USA* **94**, 3363–3367.
- Bouthenet, M. L., Souil, E., Martres, M. P., Sokoloff, P., Giros, B., and Schwartz, J. C. (1991). Localization of dopamine D3 receptor mRNA in the rat brain using *in situ* hybridization histochemistry: Comparison with dopamine D2 receptor mRNA. *Brain Res.* **564**, 203–219.
- Brederleau, A., Faigle, R., Kaplan, P., Odin, P., and Funa, K. (2002). Bone morphogenetic proteins but not growth differentiation factors induce dopaminergic differentiation in mesencephalic precursors. *Mol. Cell. Neurosci.* **21**, 367–378.

- Carlsson, A. (1995). The dopamine theory revisited. In "Schizophrenia" (S. Hirsh and D. R. Weinberger, Eds.), pp. 373–400. Blackwell, Cambridge.
- Carlsson, A., and Lindqvist, M. (1963). Effect of chlorpromazine or haloperidol on formation of 3 methoxytyramine and normetanephrine in mouse brain. *Acta Pharmacol. Toxicol. (Copenh.)* **20**, 140–144.
- Carroll, P., Lewin, G. R., Koltzenburg, M., Toyka, K. V., and Thoenen, H. (1998). A role for BDNF in mechanosensation. *Nat. Neurosci.* **1**, 42–46.
- Chauhan, N. B., Siegel, G. J., and Lee, J. M. (2001). Depletion of glial cell line-derived neurotrophic factor in substantia nigra neurons of Parkinson's disease brain. *J. Chem. Neuroanat.* **21**, 277–288.
- Chen, Q. Y., Chen, Q., Feng, G. Y., Wan, C. L., Lindpaintner, K., Wang, L. J., Chen, Z. X., Gao, Z. S., Tang, J. S., Li, X. W., and He, L. (2006). Association between the brain-derived neurotrophic factor (BDNF) gene and Schizophrenia in the Chinese population. *Neurosci. Lett.* **397**(3), 285–290.
- Conner, J. M., Lauterborn, J., Yan, Q., Gall, C., and Varon, S. (1997). Distribution of brain-derived neurotrophic factor (BDNF) protein and mRNA in the normal adult rat CNS: Evidence for anterograde axonal transport. *J. Neurosci.* **17**, 2295–2313.
- Dawson, N. M., Hamid, E. H., Egan, M. F., and Meredith, G. E. (2001). Changes in the pattern of brain-derived neurotrophic factor immunoreactivity in the rat brain after acute and subchronic haloperidol treatment. *Synapse* **39**, 70–81.
- Dempster, E., Touloupoulou, T., McDonald, C., Bramon, E., Walshe, M., Filbey, F., Wickham, H., Sham, P. C., Murray, R. M., and Collier, D. A. (2005). Association between BDNF val66 met genotype and episodic memory. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **134**, 73–75.
- Deterra-Wadleigh, S., Badner, J., Berrettini, W., Yoshikawa, T., Goldin, L., Turner, G., Rollins, D., Moses, T., Sanders, A., Karkera, J., Esterling, L., Zeng, J., *et al.* (1999). A high-density genome scan detects evidence for a bipolar-disorder susceptibility locus on 13q32 and other potential loci on 1q32 and 18p11.2. *Proc. Natl. Acad. Sci. USA* **96**, 5604–5609.
- Du, J. L., and Poo, M. M. (2004). Rapid BDNF-induced retrograde synaptic modification in a developing retinotectal system. *Nature* **429**, 878–883.
- Dubertret, C., Gorwood, P., Ades, J., Feingold, J., Schwartz, J. C., and Sokoloff, P. (1998). Meta-analysis of DRD3 gene and schizophrenia: Ethnic heterogeneity and significant association in Caucasians. *Am. J. Med. Genet.* **81**, 318–322.
- Durany, N., Michel, T., Zochling, R., Boissl, K. W., Cruz-Sanchez, F. F., Riederer, P., and Thome, J. (2001). Brain-derived neurotrophic factor and neurotrophin 3 in schizophrenic psychoses. *Schizophr. Res.* **52**, 79–86.
- Egan, M. F., Kojima, M., Callicott, J. H., Goldberg, T. E., Kolachana, B. S., Bertolino, A., Zaitsev, E., Gold, B., Goldman, D., Dean, M., Lu, B., and Weinberger, D. R. (2003). The BDNF val66met polymorphism affects activity-dependent secretion of BDNF and human memory and hippocampal function. *Cell* **112**, 257–269.
- Feng, L., Wang, C. Y., Jiang, H., Oho, C., Mizuno, K., Dugich-Djordjevic, M., and Lu, B. (1999). Differential effects of GDNF and BDNF on cultured ventral mesencephalic neurons. *Brain Res. Mol. Brain Res.* **66**, 62–70.
- Ferrer, I., Goutan, E., Marin, C., Rey, M. J., and Ribalta, T. (2000). Brain-derived neurotrophic factor in Huntington disease. *Brain Res.* **866**, 257–261.
- Figurov, A., Pozzo-Miller, L. D., Olafsson, P., Wang, T., and Lu, B. (1996). Regulation of synaptic responses to high-frequency stimulation and LTP by neurotrophins in the hippocampus. *Nature* **381**, 706–709.
- Galderisi, S., Maj, M., Kirkpatrick, B., Piccardi, P., Mucci, A., Invernizzi, G., Rossi, A., Pini, S., Vita, A., Cassano, P., Stratta, P., Severino, G., *et al.* (2005). COMT Val¹⁵⁸ Met and BDNF C²⁷⁰T polymorphisms in Schizophrenia: A case-control study. *Schizophr. Res.* **73**(1), 27–30.
- Goff, D. C., and Coyle, J. T. (2001). The emerging role of glutamate in the pathophysiology and treatment of schizophrenia. *Am. J. Psychiatry* **158**, 1367–1377.

- Goggi, J., Pullar, I. A., Carney, S. L., and Bradford, H. F. (2003). Signalling pathways involved in the short-term potentiation of dopamine release by BDNF. *Brain Res.* **968**, 156–161.
- Gourion, D., Goldberger, C., Leroy, S., Bourdel, M. C., Olie, J. P., and Krebs, M. O. (2005). Age at onset of schizophrenia: Interaction between brain-derived neurotrophic factor and dopamine D3 receptor gene variants. *Neuroreport* **16**, 1407–1410.
- Gratacos, E., Perez-Navarro, E., Tolosa, E., Arenas, E., and Alberch, J. (2001). Neuroprotection of striatal neurons against kainate excitotoxicity by neurotrophins and GDNF family members. *J. Neurochem.* **78**, 1287–1296.
- Grillo, R., Ottoni, G., Leke, R., Souza, D., Portela, L., and Lara, D. (2007). Reduced BDNF levels in schizophrenic patients on clozapine or typical antipsychotics. *J. Psychiatr. Res.* **41**(1–2), 31–35.
- Grimm, J. W., Lu, L., Hayashi, T., Hope, B. T., Su, T. P., and Shaham, Y. (2003). Time-dependent increases in brain-derived neurotrophic factor protein levels within the mesolimbic dopamine system after withdrawal from cocaine: Implications for incubation of cocaine craving. *J. Neurosci.* **23**, 742–747.
- Guillin, O., Diaz, J., Carroll, P., Griffon, N., Schwartz, J. C., and Sokoloff, P. (2001). BDNF controls dopamine D3 receptor expression and triggers behavioural sensitization. *Nature* **411**, 86–89.
- Guillin, O., Griffon, N., Diaz, J., Le Foll, B., Bezard, E., Gross, C., Lammers, C., Stark, H., Carroll, P., Schwartz, J. C., and Sokoloff, P. (2004). Brain-derived neurotrophic factor and the plasticity of the mesolimbic dopamine pathway. *Int. Rev. Neurobiol.* **59**, 425–444.
- Gurevich, E. V., Bordelon, Y., Shapiro, R. M., Arnold, S. E., Gur, R. E., and Joyce, J. N. (1997). Mesolimbic dopamine D3 receptors and use of antipsychotics in patients with schizophrenia. A postmortem study. *Arch. Gen. Psychiatry* **54**, 225–232.
- Hariri, A. R., Goldberg, T. E., Mattay, V., Kolachana, B. S., Callicott, J., Egan, M. F., and Weinberger, D. (2003). Brain-derived neurotrophic factor val66met polymorphism affects human memory-related hippocampal activity and predicts memory performance. *J. Neurosci.* **23**, 6690–6694.
- Harris, S. E., Fox, H., Wright, A. F., Hayward, C., Starr, J. M., Whalley, L. J., and Deary, I. J. (2006). The brain-derived neurotrophic factor Val66Met polymorphism is associated with age-related change in reasoning skills. *Mol. Psychiatry* **11**(5), 505–513.
- Hashimoto, T., and Lewis, D. A. (2006). BDNF Val66Met polymorphism and GAD67 mRNA expression in the prefrontal cortex of subjects with schizophrenia. *Am. J. Psychiatry* **163**, 534–537.
- Hashimoto, T., Bergen, S. E., Nguyen, Q. L., Xu, B., Monteggia, L. M., Pierri, J. N., Sun, Z., Sampson, A. R., and Lewis, D. A. (2005). Relationship of brain-derived neurotrophic factor and its receptor TrkB to altered inhibitory prefrontal circuitry in schizophrenia. *J. Neurosci.* **25**, 372–383.
- Hawi, Z., Straub, R. E., O'Neill, A., Kendler, K. S., Walsh, D., and Gill, M. (1998). No linkage or linkage disequilibrium between brain-derived neurotrophic factor (BDNF) dinucleotide repeat polymorphism and schizophrenia in Irish families. *Psychiatry Res.* **81**, 111–116.
- Hong, C. J., Yu, Y. W., Lin, C. H., and Tsai, S. J. (2003). An association study of a brain-derived neurotrophic factor Val66Met polymorphism and clozapine response of schizophrenic patients. *Neurosci. Lett.* **349**, 206–208.
- Horger, B. A., Iyasere, C. A., Berhow, M. T., Messer, C. J., Nestler, E. J., and Taylor, J. R. (1999). Enhancement of locomotor activity and conditioned reward to cocaine by brain-derived neurotrophic factor. *J. Neurosci.* **19**, 4110–4122.
- Howells, D. W., Porritt, M. J., Wong, J. Y., Batchelor, P. E., Kalnins, R., Hughes, A. J., and Donnan, G. A. (2000). Reduced BDNF mRNA expression in the Parkinson's disease substantia nigra. *Exp. Neurol.* **166**, 127–135.
- Huang, T. L., and Lee, C. T. (2005). Associations between serum brain-derived neurotrophic factor levels and clinical phenotypes in schizophrenia patients. *J. Psychiatr. Res.* **40**(7), 664–668.
- Huang, Z. J., Kirkwood, A., Pizzorusso, T., Porciatti, V., Morales, B., Bear, M. F., Maffei, L., and Tonegawa, S. (1999). BDNF regulates the maturation of inhibition and the critical period of plasticity in mouse visual cortex. *Cell* **98**, 739–755.

- Hung, H. C., and Lee, E. H. (1996). The mesolimbic dopaminergic pathway is more resistant than the nigrostriatal dopaminergic pathway to MPTP and MPP⁺ toxicity: Role of BDNF gene expression. *Brain Res. Mol. Brain Res.* **41**, 14–26.
- Hyman, C., Hofer, M., Barde, Y. A., Juhasz, M., Yancopoulos, G. D., Squinto, S. P., and Lindsay, R. M. (1991). BDNF is a neurotrophic factor for dopaminergic neurons of the substantia nigra. *Nature* **350**, 230–232.
- Jockers-Scherubl, M. C., Danker-Hopfe, H., Mahlberg, R., Selig, F., Rentzsch, J., Schurer, F., Lang, U. E., and Hellweg, R. (2004). Brain-derived neurotrophic factor serum concentrations are increased in drug-naïve schizophrenic patients with chronic cannabis abuse and multiple substance abuse. *Neurosci. Lett.* **371**, 79–83.
- Kafitz, K. W., Rose, C. R., Thoenen, H., and Konnerth, A. (1999). Neurotrophin-evoked rapid excitation through TrkB receptors. *Nature* **401**, 918–921.
- Kalb, R. (2005). The protean actions of neurotrophins and their receptors on the life and death of neurons. *Trends Neurosci.* **28**, 5–11.
- Kanning, K. C., Hudson, M., Amieux, P. S., Wiley, J. C., Bothwell, M., and Schecterson, L. C. (2003). Proteolytic processing of the p75 neurotrophin receptor and two homologs generates C-terminal fragments with signaling capability. *J. Neurosci.* **23**, 5425–5436.
- Karege, F., Schwald, M., and Cisse, M. (2002). Postnatal developmental profile of brain-derived neurotrophic factor in rat brain and platelets. *Neurosci. Lett.* **328**, 261–264.
- Kerr, B. J., Bradbury, E. J., Bennett, D. L., Trivedi, P. M., Dassan, P., French, J., Shelton, D. B., McMahon, S. B., and Thompson, S. W. (1999). Brain-derived neurotrophic factor modulates nociceptive sensory inputs and NMDA-evoked responses in the rat spinal cord. *J. Neurosci.* **19**, 5138–5148.
- Koob, G. (1992). Dopamine, addiction and reward. *Semin. Neurosci.* **4**, 139–148.
- Korte, M., Carroll, P., Wolf, E., Brem, G., Thoenen, H., and Bonhoeffer, T. (1995). Hippocampal long-term potentiation is impaired in mice lacking brain-derived neurotrophic factor. *Proc. Natl. Acad. Sci. USA* **92**, 8856–8860.
- Kovalchuk, Y., Hanse, E., Kafitz, K. W., and Konnerth, A. (2002). Postsynaptic induction of BDNF-mediated long-term potentiation. *Science* **295**, 1729–1734.
- Krebs, M. O., Guillin, O., Bourdell, M. C., Schwartz, J. C., Olie, J. P., Poirier, M. F., and Sokoloff, P. (2000). Brain derived neurotrophic factor (BDNF) gene variants association with age at onset and therapeutic response in schizophrenia. *Mol. Psychiatry* **5**, 558–562.
- Kunugi, H., Ueki, A., Otsuka, M., Isse, K., Hirasawa, H., Kato, N., Nabika, T., Kobayashi, S., and Nanko, S. (2001). A novel polymorphism of the brain-derived neurotrophic factor (BDNF) gene associated with late-onset Alzheimer's disease. *Mol. Psychiatry* **6**, 83–86.
- Laruelle, M. (2000). The role of endogenous sensitization in the pathophysiology of schizophrenia: Implications from recent brain imaging studies. *Brain Res. Brain Res. Rev.* **31**, 371–384.
- Laruelle, M. (2005). Glutamate and dopamine crosstalk in schizophrenia: Insight from imaging studies. *Biol. Psychiatry* **57**, 1S.
- Lee, J. L., Everitt, B. J., and Thomas, K. L. (2004). Independent cellular processes for hippocampal memory consolidation and reconsolidation. *Science* **304**, 839–843.
- Lee, R., Kermani, P., Teng, K. K., and Hempstead, B. L. (2001). Regulation of cell survival by secreted proneurotrophins. *Science* **294**, 1945–1948.
- Levesque, D., Martres, M. P., Diaz, J., Griffon, N., Lammers, C. H., Sokoloff, P., and Schwartz, J. C. (1995). A paradoxical regulation of the dopamine D3 receptor expression suggests the involvement of an anterograde factor from dopamine neurons. *Proc. Natl. Acad. Sci. USA* **92**, 1719–1723.
- Levi-Montalcini, R., and Cohen, S. (1960). Effects of the extract of the mouse submaxillary salivary glands on the sympathetic system of mammals. *Ann. N. Y. Acad. Sci.* **85**, 324–341.
- Levivier, M., Przedborski, S., Bencsics, C., and Kang, U. J. (1995). Intrastriatal implantation of fibroblasts genetically engineered to produce brain-derived neurotrophic factor prevents

- degeneration of dopaminergic neurons in a rat model of Parkinson's disease. *J. Neurosci.* **15**, 7810–7820.
- Lewis, D. A., and Lieberman, J. (2000). Catching up on schizophrenia: Natural history and neurobiology. *Neuron* **28**, 325–334.
- Linden, A. M., Vaisanen, J., Lakso, M., Nawa, H., Wong, G., and Castren, E. (2000). Expression of neurotrophins BDNF and NT-3, and their receptors in rat brain after administration of antipsychotic and psychotropic agents. *J. Mol. Neurosci.* **14**, 27–37.
- Linnarsson, S., Bjorklund, A., and Ernfors, P. (1997). Learning deficit in BDNF mutant mice. *Eur. J. Neurosci.* **9**, 2581–2587.
- Liou, Y. J., Liao, D. L., Chen, J. Y., Wang, Y. C., Lin, C. C., Bai, Y. M., Yu, S. C., Lin, M. W., and Lai, I. C. (2004). Association analysis of the dopamine D3 receptor gene ser9gly and brain-derived neurotrophic factor gene val66met polymorphisms with antipsychotic-induced persistent tardive dyskinesia and clinical expression in Chinese schizophrenic patients. *Neuromolecular Med.* **5**, 243–251.
- Lipska, B. K., Khaing, Z. Z., Weickert, C. S., and Weinberger, D. R. (2001). BDNF mRNA expression in rat hippocampus and prefrontal cortex: Effects of neonatal ventral hippocampal damage and antipsychotic drugs. *Eur. J. Neurosci.* **14**, 135–144.
- Lohof, A. M., Ip, N. Y., and Poo, M. M. (1993). Potentiation of developing neuromuscular synapses by the neurotrophins NT-3 and BDNF. *Nature* **363**, 350–353.
- Lu, B., Pang, P. T., and Woo, N. H. (2005). The yin and yang of neurotrophin action. *Nat. Rev. Neurosci.* **6**, 603–614.
- Metsis, M., Timmusk, T., Arenas, E., and Persson, H. (1993). Differential usage of multiple brain-derived neurotrophic factor promoters in the rat brain following neuronal activation. *Proc. Natl. Acad. Sci. USA* **90**, 8802–8806.
- Minichiello, L., Korte, M., Wolf, D., Kuhn, R., Unsicker, K., Cestari, V., Rossi-Arnaud, C., Lipp, H. P., Bonhoeffer, T., and Klein, R. (1999). Essential role for TrkB receptors in hippocampus-mediated learning. *Neuron* **24**, 401–414.
- Mogi, M., Togari, A., Kondo, T., Mizuno, Y., Komure, O., Kuno, S., Ichinose, H., and Nagatsu, T. (1999). Brain-derived growth factor and nerve growth factor concentrations are decreased in the substantia nigra in Parkinson's disease. *Neurosci. Lett.* **270**, 45–48.
- Monteggia, L. M., Barrot, M., Powell, C. M., Berton, O., Galanis, V., Gemelli, T., Meuth, S., Nagy, A., Greene, R. W., and Nestler, E. J. (2004). Essential role of brain-derived neurotrophic factor in adult hippocampal function. *Proc. Natl. Acad. Sci. USA* **101**, 10827–10832.
- Muglia, P., Vicente, A. M., Verga, M., King, N., Macciardi, F., and Kennedy, J. L. (2003). Association between the BDNF gene and schizophrenia. *Mol. Psychiatry* **8**, 146–147.
- Nakao, N., Kokaia, Z., Odin, P., and Lindvall, O. (1995). Protective effects of BDNF and NT-3 but not PDGF against hypoglycemic injury to cultured striatal neurons. *Exp. Neurol.* **131**, 1–10.
- Nanko, S., Kunugi, H., Hirasawa, H., Kato, N., Nabika, T., and Kobayashi, K. (2003). Brain-derived neurotrophic factor gene and schizophrenia: Polymorphism screening and association analysis. *Schizophr. Res.* **62**, 281–283.
- Neves-Pereira, M., Cheung, J. K., Pasdar, A., Zhang, F., Breen, G., Yates, P., Sinclair, M., Crombie, C., Walker, N., and St Clair, D. M. (2005). BDNF gene is a risk factor for schizophrenia in a Scottish population. *Mol. Psychiatry* **10**, 208–212.
- Nibuya, M., Morinibu, S., and Duman, R. S. (1995). Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatment. *J. Neurosci.* **15**, 7539–7547.
- Parain, K., Murer, M. G., Yan, Q., Faucheux, B., Agid, Y., Hirsch, E., and Raisman-Vozari, R. (1999). Reduced expression of brain-derived neurotrophic factor protein in Parkinson's disease substantia nigra. *Neuroreport* **10**, 557–561.

- Patterson, S. L., Abel, T., Deuel, T. A., Martin, K. C., Rose, J. C., and Kandel, E. R. (1996). Recombinant BDNF rescues deficits in basal synaptic transmission and hippocampal LTP in BDNF knockout mice. *Neuron* **16**, 1137–1145.
- Petersen, A., Larsen, K. E., Behr, G. G., Romero, N., Przedborski, S., Brundin, P., and Sulzer, D. (2001). Expanded CAG repeats in exon 1 of the Huntington's disease gene stimulate dopamine-mediated striatal neuron autophagy and degeneration. *Hum. Mol. Genet.* **10**, 1243–1254.
- Pezawas, L., Verchinski, B., Mattay, V., Callicott, J., Kolachana, B., Straub, R. E., Egan, M. F., Meyer-Lindenberg, A., and Weinberger, D. (2004). The brain-derived neurotrophic factor val66met polymorphism and variation in human cortical morphology. *J. Neurosci.* **24**, 10099–10102.
- Pirildar, S., Gonul, A. S., Taneli, F., and Akdeniz, F. (2004). Low serum levels of brain-derived neurotrophic factor in patients with schizophrenia do not elevate after antipsychotic treatment. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **28**, 709–713.
- Pröschel, M., Saunders, A., Roses, A. D., and Muller, C. R. (1992). Dinucleotide repeat polymorphism at the human gene for the brain-derived neurotrophic factor (BDNF). *Hum. Mol. Genet.* **1**, 353.
- Rosa, A., Cuesta, M. J., Fajó-Vilas, M., Peralta, V., Zarzuela, A., and Fananas, L. (2006). The Val66Met polymorphism of the brain-derived neurotrophic factor gene is associated with risk factor for psychosis: Evidence from a family-based association study. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **141**, 135–138.
- Rybakowski, J. K., Borkowska, A., Skibinska, M., Szczepankiewicz, A., Kapelski, P., Leszczynska-Rodziewicz, A., Czernski, P. M., and Hauser, J. (2006). Prefrontal cognition in schizophrenia and bipolar illness in relation to Val66Met polymorphism of the brain-derived neurotrophic factor gene. *Psychiatry Clin. Neurosci.* **60**, 70–76.
- Sabate, O., Campion, D., d'Amato, T., Martres, M. P., Sokoloff, P., Giros, B., Leboyer, M., Jay, M., Guedj, F., Thibaut, F., *et al.* (1994). Failure to find evidence for linkage or association between the dopamine D3 receptor gene and schizophrenia. *Am. J. Psychiatry* **151**, 107–111.
- Sasaki, T., Dai, X. Y., Kuwata, S., Fukuda, R., Kunugi, H., Hattori, M., and Nanko, S. (1997). Brain-derived neurotrophic factor gene and schizophrenia in Japanese subjects. *Am. J. Med. Genet.* **74**, 443–444.
- Seroogy, K. B., Lundgren, K. H., Tran, T. M., Guthrie, K. M., Isackson, P. J., and Gall, C. M. (1994). Dopaminergic neurons in rat ventral midbrain express brain-derived neurotrophic factor and neurotrophin-3 mRNAs. *J. Comp. Neurol.* **342**, 321–334.
- Shen, R. Y., Altar, C. A., and Chiodo, L. A. (1994). Brain-derived neurotrophic factor increases the electrical activity of pars compacta dopamine neurons *in vivo*. *Proc. Natl. Acad. Sci. USA* **91**, 8920–8924.
- Shimizu, E., Hashimoto, K., Watanabe, H., Komatsu, N., Okamura, N., Koike, K., Shinoda, N., Nakazato, M., Kumakiri, C., Okada, S., and Iyo, M. (2003). Serum brain-derived neurotrophic factor (BDNF) levels in schizophrenia are indistinguishable from controls. *Neurosci. Lett.* **351**, 111–114.
- Shults, C. W., Kimber, T., and Altar, C. A. (1995). BDNF attenuates the effects of intrastriatal injection of 6-hydroxydopamine. *Neuroreport* **6**, 1109–1112.
- Sklair-Tavron, L., Shi, W.-X., Lane, S., Harris, H. W., Bunney, B. S., and Nestler, E. J. (1996). Chronic morphine induces visible changes in the morphology of mesolimbic dopamine neurons. *Proc. Natl. Acad. Sci. USA* **93**, 11202–11207.
- Sokoloff, P., Giros, B., Martres, M. P., Bouthenet, M. L., and Schwartz, J. C. (1990). Molecular cloning and characterization of a novel dopamine receptor (D3) as a target for neuroleptics. *Nature* **347**, 146–151.
- Sokoloff, P., Diaz, G., Le Foll, B., Guillin, O., Leriche, L., Bezard, E., and Gross, C. (2006). The dopamine D3 receptor: A target for the treatment of neuropsychiatric disorders. *CNS Neurol. Disord. Drug Targets* **5**, 25–43.

- Szczepankiewicz, A., Skibinska, M., Czernski, P. M., Kapelski, P., Leszczynska-Rodziewicz, A., Slopian, A., Dmitrzak-Weglarczyk, M., Rybakowski, F., Rybakowski, J. K., and Hauser, J. (2005). No association of the brain-derived neurotrophic factor (BDNF) gene C-270T polymorphism with schizophrenia. *Schizophr. Res.* **76**, 187–193.
- Szekeres, G., Juhasz, A., Rimanoczy, A., Keri, S., and Janka, Z. (2003). The C270T polymorphism of the brain-derived neurotrophic factor gene is associated with schizophrenia. *Schizophr. Res.* **65**, 15–18.
- Takahashi, M., Shirakawa, O., Toyooka, K., Kitamura, N., Hashimoto, T., Maeda, K., Koizumi, S., Wakabayashi, K., Takahashi, H., Someya, T., and Nawa, H. (2000). Abnormal expression of brain-derived neurotrophic factor and its receptor in the corticolimbic system of schizophrenic patients. *Mol. Psychiatry* **5**, 293–300.
- Tan, Y. L., Zhou, D. F., Cao, L. Y., Zou, Y. Z., Wu, G. Y., and Zhang, X. Y. (2005a). Effect of the BDNF Val66Met genotype on episodic memory in schizophrenia. *Schizophr. Res.* **77**, 355–356.
- Tan, Y. L., Zhou, D. F., Cao, L. Y., Zou, Y. Z., and Zhang, X. Y. (2005b). Decreased BDNF in serum of patients with chronic schizophrenia on long-term treatment with antipsychotics. *Neurosci. Lett.* **382**, 27–32.
- Tan, Y. L., Zhou, D. F., and Zhang, X. Y. (2005c). Decreased plasma brain-derived neurotrophic factor levels in schizophrenic patients with tardive dyskinesia: Association with dyskinetic movements. *Schizophr. Res.* **74**, 263–270.
- Thoenen, H. (1995). Neurotrophins and neuronal plasticity. *Science* **270**, 593–598.
- Timmusk, T., Palm, K., Metsis, M., Reintam, T., Paalme, V., Saarma, M., and Persson, H. (1993). Multiple promoters direct tissue-specific expression of the rat BDNF gene. *Neuron* **10**, 475–489.
- Toyooka, K., Asama, K., Watanabe, Y., Muratake, T., Takahashi, M., Someya, T., and Nawa, H. (2002). Decreased levels of brain-derived neurotrophic factor in serum of chronic schizophrenic patients. *Psychiatry Res.* **110**, 249–257.
- Vicario-Abejon, C., Collin, C., McKay, R. D., and Segal, M. (1998). Neurotrophins induce formation of functional excitatory and inhibitory synapses between cultured hippocampal neurons. *J. Neurosci.* **18**, 7256–7271.
- Virgos, C., Martorell, L., Valero, J., Figuera, L., Civeira, F., Joven, J., Labad, A., and Vilella, E. (2001). Association study of schizophrenia with polymorphisms at six candidate genes. *Schizophr. Res.* **49**, 65–71.
- Volk, D., and Lewis, D. A. (2002). Impaired prefrontal inhibition in schizophrenia: Relevance for cognitive dysfunction. *Physiol. Behav.* **77**, 501–505.
- von Bartheld, C. S., Byers, M. R., Williams, R., and Bothwell, M. (Bartheld 1996). Anterograde transport of neurotrophins and axodendritic transfer in the developing visual system. *Nature* **379**, 830–833.
- Wassink, T. H., Nelson, J. J., Crowe, R. R., and Andreasen, N. C. (1999). Heritability of BDNF alleles and their effect on brain morphology in schizophrenia. *Am. J. Med. Genet.* **88**, 724–728.
- Weickert, C. S., Hyde, T. M., Lipska, B. K., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2003). Reduced brain-derived neurotrophic factor in prefrontal cortex of patients with schizophrenia. *Mol. Psychiatry* **8**, 592–610.
- Weickert, C. S., Liggins, D. L., Romanczyk, T., Ungaro, G., Hyde, T. M., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2005). Reductions in neurotrophin receptor mRNAs in the prefrontal cortex of patients with schizophrenia. *Mol. Psychiatry* **10**, 637–650.
- Woo, N. H., Teng, H. K., Siao, C. J., Chiaruttini, C., Pang, P. T., Milner, T. A., Hempstead, B. L., and Lu, B. (2005). Activation of p75NTR by proBDNF facilitates hippocampal long-term depression. *Nat. Neurosci.* **8**, 1069–1077.
- Zuch, C., David, D., Ujhelyi, L., Hudson, J., Gerhardt, G., Kaplan, P., and Bickford, P. (2004). Beneficial effect of intraventricular administered BMP-7 following 6-hydroxydopamine lesion. *Brain Res.* **1010**, 10–16.

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SCHIZOPHRENIA SUSCEPTIBILITY GENES: IN SEARCH OF A MOLECULAR LOGIC AND NOVEL DRUG TARGETS FOR A DEVASTATING DISORDER

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- I. The Genetic Component of Schizophrenia
- II. Genes Identified Through Systematic Follow-Up of Linkage Signals
 - A. Proline Dehydrogenase
 - B. Dystrobrevin-Binding Protein 1
 - C. Neuregulin 1
 - D. G72
 - E. Disrupted in Schizophrenia 1
 - F. Carboxyl-Terminal PDZ Ligand of Neuronal Nitric Oxide Synthase
 - G. ZDHHC8
 - H. Trace Amine Receptor 6
 - I. Epsin 4
 - J. (GABA)_A Receptor Subunit Gene Cluster
- III. Other Candidate Genes
 - A. Catechol-*O*-Methyltransferase
 - B. Regulator of G-Protein Signaling 4
 - C. Calcineurin Gamma Catalytic Subunit
 - D. AKT1
- IV. Areas of Caution in the Interpretation and Generalization of Genetic Findings
- V. Future Directions of the Genetic Research: Advancing Our Understanding of How the Specific Genetic Factors Contribute Biologically to the Disease Process
 - A. Animal Models
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Schizophrenia is a devastating psychiatric disorder that affects approximately one percent of the population worldwide. We argue that the efforts to decipher the genetic causes of schizophrenia have reached another turning point and describe evidence supporting some of the major recent genetic findings in the field. In addition, we identify some general areas of caution in the interpretation of these findings and addresses the promise this recently acquired knowledge holds for the generation of reliable animal models, characterization of genetic

interactions, dissection of the disease pathophysiology and development of novel, mechanism-based treatments for the patients.

I. The Genetic Component of Schizophrenia

Schizophrenia is a severe psychiatric disorder with a lifetime prevalence of ~1% in most studied populations (Karayiorgou and Gogos, 1997). Schizophrenia is characterized by so called “positive symptoms” including delusions and hallucinations, “negative symptoms” including blunted emotions and social isolation, as well as by cognitive deficits. Although cognitive impairment has always been regarded as a hallmark feature of schizophrenia, only recently it has been recognized as an enduring, core deficit, a strong indicator of specific genetic liability to the disease and a primary target for pharmacotherapy. Individuals with schizophrenia show varying degrees of deficiency in a diverse range of cognitive domains such as working memory, short-term and episodic memory, attention, executive functions, and learning (Bowie and Harvey, 2005; Goldman-Rakic, 1994; Green *et al.*, 2004).

Similar to many common, complex disorders, schizophrenia is a multifactorial disorder characterized by the contribution of multiple risk genes, which could act in conjunction with epigenetic and environmental processes (Karayiorgou and Gogos, 1997). More than 20 genome-wide scans aiming to localize genes for this disorder have been reported to date and two meta-analyses (Badner and Gershon, 2002; Lewis *et al.*, 2003) implicated under moderate stringency, ~12 regions of the genome as likely to contain schizophrenia susceptibility genes (2p, 5q, 3p, 11q, 2q, 1q, 22q, 8p, 6p, 20p, 13q, and 14q). This is most likely to be an underestimate because it is expected that many schizophrenia susceptibility genes will be undetectable by traditional linkage studies. The ultimate validation of the linkage results is gene identification, which in turn represents an important milestone for understanding the disease pathophysiology. Gene identification has proven to be an extraordinarily difficult task, partly because no single gene is necessary or sufficient to cause the disease but instead, many susceptibility genes with small effects act in combinations to increase the risk of illness. Phenotypic heterogeneity has also contributed to the difficulties associated with genetic research in schizophrenia. Phenotypic heterogeneity is to be expected due to the complexity of the brain, but the majority of genetic studies by relying on a categorical binary diagnosis (“affected” vs “unaffected”) do not take into account the possible differences in representation among different samples of the various components of the illness.

Nevertheless, in the past 4 years, significant advances in gene discovery have taken place driven by the completion of the sequencing of the human genome, the increasingly available technology for high-throughput genomic analysis, and the development of new analytical and bioinformatics tools. Several new susceptibility

genes have been proposed, each supported by varying degrees of evidence. Candidate genes have been identified, for the most part, through systematic follow-up of linkage signals involving genotyping of relatively large numbers of markers, including single nucleotide polymorphisms (SNPs) and linkage disequilibrium (LD) assays or through multipronged candidate gene approaches involving analysis of expression patterns and biological functions. Far from being a mere academic exercise, this newly acquired knowledge base provides a novel framework for mechanism-based drug discovery efforts. In some cases, susceptibility genes could themselves provide new drug targets. Alternatively, the identification of these genes will lead to improved understanding of the basis of schizophrenia pathogenesis and eventual detailed characterization of the affected molecular pathways.

Here, we discuss the available genetic and biological data behind these strong candidate genes, the statistical support for these findings, and future directions of the genetic research. In this context, we discuss the development of animal models, the characterization of susceptibility gene interactions, the understanding of the disease pathophysiology, and the development of mechanism-based therapies.

II. Genes Identified Through Systematic Follow-Up of Linkage Signals

In this section, we discuss the genetic data for recently identified strong positional candidate genes (in chronological order of appearance of the reports), as well as their possible biological functions. The first report of a strong positional candidate schizophrenia gene identified by a systematic fine-mapping approach within a region implicated by linkage analysis was published in 2002 (Liu *et al.*, 2002a) followed in the same year by three additional reports describing new susceptibility genes identified through similar approaches (Chumakov *et al.*, 2002; Stefansson *et al.*, 2002; Straub *et al.*, 2002). Additional genes have been reported based on systematic follow-up analysis of linkage peaks (Brzustowicz *et al.*, 2004; Duan *et al.*, 2004; Hennah *et al.*, 2003; Mukai *et al.*, 2004; Petryshen *et al.*, 2005a; Pimm *et al.*, 2005; Table I).

A. PROLINE DEHYDROGENASE

The gene is located on chromosome 22q11, a region implicated by some linkage studies (Badner and Gershon, 2002; Lewis *et al.*, 2003) and also frequently deleted in patients with schizophrenia (Karayiorgou *et al.*, 1995). Several studies have now established conclusively that the risk of schizophrenia for a patient with a 22q11 microdeletion is ~25–31 times the general population risk of 1% (Murphy *et al.*, 1999; Pulver *et al.*, 1994) and that the rate of 22q11 microdeletions

TABLE I
SCHIZOPHRENIA CANDIDATE GENES: CHROMOSOMAL LOCATIONS AND POTENTIAL FUNCTION

Gene symbol	Locus	Function	References
<i>PRODH</i>	22q11	L-Proline metabolism; influence on glutamatergic transmission, mitochondrial function	Liu <i>et al.</i> (2002a)
<i>DTNBP1</i>	6p22	Member of DPC and biogenesis of lysosomal related organelle complex; potential presynaptic effects on glutamate release	Straub <i>et al.</i> (2002)
<i>NRG1</i>	8p12	Broad involvement in neuronal function and survival	Stefansson <i>et al.</i> (2002)
<i>G72</i>	13q34	Potential activation of D-amino acid oxidase and indirect effects on glutamatergic signaling	Chumakov <i>et al.</i> (2002)
<i>DISC1</i>	1q42	Multifunctional; possible involvement in cell migration and phosphodiesterase signaling	Millar <i>et al.</i> (2000), Hennah <i>et al.</i> (2003)
<i>CAPOX</i>	1q22	Potential regulator of NMDA receptor-coupled nitric oxide signaling	Brzustowicz <i>et al.</i> (2004)
<i>ζDHHCB</i>	22q11	Palmitoylation of PSD-95 and other substrates; implications for synaptic assembly and function	Liu <i>et al.</i> (2002b), Mukai <i>et al.</i> (2004)
<i>TAAR6</i>	6q23	G-protein-coupled receptor for trace amines	Duan <i>et al.</i> (2004)
<i>EPN4</i>	5q33	Potential role in reuptake and storage of neurotransmitters	Pimm <i>et al.</i> (2005)
<i>GABA_A</i> receptors	5q34	GABAergic transmission	Petryshen <i>et al.</i> (2005a)
<i>COMT</i>	22q11	Regulation of extracellular dopamine levels in prefrontal cortex	Egan <i>et al.</i> (2001), Shifman <i>et al.</i> (2002), Paterlini <i>et al.</i> (2005)
<i>RGS4</i>	1q23	Regulator of signal transduction via dopamine, metabotropic glutamate, and muscarinic receptors	Chowdari <i>et al.</i> (2002)
<i>PPP3CC</i>	8p21	Subunit-specific function unknown; potential involvement in synaptic plasticity and D1 receptor signaling	Gerber <i>et al.</i> (2003)
<i>AKT1</i>	14q32	Multifunctional; possible involvement in D2 and GABA _B receptor signaling	Emamian <i>et al.</i> (2004)

in schizophrenia, although relatively low, is ~ 12 – 80 times the estimated general population rate (Karayiorgou *et al.*, 1995). Individual genes from this locus have been examined in systematic fine-mapping efforts (Karayiorgou and Gogos, 2004). LD analysis using 72 SNPs in family samples identified an over-transmission of a haplotypic variant located at the 3' end of the proline dehydrogenase (*PRODH*) gene (Liu *et al.*, 2002a,b). This finding was replicated in 3 independent family samples, including a very large collection of 528 families from China (Li *et al.*, 2004a) and 274 families of Ashkenazi Jewish origin (Fallin *et al.*, 2005). One negative family study has also been reported (Williams *et al.*, 2003a). In addition, 3' end variants of the gene were also identified as a risk factor for development of psychotic symptoms during adolescence in children with 22q11 microdeletions (Gothelf *et al.*, 2005). The implicated variants are consistently located at the 3' end of the gene, but their functional consequences are still unknown. However, the Liu *et al.* (2002a) study identified additional rare variants of the *PRODH* gene, which are present either exclusively or in higher frequencies in schizophrenic patients, are generated through gene conversion from a nearby pseudogene (Liu *et al.*, 2002a) and affect highly conserved amino acids leading to drastic reductions in enzymatic activity (Bender *et al.*, 2005). The same variants were described in schizophrenic patients in an independent study, which also identified a small deletion encompassing the *PRODH* gene (and its neighboring gene *DGCR6*) in a schizophrenic patient (Jacquet *et al.*, 2002). *PRODH* encodes an enzyme that metabolizes L-proline, a putative neuromodulatory amino acid that could directly influence glutamatergic transmission, which is believed to play a central role in the pathophysiology of schizophrenia (Paterlini *et al.*, 2005). A mutation in the mouse orthologue of the human *PRODH* gene in the Pro/Re hyperprolinemic mouse strain has been described (Gogos *et al.*, 1999). These mice demonstrate an increased neurotransmitter release and abnormal plasticity at glutamatergic synapses, as well as distinct abnormalities in dopamine turnover and signaling in the frontal cortex (Paterlini *et al.*, 2005) reminiscent of schizophrenia in humans.

B. DYSTROBREVIN-BINDING PROTEIN 1

Fine-mapping efforts undertaken as a follow-up to evidence for linkage on chromosome 6p24–22 in a sample of Irish families, led to identification of dystrobrevin-binding protein 1 (*DTNBPI*) gene (dysbindin; Straub *et al.*, 2002) as a schizophrenia candidate gene. Several replication studies have been reported but most replication samples used ($N = 9$) were case-control samples (Funke *et al.*, 2004; Morris *et al.*, 2003a; Numakawa *et al.*, 2004; Van Den Bogaert *et al.*, 2003; Williams *et al.*, 2004). Replication of this association has also been attempted in seven family samples, with replications observed in five of them (Fallin *et al.*, 2005;

Hall *et al.*, 2004; Kirov *et al.*, 2004; Schwab *et al.*, 2003; Tang *et al.*, 2003). In the positive studies, there are inconsistencies among the implicated alleles or haplotypes. These inconsistencies may be a product of population stratification or multiple testing. Alternatively, they could be explained by presence of distinct variations affecting different functional elements within the gene that have emerged independently on a more recent ancestral background. In addition, as this locus has not been extensively analyzed outside the confines of the *DTNBP1* gene, the possibility of a neighboring gene as a sole or partial source of the association signal cannot be excluded. Initial expression and functional studies provide some additional support for a role of *DTNBP1* in schizophrenia. *DTNBP1* has a widespread distribution in the brain, including expression in pyramidal neurons in the hippocampus and dorsal lateral prefrontal cortex (DLPFC). *DTNBP1* expression appears to be decreased in schizophrenia in both DLPFC and excitatory pathways of hippocampus (Talbot *et al.*, 2004; Weickert *et al.*, 2004). Interestingly, a substantial fraction of *DTNBP1* is presynaptically localized, and preliminary *in vitro* evidence suggests that knockdown of endogenous dysbindin protein results in the reduction of glutamate release, suggesting that dysbindin might influence exocytotic glutamate release (Numakawa *et al.*, 2004). *DTNBP1* is a member of the biogenesis of lysosome-related organelles complex (BLOC; Li *et al.*, 2003), as well as the dystrophin protein complex (DPC; Benson *et al.*, 2001).

C. NEUREGULIN 1

A broad region on chromosome 8p12-21 has been implicated in schizophrenia by multiple linkage studies, including a study of 33 extended Icelandic families. Fine mapping in the same set of families detected an association between schizophrenia and several haplotypes at the neuregulin 1 (*NRG1*) locus. A core haplotype at the 5' end of the gene comprising several markers within a 290-kb block of LD showed highly significant association with schizophrenia (Stefansson *et al.*, 2002). Several replication studies have been reported. Eight of the replication samples used were case-control samples (Corvin *et al.*, 2004; Iwata *et al.*, 2004; Li *et al.*, 2004b; Petryshen *et al.*, 2005b; Stefansson *et al.*, 2003; Tang *et al.*, 2004; Williams *et al.*, 2003b; Zhao *et al.*, 2004) and eight family samples. In the family samples, less than half show some evidence for association, but often with haplotypes other than the one originally described (Duan *et al.*, 2005a; Fallin *et al.*, 2005; Hall *et al.*, 2004; Li *et al.*, 2004b; Petryshen *et al.*, 2005b; Thiselton *et al.*, 2004; Yang *et al.*, 2003; Zhao *et al.*, 2004). Dramatic differences in the frequency of haplotypes reported between different samples (ranging from 1% to 10%; Li *et al.*, 2004b; Zhao *et al.*, 2004) could indicate either substantial heterogeneity in the LD structure across the *NRG1* locus or presence of multiple risk

alleles. In the absence of any functional significance for any of the implicated haplotypes, it is difficult to interpret further the genetic data that is published at the time of this writing. The *NRG1* gene encodes a well-characterized protein involved in a wide variety of neuronal functions, ranging from neuronal survival to myelination and synaptic plasticity (Corfas *et al.*, 2004).

D. G72

A strong and consistent linkage signal for both schizophrenia and bipolar disorder has been identified on chromosome 13q32–34 (Blouin *et al.*, 1998; Detera-Wadleigh *et al.*, 1999) and has prompted fine-mapping efforts. Significant association with schizophrenia was described for several SNPs and haplotypes at the G72 locus in a French-Canadian case-control sample. The association for two SNPs was replicated in a Russian case-control cohort (Chumakov *et al.*, 2002). Consistent with the linkage studies results, an association between variants at the G72 locus and bipolar disorder has also been described (Hattori *et al.*, 2003). G72 association with schizophrenia has been observed in several additional samples including case-control (Korostishevsky *et al.*, 2004; Schumacher *et al.*, 2004; Wang *et al.*, 2004) and family-based samples (Addington *et al.*, 2004; Zou *et al.*, 2005) with evidence for allelic heterogeneity. Negative studies have also been reported (Mulle *et al.*, 2005). Expression and functional studies suggested a potential interaction of G72 with D-amino acid oxidase that modulates its enzymatic activity and thus could indirectly affect glutamatergic signaling (Chumakov *et al.*, 2002; Mothet *et al.*, 2000). However, this interaction remains to be demonstrated *in vivo*.

E. DISRUPTED IN SCHIZOPHRENIA 1

Disrupted in schizophrenia 1 (*DISC1*) is one of two genes isolated from a chromosome 1q42 translocation breakpoint previously shown to segregate with psychopathology in a large Scottish family. The other gene is *DISC2* and is a noncoding, presumably regulatory RNA (Millar *et al.*, 2000). *DISC1* was originally described 5 years ago, but interest in it was renewed only recently when large-scale linkage (Ekelund *et al.*, 2001, 2004) and follow-up systematic association studies in Finnish families identified *DISC1* as a positional candidate from the 1q42 locus (Hennah *et al.*, 2003). *DISC1* association with schizophrenia has been observed in some additional samples with evidence for allelic heterogeneity, but negative studies have also been reported (Fallin *et al.*, 2005; Hennah *et al.*, 2003; Hodgkinson *et al.*, 2004). *DISC1* association with schizophrenia-related endophenotypes has been also reported. In one preliminary imaging study, variation in the *DISC1* gene

was associated with altered hippocampal structure and function in healthy subjects (Callicott *et al.*, 2005). An independent study implicated *DISC1* variation in visual working memory performance (Hennah *et al.*, 2005). A family afflicted with schizophrenia and schizoaffective disorder was shown to segregate a rare frame-shift variant of the gene (Sachs *et al.*, 2005). *DISC1* is a complex gene with poorly understood involvement in development and synaptic plasticity. It is associated with numerous cytoskeletal proteins, and it could be involved in centrosomal and microtubule function, cell migration, neurite outgrowth, membrane trafficking of receptors, mitochondrial function, and possibly phosphodiesterase signaling (Morris *et al.*, 2003b).

F. CARBOXYL-TERMINAL PDZ LIGAND OF NEURONAL NITRIC OXIDE SYNTHASE

Brzustowicz *et al.* (2000) have previously reported evidence for linkage at 1q22. Fine mapping using 14 microsatellite markers and 15 SNPs from a subregion of the linkage locus (Brzustowicz *et al.*, 2004) produced nominally significant evidence of LD between schizophrenia and a subset of markers located within the genomic region of carboxyl-terminal PDZ ligand of neuronal nitric oxide synthase (*CAPON*). Abnormal expression pattern of this gene was observed in brains from individuals with schizophrenia (Xu *et al.*, 2005) making it a prime positional candidate from the schizophrenia susceptibility locus on 1q22. Two case-control replication studies (one positive and one negative) have been reported (Puri *et al.*, 2005; Zheng *et al.*, 2005). *CAPON* is involved in NMDA receptor-coupled nitric oxide signaling (Jaffrey *et al.*, 1998).

G. ZDHHC8

Involvement of this gene was identified in the same LD screen of the 22q11 locus that led to the discovery of the *PRODH*-schizophrenia association (Liu *et al.*, 2002a,b). It was shown that one of the *ZDHHC8* risk alleles (at SNP rs175174), located in intron 4, affects the ratio of an intron 4-containing unspliced form (that encodes a putative truncated form of the protein) over the fully spliced active form (Mukai *et al.*, 2004). The presence of the risk allele rs175174-A results in the production of relatively higher levels (~25%) of the unspliced inactive form (Mukai *et al.*, 2004). Other variants of the gene (affecting distinct aspects of its complex splicing or its expression level) might modulate the disease risk in other nondeleted patient samples. One positive and one negative family-based study of nondeleted patients have been reported so far (Chen *et al.*, 2004a; Glaser *et al.*, 2005). The general involvement of this gene in schizophrenia awaits analysis of additional family samples, but the effect of the gene is predicted to be much stronger in individuals with 22q11 deletions and schizophrenia, where a 50%

(or ~65% when the nondeleted allele carries the risk SNP rs175174 variant) decrease in ZDHHC8 activity levels is predicted. *ZDHHC8* is predicted to encode a transmembrane palmitoyltransferase that modifies PSD-95 among other targets and could play an important role in excitatory synaptic transmission (el-Husseini and Bredt, 2002).

H. TRACE AMINE RECEPTOR 6

A broad area on chromosome 6q (6q13-26) has been implicated in schizophrenia in linkage studies using European ancestry and African-American schizophrenia pedigrees (Levinson *et al.*, 2000). Two-stage SNP-based fine-mapping efforts focusing on band q23.2 identified trace amine receptor 6 (*TAAR6*) as a prime positional candidate (Duan *et al.*, 2004) for the schizophrenia susceptibility locus on 6q23.2. Two negative replication studies have been reported (Duan *et al.*, 2005b; Ikeda *et al.*, 2005). An independent study implicated the trace amine receptor genes at 6q23.2 in susceptibility to bipolar disorder (Abou Jamra *et al.*, 2005). *TAAR6* is a GPCR widely expressed in the brain (Borowsky *et al.*, 2001).

I. EPSIN 4

Chromosome 5q33 is a region that has previously shown strong evidence of linkage to schizophrenia in four independent linkage studies. Four adjacent markers (and associated haplotypes) at the 5' end of the *Epsin 4* (*EPN4*) gene, which is located in this region, showed significant evidence of LD with schizophrenia in a case-control fine-mapping study (Pimm *et al.*, 2005). The *Epsin 4* gene encodes the clathrin-associated protein enthoprotin, which has a role in transport and stability of neurotransmitter vesicles at the synapses and within neurons. No replication studies (especially family based) have been reported yet.

J. (GABA)_A RECEPTOR SUBUNIT GENE CLUSTER

Chromosome 5q31-35 was implicated in Portuguese schizophrenia families (Sklar *et al.*, 2004) and was supported by subsequent meta-analysis. A group of γ -aminobutyric acid (GABA)_A receptor subunit genes (*GABRA1*, *GABRA6*, *GABRB2*, *GABRG2*, and *GABRP*) that map within this linkage peak were examined in Portuguese patients, and associations with SNPs and haplotypes in *GABRA1*, *GABRP*, and *GABRA6* were detected (Petryshen *et al.*, 2005a). The *GABRA1* and *GABRP* findings were replicated in an independent German family-based sample (Petryshen *et al.*, 2005a). These genes are plausible candidates based on prior evidence for GABA system involvement in schizophrenia (Lewis *et al.*, 2005).

III. Other Candidate Genes

The candidacy of the genes described in this section is based on convergent genetic and biological evidence rather than on positional cloning. Interestingly, many of these genes are located in the general vicinity of linkage signals. Although far from proven, the recurrent observation of clustering of candidate susceptibility genes could indicate that more than one gene could contribute to at least some of the linkage signals in psychiatric disorders.

A. CATECHOL-*O*-METHYLTRANSFERASE

The gene is located in the 22q11 region between the *PRODH* and *2DHHC8* genes. In addition to being a positional candidate gene, catechol-*O*-methyltransferase (*COMT*) is also an attractive functional candidate gene since it is involved in the breakdown of dopamine. One variant in particular, in codon 158 that affects enzymatic activity depending on presence of Val (high activity) or Met (low activity), has been examined extensively in studies testing directly for association with schizophrenia. It has been proposed that the high-activity Val allele increases the risk for schizophrenia but the genetic association results are equivocal (Fan *et al.*, 2005; Glatt *et al.*, 2003; Lohmueller *et al.*, 2003; Munafo *et al.*, 2005; Shifman *et al.*, 2002; Tsai *et al.*, 2006; Williams *et al.*, 2005). The same allele was shown in some studies to impair executive function, which is affected in schizophrenic patients (Egan *et al.*, 2001; Ho *et al.*, 2005). Studies in animal models, however, suggested that low activity of this enzyme could be a risk factor for schizophrenia by failing to buffer the effect of other primary mutations that affect dopamine turnover and signaling in the cortex (Paterlini *et al.*, 2005). This prediction was supported by the results of a longitudinal follow-up study of children with 22q11 microdeletions, which revealed that the low-activity form of the enzyme (Met158) is a risk factor for decline in prefrontal cortical volume and cognition, as well as for the consequent development of psychotic symptoms during adolescence, in these children (Gothelf *et al.*, 2005). Therefore, the contribution of *COMT* to schizophrenia, in general, is likely to be complex.

B. REGULATOR OF G-PROTEIN SIGNALING 4

The gene maps to 1q21–22, 0.7 Mb from *CAPON* (see Section II). Regulator of G-protein signaling 4 (*RGS4*) was initially identified as the only transcript (out of 7800 sampled by Mirnics *et al.*, 2000) consistently reduced in the DLPFC of individuals with schizophrenia. Subsequently, Chowdari *et al.* (2002) genotyped 13 SNPs across a 300-kb segment spanning the gene in several independent

datasets and found weak evidence for association with schizophrenia in each of the samples within a haplotype block stretching from intron 1 to several kb upstream of the transcription start site. However, the pattern of allelic association was not consistent among samples. Independent replication efforts have been reported including both positive and negative studies (Chen *et al.*, 2004b; Fallin *et al.*, 2005; Sobell *et al.*, 2005; Zhang *et al.*, 2005). *RGS4* is one of 19 human *RGS* transcripts and is abundant in the cerebral cortex (Larminie *et al.*, 2004). *RGS4* encodes for a GTPase activator, which desensitizes Gi/Go and Gq, thus negatively modulating G-protein-mediated signaling via dopamine, metabotropic glutamate, and muscarinic receptors (Ross and Wilkie, 2000).

C. CALCINEURIN GAMMA CATALYTIC SUBUNIT

PPP3CC which encodes the calcineurin gamma catalytic subunit is located at 8p21.3, 10 Mb from *NRG1*, but adjacent to previously described linkage signals (Gerber *et al.*, 2003). Forebrain-specific calcineurin knockout mice were reported to have a spectrum of behavioral abnormalities related to altered behaviors observed in schizophrenia patients (Miyakawa *et al.*, 2003) supporting the proposal that alterations in calcineurin signaling contribute to schizophrenia pathogenesis. In further support of this proposal, *PPP3CC* was found to be downregulated in the hippocampus of individuals with schizophrenia (Eastwood, 2005). The genetic association was not replicated, however, in a sample of Ashkenazi Jewish nuclear families (Fallin *et al.*, 2005). Calcineurin is a multifunctional calcium-dependent serine/threonine phosphatase, known to be centrally involved in many aspects of synaptic plasticity. It has particular roles in glutamate and dopamine signaling and their interactions, including regulation of DARPP32, a molecular node of convergence between dopamine receptor 1 and NMDA receptor signaling pathways (Miyakawa *et al.*, 2003; Winder and Sweatt, 2001).

D. AKT1

AKT-GSK3 β signaling is a target of lithium and as such has been implicated in the pathogenesis of mood disorders. Evidence was provided that this signaling pathway also has a role in schizophrenia (Emamian *et al.*, 2004), including convergent evidence for a decrease in AKT1 protein levels and levels of substrate phosphorylation in the peripheral lymphocytes and brains of individuals with schizophrenia; a significant association between schizophrenia and an *Akt1* haplotype associated with lower AKT1 protein levels; and a greater sensitivity to the sensorimotor gating-disruptive effect of amphetamine, conferred by AKT1 deficiency. The genetic association has been confirmed thus far in two

independent populations, namely in a combined sample of ~1000 cases and 1000 controls from Japan (Ikeda *et al.*, 2004; Ohtsuki *et al.*, 2004) and in a sample of European sib-pair families (Schwab *et al.*, 2005). Dopamine plays an important role in the etiology of schizophrenia, and despite claims that most behavioral actions of DA are associated with the modulation of adenylate cyclase and PKA activity (Greengard, 2001), investigations have uncovered that stimulation of D2 class receptors also results in a adenosine 3',5'-monophosphate (cAMP)-independent dephosphorylation/inactivation of Akt (Beaulieu *et al.*, 2004) associated with the expression of DA-dependent behaviors (Beaulieu *et al.*, 2004; Emamian *et al.*, 2004). These novel results suggest a model in which both cAMP-dependent and cAMP-independent events play important and perhaps cooperative functions mediate schizophrenia-related DA actions. Consistent with this model, administration of haloperidol or the selective D2 class receptor antagonist raclopride has been shown to prevent the regulation of Akt by DA or enhance Akt phosphorylation in animal models (Beaulieu *et al.*, 2004; Emamian *et al.*, 2004) and thus, in principle, could compensate for an impaired function of this signaling pathway in schizophrenia. Moreover, two other drugs used in the management of psychosis, lithium, and clozapine also act as enhancers of Akt signaling *in vivo* (Beaulieu *et al.*, 2004) or *in vitro* (Chalecka-Franaszek and Chuang, 1999; Kang *et al.*, 2004). Akt signaling has also been implicated in GABAergic transmission (Wang *et al.*, 2003) and outside the CNS has been implicated in the regulation of multiple biological processes ranging from glycogenesis to embryonic development, apoptosis, and cell proliferation (Scheid and Woodgett, 2001).

IV. Areas of Caution in the Interpretation and Generalization of Genetic Findings

A combination of criteria that include the degree of statistical significance, the reproducibility of the associations in independent samples, the identification of independent rare risk alleles, and the consistent findings from animal model studies and endophenotype-based studies in humans need to be considered in assessing the degree of confidence assigned to each of the findings outlined in the sections above. On the basis of these criteria, support for at least some of the findings described above (such as *PRODH*, *DTNBP1*, *NRG1*, *G72*, *DISC1*, or *COMT*) appears to be quite strong. However, it should be emphasized that, even for the stronger findings, it is too early to draw firm conclusions about their generalization among different samples and populations primarily due to uncertainties pertaining to the extent of coverage of the implicated loci, consistency of the risk allele or risk haplotype across studies, the structure of the samples used in the original and replication studies, publication bias against negative reports, and supporting biological data. Because of these issues, claims of "replication" should

be taken with caution and the reader is encouraged to undertake a careful analysis of the properties of the employed samples and the methods used in order to determine the validity of such claims.

What are the general areas of caution in the interpretation of these findings? In general, the statistical burden of proof is lower for genes identified through systematic follow-up of linkage signals compared to genes picked in essentially random fashion irrespective of their location relative to linkage signals (this is discussed in detail in Freimer and Sabatti, 2004). Another important issue of concern regards the structure of the tested replication samples.

It is becoming increasingly clear that unreliable results may be obtained when allele frequencies differ notably among subpopulations not represented equally between cases and controls (Campbell *et al.*, 2005). Therefore, the possibility that original or replication studies using case-control samples are false positives (or negatives) is a major source of concern. This issue is relevant to all common, complex disorders, but it is likely to be more pronounced in genetic studies of psychiatric disorders, which are confounded by a larger degree of phenotypic heterogeneity. Even more alarmingly, several of the original or “replication” samples have been used repeatedly in genetic association studies making the issue of multiple testing corrections highly relevant. These are not merely theoretical considerations as they can lead to striking inconsistencies among variant alleles and haplotypes implicated in various replication studies (inconsistencies are sometimes observed even in more reliable family-based samples and can be explained in some instances by presence of distinct variations affecting different functional elements within the gene that have emerged independently on a more recent ancestral background). Publication bias almost certainly affects the level of confidence ascribed to any given susceptibility gene, primarily because negative studies are more likely to accumulate with considerable delay or not at all (negative studies are less likely to be submitted for publication and when they are submitted they are less likely to be published in the same journals where the original discovery was reported).

V. Future Directions of the Genetic Research: Advancing Our Understanding of How the Specific Genetic Factors Contribute Biologically to the Disease Process

The recent gene discovery studies promise to provide researchers with important new clues regarding the genetic causes of schizophrenia. As additional genes are identified through linkage or genome-wide association studies (which are now starting to be implemented) a central goal of future research will be to understand the functional implications and interactions of the susceptibility genes and their variants in the context of schizophrenia.

In the absence of well-defined and fully penetrant mutations, similar to the ones found in Mendelian disorders, it is important to balance human genetic and hard biological evidence against the need for timely identification of targets and improvements in therapy. Genetic studies of endophenotypes (Egan *et al.*, 2001) (provided they are designed to avoid all the pitfalls described above, which are associated with genetic studies of the clinical syndrome) and most importantly hard biological data from animal model studies promises to advance our understanding of the disease pathophysiology in the next few years.

A. ANIMAL MODELS

Identification of susceptibility genes will permit incisive studies to illuminate the physiological and biochemical etiology of the disease by examining the gene products in the context of a model organism and their impact on the development of the disorder. Such studies could also provide a critical resource for testing new mechanism-based candidate therapies. It remains a challenge, however, to define the optimal means to harness such model organisms in investigative strategies designed to understand and manipulate candidate factors predisposing to schizophrenia, a uniquely human disorder. For example, generation of *bona fide* mouse models for most psychiatric disorders is highly unlikely due to constraints imposed by the complex polygenic nature of human psychiatric disorders, by the magnitude and pattern of change during hominid brain evolution and by uncertainty regarding the clinical features of the human syndromes. On the other hand, mouse models of “susceptibility genes” identified through forward genetic studies in humans hold tremendous promise in understanding the function of the genes in the context of simple cellular pathways or even at the level of simple neural circuits and behavior.

Even in this context, there are several important factors that need to be considered in generating such models and in designing and interpreting their analysis. Possibly the most important consideration concerns the nature of the susceptibility allele. For example, it is critical to know whether the risk allele constitutes a hypomorph or a gain of function to predict whether a mouse knockout allele can model it accurately. A related consideration concerns potential broad expression and pleiotropic effects of particular susceptibility genes. Several of the strong candidate genes (i.e., *NRG1*) appear to participate in virtually all aspects of brain development, maturation, and function and modulate signaling through a large number of neurotransmitter receptors (Corfas *et al.*, 2004). Given this complexity, when modeling such genes using, for example, mouse knockout approaches, one must consider carefully which of a large number of alternative phenotypes might provide a critical link between the genetic risk variant and susceptibility to schizophrenia. Therefore, one very important goal

of human genetic research will be: (1) to define as accurately as possible the risk alleles or haplotypes and (2) to decipher the functional implications of the risk alleles or haplotypes (Pastinen and Hudson, 2004) in order to model them in mice as closely as possible. Given our present limited understanding of the functional impact of human genetic variation, accurate mouse models of risk alleles have been reported for only a small number of schizophrenia susceptibility genes (Huotari *et al.*, 2002; Paterlini *et al.*, 2005; Paylor *et al.*, 2001). Provided that reliable mouse models are available, mechanistic insights into the mode of contribution of these genes, as well as their interactions can be obtained through a heuristic progression starting from the molecular level to the cellular and synaptic level to the systems level and culminating at the behavioral level.

B. GENETIC INTERACTIONS

The genetic complexity of common psychiatric disorders has been repeatedly inferred from the pattern of inheritance and the inability of the research community to identify consistent linkage signals. It is believed that genetic interactions among susceptibility genes (especially epistasis) lie at the core of this complexity. Generally speaking, epistasis is a phenomenon whereby the effects of a given gene on a biological trait are masked or enhanced by one or more other genes. It has been speculated that this type of genetic buffering leads to phenotypes that are stable in the presence of mutations (Moore, 2005). It has also been argued that for a phenotype to be buffered against the effects of mutations, it must have an underlying genetic architecture that is composed of networks of genes that are redundant and robust (Moore, 2005). As a result, substantial effects on the phenotype are observed only when there are multiple mutational hits to the gene network.

The biological basis of these epistatic interactions remains elusive in psychiatric disorders, but two studies provided some important relevant insights (Millar *et al.*, 2005; Paterlini *et al.*, 2005). One study, using animal models, demonstrated a clear epistatic interaction between the *prodh* and *comt* genes at the level of transcription and behavior that is likely to represent a *comt*-modulated homeostatic response to abnormal dopaminergic signaling in the frontal cortex that emerges as a result of *prodh* deficiency (Paterlini *et al.*, 2005). This is an intriguing finding, because dopaminergic dysregulation in schizophrenia is well established, based primarily on the therapeutic effect of dopamine receptor antagonists (Seeman, 1987). Moreover, based on clinical and preclinical observations, it has been suggested that this dopaminergic dysregulation emerges as a secondary result of other primary deficits, including impaired glutamate transmission. In any case, it is conceivable that similar patterns of genetic interactions that involve impaired synaptic function and impaired homeostasis or compensation

(genomic buffering) might also account for the epistatic component of the genetic risk of psychiatric disorders in general.

A second study used the two-hybrid system to identify a molecular interaction between the DISC1 protein and phosphodiesterase 4B (PDE4B; Millar *et al.*, 2005). Although the gene encoding for PDE4B is not included in the list of strong candidate genes outlined above, the authors showed it is disrupted by a balanced translocation in a subject diagnosed with schizophrenia and a relative with chronic psychiatric illness. The PDEs inactivate cAMP, a second messenger implicated in learning, memory, and mood. It was shown that DISC1 interacts with the UCR2 domain of PDE4B and that elevation of cellular cAMP leads to dissociation of PDE4B from DISC1 and an increase in PDE4B activity. The authors proposed a genetic interaction model whereby DISC1 sequesters PDE4B in resting cells and releases it in an activated state in response to elevated cAMP.

Methods of analysis designed to probe epistasis are clearly of growing importance in the genetic dissection of complex disorders and currently a variety of methods exist to detect or control for the presence of epistasis. These methods are limited by the need for large sample sizes that ensure adequate power to detect gene interactions. Moreover, direct biological inference from the results of statistical tests is very difficult because statistical interaction does not necessarily imply interaction at the biological level (Thompson, 1991). In all, the degree to which statistical modeling can provide insights into the underlying disease mechanisms is likely to be limited, and might require prior knowledge of the underlying etiology. The question of true biological interaction remains of extreme importance in the field of complex psychiatric genetics, but might ultimately be better answered primarily via a combination of molecular and animal model-based approaches.

C. UNDERSTANDING DISEASE PATHOPHYSIOLOGY

There are two major pharmacological hypotheses regarding the pathophysiology of schizophrenia—the dopaminergic and glutamatergic hypotheses. The dopaminergic hypothesis is based primarily on the observation that all drugs with efficacy in treating symptoms of schizophrenia share the property of dopamine D2 receptor antagonism and also on the fact that indirect dopaminergic agonists, such as cocaine and amphetamine, have psychotomimetic properties (Seeman, 1987). The glutamatergic hypothesis arose from the finding that phencyclidine (PCP), a potent psychotomimetic drug, is an antagonist of the NMDA receptor, and posits that a major underlying cause of schizophrenia is abnormal glutamatergic transmission, particularly in the prefrontal cortex, limbic areas, and striatum (Coyle, 1996). Since the glutamatergic and dopaminergic systems

are known to have complex interactions, the two hypotheses are not incompatible, and it is likely that primary changes in one system would lead to associated alterations in the other.

Although efforts for synthesis of the initial genetic findings in the context of specific neurotransmitter systems have been reported (Moghaddam, 2003), it might be premature at this point to claim that the existing genetic data support the critical involvement of one neurotransmitter system over another. The genes outlined in this review article are involved in several neurotransmitter systems. For some of them (such as *PRODH*, *G72*, *DTNBP1*, *NRG1*, *ζDHH8*, *CAPON*), there is variable degree of experimental evidence that they are involved in excitatory glutamatergic pathways. For other genes, there is clear evidence of involvement in dopamine (*COMT*, *AKT1*), GABA (5q *GABA* receptor cluster, *AKT1*), or trace amine signaling (*TAAR6*). However, even for genes that primarily act via disruption of excitatory synaptic function, the final effect could be mediated by abnormal dopamine signaling (Paterlini *et al.*, 2005; see previous paragraph). Finally, many of the genes (such as *NRG1*, *DISC1*, *PPP3CC*, *RGS4*, *AKT1*) appear to have pleiotropic effects that are not restricted to a particular type of synaptic transmission and involve several aspects of neuronal biology. While it is too early to determine from the existing genetic evidence how neurotransmitter systems might be primarily affected in the disease, identification of additional genes and further functional studies will help elucidate this issue. However, it is equally likely that the final cumulative effect of the risk variants will emerge from, and be determined by, the pattern of expression, as well as the pattern of interaction among these genes, and it could be restricted to specific brain regions, specific cell types, or both, rather than specific neurotransmitter systems. This regional or cellular selectivity could underlie the differences and commonalities among common psychiatric disorders, as well as their distinction from other common and serious CNS conditions, such as mental retardation, or epilepsy that employ common neurotransmitter systems.

D. MECHANISM-BASED THERAPIES

It is common place to state that understanding of the function and the interactions among individual susceptibility elements could eventually lead to design of highly effective targeted therapies for patients with specific genetic predisposition with fewer side effects and more positive long-term disease outcomes. However, optimism should perhaps be tempered by experience gained from study of simple genetic conditions where knowledge of the genes and protein alterations is often available, and yet it has proven highly challenging to translate this detailed knowledge into creation of therapies. It is conceivable that despite their more complex etiology, multifactorial disorders like schizophrenia may be

more amenable to mechanism-based therapeutic intervention. So far, it appears that genetic alterations contributing to schizophrenia consist largely of common but relatively subtle variations presumably affecting transcript expression or processing and in some cases protein function. The disease risk associated with such variations is usually very low, but because the risk alleles are so common, a low disease risk corresponds to a large population attributable risk (which means that if the population were monomorphic for the nonrisk allele, the prevalence of the disease would be considerably lower). Directed therapies, therefore, might only need to provide relatively modest modulation of appropriate molecular targets to reach an effective threshold in a large fraction of patients, in contrast to simple conditions, in which compensation for more pronounced functional alterations might be required.

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References

- Abou Jamra, R., Sircar, I., Becker, T., Freudenberg-Hua, Y., Ohlraun, S., Freudenberg, J., Brockschmidt, F., Schulze, T. G., Gross, M., Spira, F., Deschner, M., Schmal, C., *et al.* (2005). A family-based and case-control association study of trace amine receptor genes on chromosome 6q23 in bipolar affective disorder. *Mol. Psychiatry* **10**, 618–620.
- Addington, A. M., Gornick, M., Sporn, A. L., Gogtay, N., Greenstein, D., Lenane, M., Gochman, P., Baker, N., Balkissoon, R., Vakkalanka, R. K., Weinberger, D. R., Straub, R. E., *et al.* (2004). Polymorphisms in the 13q33.2 gene G72/G30 are associated with childhood-onset schizophrenia and psychosis not otherwise specified. *Biol. Psychiatry* **55**, 976–980.
- Badner, J. A., and Gershon, E. S. (2002). Meta-analysis of whole-genome linkage scans of bipolar disorder and schizophrenia. *Mol. Psychiatry* **7**, 405–411.
- Beaulieu, J. M., Sotnikova, T. D., Yao, W. D., Kockeritz, L., Woodgett, J. R., Gainetdinov, R. R., and Caron, M. G. (2004). Lithium antagonizes dopamine-dependent behaviors mediated by an AKT/glycogen synthase kinase 3 signaling cascade. *Proc. Natl. Acad. Sci. USA* **101**, 5099–5104.
- Bender, H. U., Almashanu, S., Steel, G., Hu, C. A., Lin, W. W., Willis, A., Pulver, A., and Valle, D. (2005). Functional consequences of PRODH missense mutations. *Am. J. Hum. Genet.* **76**, 409–420.
- Benson, M. A., Newey, S. E., Martin-Rendon, E., Hawkes, R., and Blake, D. J. (2001). Dysbindin, a novel coiled-coil-containing protein that interacts with the dystrobrevins in muscle and brain. *J. Biol. Chem.* **276**, 24232–24241.

- Blouin, J. L., Dombroski, B. A., Nath, S. K., Lasseter, V. K., Wolyniec, P. S., Nestadt, G., Thornquist, M., Ullrich, G., McGrath, J., Kasch, L., Lamacz, M., Thomas, M. G., *et al.* (1998). Schizophrenia susceptibility loci on chromosomes 13q32 and 8p21. *Nat. Genet.* **20**, 70–73.
- Borowsky, B., Adham, N., Jones, K. A., Raddatz, R., Artymyshyn, R., Ogozalek, K. L., Durkin, M. M., Lakhani, P. P., Bonini, J. A., Pathirana, S., Boyle, N., Pu, X., *et al.* (2001). Trace amines: Identification of a family of mammalian G protein-coupled receptors. *Proc. Natl. Acad. Sci. USA* **98**, 8966–8971.
- Bowie, C. R., and Harvey, P. D. (2005). Cognition in schizophrenia: Impairments, determinants, and functional importance. *Psychiatr. Clin. North Am.* **28**, 613–633.
- Brzustowicz, L. M., Hodgkinson, K. A., Chow, E. W., Honer, W. G., and Bassett, A. S. (2000). Location of a major susceptibility locus for familial schizophrenia on chromosome 1q21-q22. *Science* **288**, 678–682.
- Brzustowicz, L. M., Simone, J., Mohseni, P., Hayter, J. E., Hodgkinson, K. A., Chow, E. W., and Bassett, A. S. (2004). Linkage disequilibrium mapping of schizophrenia susceptibility to the CAPON region of chromosome 1q22. *Am. J. Hum. Genet.* **74**, 1057–1063.
- Callicott, J. H., Straub, R. E., Pezawas, L., Egan, M. F., Mattay, V. S., Hariri, A. R., Verchinski, B. A., Meyer-Lindenberg, A., Balkissoon, R., Kolachana, B., Goldberg, T. E., and Weinberger, D. R. (2005). Variation in DISC1 affects hippocampal structure and function and increases risk for schizophrenia. *Proc. Natl. Acad. Sci. USA* **102**, 8627–8632.
- Campbell, C. D., Ogburn, E. L., Lunetta, K. L., Lyon, H. N., Freedman, M. L., Groop, L. C., Altshuler, D., Ardlie, K. G., and Hirschhorn, J. N. (2005). Demonstrating stratification in a European American population. *Nat. Genet.* **37**, 868–872.
- Chalecka-Franaszek, E., and Chuang, D. M. (1999). Lithium activates the serine/threonine kinase Akt-1 and suppresses glutamate-induced inhibition of Akt-1 activity in neurons. *Proc. Natl. Acad. Sci. USA* **96**, 8745–8750.
- Chen, W. Y., Shi, Y. Y., Zheng, Y. L., Zhao, X. Z., Zhang, G. J., Chen, S. Q., Yang, P. D., and He, L. (2004a). Case-control study and transmission disequilibrium test provide consistent evidence for association between schizophrenia and genetic variation in the 22q11 gene ZDHHC8. *Hum. Mol. Genet.* **13**, 2991–2995.
- Chen, X., Dunham, C., Kendler, S., Wang, X., O'Neill, F. A., Walsh, D., and Kendler, K. S. (2004b). Regulator of G-protein signaling 4 (RGS4) gene is associated with schizophrenia in Irish high density families. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **129**, 23–26.
- Chowdari, K. V., Mirnics, K., Semwal, P., Wood, J., Lawrence, E., Bhatia, T., Deshpande, S. N., Thelma, B. K., Ferrell, R. E., Middleton, F. A., Devlin, B., Levitt, P., *et al.* (2002). Association and linkage analyses of RGS4 polymorphisms in schizophrenia. *Hum. Mol. Genet.* **11**, 1373–1380.
- Chumakov, I., Blumenfeld, M., Guerassimenko, O., Cavarec, L., Palicio, M., Abderrahim, H., Bougueleret, L., Barry, C., Tanaka, H., La Rosa, P., Puech, A., Tahri, N., *et al.* (2002). Genetic and physiological data implicating the new human gene G72 and the gene for D-amino acid oxidase in schizophrenia. *Proc. Natl. Acad. Sci. USA* **99**, 13675–13680.
- Corfas, G., Roy, K., and Buxbaum, J. D. (2004). Neuregulin 1-erbB signaling and the molecular/cellular basis of schizophrenia. *Nat. Neurosci.* **7**, 575–580.
- Corvin, A. P., Morris, D. W., McGhee, K., Schwaiger, S., Scully, P., Quinn, J., Meagher, D., Clair, D. S., Waddington, J. L., and Gill, M. (2004). Confirmation and refinement of an 'at-risk' haplotype for schizophrenia suggests the EST cluster, Hs.97362, as a potential susceptibility gene at the Neuregulin-1 locus. *Mol. Psychiatry* **9**, 208–213.
- Coyle, J. T. (1996). The glutamatergic dysfunction hypothesis for schizophrenia. *Harv. Rev. Psychiatry* **3**, 241–253.
- Detera-Wadleigh, S. D., Badner, J. A., Berrettini, W. H., Yoshikawa, T., Goldin, L. R., Turner, G., Rollins, D. Y., Moses, T., Sanders, A. R., Karkera, J. D., Esterling, L. E., Zeng, J., *et al.* (1999).

- A high-density genome scan detects evidence for a bipolar-disorder susceptibility locus on 13q32 and other potential loci on 1q32 and 18p11.2. *Proc. Natl. Acad. Sci. USA* **96**, 5604–5609.
- Duan, J., Martinez, M., Sanders, A. R., Hou, C., Saitou, N., Kitano, T., Mowry, B. J., Crowe, R. R., Silverman, J. M., Levinson, D. F., and Gejman, P. V. (2004). Polymorphisms in the trace amine receptor 4 (TRAR4) gene on chromosome 6q23.2 are associated with susceptibility to schizophrenia. *Am. J. Hum. Genet.* **75**, 624–638.
- Duan, J., Martinez, M., Sanders, A. R., Hou, C., Krasner, A. J., Schwartz, D. B., and Gejman, P. V. (2005a). Neuregulin 1 (NRG1) and schizophrenia: Analysis of a US family sample and the evidence in the balance. *Psychol. Med.* **35**, 1599–1610.
- Duan, S., Du, J., Xu, Y., Xing, Q., Wang, H., Wu, S., Chen, Q., Li, X., Li, X., Shen, J., Feng, G., and He, L. (2005b). Failure to find association between TRAR4 and schizophrenia in the Chinese Han population. *J. Neural Transm.* [Epub ahead of print: Aug 3, 2005; PMID: 16075187].
- Eastwood, S. L. (2005). Decreased hippocampal expression of the susceptibility gene PPP3CC and other calcineurin subunits in schizophrenia. *Biol. Psychiatry* **57**, 702–710.
- Egan, M. F., Goldberg, T. E., Kolachana, B. S., Callicott, J. H., Mazzanti, C. M., Straub, R. E., Goldman, D., and Weinberger, D. R. (2001). Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia. *Proc. Natl. Acad. Sci. USA* **98**, 6917–6922.
- Ekelund, J., Hovatta, I., Parker, A., Paunio, T., Varilo, T., Martin, R., Suhonen, J., Ellonen, P., Chan, G., Sinsheimer, J. S., Sobel, E., Juvonen, H., *et al.* (2001). Chromosome 1 loci in Finnish schizophrenia families. *Hum. Mol. Genet.* **10**, 1611–1617.
- Ekelund, J., Hennah, W., Hiekkalinna, T., Parker, A., Meyer, J., Lonnqvist, J., and Peltonen, L. (2004). Replication of 1q42 linkage in Finnish schizophrenia pedigrees. *Mol. Psychiatry* **9**, 1037–1041.
- el-Husseini, Ael-D., and , and Bredt, D. S. (2002). Protein palmitoylation: A regulator of neuronal development and function. *Nat. Rev. Neurosci.* **3**, 791–802.
- Emamian, E. S., Hall, D., Birnbaum, M. J., Karayiorgou, M., and Gogos, J. A. (2004). Convergent evidence for impaired AKT1-GSK3 β signaling in schizophrenia. *Nat. Genet.* **36**, 131–137.
- Fallin, M. D., Lasseter, V. K., Avramopoulos, D., Nicodemus, K. K., Wolyniec, P. S., McGrath, J. A., Steel, G., Nestadt, G., Liang, K. Y., Haganir, R. L., Valle, D., and Pulver, A. E. (2005). Bipolar I disorder and schizophrenia: A 440-single-nucleotide polymorphism screen of 64 candidate genes among Ashkenazi Jewish case-parent trios. *Am. J. Hum. Genet.* **77**, 918–936.
- Fan, J. B., Zhang, C. S., Gu, N. F., Li, X. W., Sun, W. W., Wang, H. Y., Feng, G. Y., St. Clair, D., and He, L. (2005). Catechol-O-methyltransferase gene Val/Met functional polymorphism and risk of schizophrenia: A large-scale association study plus meta-analysis. *Biol. Psychiatry* **57**, 139–144.
- Freimer, N., and Sabatti, C. (2004). The use of pedigree, sib-pair and association studies of common diseases for genetic mapping and epidemiology. *Nat. Genet.* **36**, 1045–1051.
- Funke, B., Finn, C. T., Plocik, A. M., Lake, S., DeRosse, P., Kane, J. M., Kucherlapati, R., and Malhotra, A. K. (2004). Association of the DTNBP1 locus with schizophrenia in a U.S. population. *Am. J. Hum. Genet.* **75**, 891–898.
- Gerber, D. J., Hall, D., Miyakawa, T., Demars, S., Gogos, J. A., Karayiorgou, M., and Tonegawa, S. (2003). Evidence for association of schizophrenia with genetic variation in the 8p21.3 gene, PPP3CC, encoding the calcineurin gamma subunit. *Proc. Natl. Acad. Sci. USA* **100**, 8993–8998.
- Glaser, B., Schumacher, J., Williams, H. J., Jamra, R. A., Ianakiev, N., Milev, R., Ohlraun, S., Schulze, T. G., Czerski, P. M., Hauser, J., Jonsson, E. G., Sedvall, G. C., *et al.* (2005). No association between the putative functional ZDHHC8 single nucleotide polymorphism rs175174 and schizophrenia in large European samples. *Biol. Psychiatry* **58**, 78–80.
- Glatt, S. J., Faraone, S. V., and Tsuang, M. T. (2003). Association between a functional catechol O-methyltransferase gene polymorphism and schizophrenia: Meta-analysis of case-control and family-based studies. *Am. J. Psychiatry* **160**, 469–476.

- Goldman-Rakic, P. S. (1994). Working memory dysfunction in schizophrenia. *J. Neuropsychiatry Clin. Neurosci.* **6**, 348–357.
- Gogos, J. A., Santha, M., Takacs, Z., Beck, K. D., Luine, V., Lucas, L. R., Nadler, J. V., and Karayiorgou, M. (1999). The gene encoding proline dehydrogenase modulates sensorimotor gating in mice. *Nat. Genet.* **21**, 434–439.
- Gothelf, D., Eliez, S., Thompson, T., Hinard, C., Penniman, L., Feinstein, C., Kwon, H., Jin, S., Jo, B., Antonarakis, S. E., Morris, M. A., and Reiss, A. L. (2005). COMT genotype predicts longitudinal cognitive decline and psychosis in 22q11.2 deletion syndrome. *Nat. Neurosci.* **8**, 1500–1502.
- Green, M. F., Nuechterlein, K. H., Gold, J. M., Barch, D. M., Cohen, J., Essock, S., Fenton, W. S., Frese, F., Goldberg, T. E., Heaton, R. K., Keefe, R. S., Kern, R. S., *et al.* (2004). Approaching a consensus cognitive battery for clinical trials in schizophrenia: The NIMH-MATRICES conference to select cognitive domains and test criteria. *Biol. Psychiatry* **56**, 301–307.
- Greengard, P. (2001). The neurobiology of slow synaptic transmission. *Science* **294**, 1024–1030.
- Hall, D., Gogos, J. A., and Karayiorgou, M. (2004). The contribution of three strong candidate schizophrenia susceptibility genes in demographically distinct populations. *Genes Brain Behav.* **3**, 240–248.
- Hattori, E., Liu, C., Badner, J. A., Bonner, T. I., Christian, S. L., Maheshwari, M., Detera-Wadleigh, S. D., Gibbs, R. A., and Gershon, E. S. (2003). Polymorphisms at the G72/G30 gene locus, on 13q33, are associated with bipolar disorder in two independent pedigree series. *Am. J. Hum. Genet.* **72**, 1131–1140.
- Hennah, W., Varilo, T., Kestila, M., Paunio, T., Arajärvi, R., Haukka, J., Parker, A., Martin, R., Levitzky, S., Partonen, T., Meyer, J., Lonnqvist, J., *et al.* (2003). Haplotype transmission analysis provides evidence of association for DISC1 to schizophrenia and suggests sex-dependent effects. *Hum. Mol. Genet.* **12**, 3151–3159.
- Hennah, W., Tuulio-Henriksson, A., Paunio, T., Ekelund, J., Varilo, T., Partonen, T., Cannon, T. D., Lonnqvist, J., and Peltonen, L. (2005). A haplotype within the DISC1 gene is associated with visual memory functions in families with a high density of schizophrenia. *Mol. Psychiatry* **10**, 1097–1103.
- Ho, B. C., Wassink, T. H., O'Leary, D. S., Sheffield, V. C., and Andreasen, N. C. (2005). Catechol-O-methyl transferase Val158Met gene polymorphism in schizophrenia: Working memory, frontal lobe MRI morphology and frontal cerebral blood flow. *Mol. Psychiatry* **10**, 287–298.
- Hodgkinson, C. A., Goldman, D., Jaeger, J., Persaud, S., Kane, J. M., Lipsky, R. H., and Malhotra, A. K. (2004). Disrupted in schizophrenia 1 (DISC1): Association with schizophrenia, schizoaffective disorder and bipolar disorder. *Am. J. Hum. Genet.* **75**, 862–872.
- Huotari, M., Gogos, J. A., Karayiorgou, M., Koponen, O., Forsberg, M., Raasmaja, A., Hyttinen, J., and Mannisto, P. T. (2002). Brain catecholamine metabolism in catechol-O-methyltransferase (COMT)-deficient mice. *Eur. J. Neurosci.* **15**, 246–256.
- Ikeda, M., Iwata, N., Suzuki, T., Kitajima, T., Yamanouchi, Y., Kinoshita, Y., Inada, T., and Ozaki, N. (2004). Association of AKT1 with schizophrenia confirmed in a Japanese population. *Biol. Psychiatry* **56**, 698–700.
- Ikeda, M., Iwata, N., Suzuki, T., Kitajima, T., Yamanouchi, Y., Kinoshita, Y., Inada, T., and Ozaki, N. (2005). No association of haplotype-tagging SNPs in TRAR4 with schizophrenia in Japanese patients. *Schizophr. Res.* **78**, 127–130.
- Iwata, N., Suzuki, T., Ikeda, M., Kitajima, T., Yamanouchi, Y., Inada, T., and Ozaki, N. (2004). No association with the neuregulin 1 haplotype to Japanese schizophrenia. *Mol. Psychiatry* **9**, 126–127.
- Jacquet, H., Raux, G., Thibaut, F., Hecketsweiler, B., Houy, E., Demilly, C., Haouzir, S., Allio, G., Fouldrin, G., Drouin, V., Bou, J., Petit, M., *et al.* (2002). PRODH mutations and hyperprolinaemia in a subset of schizophrenic patients. *Hum. Mol. Genet.* **11**, 2243–2249.

- Jaffrey, S. R., Snowman, A. M., Eliasson, M. J., Cohen, N. A., and Snyder, S. H. (1998). CAPON: A protein associated with neuronal nitric oxide synthase that regulates its interactions with PSD95. *Neuron* **20**, 115–124.
- Kang, U. G., Seo, M. S., Roh, M. S., Kim, Y., Yoon, S. C., and Kim, Y. S. (2004). The effects of clozapine on the GSK-3-mediated signaling pathway. *FEBS Lett.* **560**, 115–119.
- Karayiorgou, M., and Gogos, J. A. (1997). A turning point in schizophrenia genetics. *Neuron* **19**, 967–979.
- Karayiorgou, M., and Gogos, J. A. (2004). The molecular genetics of the 22q11-associated schizophrenia. *Mol. Brain Res.* **132**, 95–104.
- Karayiorgou, M., Morris, M. A., Morrow, B., Shprintzen, R. J., Goldberg, R., Borrow, J., Gos, A., Nestadt, G., Wolyniec, P. S., Lasseter, V. K., Eisen, H., Childs, B., *et al.* (1995). Schizophrenia susceptibility associated with interstitial deletions of chromosome 22q11. *Proc. Natl. Acad. Sci. USA* **92**, 7612–7616.
- Kirov, G., Ivanov, D., Williams, N. M., Preece, A., Nikolov, I., Milev, R., Koleva, S., Dimitrova, A., Toncheva, D., O'Donovan, M. C., and Owen, M. J. (2004). Strong evidence for association between the dystrobrevin binding protein 1 gene (DTNBP1) and schizophrenia in 488 parent-offspring trios from Bulgaria. *Biol. Psychiatry* **55**, 971–975.
- Korostishevsky, M., Kaganovich, M., Cholostoy, A., Ashkenazi, M., Ratner, Y., Dahary, D., Bernstein, J., Bening-Abu-Shach, U., Ben-Asher, E., Lancet, D., Ritsner, M., and Navon, R. (2004). Is the G72/G30 locus associated with schizophrenia? single nucleotide polymorphisms, haplotypes, and gene expression analysis. *Biol. Psychiatry* **56**, 169–176.
- Larminie, C., Murdock, P., Walhin, J. P., Duckworth, M., Blumer, K. J., Scheideler, M. A., and Garnier, M. (2004). Selective expression of regulators of G-protein signaling (RGS) in the human central nervous system. *Mol. Brain Res.* **122**, 24–34.
- Levinson, D. F., Holmans, P., Straub, R. E., Owen, M. J., Wildenauer, D. B., Gejman, P. V., Pulver, A. E., Laurent, C., Kendler, K. S., Walsh, D., Norton, N., Williams, N. M., *et al.* (2000). Multicenter linkage study of schizophrenia candidate regions on chromosomes 5q, 6q, 10p, and 13q: Schizophrenia linkage collaborative group III. *Am. J. Hum. Genet.* **67**, 652–663.
- Lewis, C. M., Levinson, D. F., Wise, L. H., DeLisi, L. E., Straub, R. E., Hovatta, I., Williams, N. M., Schwab, S. G., Pulver, A. E., Faraone, S. V., Brzustowicz, L. M., Kaufmann, C. A., *et al.* (2003). Genome scan meta-analysis of schizophrenia and bipolar disorder, part II: Schizophrenia. *Am. J. Hum. Genet.* **73**, 34–48.
- Lewis, D. A., Hashimoto, T., and Volk, D. W. (2005). Cortical inhibitory neurons and schizophrenia. *Nat. Rev. Neurosci.* **6**, 312–324.
- Li, T., Stefansson, H., Gudfinnsson, E., Cai, G., Liu, X., Murray, R. M., Steinthorsdottir, V., Januel, D., Gudnadottir, V. G., Petursson, H., Ingason, A., Gulcher, J. R., *et al.* (2004b). Identification of a novel neuregulin 1 at-risk haplotype in Han schizophrenia Chinese patients, but no association with the Icelandic/Scottish risk haplotype. *Mol. Psychiatry* **9**, 698–704.
- Li, T., Ma, X., Sham, P. C., Sun, X., Hu, X., Wang, Q., Meng, H., Deng, W., Liu, X., Murray, R. M., and Collier, D. A. (2004a). Evidence for association between novel polymorphisms in the PRODH gene and schizophrenia in a Chinese population. *Am. J. Med. Genet.* **129B**, 13–15.
- Li, W., Zhang, Q., Oiso, N., Novak, E. K., Gautam, R., O'Brien, E. P., Tinsley, C. L., Blake, D. J., Spritz, R. A., Copeland, N. G., Jenkins, N. A., Amato, D., *et al.* (2003). Hermansky-Pudlak syndrome type 7 (HPS-7) results from mutant dysbindin, a member of the biogenesis of lysosome-related organelles complex 1 (BLOC-1). *Nat. Genet.* **35**, 84–89.
- Liu, H., Heath, S. C., Sobin, C., Roos, J. L., Galke, B. L., Blundell, M. L., Lenane, M., Robertson, B., Wijsman, E. M., Rapoport, J. L., Gogos, J. A., and Karayiorgou, M. (2002a). Genetic variation at the 22q11 PRODH2/DGCR6 locus presents an unusual pattern and increases susceptibility to schizophrenia. *Proc. Natl. Acad. Sci. USA* **99**, 3717–3722.

- Liu, H., Abecasis, G. R., Heath, S. C., Knowles, A., Demars, S., Chen, Y. J., Roos, J. L., Rapoport, J. L., Gogos, J. A., and Karayiorgou, M. (2002b). Genetic variation in the 22q11 locus and susceptibility to schizophrenia. *Proc. Natl. Acad. Sci. USA* **99**, 16859–16864.
- Lohmueller, K. E., Pearce, C. L., Pike, M., Lander, E. S., and Hirschhorn, J. N. (2003). Meta-analysis of genetic association studies supports a contribution of common variants to susceptibility to common disease. *Nat. Genet.* **33**, 177–182.
- Millar, J. K., Wilson-Annan, J. C., Anderson, S., Christie, S., Taylor, M. S., Semple, C. A., Devon, R. S., Clair, D. M., Muir, W. J., Blackwood, D. H., and Porteous, D. J. (2000). Disruption of two novel genes by a translocation co-segregating with schizophrenia. *Hum. Mol. Genet.* **9**, 1415–1423.
- Millar, J. K., Pickard, B. S., Mackie, S., James, R., Christie, S., Buchanan, S. R., Malloy, M. P., Chubb, J. E., Huston, E., Baillie, G. S., Thomson, P. A., Hill, E. V., *et al.* (2005). DISC1 and PDE4B are interacting genetic factors in schizophrenia that regulate cAMP signaling. *Science* **310**, 1187–1191.
- Mirmics, K., Middleton, F. A., Marquez, A., Lewis, D. A., and Levitt, P. (2000). Molecular characterization of schizophrenia viewed by microarray analysis of gene expression in prefrontal cortex. *Neuron* **28**, 53–67.
- Miyakawa, T., Leiter, L. M., Gerber, D. J., Gainetdinov, R. R., Sotnikova, T. D., Zeng, H., Caron, M. G., and Tonegawa, S. (2003). Conditional calcineurin knockout mice exhibit multiple abnormal behaviors related to schizophrenia. *Proc. Natl. Acad. Sci. USA* **100**, 8982–8987.
- Moghaddam, B. (2003). Bringing order to the glutamate chaos in schizophrenia. *Neuron* **40**, 881–884.
- Moore, J. H. (2005). A global view of epistasis. *Nat. Genet.* **37**, 13–14.
- Morris, D. W., McGhee, K. A., Schwaiger, S., Scully, P., Quinn, J., Meagher, D., Waddington, J. L., Gill, M., and Corvin, A. P. (2003a). No evidence for association of the dysbindin gene [DTNBP1] with schizophrenia in an Irish population-based study. *Schizophr. Res.* **60**, 167–172.
- Morris, J. A., Kandpal, G., Ma, L., and Austin, C. P. (2003b). DISC1 (Disrupted-In-Schizophrenia 1) is a centrosome-associated protein that interacts with MAP1A, MIPT3, ATF4/5 and NUDEL: Regulation and loss of interaction with mutation. *Hum. Mol. Genet.* **12**, 1591–1608.
- Mothet, J. P., Parent, A. T., Wolosker, H., Brady, R. O., Jr., Linden, D. J., Ferris, C. D., Rogawski, M. A., and Snyder, S. H. (2000). D-serine is an endogenous ligand for the glycine site of the N-methyl-D-aspartate receptor. *Proc. Natl. Acad. Sci. USA* **97**, 4926–4931.
- Mukai, J., Liu, H., Burt, R. A., Swor, D. E., Lai, W. S., Karayiorgou, M., and Gogos, J. A. (2004). Evidence that the gene encoding ZDHHC8 contributes to the risk of schizophrenia. *Nat. Genet.* **36**, 725–731.
- Mulle, J. G., Chowdari, K. V., Nimgaonkar, V., and Chakravarti, A. (2005). No evidence for association to the G72/G30 locus in an independent sample of schizophrenia families. *Mol. Psychiatry* **10**, 431–433.
- Munafo, M. R., Bowes, L., Clark, T. G., and Flint, J. (2005). Lack of association of the COMT (Val158/108 Met) gene and schizophrenia: A meta-analysis of case-control studies. *Mol. Psychiatry* **10**, 765–770.
- Murphy, K. C., Jones, L. A., and Owen, M. J. (1999). High rates of schizophrenia in adults with velocardio-facial syndrome. *Arch. Gen. Psychiatry* **56**, 940–945.
- Numakawa, T., Yagasaki, Y., Ishimoto, T., Okada, T., Suzuki, T., Iwata, N., Ozaki, N., Taguchi, T., Tatsumi, M., Kamijima, K., Straub, R. E., Weinberger, D. R., *et al.* (2004). Evidence of novel neuronal functions of dysbindin, a susceptibility gene for schizophrenia. *Hum. Mol. Genet.* **13**, 2699–2708.
- Ohtsuki, T., Inada, T., and Arinami, T. (2004). Failure to confirm association between AKT1 haplotype and schizophrenia in a Japanese case-control population. *Mol. Psychiatry* **9**, 981–983.
- Pastinen, T., and Hudson, T. J. (2004). Cis-acting regulatory variation in the human genome. *Science* **306**, 647–650.

- Paterlini, M., Zakharenko, S. S., Lai, W. S., Qin, J., Zhang, H., Mukai, J., Westphal, K. G., Olivier, B., Sulzer, D., Pavlidis, P., Siegelbaum, S. A., Karayiorgou, M., *et al.* (2005). Transcriptional and behavioral interaction between 22q11.2 orthologs modulates schizophrenia-related phenotypes in mice. *Nat. Neurosci.* **8**, 1586–1594.
- Paylor, R., McIlwain, K. L., McAninch, R., Nellis, A., Yuva-Paylor, L. A., Baldini, A., and Lindsay, E. A. (2001). Mice deleted for the DiGeorge/velocardiofacial syndrome region show abnormal sensorimotor gating and learning and memory impairments. *Hum. Mol. Genet.* **10**, 2645–2650.
- Petryshen, T. L., Middleton, F. A., Tahl, A. R., Rockwell, G. N., Purcell, S., Aldinger, K. A., Kirby, A., Morley, C. P., McGann, L., Gentile, K. L., Waggoner, S. G., Medeiros, H. M., *et al.* (2005a). Genetic investigation of chromosome 5q GABA(A) receptor subunit genes in schizophrenia. *Mol. Psychiatry* **10**, 1074–1088.
- Petryshen, T. L., Middleton, F. A., Kirby, A., Aldinger, K. A., Purcell, S., Tahl, A. R., Morley, C. P., McGann, L., Gentile, K. L., Rockwell, G. N., Medeiros, H. M., Carvalho, C., *et al.* (2005b). Support for involvement of neuregulin 1 in schizophrenia pathophysiology. *Mol. Psychiatry* **10**, 366–374.
- Pimm, J., McQuillin, A., Thirumalai, S., Lawrence, J., Quedsted, D., Bass, N., Lamb, G., Moorey, H., Datta, S. R., Kalsi, G., Badacsonyi, A., Kelly, K., *et al.* (2005). The Epsin 4 gene on chromosome 5q, which encodes the clathrin-associated protein enthoprotin, is involved in the genetic susceptibility to schizophrenia. *Am. J. Hum. Genet.* **76**, 902–907.
- Pulver, A. E., Nestadt, G., Goldberg, R., Shprintzen, R. J., Lamacz, M., Wolyniec, P. S., Morrow, B., Karayiorgou, M., Antonarakis, S. E., and Housman, D. (1994). Psychotic illness in patients diagnosed with velo-cardio-facial syndrome and their relatives. *J. Nerv. Ment. Dis.* **182**, 476–478.
- Puri, V., McQuillin, A., Thirumalai, S., Lawrence, J., Krasucki, R., Choudhury, K., Datta, S., Kerwin, S., Quedsted, D., Bass, N., Pimm, J., Lamb, G., *et al.* (2005). Failure to confirm allelic association between markers at the CAPON gene locus and schizophrenia in a British sample. *Biol. Psychiatry*. [Epub ahead of print: Sep 30, 2005; PMID: 16202394].
- Ross, E. M., and Wilkie, T. M. (2000). GTPase-activating proteins for heterotrimeric G proteins: Regulators of G protein signaling (RGS) and RGS-like proteins. *Annu. Rev. Biochem.* **69**, 795–827.
- Sachs, N. A., Sawa, A., Holmes, S. E., Ross, C. A., DeLisi, L. E., and Margolis, R. L. (2005). A frameshift mutation in disrupted in Schizophrenia 1 in an American family with schizophrenia and schizoaffective disorder. *Mol. Psychiatry* **10**, 758–764.
- Scheid, M. P., and Woodgett, J. R. (2001). PKB/AKT: Functional insights from genetic models. *Nat. Rev. Mol. Cell. Biol.* **2**, 760–768.
- Schumacher, J., Jamra, R. A., Freudenberger, J., Becker, T., Ohlraun, S., Otte, A. C., Tullius, M., Kovalenko, S., Bogaert, A. V., Maier, W., Rietschel, M., Propping, P., *et al.* (2004). Examination of G72 and D-amino-acid oxidase as genetic risk factors for schizophrenia and bipolar affective disorder. *Mol. Psychiatry* **9**, 203–207.
- Schwab, S. G., Knapp, M., Mondabon, S., Hallmayer, J., Borrmann-Hassenbach, M., Albus, M., Lerer, B., Rietschel, M., Trixler, M., Maier, W., and Wildenauer, D. B. (2003). Support for association of schizophrenia with genetic variation in the 6p22.3 gene, dysbindin, in sib-pair families with linkage and in an additional sample of triad families. *Am. J. Hum. Genet.* **72**, 185–190.
- Schwab, S. G., Hoefgen, B., Hanses, C., Hassenbach, M. B., Albus, M., Lerer, B., Trixler, M., Maier, W., and Wildenauer, D. B. (2005). Further evidence for association of variants in the AKT1 gene with schizophrenia in a sample of European sib-pair families. *Biol. Psychiatry* **58**, 446–450.
- Seeman, P. (1987). Dopamine receptors and the dopamine hypothesis of schizophrenia. *Synapse* **1**, 133–152.
- Shifman, S., Bronstein, M., Sternfeld, M., Pisante-Shalom, A., Lev-Lehman, E., Weizman, A., Reznik, I., Spivak, B., Grisaru, N., Karp, L., Schiffer, R., Kotler, M., *et al.* (2002). A highly

- significant association between a COMT haplotype and schizophrenia. *Am. J. Hum. Genet.* **71**, 1296–1302.
- Sklar, P., Pato, M. T., Kirby, A., Petryshen, T. L., Medeiros, H., Carvalho, C., Macedo, A., Dourado, A., Coelho, I., Valente, J., Soares, M. J., Ferreira, C. P., *et al.* (2004). Genome-wide scan in Portuguese Island families identifies 5q31–5q35 as a susceptibility locus for schizophrenia and psychosis. *Mol. Psychiatry* **9**, 213–218.
- Sobell, J. L., Richard, C., Wirshing, D. A., and Heston, L. L. (2005). Failure to confirm association between RGS4 haplotypes and schizophrenia in Caucasians. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **139**, 3–7.
- Stefansson, H., Sigurdsson, E., Steinthorsdottir, V., Bjornsdottir, S., Sigmundsson, T., Ghosh, S., Brynjolfsson, J., Gunnarsdottir, S., Ivarsson, O., Chou, T. T., Hjaltason, O., Birgisdottir, B., *et al.* (2002). Neuregulin 1 and susceptibility to schizophrenia. *Am. J. Hum. Genet.* **71**, 877–892.
- Stefansson, H., Sarginson, J., Kong, A., Yates, P., Steinthorsdottir, V., Gudfinnsson, E., Gunnarsdottir, S., Walker, N., Petursson, H., Crombie, C., Ingason, A., Gulcher, J. R., *et al.* (2003). Association of neuregulin 1 with schizophrenia confirmed in a Scottish population. *Am. J. Hum. Genet.* **72**, 83–87.
- Straub, R. E., Jiang, Y., MacLean, C. J., Ma, Y., Webb, B. T., Myakishev, M. V., Harris-Kerr, C., Wormley, B., Sadek, H., Kadambi, B., Cesare, A. J., Gibberman, A., *et al.* (2002). Genetic variation in the 6p22.3 gene DTNBP1, the human ortholog of the mouse dysbindin gene, is associated with schizophrenia. *Am. J. Hum. Genet.* **71**, 337–348.
- Talbot, K., Eidem, W. L., Tinsley, C. L., Benson, M. A., Thompson, E. W., Smith, R. J., Hahn, C. G., Siegel, S. J., Trojanowski, J. Q., Gur, R. E., Blake, D. J., and Arnold, S. E. (2004). Dysbindin-1 is reduced in intrinsic, glutamatergic terminals of the hippocampal formation in schizophrenia. *J. Clin. Invest.* **113**, 1353–1363.
- Tang, J. X., Zhou, J., Fan, J. B., Li, X. W., Shi, Y. Y., Gu, N. F., Feng, G. Y., Xing, Y. L., Shi, J. G., and He, L. (2003). Family-based association study of DTNBP1 in 6p22.3 and schizophrenia. *Mol. Psychiatry* **8**, 717–718.
- Tang, J. X., Chen, W. Y., He, G., Zhou, J., Gu, N. F., Feng, G. Y., and He, L. (2004). Polymorphisms within 5' end of the Neuregulin 1 gene are genetically associated with schizophrenia in the Chinese population. *Mol. Psychiatry* **9**, 11–12.
- Thiselton, D. L., Webb, B. T., Neale, B. M., Ribble, R. C., O'Neill, F. A., Walsh, D., Riley, B. P., and Kendler, K. S. (2004). No evidence for linkage or association of neuregulin-1 (NRG1) with disease in the Irish study of high-density schizophrenia families (ISHDSF). *Mol. Psychiatry* **9**, 777–783.
- Thompson, W. D. (1991). Effect modification and the limits of biological inference from epidemiologic data. *J. Clin. Epidemiol.* **44**, 221–232.
- Tsai, S. J., Hong, C. J., Hou, S. J., and Yen, F. C. (2006). Lack of association of catechol-O-methyltransferase gene Val108/158Met polymorphism with schizophrenia: A family-based association study in a Chinese population. *Mol. Psychiatry* **11**, 2–3.
- Van Den Bogaert, A., Schumacher, J., Schulze, T. G., Otte, A. C., Ohlraun, S., Kovalenko, S., Becker, T., Freudenberger, J., Jonsson, E. G., Mattila-Evendén, M., Sedvall, G. C., Czernski, P. M., *et al.* (2003). The DTNBP1 (dysbindin) gene contributes to schizophrenia, depending on family history of the disease. *Am. J. Hum. Genet.* **73**, 1438–1443.
- Wang, Q., Liu, L., Pei, L., Ju, W., Ahmadian, G., Lu, J., Wang, Y., Liu, F., and Wang, Y. T. (2003). Control of synaptic strength, a novel function of Akt. *Neuron* **38**, 915–928.
- Wang, X., He, G., Gu, N., Yang, J., Tang, J., Chen, Q., Liu, X., Shen, Y., Qian, X., Lin, W., Duan, Y., Feng, G., and He, L. (2004). Association of G72/G30 with schizophrenia in the Chinese population. *Biochem. Biophys. Res. Commun.* **319**, 1281–1286.
- Weickert, C. S., Straub, R. E., McClintock, B. W., Matsumoto, M., Hashimoto, R., Hyde, T. M., Herman, M. M., Weinberger, D. R., and Kleinman, J. E. (2004). Human dysbindin (DTNBP1)

- gene expression in normal brain and in schizophrenic prefrontal cortex. *Arch. Gen. Psychiatry* **61**, 544–555.
- Williams, H. J., Williams, N., Spurlock, G., Norton, N., Ivanov, D., McCreadie, R. G., Preece, A., Sharkey, V., Jones, S., Zammit, S., Nikolov, I., Kehaiov, I., *et al.* (2003a). Association between PRODH and schizophrenia is not confirmed. *Mol. Psychiatry* **8**, 644–645.
- Williams, N. M., Preece, A., Spurlock, G., Norton, N., Williams, H. J., Zammit, S., O'Donovan, M. C., and Owen, M. J. (2003b). Support for genetic variation in neuregulin 1 and susceptibility to schizophrenia. *Mol. Psychiatry* **8**, 485–487.
- Williams, N. M., Preece, A., Morris, D. W., Spurlock, G., Bray, N. J., Stephens, M., Norton, N., Williams, H., Clement, M., Dwyer, S., Curran, C., Wilkinson, J., *et al.* (2004). Identification in 2 independent samples of a novel schizophrenia risk haplotype of the dystrobrevin binding protein gene (DTNBP1). *Arch. Gen. Psychiatry* **61**, 336–344.
- Williams, H. J., Glaser, B., Williams, N. M., Norton, N., Zammit, S., MacGregor, S., Kirov, G. K., Owen, M. J., and O'Donovan, M. C. (2005). No association between schizophrenia and polymorphisms in COMT in two large samples. *Am. J. Psychiatry* **162**, 1736–1738.
- Winder, D. G., and Sweatt, J. D. (2001). Roles of serine/threonine phosphatases in hippocampal synaptic plasticity. *Nat. Rev. Neurosci.* **2**, 461–474.
- Xu, B., Wratten, N., Charych, E. I., Buyske, S., Firestein, B. L., and Brzustowicz, L. M. (2005). Increased expression in dorsolateral prefrontal cortex of CAPON in schizophrenia and bipolar disorder. *PLoS Med* **2**, e263[PMID: 16146415].
- Yang, J. Z., Si, T. M., Ruan, Y., Ling, Y. S., Han, Y. H., Wang, X. L., Zhou, M., Zhang, H. Y., Kong, Q. M., Liu, C., Zhang, D. R., Yu, Y. Q., *et al.* (2003). Association study of neuregulin 1 gene with schizophrenia. *Mol. Psychiatry* **8**, 706–709.
- Zhang, F., St. Clair, D., Liu, X., Sun, X., Sham, P. C., Crombie, C., Ma, X., Wang, Q., Meng, H., Deng, W., Yates, P., Hu, X., *et al.* (2005). Association analysis of the RGS4 gene in Han Chinese and Scottish populations with schizophrenia. *Genes Brain Behav.* **4**, 444–448.
- Zhao, X., Shi, Y., Tang, J., Tang, R., Yu, L., Gu, N., Feng, G., Zhu, S., Liu, H., Xing, Y., Zhao, S., Sang, H., *et al.* (2004). A case control and family based association study of the neuregulin 1 gene and schizophrenia. *J. Med. Genet.* **41**, 31–34.
- Zheng, Y., Li, H., Qin, W., Chen, W., Duan, Y., Xiao, Y., Li, C., Zhang, J., Li, X., Feng, G., and He, L. (2005). Association of the carboxyl-terminal PDZ ligand of neuronal nitric oxide synthase gene with schizophrenia in the Chinese Han population. *Biochem. Biophys. Res. Commun.* **328**, 809–815.
- Zou, F., Li, C., Duan, S., Zheng, Y., Gu, N., Feng, G., Xing, Y., Shi, J., and He, L. (2005). A family-based study of the association between the G72/G30 genes and schizophrenia in the Chinese population. *Schizophr. Res.* **73**, 257–261.

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